

SUPPLEMENTARY DATA

Steric analysis of epoxyalcohol and trihydroxy derivatives of 9-hydroperoxy-linoleic acid from hematin and enzymatic synthesis

by

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Table S1. ^1H NMR spectrum of 9S,10S,13S-trihydroxyoctadecenoate methyl ester, DMP derivative. (Trihydroxy I, generated from the acid hydrolysis of 9S,10S-*trans*-epoxy-13S-hydroxy-octadec-11E-enoic acid)

Table S2. ^1H NMR spectrum of 9S,12R,13S-trihydroxyoctadecenoate methyl ester, DMP derivative. (Trihydroxy IIa, generated from the acid hydrolysis of 9S,10S-*trans*-epoxy-13S-hydroxy-octadec-11E-enoic acid)

Table S3. ^1H NMR spectrum of 9S,10R,13S-trihydroxyoctadecenoate methyl ester, DMP derivative. (Trihydroxy IIb, generated from the acid hydrolysis of 9S,10S-*trans*-epoxy-13S-hydroxy-octadec-11E-enoic acid)

Table S4. ^1H NMR spectrum of 9S,12S,13S-trihydroxyoctadecenoate methyl ester, DMP derivative. (Trihydroxy III, pinellic acid, a minor acid hydrolysis product of 9S,10S-*trans*-epoxy-13S-hydroxyoctadec-11E-enoic acid)

Table S5. ^1H NMR spectrum of 9S,12S,13S-trihydroxyoctadecenoate methyl ester, DMP derivative. (Pinellic acid generated from 9S-HPODE in beetroot extract)

Table S6. ^1H NMR spectrum of 9R-hydroxy-10E-12R,13S-*cis*-epoxyoctadecenoate methyl ester. (Main epoxyalcohol generated by short (5 min) incubation of 9R-HPODE in beetroot)

Table S7. ^1H NMR spectrum of 9R,12S,13S-trihydroxyoctadecenoate methyl ester, DMP derivative. (Main trihydroxy product of 60 min incubation of 9R-HPODE in beetroot)

Table S8. ^1H NMR spectrum of 9-hydroxy-11E-13-oxo-octadecenoate methyl ester. (Major δ -ketol formed during mild acid hydrolysis of 9S,10S-*trans*-epoxy-13S-hydroxy-octadec-11E-enoic acid).

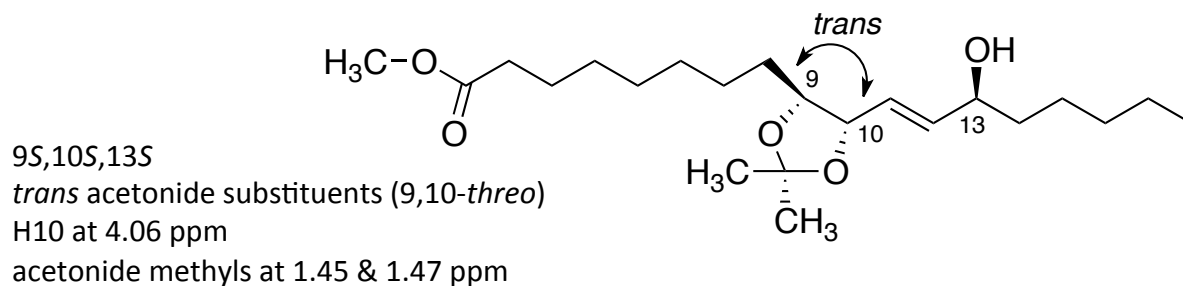
Table S9. ^1H -NMR spectrum of 9S,10S-*trans*-epoxy-11S-*erythro*-hydroxy-octadec-12Z-enoate methyl ester.

Table S10. ^1H -NMR spectrum of 9S,10S-*trans*-epoxy-11R-*threo*-hydroxy-octadec-12Z-enoate methyl ester.

and

Assignment of *erythro* versus *threo* 9,10-*trans*-epoxy-11-hydroxy-octadecenoates.

Table S1. ¹H NMR of 9(S),10(S),13(S)-trihydroxyoctadecenoate methyl ester, DMP derivative. The spectrum (600MHz) was acquired at room temperature in d₆-benzene solvent. Trihydroxy I is generated from the acid hydrolysis of 9(S),10(S)-epoxy-13(S)-hydroxyoctadecenoic acid.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.76	dd	H12 $J_{11,12}=15.5\text{Hz}$, $J_{12,13}=5.5\text{Hz}$
5.69	dd	H11 $J_{11,12}=15.5\text{Hz}$, $J_{10,11}=6.8\text{Hz}$
4.06	dd	H10 $J_{10,11}=7.5\text{Hz}$, $J_{9,10}=7.7\text{Hz}$
3.91	m	H9
3.72	m	H13
3.35	s	-OCH ₃

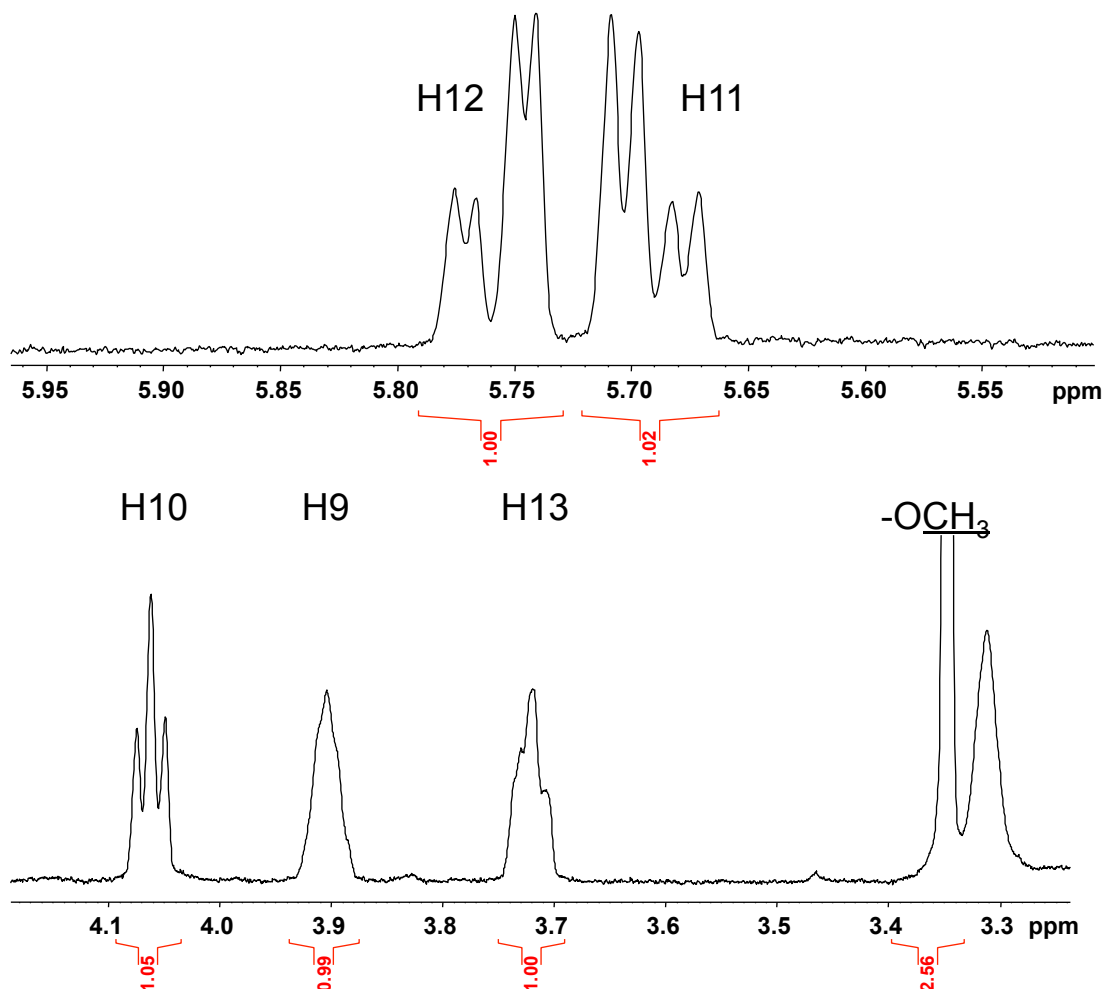
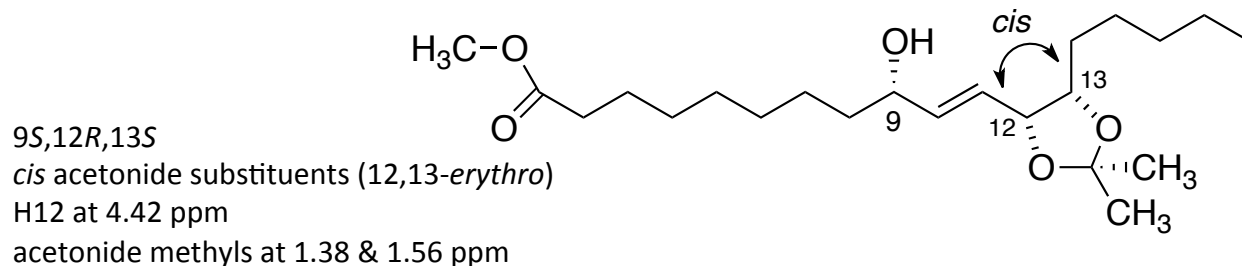


Table S2. ^1H NMR spectrum of 9(*S*),12(*R*),13(*S*)-trihydroxyoctadecenoate methyl ester, DMP derivative. The spectrum (600MHz) was acquired at room temperature in d_6 -benzene solvent. Trihydroxy **IIa** is generated from the acid hydrolysis of 9(*S*),10(*S*)-epoxy, 13(*S*)-hydroxyoctadecenoic acid.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.68	dd	H10 $J_{10,11}=15.5\text{Hz}$, $J_{9,10}=7.4\text{Hz}$
5.62	dd	H11 $J_{10,11}=15.5\text{Hz}$, $J_{11,12}=6.1\text{Hz}$
4.42	dd	H12 $J_{11,12}=6.8\text{Hz}$, $J_{12,13}=6.8\text{Hz}$
4.02	m	H13
3.87	m	H9
3.35	s	-OCH ₃

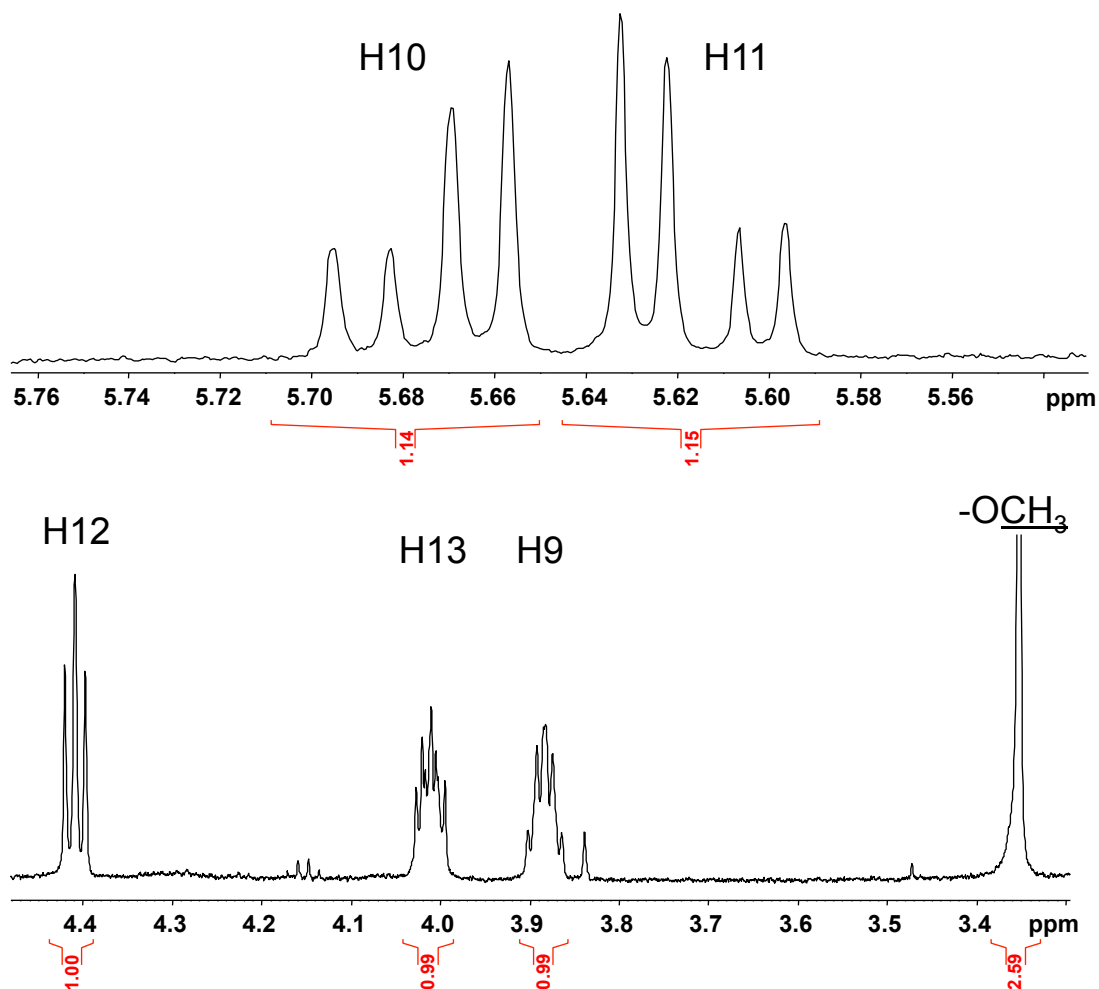
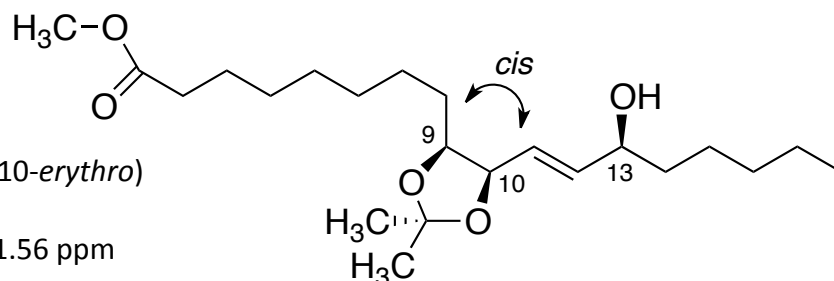


Table S3. ^1H NMR spectrum of 9(*S*),10(*R*),13(*S*)-trihydroxyoctadecenoate methyl ester, DMP derivative. The spectrum (600MHz) was acquired at room temperature in d_6 -benzene solvent. Trihydroxy IIb is generated from the acid hydrolysis of 9(*S*),10(*S*)-epoxy, 13(*S*)-hydroxyoctadecenoic acid.

9*S*,10*R*,13*S*
cis acetonide substituents (9,10-*erythro*)
 H10 at 4.42 ppm
 acetonide methyls at 1.38 & 1.56 ppm



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.68	dd	H12 $J_{11,12}=15.8\text{Hz}$, $J_{12,13}=7.6\text{Hz}$
5.62	dd	H11 $J_{11,12}=15.4\text{Hz}$, $J_{10,11}=5.8\text{Hz}$
4.42	dd	H10 $J_{10,11}=6.7\text{Hz}$, $J_{9,10}=7.0\text{Hz}$
4.01	m	H9
3.90	m	H13
3.35	s	-OCH ₃

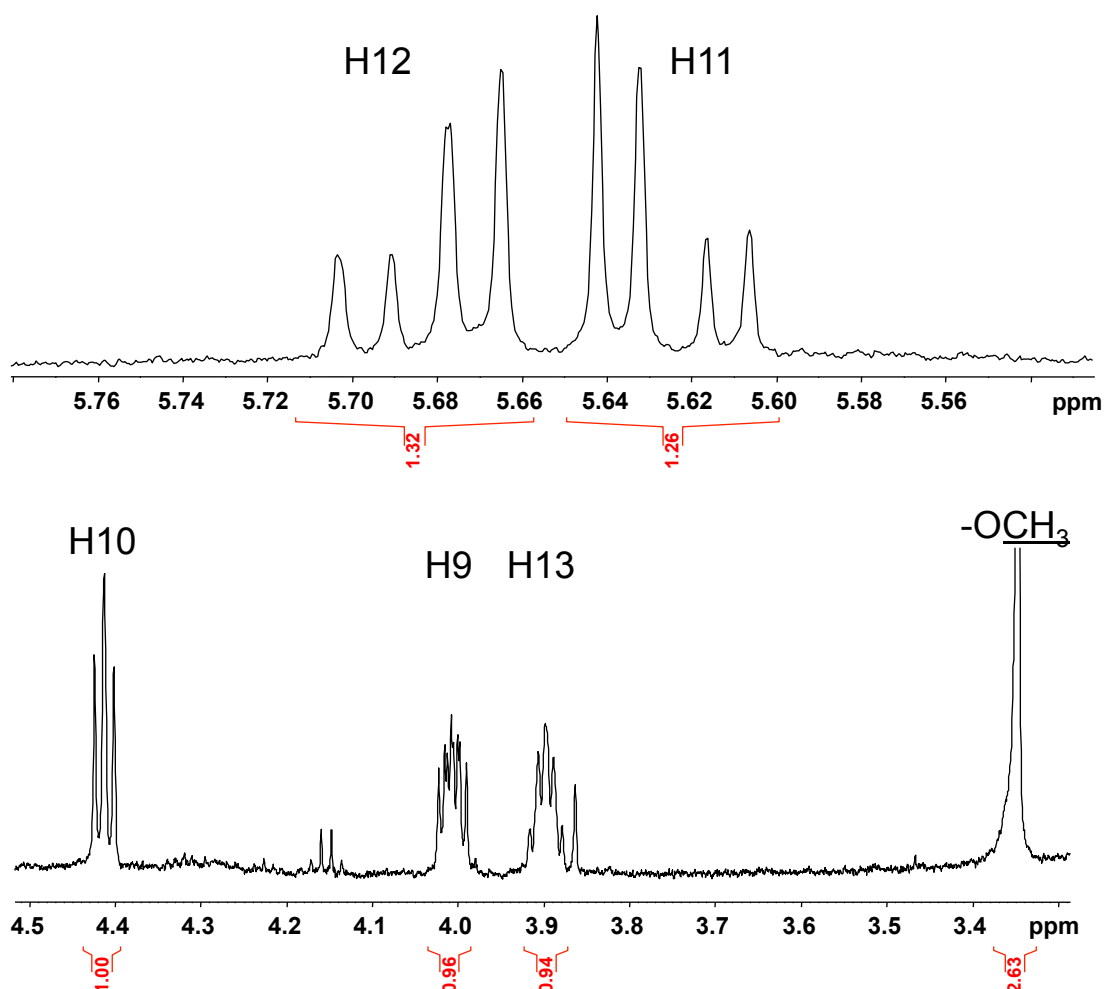
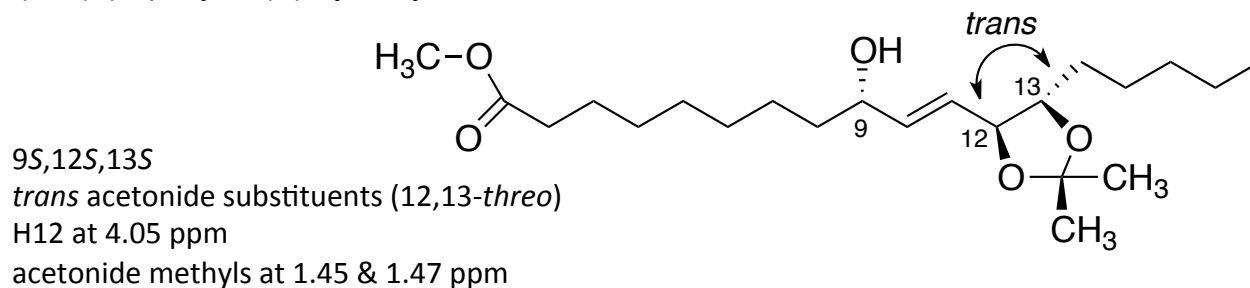


Table S4. ^1H NMR spectrum of 9(S),12(S),13(S)-trihydroxyoctadecenoate (pinellic acid) methyl ester, DMP derivative. The spectrum (600MHz) was acquired at room temperature in d_6 -benzene solvent. Trihydroxy III is generated from the acid hydrolysis of 9(S),10(S)-epoxy-13(S)-hydroxyoctadecenoic acid.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.75	dd	H10 $J_{10,11}=15.5\text{Hz}$, $J_{9,10}=5.5\text{Hz}$
5.68	dd	H11 $J_{10,11}=15.5\text{Hz}$, $J_{11,12}=6.8\text{Hz}$
4.06	dd	H12 $J_{11,12}=7.6\text{Hz}$, $J_{12,13}=7.6\text{Hz}$
3.88	m	H13
3.73	m	H9
3.35	s	-OCH ₃

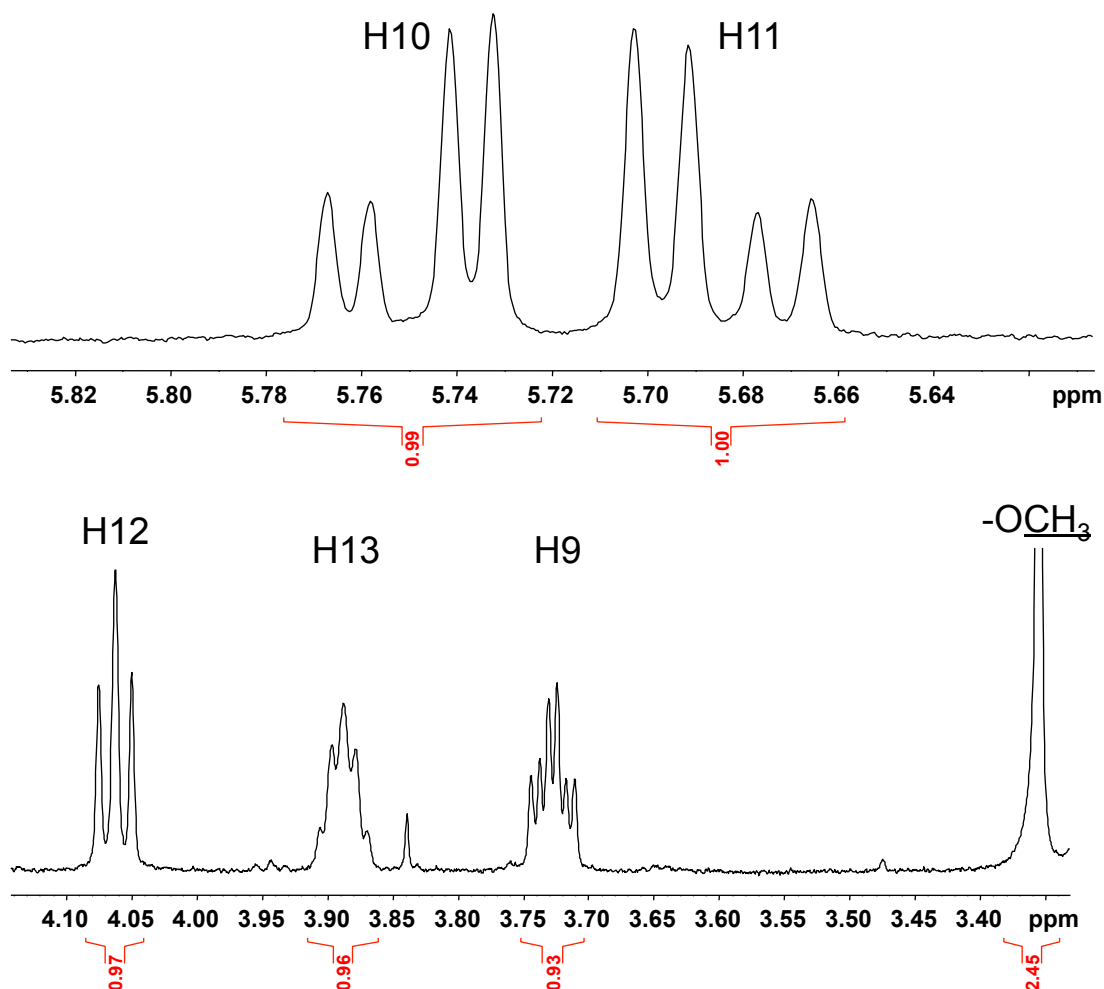
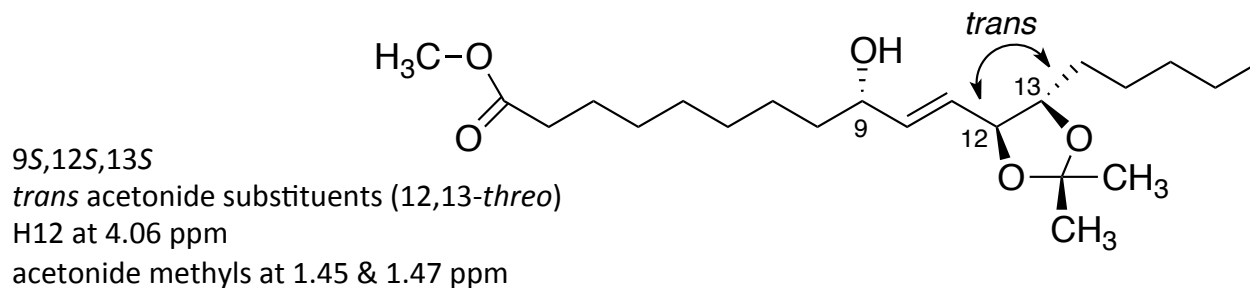


Table S5. ^1H NMR spectrum of 9(*S*),12(*S*),13(*S*)-trihydroxyoctadecenoate methyl ester, DMP derivative. The spectrum (600MHz) was acquired at room temperature in d_6 -benzene solvent. Pinellic acid is generated from the incubation of 9(*S*)-HPODE with beetroot extract.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.75	ddd	H10 $J_{10,11}=15.5\text{Hz}$, $J_{9,10}=5.5\text{Hz}$, $J_{10,12} = 0.3\text{Hz}$
5.68	ddd	H11 $J_{10,11} = 15.5\text{Hz}$, $J_{11,12} = 6.9\text{Hz}$, $J_{9,11} = 0.9\text{Hz}$
4.06	dd	H12 $J_{11,12} = 7.3\text{Hz}$, $J_{12,13} = 7.1\text{Hz}$
3.89	m	H13
3.72	m(dt)	H9
3.36	s	-OCH ₃

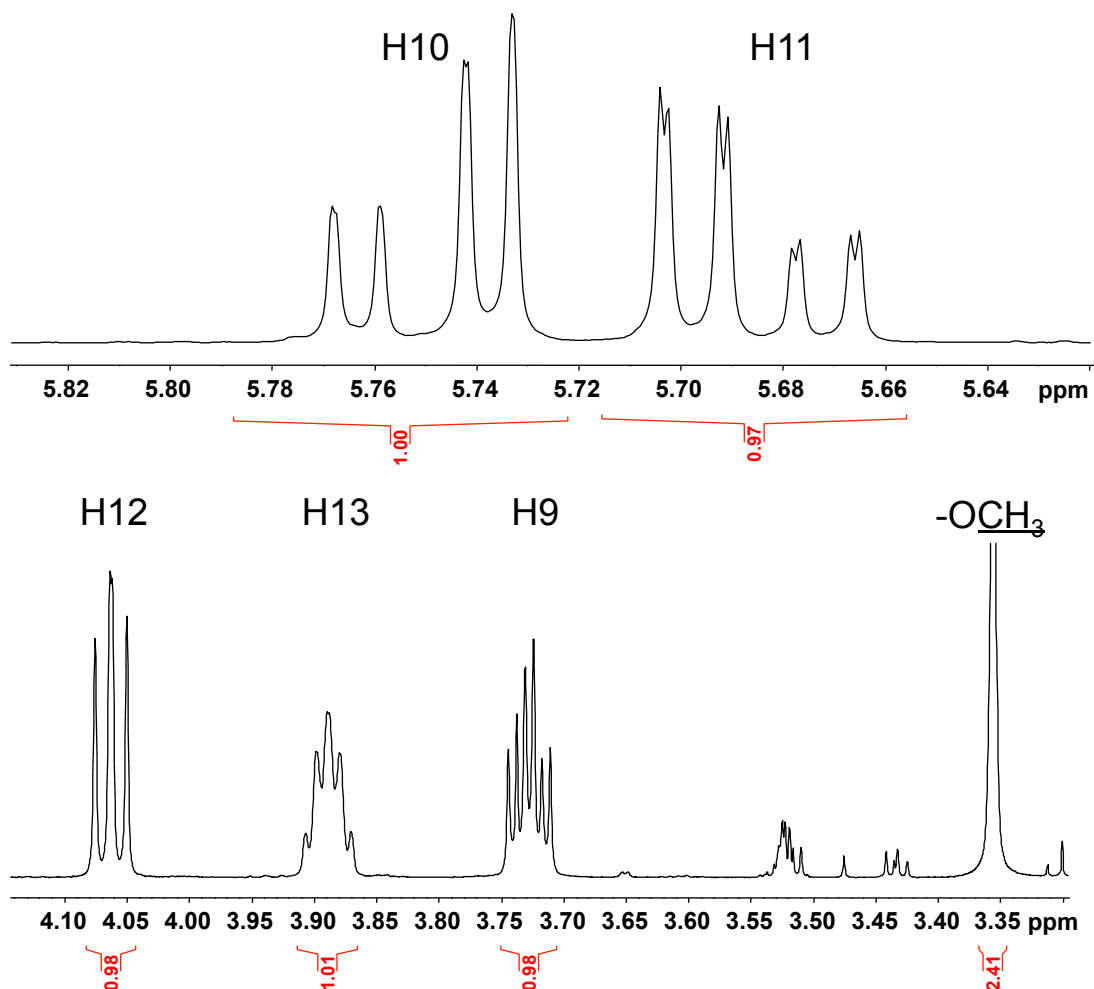
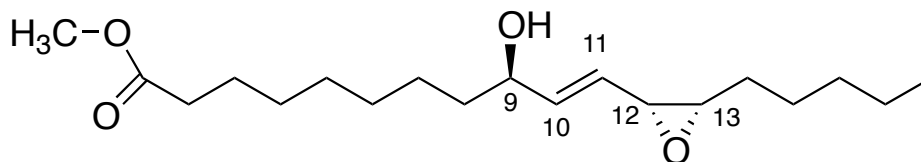


Table S6. ¹H NMR spectrum of 9(*R*)-hydroxy-12(*R*),13(*S*)-epoxy-octadecenoate methyl ester. The spectrum (600MHz) was acquired at room temperature in d₆-benzene solvent. The epoxyhydroxy is generated from the short duration incubation of 9(*R*)-HPODE with beetroot extract.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.79	dd	H12 $J_{11,12}=15.5\text{Hz}$, $J_{12,13}=5.8\text{Hz}$
5.58	ddd	H11 $J_{11,12}=15.5\text{Hz}$, $J_{10,11}=7.4\text{Hz}$, $J_{11,13}=1.3\text{Hz}$
3.86	m	H13
3.35	s	-OCH ₃
3.25	dd	H10 $J_{10,11}=7.2\text{Hz}$, $J_{9,10}=4.4\text{Hz}$
2.86	m	H9
2.09	t	H2

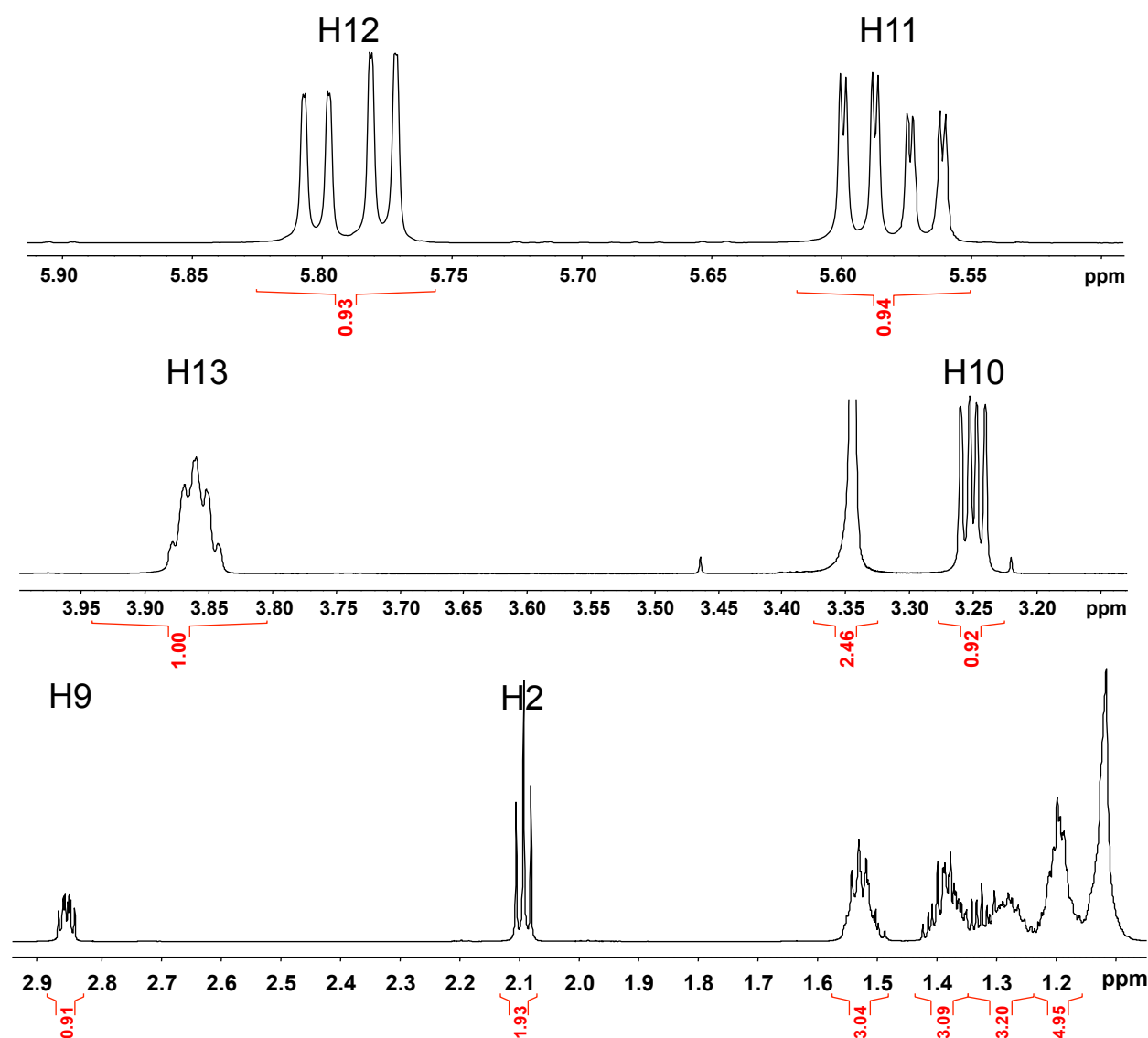
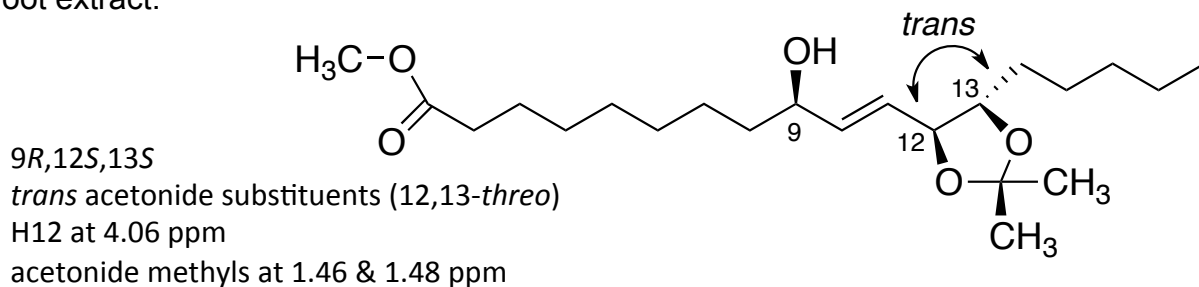


Table S7. ^1H NMR spectrum of 9(*R*),12(*S*),13(*S*)-trihydroxyoctadecenoate methyl ester, DMP derivative. The spectrum (600MHz) was acquired at room temperature in d_6 -benzene solvent. The trihydroxy is generated from the incubation of 9(*R*)-HPODE with beetroot extract.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.73	dd	H10 $J_{10,11}=15.6\text{Hz}$, $J_{9,10}=5.8\text{Hz}$
5.66	dd	H11 $J_{10,11}=16.3\text{Hz}$, $J_{11,12}=6.9\text{Hz}$
4.06	dd	H12 $J_{11,12}=7.3\text{Hz}$, $J_{12,13}=7.3\text{Hz}$
3.87	m	H13
3.72	m(dt)	H9
3.36	s	-OCH ₃

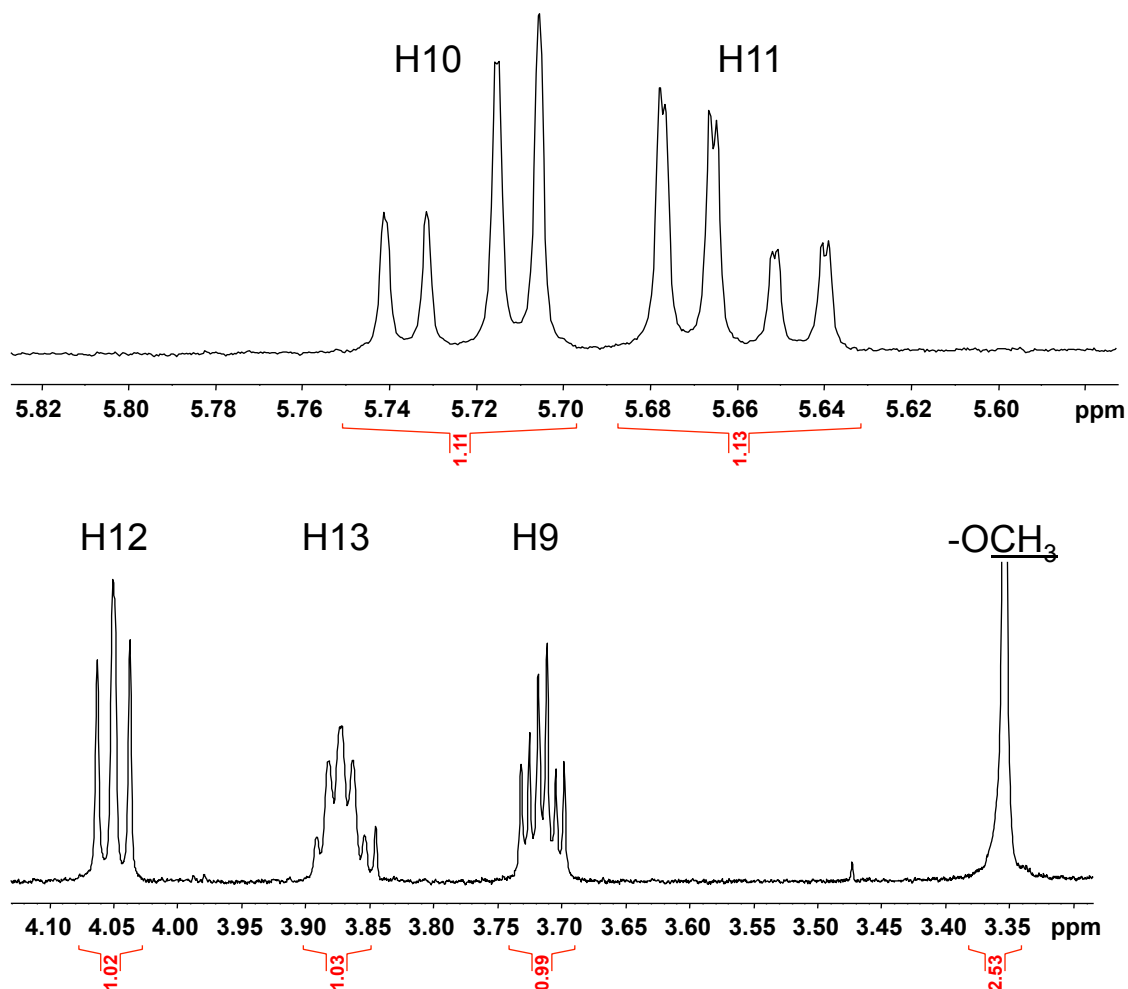
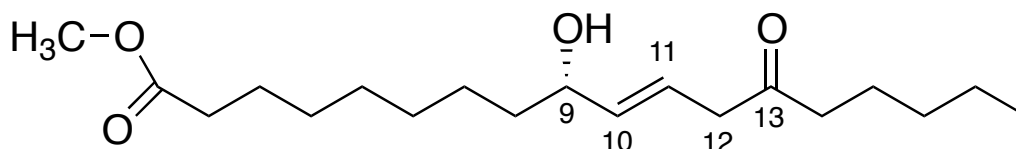


Table S8. ^1H NMR spectrum of 9(*S*)-hydroxy,11-*trans*-13-ketooctadecenoic methyl ester. The spectrum (600MHz) was acquired at room temperature in d_6 -benzene solvent. The δ -ketol by-product is generated during the acid hydrolysis of 9(*S*),10(*S*)-*trans*-epoxy-13(*S*)-hydroxyoctadecenoic acid.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.74	dt	H11 $J_{10,11} = 15.5\text{Hz}$, $J_{11,12} = 7.0\text{Hz}$, $J_{9,11} = 1.1\text{Hz}$
5.40	dd	H10 $J_{10,11} = 15.5\text{Hz}$, $J_{9,10} = 6.6\text{Hz}$, $J_{10,12} = 1.7\text{Hz}$
3.87	m	H9
2.73	d	H12 $J_{11,12} = 7.0\text{Hz}$

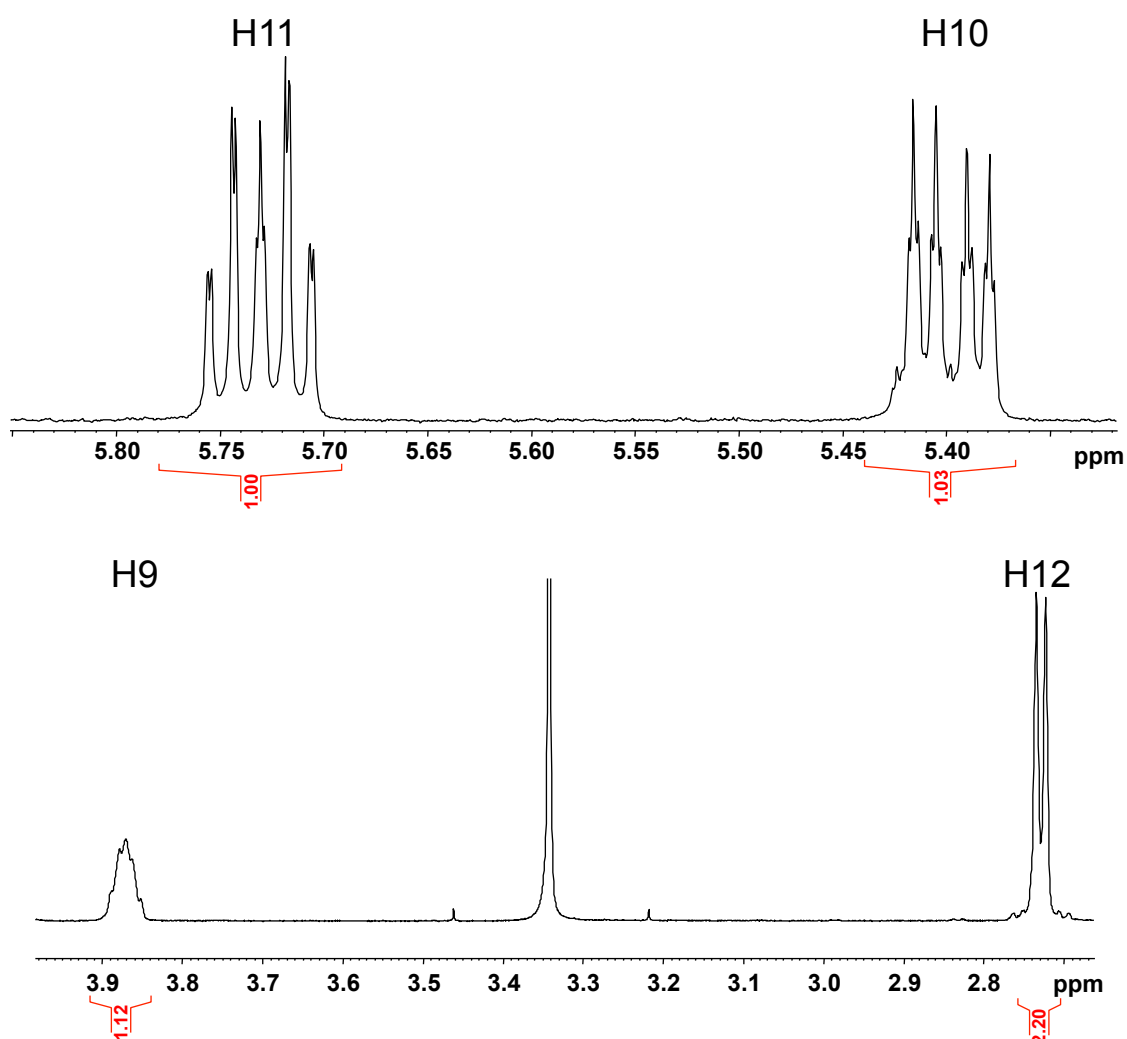
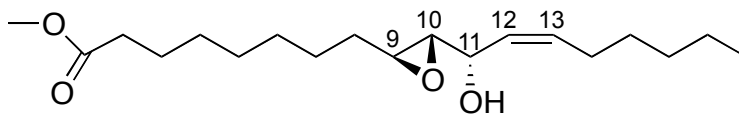


Table S9 . ¹H-NMR spectrum of 9S,10S-trans-epoxy-11S-erythro-hydroxy-octadec-12Z-enoate methyl ester. The spectrum (600MHz) was acquired at room temperature in d6- benzene solvent.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.45-5.52	dd/dt (m)	H12,13 $J_{12,13}=11.1\text{Hz}$
4.54	(dt) ddd	H11 $J_{-OH,11} \approx J_{10,11} = 2.5\text{-}3\text{Hz}, J_{11,12} = 6.7\text{Hz}$
3.35	s	-OCH ₃
2.94	dt	H9 $J_{8,9} = 5.6\text{Hz}, J_{9,10} = 2.2\text{Hz}$
2.69	dd	H10 $J_{9,10} = 2.2\text{Hz}, J_{10,11} = 3.12\text{Hz}$
2.09	t	H2 $J_{2,3} = 7.5\text{Hz}$
1.94	m	H14
1.63	d	11-OH $J = 2.5\text{Hz}$
1.52	p	H3

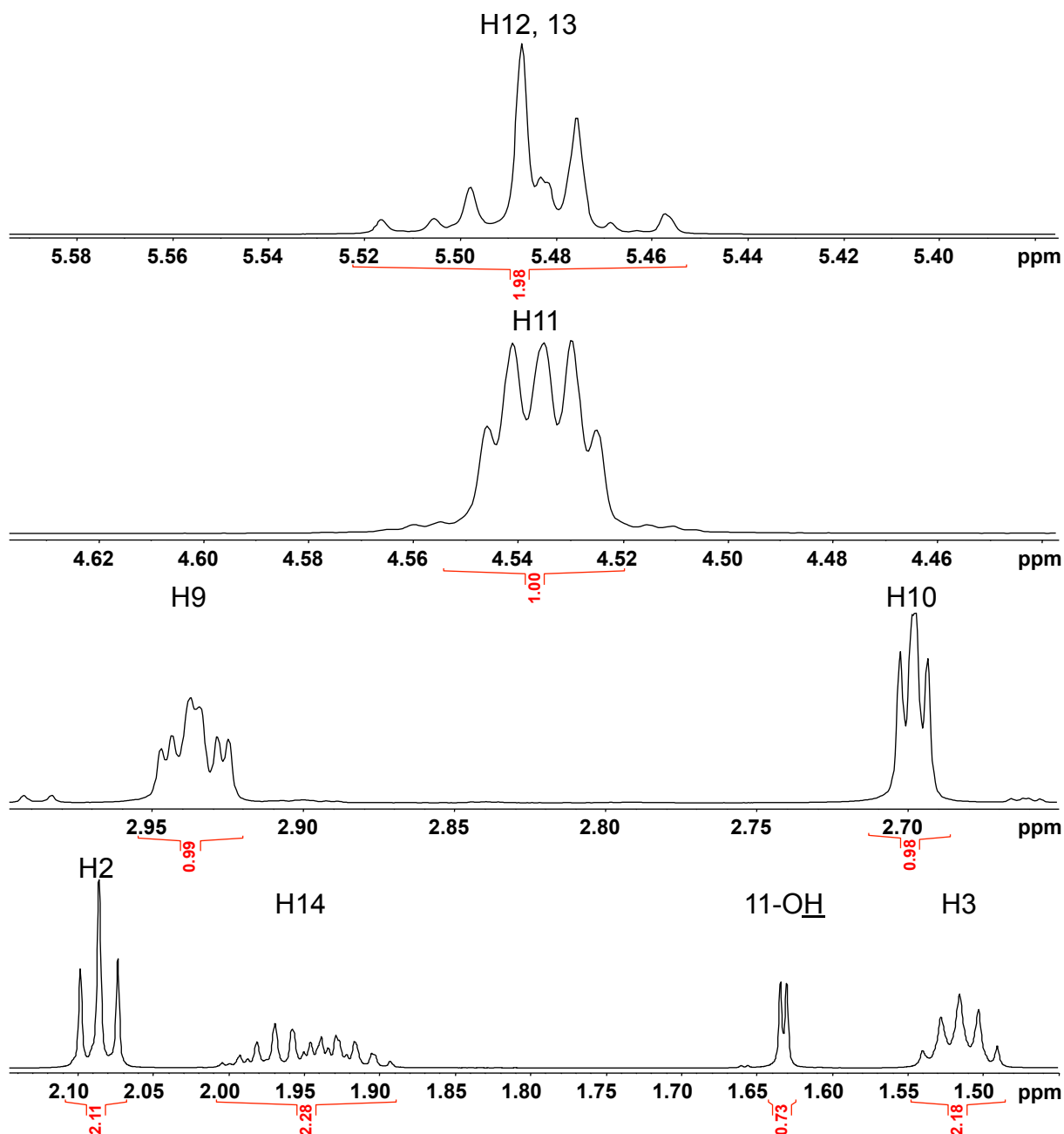
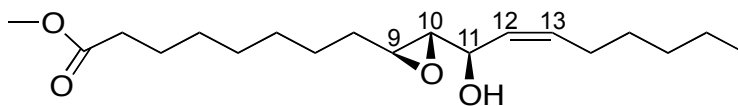
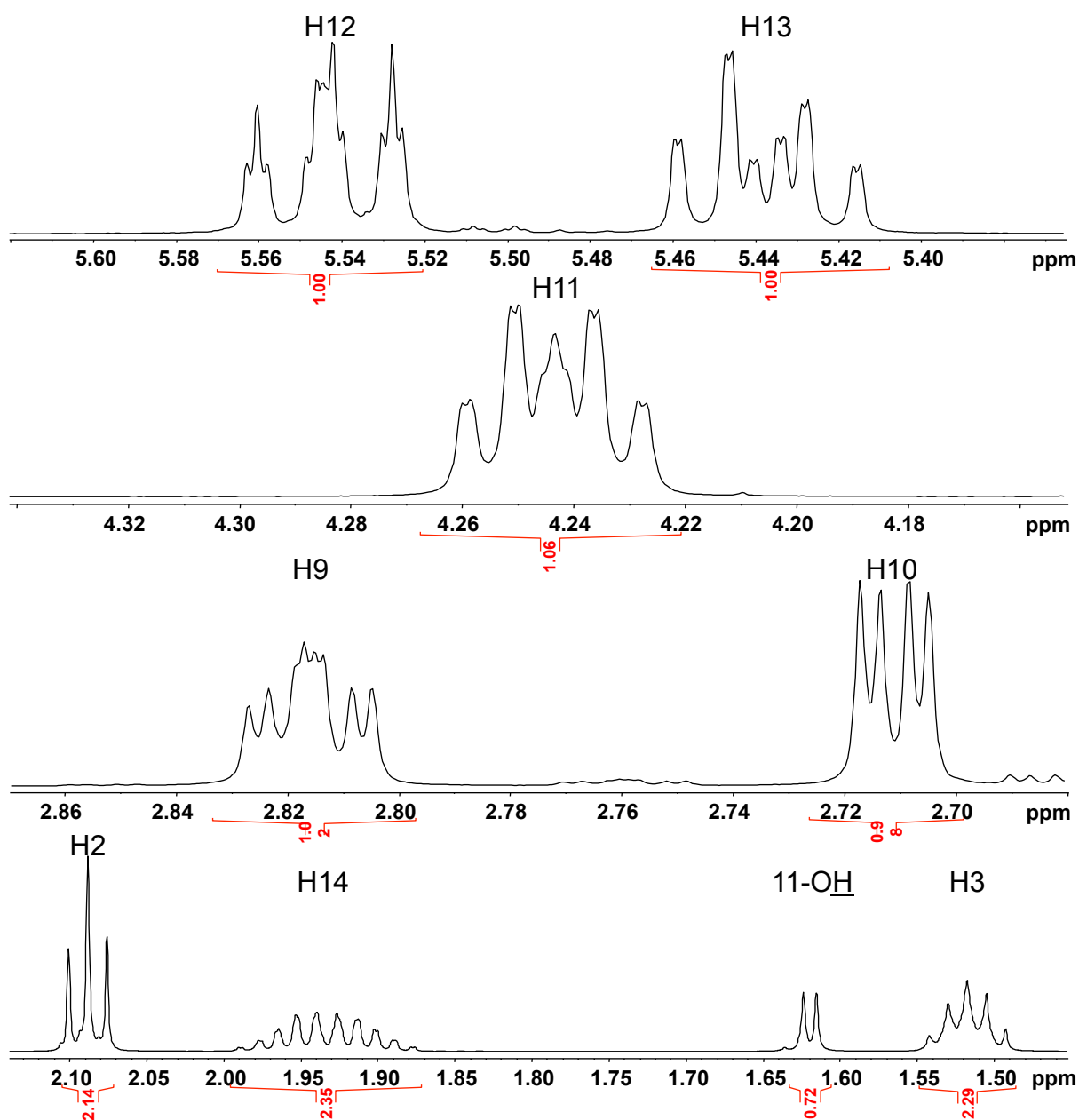


Table S10 . ¹H-NMR spectrum of 9*S*,10*S*-*trans*-epoxy-11*R*-*threo*-hydroxy-octadec-12*Z*-enoate methyl ester. The spectrum (600MHz) was acquired at room temperature in d₆- benzene solvent.



Chemical shift (ppm)	Multiplicity	Assignment, coupling constant
5.54	dd (+ 1.5 Hz allylic coupling)	H12 $J_{11,12}=8.6\text{Hz}$, $J_{12,13}=11.0\text{Hz}$
5.44	dt	H13 $J_{12,13}=11.0\text{Hz}$, $J_{13,14}=7.5\text{Hz}$
4.25	(dt) ddd	H11 $J_{\text{-OH},11} \approx J_{10,11} = 5.2\text{Hz}$, $J_{11,12} = 8.6\text{Hz}$
3.35	s	-OCH ₃
2.82	dt	H9 $J_{8,9} = 5.6\text{Hz}$, $J_{9,10} = 2.2\text{Hz}$
2.71	dd	H10 $J_{9,10} = 2.2\text{Hz}$, $J_{10,11} = 5.2\text{Hz}$
2.09	t	H2 $J_{2,3} = 7.5\text{Hz}$
1.94	m	H14
1.62	d	11-OH $J = 5.2\text{Hz}$
1.52	p	H3



Assignment of *erythro* versus *threo* 9,10-*trans*-epoxy-11-hydroxy-octadecenoates.

COSY analysis confirmed the overall structures for the two diastereomers in Supplementary Tables S9 and S10 as 9,10-epoxy-11-hydroxy-octadec-12Z-enoates. In both diastereomers, the coupling constant for the epoxide protons, $J = 2.2$ Hz, established the *trans* epoxide configuration. The *erythro* versus *threo* assignment is confirmed on the basis of three criteria (Bernart and Gerwick, 1994):

1) According to precedents for *erythro* and *threo* epoxyalcohols of this type, the mobility of the two diastereomers on silica (SP-HPLC or TLC) is in the order of the more mobile *erythro* followed by *threo* (Corey and Mehrotra, 1983; Corey et al., 1983; Dix and Marnett, 1985; Gardner and Crawford, 1981; Vasiljeva et al., 1993). Our *erythro* product eluted at 9.3 min on SP-HPLC (Fig. 1A, main text), while the *threo* diastereomer eluted later at 11 min (co-chromatographing with the first of the allylic epoxyalcohols (i.e. 9S,10S-*cis*-epoxy-10E-13-hydroxy-octadecenoate).

2) The α -hydroxy proton in the *erythro* isomer is the further downfield. We observed values of 4.535 ppm (*erythro*) and 4.35 ppm (*threo*).

3) The $^1\text{H-NMR}$ coupling constant for the 10-epoxide and 11-hydroxyl protons should be ~ 3.2 Hz for the *erythro* isomer (observed here as $J_{10,11} = 3.1$ Hz, Fig. S9) and at least 5 Hz for *threo* ($J_{10,11} = 5.2$ Hz, Table S10) (Mercier and Agoh, 1974).

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