



Figure S1. The growth phenotypes of *M. alpina* grown on medium containing glucose (A), phenylalanine (B), phenylacetate (C), phenylalanine and p-chlorophenylalanine (D), tyrosine (E), or glucose and phenylalanine (F) as the carbon source. The data shown are the results of three independent experiments.

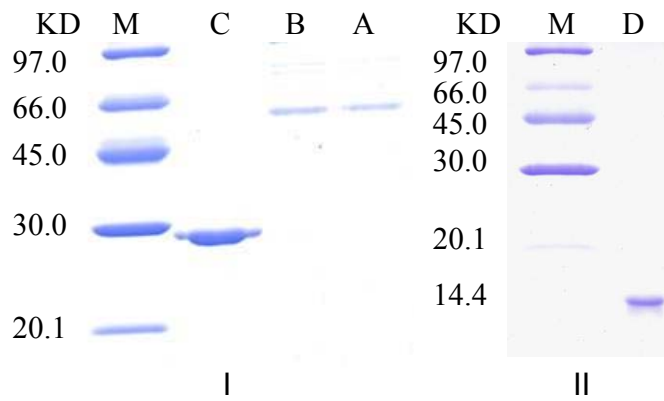


Fig. S2. SDS-polyacrylamide gel electrophoresis. The protein patterns are: (A) PAH-1; (B) PAH-2; (C) DHPR; (D) PCD; Proteins were denatured at 100°C for 5 min in 0.1% SDS and 1% 2-mercaptoethanol before being loaded in a 5% (wt/vol) stacking gel and separated in a 12% (I) or 15% (II) (wt/vol) separation gel. The gel was stained with coomassie bright blue R250. The molecular weight markers from GE Healthcare are indicated at each side of the panel.

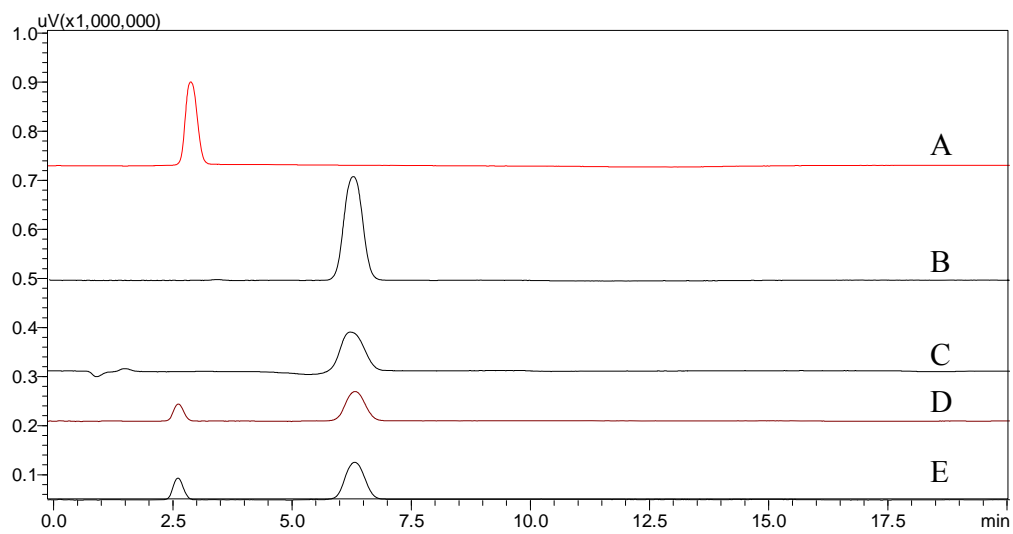


Figure S3. HPLC chromatographs of the reaction products: Tyrosine standard (A); Phenylalanine standard (B); Blank reaction (C); PAH-1 reaction (D); PAH-2 reaction (E).

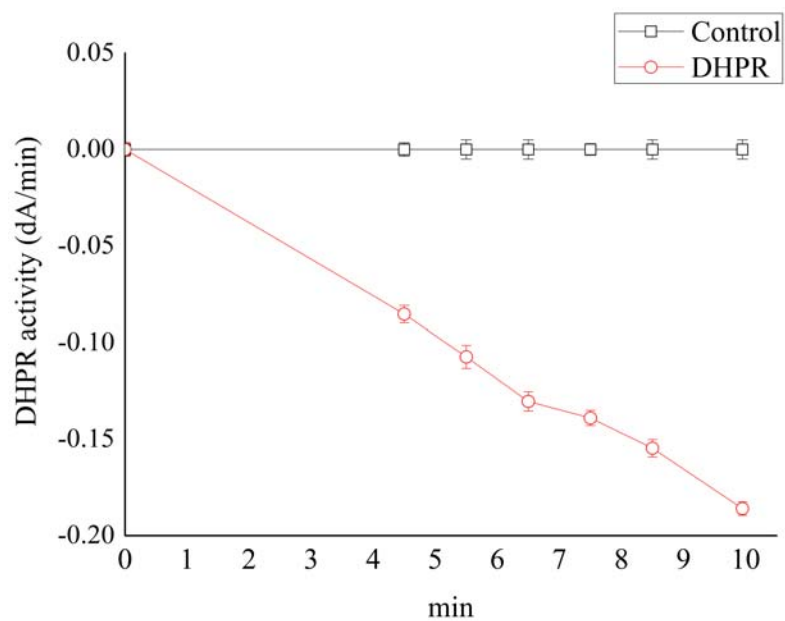


Figure S4. The purified DHPR protein shows DHPR enzyme activity. The open round circle (\circ) indicates the addition of a recombinant protein and the open square (\square) indicates no addition of protein. The data shown are the averages of three independent experiments.

1	Mortierella alpina PAH-1	MSAAAMHRCADTSVTP	VKTHHSTI	DKVGDQDC	MAIKAMHLSITRIBRFS	KIADWDYDFYDFM	AYDA
1	Mortierella alpina PAH-2	MSSTFCHLHKTDATFT	VKTHHSTA	DKVGDQDC	MAIKAMHLSITRIBRFS	KIADWDYDFYDFM	ATSA
1	Rhizopus oryzae PAH						
1	Soizellomyces punctatus PAH						
1	Alloomyces macrognus PAH						
1	Phycomyces blakesleeanus PAH						
1	Homo sapiens PAH						
1	Mus musculus PAH						
1	Rattus norvegicus PAH						
1	Drosophila melanogaster PAH						
1	Anopheles gambiae PAH						
1	Chlamydomonas reinhardtii PAH						
1	Physcomitrella patens PAH						
1	Pinus taeda PAH						
1	Dictyostelium discoideum PAH						
1	Caenorhabditis elegans PAH						
1	Mesorhizobium loti PAH						
1	Chromobacterium violaceum PAH						
1	Caulobacter crescentus PAH						
1	Colwellia Psychroerythraea PAH						
1	Vibrio cholerae PAH						
1	Kalstonia solanacearum PAH						
1	Pseudomonas aeruginosa PAH						
1	Homo sapiens TPH						
1	Mus musculus TPH						
1	Gallus gallus TPH						
1	Drosophila melanogaster TPH						
1	Mortierella alpina PAH-1						
1	Mortierella alpina PAH-2						
1	Rhizopus oryzae PAH						
1	Soizellomyces punctatus PAH						
1	Alloomyces macrognus PAH						
1	Phycomyces blakesleeanus PAH						
1	Homo sapiens PAH						
1	Mus musculus PAH						
1	Rattus norvegicus PAH						
1	Drosophila melanogaster PAH						
1	Anopheles gambiae PAH						
1	Chlamydomonas reinhardtii PAH						
1	Physcomitrella patens PAH						
1	Pinus taeda PAH						
1	Dictyostelium discoideum PAH						
1	Caenorhabditis elegans PAH						
1	Mesorhizobium loti PAH						
1	Chromobacterium violaceum PAH						
1	Caulobacter crescentus PAH						
1	Colwellia Psychroerythraea PAH						
1	Vibrio cholerae PAH						
1	Kalstonia solanacearum PAH						
1	Pseudomonas aeruginosa PAH						
1	Homo sapiens TPH						
1	Mus musculus TPH						
1	Gallus gallus TPH						
1	Drosophila melanogaster TPH						
74	Mortierella alpina PAH-1						
77	Mortierella alpina PAH-2						
135	Rhizopus oryzae PAH						
142	Soizellomyces punctatus PAH						
143	Alloomyces macrognus PAH						
144	Phycomyces blakesleeanus PAH						
145	Homo sapiens PAH						
146	Mus musculus PAH						
147	Rattus norvegicus PAH						
148	Drosophila melanogaster PAH						
149	Anopheles gambiae PAH						
150	Chlamydomonas reinhardtii PAH						
151	Physcomitrella patens PAH						
152	Pinus taeda PAH						
153	Dictyostelium discoideum PAH						
154	Caenorhabditis elegans PAH						
155	Mesorhizobium loti PAH						
156	Chromobacterium violaceum PAH						
157	Caulobacter crescentus PAH						
158	Colwellia Psychroerythraea PAH						
159	Vibrio cholerae PAH						
160	Kalstonia solanacearum PAH						
161	Pseudomonas aeruginosa PAH						
162	Homo sapiens TPH						
163	Mus musculus TPH						
164	Gallus gallus TPH						
165	Drosophila melanogaster TPH						

Fig. S5. Multiple amino acid sequence alignment of amino acid hydroxylases from fungi, animal, protist, plant and bacteria. Multiple alignments were performed with the ClustalW program using the BioEdit sequence alignment software. The corresponding nucleotide sequences encoding *M. alpina* PAH has been deposited in the GenBank/EMBL database with Accession No. (JN982953 and JN982954). Accession numbers for the other PAH protein sequences above are: *Rhizopus_oryzae* (RO3G_07757.1), *Spizellomyces punctatus* (SPPG_07026T0), *Allomyces macrogynus* (AMAG_15797T0), *Phycomyces blakesleeanus* (Jgi|Phybl2|161646), *Homo sapiens* (AAH26251.1), *Mus musculus* (CAA36205.1), *Rattus norvegicus* (AAA41843.1), *Drosophila melanogaster* (AAA69513.1), *Anopheles gambiae* (EAA10731.4), *Chlamydomonas reinhardtii* (HQ003816.1), *Physcomitrella patens* (HQ003815.1), *Pinus taeda* (HQ003814.1), *Dictyostelium discoideum* (EAL67992.1), *Caenorhabditis elegans* (CAA91286.1), *Mesorhizobium loti* (BAB51399.1), *Chromobacterium violaceum* (AAA23115.1), *Caulobacter crescentus* (AAK23591.1), *Colwellia Psychrerythraea* (150261591), *Vibrio cholerae* (AAF96726.1), *Ralstonia solanacearum* (CAD17143.1), and *Pseudomonas aeruginosa* (AAA25936.1). Accession numbers for the TPH protein sequences above are: *Homo sapiens* (AAA67050.1), *Mus musculus* (AAA63401.1), *Gallus gallus* (AAC60036.1), and *Drosophila melanogaster* (AAF47444.1). Accession numbers for the TH protein sequences above are: *Homo sapiens* (AAI04968.1), *Mus musculus* (AAA40434.1), *Gallus gallus* (CAB62388.1), and *Drosophila melanogaster* (CAA53802.1).



Fig. S6. Phylogenetic tree of amino acid hydroxylases from different organisms (PAH: Phenylalanine hydroxylase; TPH: Tryptophan hydroxylase; TH: Tyrosine hydroxylase). The tree was constructed using the neighbor-joining method with CLUSTAL W NJPLOT. The horizontal branch length is proportional to the amino acid substitution rate per site.

The numbers represent the frequencies with which the tree topology presented here was replicated after 1000 bootstrap iterations.

grisea 70-15(3), *Neurospora crassa* OR74A(7), *Neosartorya fischeri* NRRL 181, *Podospora anserina* DSM 980(6), *Phanerochaete chrysosporium*, *Pichia stipitis* CBS 6054(10), *Saccharomyces bayanus*(14), *Saccharomyces cerevisiae* S288C(9), *Saccharomyces mikatae*(14), *Saccharomyces paradoxus*(14), *Schizosaccharomyces pombe* 972h(23), *Sclerotinia sclerotiorum* 1980 UF-70, *Ustilago maydis* 521(11), *Yarrowia lipolytica* CLIB122(5), *Schizosaccharomyces cryophilus* OY26, *Schizosaccharomyces japonicus* yFS275 and *Schizosaccharomyces octosporus* ATCC4206 (21), and *Vanderwaltozyma polyspora* DSM 70294(22) were published. Genome sequences of *Nectria haematococca* MPVI (*Fusarium solani*) (Joint Genome Institute), *Trichoderma atroviride* P1 (Joint Genome Institute), *Trichoderma virens* Gv29-8 (Joint Genome Institute), *Batrachochytrium dendrobatidis* JAM81 (Joint Genome Institute), *Mucor circinelloides* (Joint Genome Institute), *Trichoderma reese* QM6a (Joint Genome Institute), *Neosartorya fischeri* NRRL 181 (Broad Institute), and *Phytophthora infestans* T30-4 (Broad Institute) were downloaded from the corresponding websites.

Table. S2. The status of fatty acids in *M. alpina* and *A. oryza* grown on PAH inhibitor medium^a

Medium	Cell dry weight (CDW) (g/L)	Cell lipid (mg/g CDW)	SAFA						MUFA							
			14:0	16:0	18:0	20:0	22:0	24:0	14:1	16:1	total 18:1	oleate	vaccenate	20:1	22:1	24:1
Inhibitor (<i>M. alpina</i>)	5.3±0.2	24.1±1.3	2.4±0.1	18.5±1.1	8.7±0.6	0.5±0.1	1.4±0.2	2.9±0.1		0.1±0.0	24.2±1.4	23.8±1.2	0.4±0.1	0.8±0.1		0.2±0.0
Inhibitor+Tyrosine (<i>M. alpina</i>)	5.1±0.2	24.2±1.2	2.2±0.1	19.7±0.9	7.9±0.3	0.5±0.1	1.6±0.2	4.2±0.2		0.1±0.0	24.9±1.3	24.5±1.2	0.4±0.1	0.7±0.1		0.1±0.0
Control (<i>M. alpina</i>)	6.6±0.2	35.5±1.5	1.3±0.1	16.2±1.5	11.0±0.8	0.6±0.1	1.3±0.1	1.9±0.1		0.1±0.0	11.7±1.8	11.4±1.6	0.3±0.0	0.3±0.0		0.2±0.0
Inhibitor (<i>A. oryzae</i>)	7.0±0.2	11.6±0.2	0.4±0.0	11.8±0.3	15.4±0.5	0.4±0.0	0.4±0.0	2.2±0.1		0.3±0.0	20.7±1.0	20.6±1.0	0.1±0.0	0.1±0.0		
Control (<i>A. oryzae</i>)	7.3±0.3	11.2±0.2	0.4±0.0	11.7±0.2	14.8±0.8	0.4±0.0	0.4±0.0	2.1±0.1		0.3±0.0	21.2±1.2	21.1±1.2	0.1±0.0	0.1±0.0		
Medium	ω6 PUFA					ω3 PUFA										
	18:2	20:2	18:3 n-6	20:3	20:4	18:3 n-3	20:5	22:5	22:6	24:5	24:6					
Inhibitor (<i>M. alpina</i>)	18.8±1.2	0.7±0.1			20.8±0.8		0.1±0.0									
Inhibitor+Tyrosine (<i>M. alpina</i>)	19.1±0.9	0.6±0.1			18.1±0.7		0.1±0.0									
Control (<i>M. alpina</i>)	20.8±1.7	0.8±0.1			33.9±2.1		0.1±0.0									
Inhibitor (<i>A. oryzae</i>)	46.7±2.3	0.1±0.0				1.5±0.1										
Control (<i>A. oryzae</i>)	47.0±2.0	0.1±0.0				1.5±0.1										

^aMolar percent of each fatty acid in day 6 mycelia. 14:0 myristic acid (tetradecanoic acid), 14:1 myristoleic acid (tetradecenoic acid), 16:0 palmitic acid (hexadecanoic acid), 16:1 palmitoleic acid (hexadecenoic acid), 18:0 stearic acid (octadecanoic acid), 18:1 oleic acid (9Z-octadecenoic acid) and vaccenic acid (11Z-octadecenoic acid), 18:2 linoleic acid (octadecadienoic acid), 18:3 α-linolenic acid (9Z,12Z,15Zoctadecatrienoic acid) and γ-linolenic acid (6Z,9Z,12Z-octadecatrienoic acid), 20:0 arachidic acid (eicosanoic acid), 20:1 gondoic acid (eicosenoic acid), 20:2 (Eicosadienoic acid), 20:3 dihomo-γ-linolenic acid (eicosatrienoic acid), 20:4 arachidonic acid (eicosatetraenoic acid), 20:5 EPA (eicosapentaenoic acid), 22:0 behenic acid (docosanoic acid), 22:1 erucic acid

(docosenoic acid), 22:5 DPA (docosapentaenoic acid), 22:6 DHA (docosahexaenoic acid), 24:0 lignoceric acid (tetracosanoic acid), 24:1 nervonic acid (tetracosenoic acid), 24:5 (tetracosapentaenoic acid), and 24:6 (tetracosahexaenoic acid). The data shown are the averages of three independent experiments.

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