

**SUPPLEMENTARY ONLINE DATA****The dynamic action of SecA during the initiation of protein translocation**Vicki A. M. GOLD<sup>1</sup>, Sarah WHITEHOUSE, Alice ROBSON and Ian COLLINSON<sup>2</sup>

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**MATERIALS AND METHODS****Steady-state ATPase and determination of the  $K_m$** 

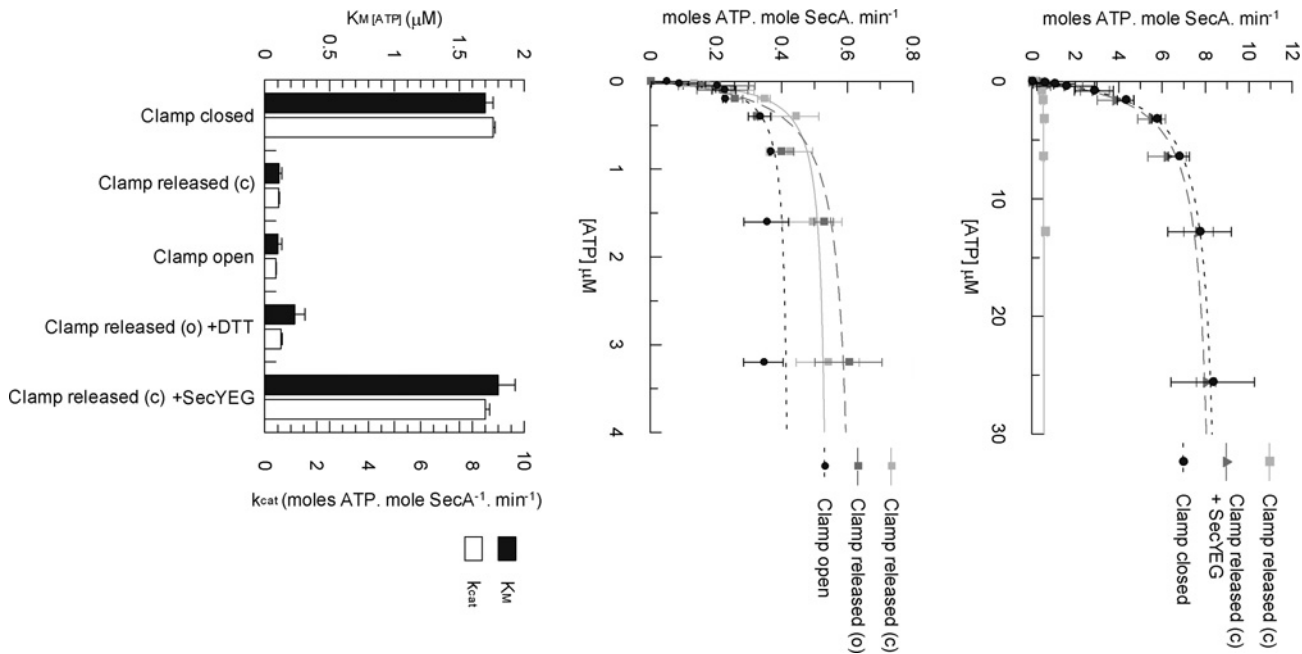
Steady-state SecA ATPase measurements were performed as described previously [1,2] with various concentrations of ATP. The  $K_m$  and  $V_{max}$  ( $k_{cat}$ ) were determined by fitting the data to the Michaelis–Menten equation (eqn S1)

$$v = \frac{V_{max} \cdot [S]}{K_m + [S]}$$

where  $v$  is equal to the enzyme velocity,  $V_{max}$  is the total capacity of the substrate-associated ATPase stimulation,  $[S]$  is the substrate concentration and  $K_m$  is the Michaelis constant.

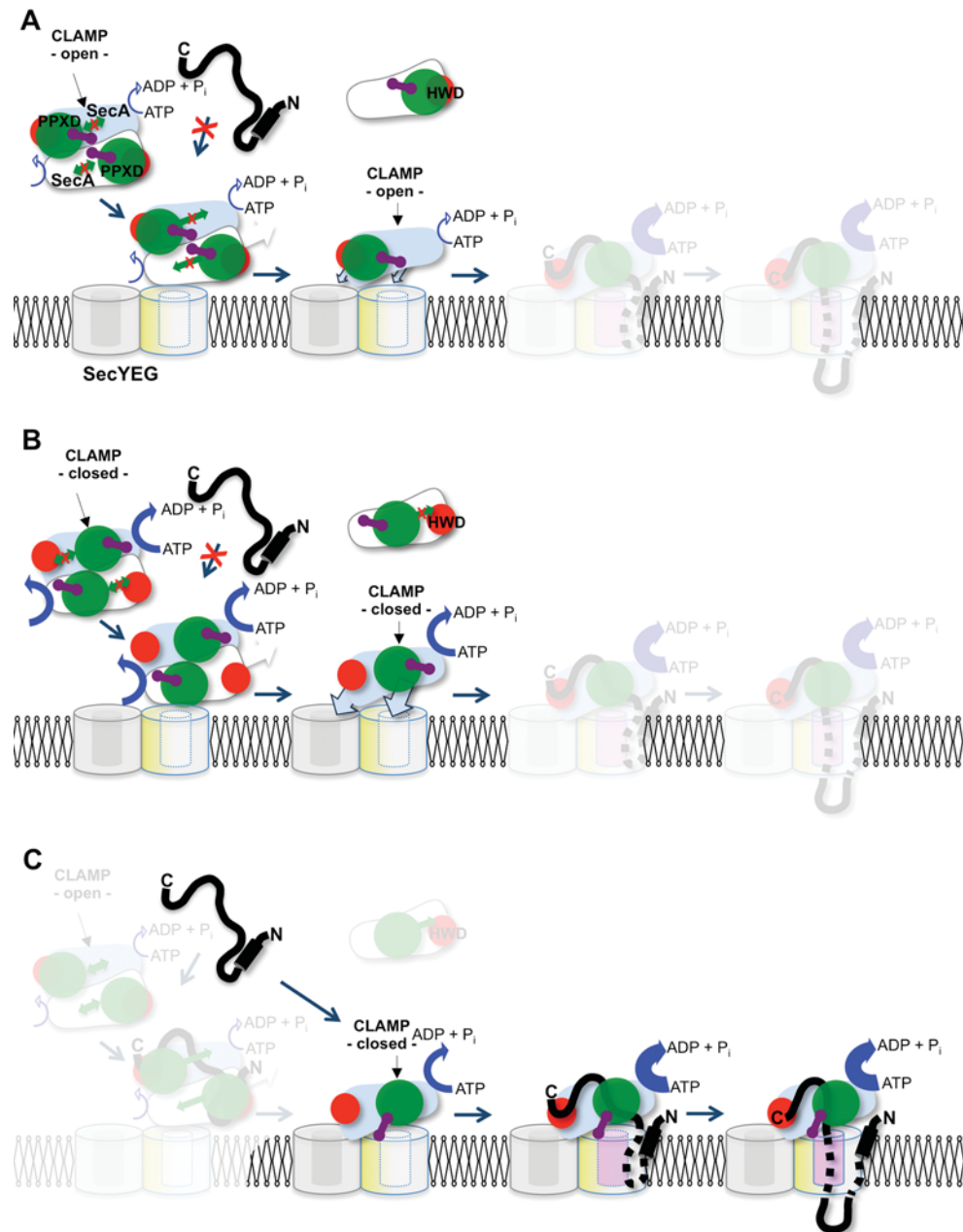
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**Table S1** Kinetic parameters

Various parameters derived from data fits (see individual Figure legends in the main text). The data for the clamp-closed SecA are shown in bold to emphasize the significant differences in this version. Errors represent S.D. values from the fit.

Condition	SecA-SecYEG <sub>soluble</sub>		SecA-SecYEG <sub>CL</sub>		SecA-ATP		SecA <sub>(2HF)</sub> -SecY <sub>K268F</sub> EG <sub>soluble</sub>		SecA-SecYEG <sub>vesicles</sub>		[SecA-pOA]-SecYEG <sub>vesicles</sub>		[SecA-SecYEG <sub>vesicles</sub> ]-pOA	
	1 <i>K<sub>d</sub></i> (μM)	2 <i>k<sub>cat</sub></i>	3 <i>K<sub>d</sub></i> (nM)	4 <i>k<sub>cat</sub></i>	5 <i>K<sub>m</sub></i> (μM)	6 <i>k<sub>cat</sub></i>	7 <i>K<sub>d</sub></i> (nM)	8 ΔFI (%)	9 <i>K<sub>d</sub></i> (μM)	10 <i>k<sub>cat</sub></i>	11 <i>K<sub>d</sub></i> (μM)	12 <i>k<sub>cat</sub></i>	13 <i>K<sub>d</sub></i> (μM)	14 <i>k<sub>cat</sub></i>
Δcys	7.6 ± 0.6	31.4 ± 0.6	3.7 ± 1.9	74.9 ± 3.5	-	-	18.4 ± 0.6	35.3 ± 0.2	1.13 ± 0.32	154.2 ± 22.0	0.26 ± 0.04	656.4 ± 31.0	0.50 ± 0.12	605.8 ± 38.3
Δcys + DTT	7.8 ± 1.4	25.0 ± 1.1	5.6 ± 0.6	82.7 ± 0.9	-	-	20.6 ± 2.1	36.0 ± 1.3	1.11 ± 0.26	149.4 ± 17.2	0.19 ± 0.03	631.6 ± 22.6	0.78 ± 0.10	604.6 ± 20.2
<b>Clamp closed</b>	<b>0.015 ± 0.011</b>	<b>23.2 ± 0.6</b>	-	-	<b>1.7 ± 0.06</b>	<b>8.8 ± 0.08</b>	<b>28.6 ± 5.0</b>	<b>20.9 ± 1.4</b>	<b>0.029 ± 0.007</b>	<b>99.7 ± 3.0</b>	<b>0.025 ± 0.007</b>	<b>131.7 ± 4.5</b>	-	-
Clamp released (c)	6.3 ± 0.7	22.5 ± 0.7	8.8 ± 2.1	59.1 ± 1.9	0.11 ± 0.02	0.54 ± 0.02	117.7 ± 12.7	23.5 ± 0.8	0.34 ± 0.20	59.4 ± 8.2	0.13 ± 0.02	253.6 ± 10.0	0.24 ± 0.06	231.6 ± 13.3
Clamp open	5.9 ± 2.1	9.9 ± 0.5	-	-	0.10 ± 0.03	0.43 ± 0.02	60.9 ± 10.0	15.0 ± 0.6	0.65 ± 0.78	22.5 ± 7.8	0.17 ± 0.18	21.1 ± 3.5	-	-
Clamp released (o)	6.0 ± 1.4	15.9 ± 0.6	6.5 ± 0.9	31.2 ± 0.4	0.23 ± 0.08	0.63 ± 0.05	59.5 ± 6.4	21.3 ± 0.6	0.65 ± 0.42	29.9 ± 6.4	0.13 ± 0.07	90.3 ± 9.7	0.35 ± 0.15	50.5 ± 4.7
Clamp released (c) + SecYEG	-	-	-	-	1.8 ± 0.13	8.5 ± 0.17	-	-	-	-	-	-	-	-



**Figure S2 The consequences of the immobilization of the PPXD within SecA to protein transport**

The model of pre-protein transport (Figure 7A of the main text) has been modified to incorporate the intra- and inter-molecular disulfide bonds, and to explain their effect on the activity. Colour co-ordination and labelling is as in Figure 7 of the main text. Stages of the mechanism precluded as a result have been fogged out. (A) When the PPXD is cross-linked (purple bar) to the HSD the clamp is permanently held open. This prevents the activation of the ATPase activity (thin blue arrows), maintains a low affinity for SecYEG (small pale blue arrows) and prevents association with the pre-protein. (B) Fixing the PPXD to NBD2, holding the clamp closed, activates the ATPase activity (blue arrows), brings about a high-affinity association with SecYEG (large pale blue arrows), but prevents the engagement of pre-protein. (C) The pre-activated cross-linked SecA–SecYEG complex is primed for translocation and capable of pre-protein intercalation and ATP-driven (thick blue arrows) translocation.

## REFERENCES

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Received 20 August 2012/15 October 2012; accepted 5 November 2012  
Published as BJ Immediate Publication 5 November 2012, doi:10.1042/BJ20121314