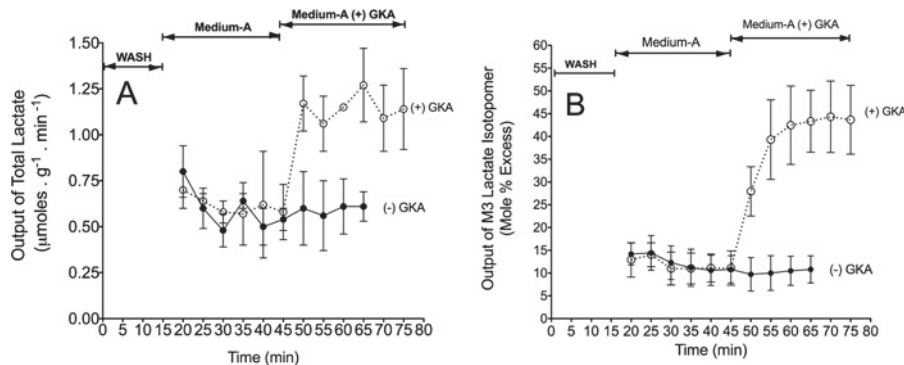


## SUPPLEMENTARY ONLINE DATA

# Effects of a glucokinase activator on hepatic intermediary metabolism: study with <sup>13</sup>C-isotopomer-based metabolomics

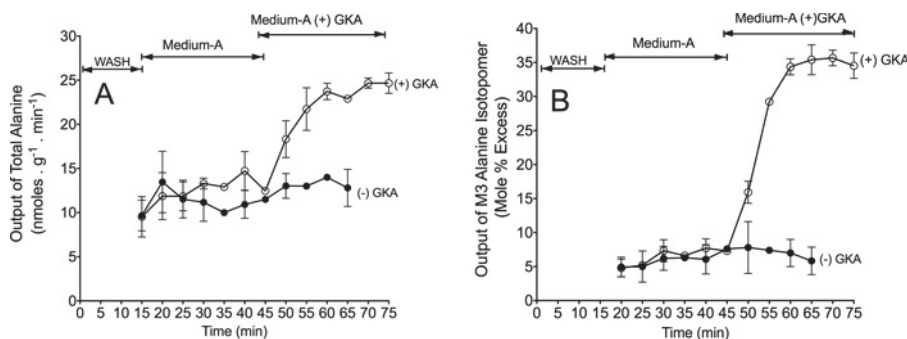
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**Figure S1** Output of total and M3 isotopomer of <sup>13</sup>C<sub>3</sub> lactate during liver perfusion with [U-<sup>13</sup>C]glucose

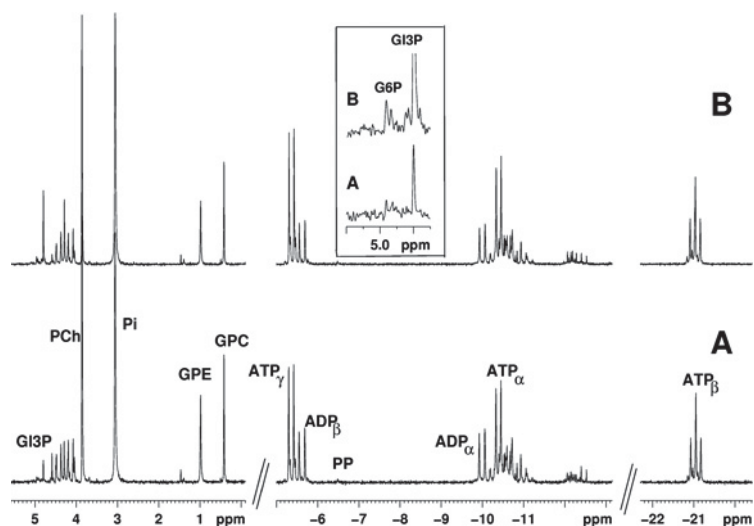
(A) Output of total lactate ( $\mu\text{mol}\cdot\text{g}^{-1}\cdot\text{min}^{-1}$ ). (B) Output of M3 lactate (lactate labelled with <sup>13</sup>C at three carbons). Liver perfusions were carried out as detailed in the Experimental section of the main text. Briefly, after a 15 min conditioning of the liver with medium containing 0.1 % DMSO and 5 mM unlabelled glucose in Krebs buffer, pH 7.4 (WASH), the perfusate was replaced with medium containing 0.1 % DMSO, 5 mM [U-<sup>13</sup>C]glucose, 0.3 mM <sup>15</sup>NH<sub>4</sub>Cl and 1 mM glutamine (Medium-A). At 45 min, perfusion was continued with Medium-A plus 3  $\mu\text{mol/l}$  Piragliatin, [(+) GKA]. Independent control perfusions without GKA [(-) GKA] were performed with perfusate containing Medium-A. Results are means  $\pm$  S.D. of three independent liver perfusions per study group. M3 lactate isotopomer was chosen as a marker of direct metabolite of glycolysis from [U-<sup>13</sup>C]glucose. M3 lactate must have been derived from [U-<sup>13</sup>C]pyruvate, the product of [U-<sup>13</sup>C]glucose glycolysis. These results demonstrate that: (i) GKA stimulated glycolysis of [U-<sup>13</sup>C]glucose within 3–5 min after its addition to the perfusate; and (ii) the activation of glucokinase by 3  $\mu\text{mol/l}$  Piragliatin is directly responsible for the remarkable elevation of total and <sup>13</sup>C-labelled lactate output from the perfusate [U-<sup>13</sup>C]glucose.



**Figure S2** Output of total and M3 isotopomer of <sup>13</sup>C<sub>3</sub> alanine during liver perfusion with [U-<sup>13</sup>C]glucose

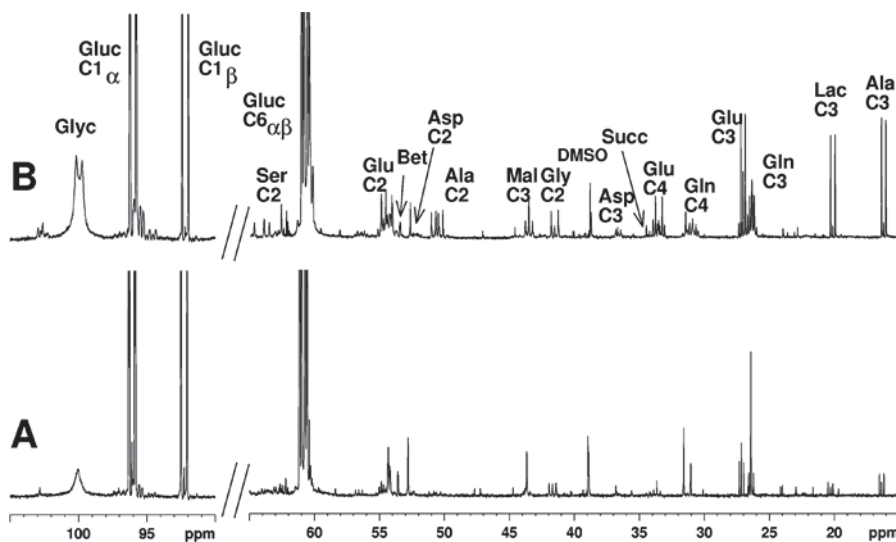
(A) Output of total alanine. (B) Output of M3 alanine (alanine labelled with <sup>13</sup>C at three carbons). Experimental details are as described in the legend to Figure S1. Results are means  $\pm$  S.D. for three independent liver perfusions per study group. M3 alanine isotopomer must have been derived from [U-<sup>13</sup>C]pyruvate, the product of [U-<sup>13</sup>C]glucose glycolysis. As indicated in the legend to Figure S1, these results demonstrate that the activation of GK by 3  $\mu\text{mol/l}$  Piragliatin was directly responsible for the remarkable elevation of total and <sup>13</sup>C-labelled alanine output from the perfusate [U-<sup>13</sup>C]glucose.

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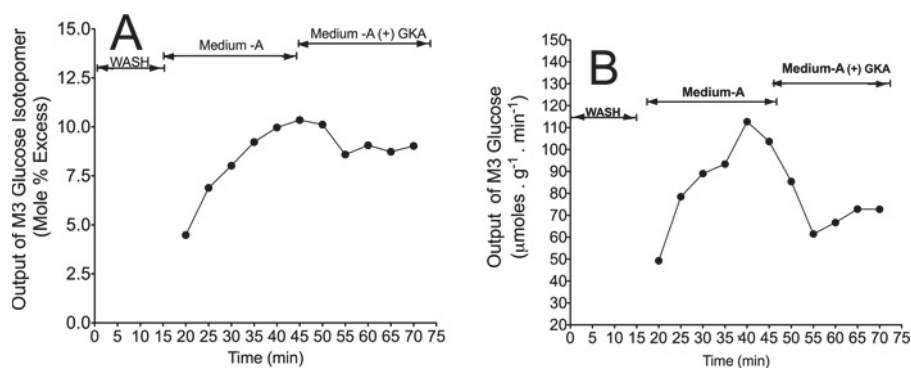
**Figure S3** Representative  $^{31}\text{P}$ -NMR spectra of freeze-clamped liver following perfusion without (A) or with (B) a GKA

Spectra demonstrate an approximately 2-fold higher peak of glycerol 3-phosphate (GI3P) following perfusion with a GKA (B). In addition, the inset represents the expansion of the G6P peak at approximately 4.7 p.p.m., which was higher following perfusion with a GKA (B) compared with the control (A). PCh, phosphocholine; GPE, glycerophosphoryl ethanolamine; GPC, glycerophosphocholine.



**Figure S4** Representative  $^{13}\text{C}$ -NMR spectrum of freeze-clamped liver following perfusion without (A) or with (B) GKA

These spectra demonstrate remarkably higher peaks of  $^{13}\text{C}$  isotomers of lactate, alanine, serine, glycine, malate, glutamate, aspartate and glycogen in liver perfused with GKA (B) compared with control (A). Bet, betaine; Gluc, glucose; Lac, lactate; Glyc, C1-glucosyl (glycogen); Mal, malate; Succ, succinate.



**Figure S5** Output of total and  $^{13}\text{C}$ -labelled glucose during liver perfusion with  $[\text{U}-^{13}\text{C}]$ -labelled pyruvate and lactate

(A) Output of M3 glucose (glucose labelled with  $^{13}\text{C}$  at three carbons). (B) Rate of M3 glucose output calculated as the rate of total glucose output ( $\mu\text{mol}\cdot\text{g}^{-1}\cdot\text{min}^{-1}$ ) multiplied by enrichment (MPE) of M3 isotopomer. Liver perfusion was carried out as detailed in the Experimental section in the main text. Briefly, after a 15 min conditioning of the liver (WASH), the perfusate was replaced with medium containing 0.1% DMSO, 2.1 mM lactate and 0.3 mM pyruvate (both  $[\text{U}-^{13}\text{C}]$  labelled), 0.3 mM  $^{15}\text{NH}_4\text{Cl}$  and 1mM glutamine (Medium-A). At 45 min, perfusion was continued with Medium-A plus 3  $\mu\text{mol/l}$  Piragliatin [(+) GKA]. The production of M3 glucose isotopomer was chosen as representative of gluconeogenesis from  $[\text{U}-^{13}\text{C}]$ pyruvate plus lactate. M3 glucose must have been directly derived from  $[\text{U}-^{13}\text{C}]$ labelled pyruvate plus lactate. In addition, LC-MS analysis demonstrated the production of M2, M4, M5 and M6 glucose, but with remarkably lower  $^{13}\text{C}$  enrichment than M3 glucose (results not shown). These data demonstrate that 3  $\mu\text{mol/l}$  Piragliatin inhibited gluconeogenesis from pyruvate plus lactate within 3–5 min after its addition to the perfusate.

**Table S1**  $^{13}\text{C}$  enrichment in the glycolytic intermediate G6P in freeze-clamped liver following perfusion without (control) or with a GKA [(+) GKA]

Results are means  $\pm$  S.D. of three livers per group.

$^{13}\text{C}$ Isotopomer	Control (MPE)	(+) GKA (MPE)
M + 1	0.4 $\pm$ 0.3	1.2 $\pm$ 1.0
M + 2	2.9 $\pm$ 0.9	2.2 $\pm$ 0.4
M + 3	5.1 $\pm$ 1.2	2.7 $\pm$ 1.6
M + 4	2.4 $\pm$ 0.8	1.8 $\pm$ 0.6
M + 5	4.4 $\pm$ 1.1	2.9 $\pm$ 1.4
M + 6	40.8 $\pm$ 6.4	57.6 $\pm$ 15.1
Sum	55.9 $\pm$ 4.9	68.7 $\pm$ 14.7

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