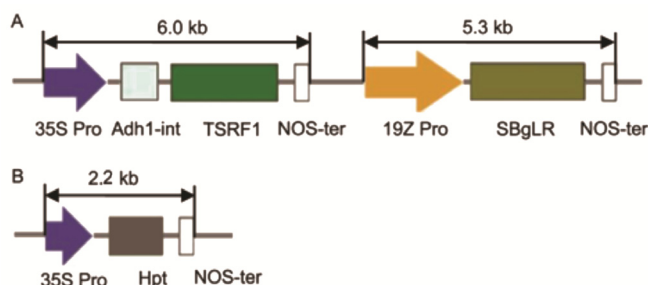
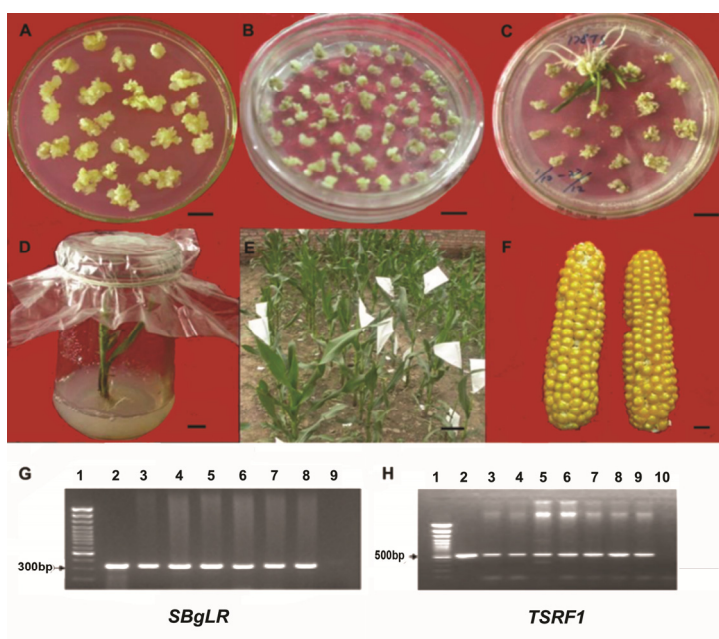


## Supplemental Information

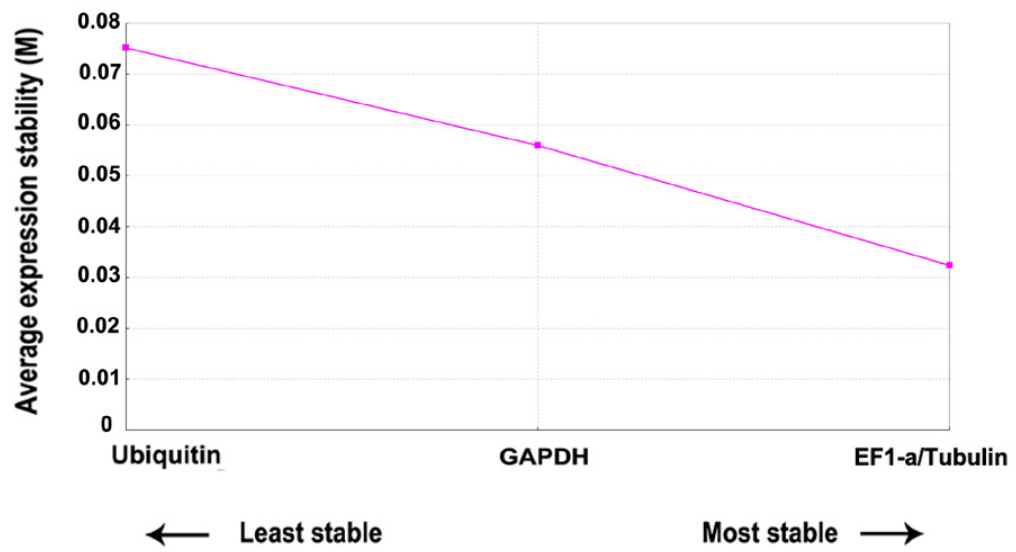
**Figure S1.** Diagram of the two constructs used in the present study (not drawn to scale). (A) pTSSB; (B) pHpt. 35S Pro, *cauliflower mosaic virus* 35S promoter; 19Z Pro, maize 19 kD zein gene promoter; Adh1-int, the first intron of maize alcohol dehydrogenase gene 1; NOS-ter, nopaline synthase terminator; Hpt, hygromycin B phosphotransferase gene.



**Figure S2.** Transgenic plants derived from maize embryogenic calli by particle bombardment-mediated co-transformation. (A) Embryonic calli of maize inbred X178; (B) Hygromycin-resistant calli after 6 weeks of selection; (C) Shoot formation from the hygromycin-resistant calli. The photo was taken 3 weeks after hygromycin-resistant calli had been placed on hygromycin-containing regeneration medium; (D) Root development from hygromycin-resistant shoot after 2 weeks culture in the rooting medium; (E) Fertile plants in the greenhouse; (F) Seed set on the ears of R<sub>0</sub> plants; (G) PCR analysis of some R<sub>0</sub> plants for *SBgLR* gene. Lane 1: DNA Ladder, Lane 2: Positive control, Lane 3-8: Transformed plants, Lane 9: Untransformed plants as negative control; (H) PCR analysis of some R<sub>0</sub> plants for *TSRF1* gene. Lane 1: DNA Ladder, Lane 2: Positive control, Lane 3-9: Transformed plants, Lane 10: Untransformed plants as negative control. Scale bars: A–D and F, 1 cm; E, 30 cm.



**Figure S3.** Average expression stability values of reference gene candidates by geNorm analysis.



**Table S1.** Primers used in this study. The primers were designed using the Primer 3.0 software (version 0.4.0) (<http://frodo.wi.mit.edu/>). The accession numbers of these genes and the amplification lengths were described. Primer sequences of each gene for different uses were listed. F means forward primer. R means reverse primer.

Gene	Accession No.	Use	Primer sequences (forward and reverse)	Amplicon length (bp)
<i>SBgLR</i>	AY377987.1	PCR/Dot blot	F: 5'-TTGGAAATTCTCAACCTTGAA-3' R: 5'-ATGAAGAATTGGAGGAACCTTGC-3'	260
		RT-PCR	F: 5'-GAACGATGAAACAACCTCCGGTT-3' R: 5'-GGCCTCAGAAACAGCGATAACTT-3'	181
<i>TSRF1</i>	AF494201.1	PCR/RT-PCR	F: 5'-ACAACATCCGAAACAGTCAC-3' R: 5'-CAGCACCCAAATCTTCAAAC-3'	472
<i>Tubulin</i>	NM_001174192.1	RT-PCR/qPCR	F: 5'-GAGCATGGCATTTCAGGCTGTCG-3' R: 5'-TCAACAAAAACAGCACGGGGGA-3'	128
<i>GAPDH</i>	NM_001112119.1	qPCR	F: 5'-CCTTCCAGGGACTGAAAGACAG -3' R: 5'-CTCCAAATCTCACGTGGCTA -3'	200
<i>Ubiquitin</i>	NM_001153555.1	qPCR	F: 5'-AGCTCCGACACCATCGACAA-3' R: 5'-TTACTGACCACCGCGGAG -3'	180
<i>EF1-α</i>	NM_001112117.1	qPCR	F: 5'-GGTGATGCTGGTATGGTGAA-3' R: 5'-TCATTTCTTCTTGGCAGCAG-3'	192
<i>ZmMYC1</i>	EU953366.1	qPCR	F: 5'-AATGAGCTGCGAGACGAGAAGCAA-3' R: 5'-AGGGTAGCCAATCACAGGCATCAT-3'	192
<i>ZmMBY59</i>	CM000780.2	qPCR	F: 5'-TGCCATCACCTTCATCCTCATCCT-3' R: 5'-TGTGGTGCCTCAATCTCCTTCCAT-3'	190
<i>ZmSDR</i>	GK000032.2	qPCR	F: 5'-TTGGAGCAGGCAATGGAGGGCAT-3' R: 5'-CTTTGGACGTTCCAGTGGATGGAT-3'	178
<i>ZmrbcS</i>	Y09214.1	qPCR	F: 5'-GCAACAAGAAGTTCGAGACGCTGT-3' R: 5'-TACACGAACCCGAGCTTGCTGAA-3'	130
<i>ZmELIP</i>	BT034060.1	qPCR	F: 5'-CCAACGCCGAGCTCTGGAA-3' R: 5'-TACGCGTTGATGAACGGTGCG-3'	97
<i>ZmPSI-N</i>	BT034060.1	qPCR	F: 5'-CCAACAAGGAGCTGAACGACAAGA-3' R: 5'-AATCTCGAGGTCGTCGCTGATGAA-3'	167
<i>ZmOEE</i>	CM000781.2	qPCR	F: 5'-GTACCAGATGAAGAAGCTGTGCCT-3' R: 5'-ATGCCGTCCTTCTCCTCGAACTT-3'	192
<i>ZmPLAS</i>	NM_001154032.1	qPCR	F: 5'-AAGATCTCGCAGGAGGAGTACCTCA-3' R: 5'-TTAGTTGACGGTGATCTTGCCGAC-3'	135
<i>ZmP5CS1</i>	BT083588.1	qPCR	F: 5'-ATCCTTGTGACCTCAGGTGCTGTT-3' R: 5'-AGTTGCGAGGAGGACACATCAAGT-3'	197
<i>ZmP5CS2</i>	CM000784.2	qPCR	F: 5'-GCAAGTTGATAGTGCCGCTGTGTT-3' R: 5'-ACTCCCTTGTCACCATTCACT-3'	195

**Table S2.** Lysine content of T<sub>2</sub> seeds. Samples were ground corn meal of 20 mature kernels from T<sub>2</sub> individual self-fertilized ears. The lysine contents are expressed as g/100 g dry seed. Data are averages of triplicate  $\pm$  standard deviations. Asterisks \*\* denote transgenic lines statistically different from control by Student *t*-test at  $p < 0.01$ .

Line number	Lysine content (g/100 g dry weight)	Increase (%)
9-10	0.308 $\pm$ 0.06	48.79 **
9-11	0.284 $\pm$ 0.01	37.20 **
10-9	0.337 $\pm$ 0.01	62.80 **
12-8	0.318 $\pm$ 0.02	53.62 **
17-8	0.287 $\pm$ 0.01	38.65 **
19-11	0.306 $\pm$ 0.05	47.83 **
20-9	0.314 $\pm$ 0.02	51.69 **
21-9	0.243 $\pm$ 0.01	17.39 **
control	0.207 $\pm$ 0.02	-

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