



Supplemental Material to:

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**Autophagy: Resetting glutamine-dependent metabolism
and oxygen consumption**

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Figure S1. Complete metabolomic profile of WT and *atg5*^{-/-} MEFs under glutamine depletion. Hierarchical cluster (single dendrogram clustered by biomolecules) analysis of 204 analyzed biomolecules with complete linkage algorithm is shown. Signal fold change was normalized to WT at 0 h set as 1. All data points were log₂ transformed and used to calculate the Self-Organizing Map. Transformed fold changes are shown in green (decreasing) and red (increasing).

Figure S2. *Atg5* regulates the metabolic response to glutamine depletion in m5-7 cells. (A) Lower intracellular glutamine levels in Dox-treated “ATG5-off” m5-7 cells. The m5-7 cells, which were derived from *atg5*^{-/-} MEFs and stably express an inducible tetracycline-off *Atg5* construct¹, were cultured in the presence and absence of Dox. The repression of ATG5 expression was confirmed by Western blot (right panel). Intracellular glutamine levels were measured and normalized to cell number (left panel). Cells were harvested at the end of glutamine deprivation (0, 6 and 24 h). (B–E) Glutamine depletion increases glucose consumption and lactate production. Dox⁺ “ATG5-off” m5-7 cells exhibited lower glucose consumption and reduced ability to produce lactate upon glutamine depletion. The amounts of (B) glucose, (C) lactate, (D) sodium and (E) potassium ions in the medium, collected individually at the end of glutamine deprivation (0, 6 and 24 h), were analyzed, normalized (5 × 10⁶ cells), and plotted. The data were collected from three independent experiments. Dox: doxycycline; *p < 0.05.

Figure S3. Glutamine depletion decreased the level of γ -glutamyl amino acids and non-essential amino acids in WT MEFs but increased them in *atg5*^{-/-} MEFs. (A) Alterations of alanine, glutamine, glutamate, aspartate, proline and asparagine levels in glutamine-starved WT and *atg5*^{-/-} MEFs. Signal fold change was normalized to WT at 0 h set as 1 and log₂-transformed. Transformed fold changes are shown in green (decreasing) and red (increasing). (B) Profiles of γ -glutamyl-leucine, γ -glutamyl-isoleucine, γ -glutamyl-valine and γ -glutamyl-threonine in WT and *atg5*^{-/-} MEFs subjected to glutamine withdrawal. Cell preparation, quantitation and data

analyses were performed as described in Figure 2. *p < 0.05; **p < 0.01; ***p < 0.001; Gln: glutamine.

Figure S4. Glutamine supplementation rescued cell proliferation. (A) Cell proliferation was inhibited by glutamine depletion and 2-DG treatment. WT and *atg5^{-/-}* MEFs were seeded in complete medium to reach 10% confluence on the day of treatment, and then grown in serum-supplemented DMEM in the presence and absence of glutamine (4 mM) or in full medium with 2-DG (10 mM) for the indicated time periods. Relative cell proliferation (determined by the cell proliferation assay in methods) with each treatment was calculated by comparing to the 0 h time point, set as 1. Results from three independent experiments are shown as the mean \pm S.D. The student's *t*-test was performed to determine the difference between (a) control (24/48 h) and -Gln (24/48 h); (b) control (24/48 h) and 2-DG (24/48 h). *p < 0.05. (B) WT and *atg5^{-/-}* MEFs were seeded in complete medium to reach 10% confluence on the day of treatment, and then grown in glutamine-depleted DMEM with the indicated supplements. Cell proliferation assays were performed at the end of indicated time periods. Results from three independent experiments are shown as the mean \pm S.D. Gln: glutamine (4 mM); dNTP: deoxynucleotides triphosphate (10 μ M); dNMP: deoxynucleotides monophosphate (10 μ M); IMP: inosine monophosphate (10 μ M); MP: methylpyruvate (10 mM).

Figure S5. Lack of autophagy induction by glutamine deprivation in tfLC3-m5-7 cells. The m5-7 cells, which were derived from *atg5^{-/-}* MEFs and stably express an inducible tetracycline-off *Atg5* construct¹, were transiently transfected with mRFP-GFP tandem fluorescent-tagged MAP1LC3 (tfLC3). (A) Suppression of ATG5 expression by Dox-treatment in m5-7 cells. ATG5 was expressed in the absence of Dox (B, D, F and H) or suppressed in the presence of doxycycline (Dox; 20 μ g/ml; (C, E, G and I)). The cells were maintained in (B and C) complete medium, (D and E) EBSS for 6 and 24 h, (F and G) medium supplemented with 2-DG (10 mM) for 6 and 24 h or (H and I) glutamine-depleted medium for 6 and 24 h. Gln, glutamine; 2-DG, 2-deoxyglucose; Scale bar, 10 microns.

Reference

1. Hosokawa N, Hara Y, Mizushima N. Generation of cell lines with tetracycline-regulated autophagy and a role for autophagy in controlling cell size. FEBS letters 2006; **580** (11):2623-2629.

- Gln

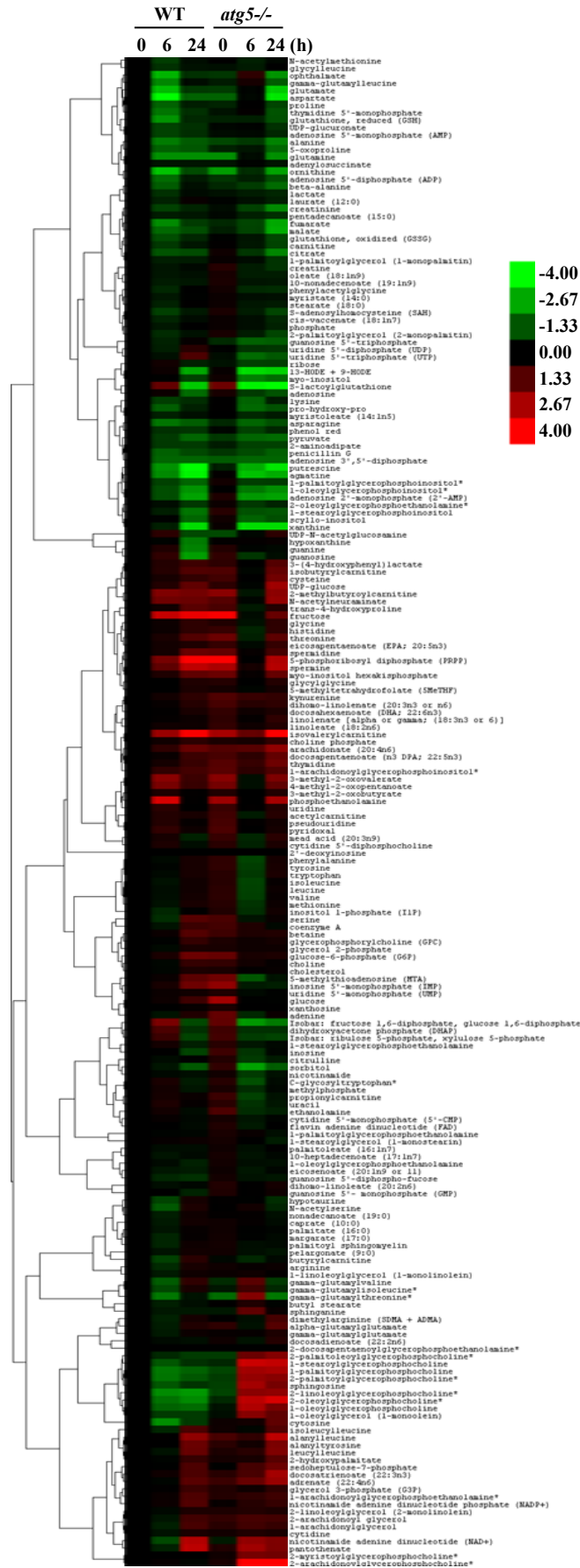


Figure S1

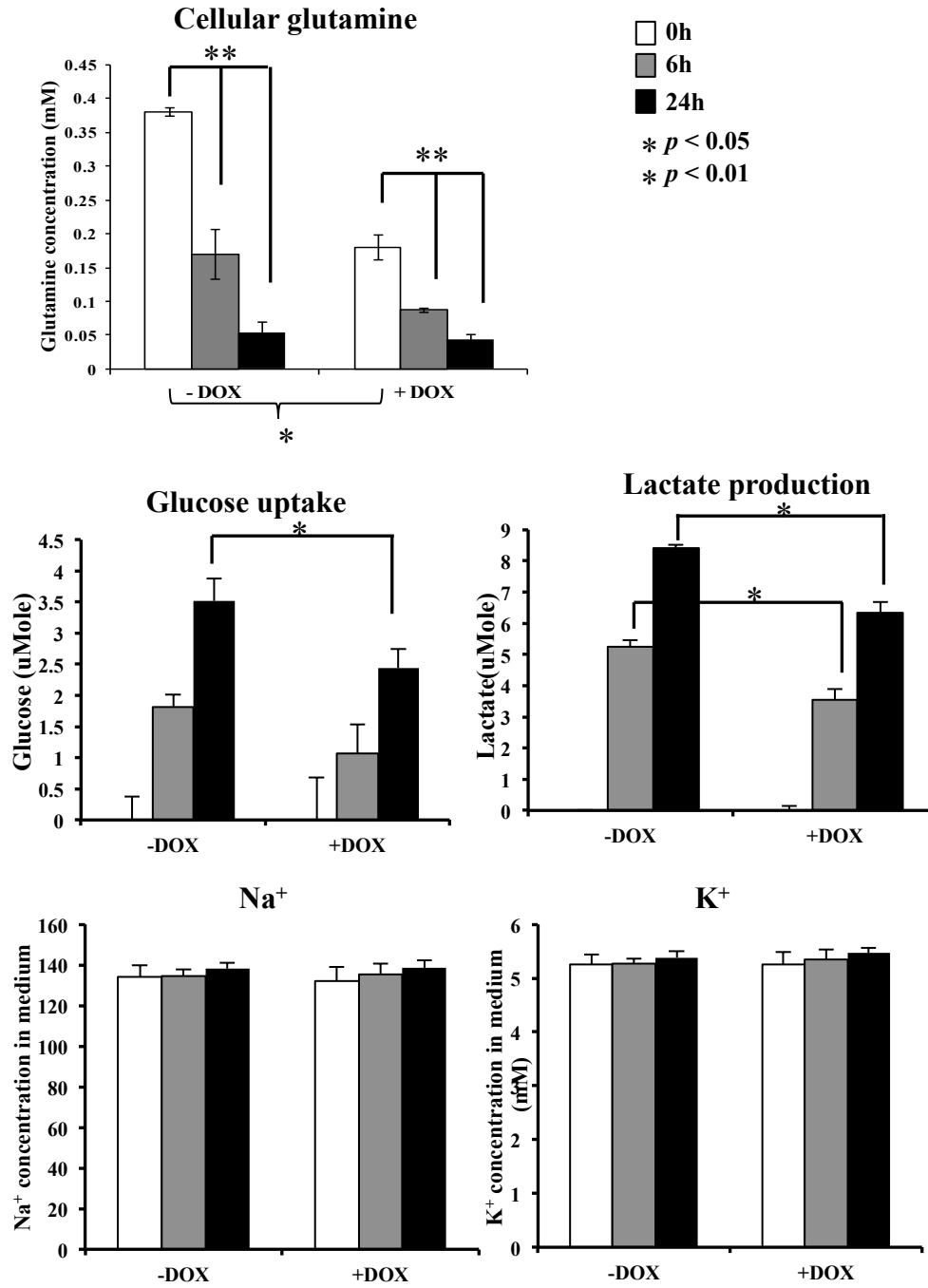
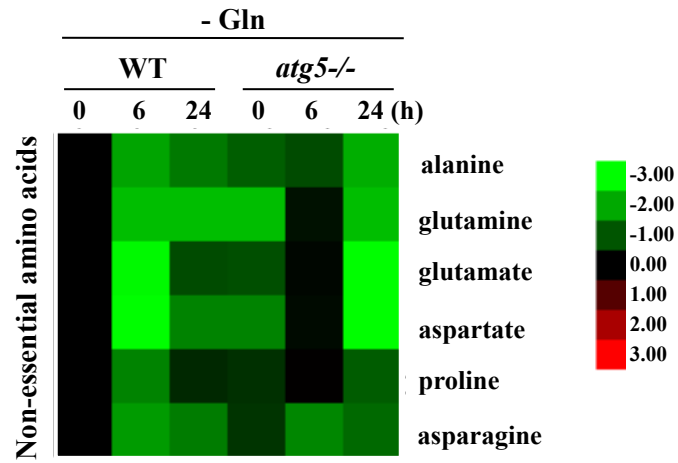


Figure S2

A



B

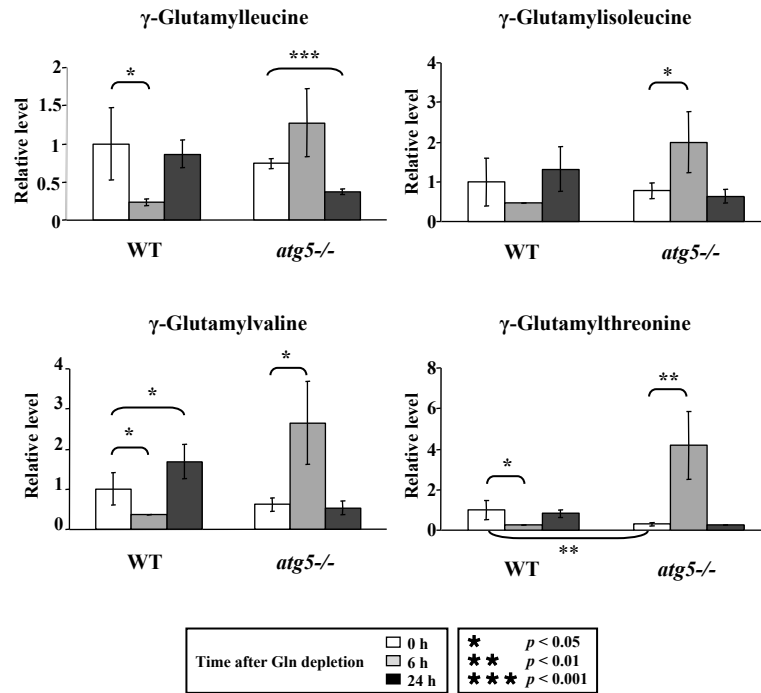
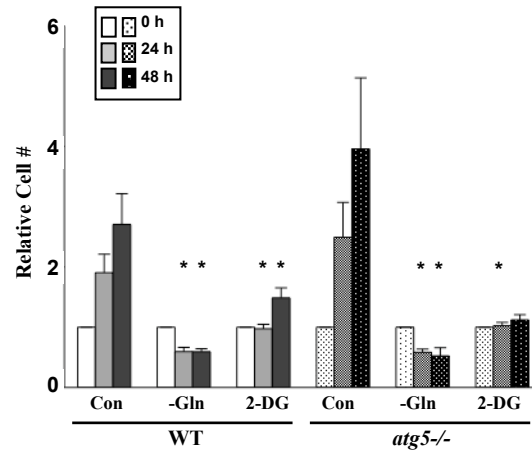


Figure S3

A



B

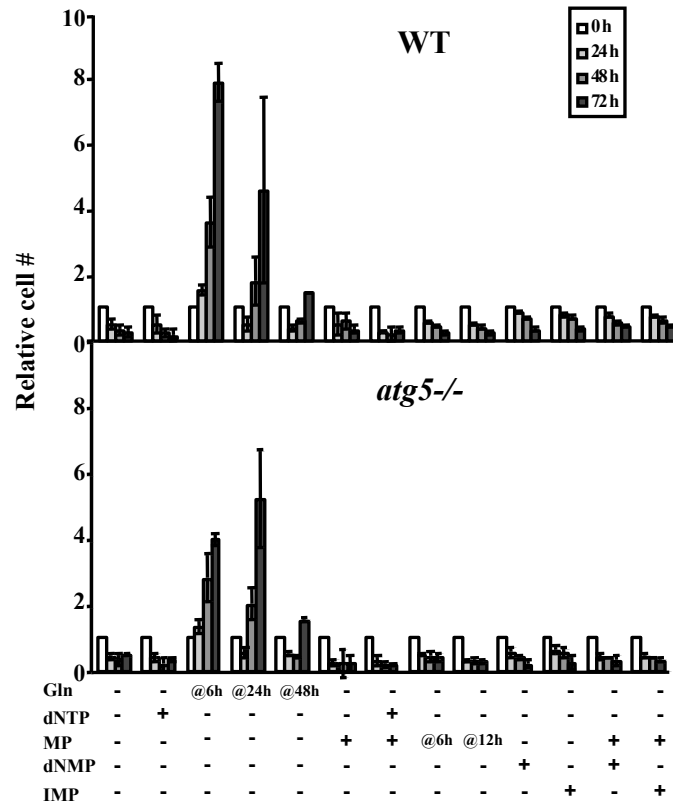


Figure S4

A

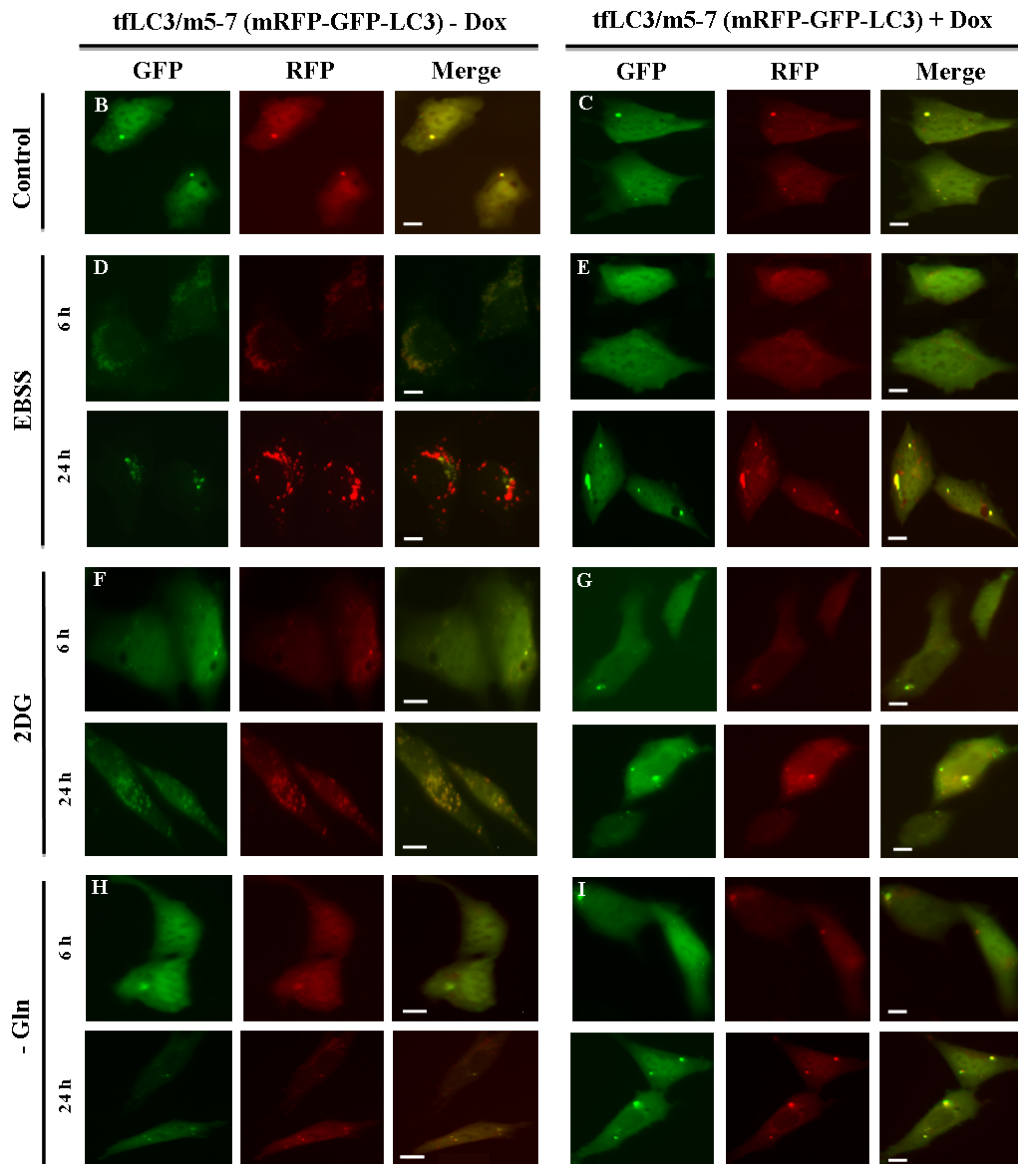
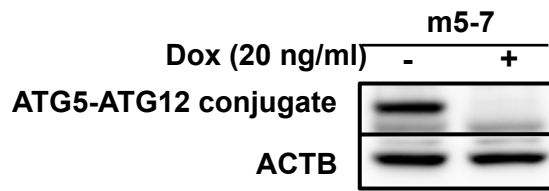


Figure S5

Supplementary Table S1. Compounds with significant differences between WT MEFs and *atg5*^{-/-} MEFs.

Super Pathway	Sub Pathway	Biochemical Name	Ratio of basal <i>atg5</i> ^{-/-} / WT)	
Upregulated				
Amino Acid	Creatine metabolism	creatine	1.53	
	Cysteine, methionine, SAM, taurine metabolism	cysteine	2.05	
	Cysteine, methionine, SAM, taurine metabolism	S-adenosylhomocysteine (SAH)	1.34	
	Cysteine, methionine, SAM, taurine metabolism	methionine	1.52	
	Glycine, serine and threonine metabolism	glycine	1.43	
	Glycine, serine and threonine metabolism	serine	2.24	
	Glycine, serine and threonine metabolism	threonine	3.23	
	Glycine, serine and threonine metabolism	betaine	1.96	
	Histidine metabolism	histidine	1.82	
	Phenylalanine & tyrosine metabolism	phenylalanine	1.67	
	Phenylalanine & tyrosine metabolism	tyrosine	1.68	
	Phenylalanine & tyrosine metabolism	3-(4-hydroxyphenyl)lactate	1.64	
	Polyamine metabolism	spermine	6.43	
	Polyamine metabolism	spermidine	2.89	
	Polyamine metabolism	5-methylthioadenosine (MTA)	3.37	
	Tryptophan metabolism	C-glycosyltryptophan*	1.52	
	Tryptophan metabolism	tryptophan	1.54	
	Urea cycle; arginine-, proline-, metabolism	citrulline	1.46	
	Valine, leucine and isoleucine metabolism	3-methyl-2-oxoalate	4.37	
	Valine, leucine and isoleucine metabolism	isoleucine	1.52	
	Valine, leucine and isoleucine metabolism	leucine	1.59	
	Valine, leucine and isoleucine metabolism	valine	1.61	
	Valine, leucine and isoleucine metabolism	4-methyl-2-oxopentanoate	3.52	
	Valine, leucine and isoleucine metabolism	2-methylbutyrylcarnitine	4.53	
	Valine, leucine and isoleucine metabolism	isovalerylcarnitine	6.16	
	Carbohydrate	Aminosugars metabolism	N-acetylneuraminate	2.53
		Nucleotide sugars, pentose metabolism	5-phosphoribosyl diphosphate (PRPP)	16.16
	Cofactors and vitamins	Folate metabolism	5-methyltetrahydrofolate (5MeTHF)	1.27
Pantothenate and CoA metabolism		pantothenate	1.44	
Pyridoxal metabolism		pyridoxal	2.27	
Riboflavin metabolism		flavin adenine dinucleotide (FAD)	1.42	
Pantothenate and CoA metabolism		coenzyme A	2.08	
Lipid	Carnitine metabolism	acetylcarnitine	2.03	

	Essential fatty acid	linolenate [alpha or gamma; (18:3n3 or 6)]	1.65
	Essential fatty acid	dihomo-linolenate (20:3n3 or n6)	2.15
	Essential fatty acid	docosapentaenoate (n3 DPA; 22:5n3)	2.18
	Essential fatty acid	docosahexaenoate (DHA; 22:6n3)	1.73
	Essential fatty acid	eicosapentaenoate (EPA; 20:5n3)	1.8
	Fatty acid metabolism (also BCAA metabolism)	propionylcarnitine	1.89
	Glycerolipid metabolism	glycerol 3-phosphate (G3P)	1.44
	Glycerolipid metabolism	phosphoethanolamine	5.52
	Glycerolipid metabolism	glycerophosphorylcholine (GPC)	1.36
	Glycerolipid metabolism	cytidine 5'-diphosphocholine	1.57
	Glycerolipid metabolism	choline	1.73
	Inositol metabolism	inositol hexaphosphate (IP6)	3.21
	Long chain fatty acid	linoleate (18:2n6)	1.74
	Long chain fatty acid	10-heptadecenoate (17:1n7)	1.26
	Long chain fatty acid	10-nonadecenoate (19:1n9)	1.33
	Long chain fatty acid	oleate (18:1n9)	1.3
	Long chain fatty acid	dihomo-linoleate (20:2n6)	1.44
	Long chain fatty acid	mead acid (20:3n9)	1.81
	Long chain fatty acid	palmitate (16:0)	1.18
	Long chain fatty acid	arachidonate (20:4n6)	2.45
	Lysolipid	1-arachidonoylglycerophosphoinositol*	2.62
	Lysolipid	1-stearoylglycerophosphoethanolamine	1.55
	Lysolipid	1-arachidonoylglycerophosphoethanolamine*	1.48
	Monoacylglycerol	2-linoleoylglycerol (2-monolinolein)	1.37
	Monoacylglycerol	2-arachidonoyl glycerol	1.73
	Monoacylglycerol	2-palmitoylglycerol (2-monopalmitin)	1.3
	Sphingolipid	palmitoyl sphingomyelin	1.24
	Sterol/Steroid	cholesterol	1.32
Nucleotide	Purine metabolism, (hypo)xanthine/inosine containing	inosine 5'-monophosphate (IMP)	4.83
	Purine metabolism, adenine containing	adenine	2.96
	Purine metabolism, guanine containing	guanosine 5'- monophosphate (GMP)	1.4
	Purine metabolism, guanine containing	guanosine 5'-diphospho-fucose	1.24
	Pyrimidine metabolism, cytidine containing	cytidine 5'-monophosphate (5'-CMP)	1.26
	Pyrimidine metabolism, uracil containing	pseudouridine	2.13
	Pyrimidine metabolism, uracil containing	uridine	1.88
	Pyrimidine metabolism, uracil containing	uridine 5'-monophosphate (UMP)	1.43

Downregulated				
Amino acid	Alanine and aspartate metabolism	alanine	0.46	
	Alanine and aspartate metabolism	aspartate	0.34	
	Glutamate metabolism	glutamine	0.21	
	Glutamate metabolism	glutamate	0.52	
	Glutathione metabolism	5-oxoproline	0.7	
	Glutathione metabolism	glutathione, reduced (GSH)	0.69	
	Urea cycle; arginine-, proline-, metabolism	ornithine	0.18	
	Urea cycle; arginine-, proline-, metabolism	proline	0.67	
	Energy	Krebs cycle	malate	0.72
	Nucleotide	Purine metabolism, adenine containing	adenosine 5'-monophosphate (AMP)	0.88
Purine metabolism, adenine containing		adenosine 3',5'-diphosphate	0.37	
Purine metabolism, adenine containing		adenylosuccinate	0.8	
Peptide	gamma-glutamyl	gamma-glutamylthreonine*	0.31	

*indicates a biochemical that has not been confirmed with an authentic standard but whose spectral properties strongly support its identity

Supplementary Table S2. Real-time PCR primer pairs

Gene	Forward primer	Reverse primer
<i>Aco1</i>	CCATCCGTGATGTTAGGAGCAG	GACAGGTAAGGCATGACTCCAC
<i>Atg5</i>	CAGAAGGTTATGAGACAAGAAGATG	TGGATGGACAGTGTAGAAGGTC
<i>Cs</i>	ATGCAGAGGGAATGAACCGAGC	GAGTCAATGGCTCCGATACTGC
<i>Flnb</i>	GAACTCACACGCAGGATGCTGT	GGCGGCTTTTATTCTCACCATCG
<i>Gapdh</i>	CCCCTTCATTGACCTCAACTA	CTCCTGGAAGATGGTGATGG
<i>Gpt2</i>	CATTGCGGCAAGCCAAAGACCA	GCTTCTCTCCCAGGCCAAAGTG
<i>Idh1</i>	GCTGGCTTTGTATCTCA	CGCCATCATACTTCTTCA
<i>Idh2</i>	GGCTGTCAAGTGTGCCACAATC	TTGGCTCTCTGAAGACGGTTCC
<i>Ldhb</i>	CCTCAGATCGTCAAGTACAGCC	ATCCGCTTCCAATCACACGGTG
<i>Mdh1</i>	TTCTGGACGGTGTCTGATGGA	TAGGACAGCCACATCCAGGTCT
<i>Me1</i>	ATGGAGAAGGAAGGTTTATCAAAG	GGCTTCTAGGTTCTTCATTTCTTC
<i>Me2</i>	GGCTAAGAGCTGTTACCACTCC	CGTAAACGCCATTCCCTTGT
<i>Myc</i>	TCGCTGCTGCCTCCGAGTCC	GGTTTGCCTCTTCTCCACAGAC
<i>Ogdh</i>	GGTGTGTCATCAATCAGCCTGAGT	ATCCAGCCAGTGCTTGATGTGC
<i>Sdha</i>	GAGATACGCACCTGTTGCCAAG	GGTAGACGTGATCTTTCTCAGGG
<i>Sdhb</i>	TGCGGACCTATGGTGTGGATG	CCAGAGTATTGCCTCCGTTGATG
<i>Slc1a5</i>	CTGCCTGTGAAGGACATCTCCT	CTCGGCATCTTGGTTCGATCCA
<i>Slc3a2</i>	GAGCGTACTGAATCCCTAGTCAC	GCTGGTAGAGTCGGAGAAGATG
<i>Slc7a5</i>	GGTCTCTGTTACGTCCTCAAG	GAACACCAGTGATGGCACAGGT
<i>Sucla2</i>	GGTGTCTCTGTTCCCAAAGGCT	TTTCTCTGCCGCCAGCCAAAA
<i>Suclg1</i>	GTCTTACACAGCCTCTCGGAAAC	ACTCCAAAGCCTGCTGACTGTG
<i>Suclg2</i>	AGCTCAAGGTGCCACTGGTAGT	GCTTTCTTGGCTGCATCCTCCA
<i>Cdkn1a</i>	TCGCTGTCTTGCACTCTGGTGT	CCAATCTGCGCTTGGAGTGATAG
<i>Bbc3</i>	ACCGCTCCACCTGCCGTAC	ACGGGCGACTCTAAGTGCTGC
<i>Pmaip1</i>	GGAAGTCGCAAAGAGCAGGATG	CTGCCGTAAATTCACCTTGTCTCC