

Figure S1. Up-regulation of CD44 on pmel-1 T cells after culture. WT, T-bet^{-/-}, Eomes^{-/-}, or T-bet/Eomes DKO pmel-1 T cells which have been cultured in Th1 conditions for 3 days. The expression of CD44 on these cells was analyzed by flow cytometry. Data represents three independent experiments. One mouse from each strain was used in each experiment.

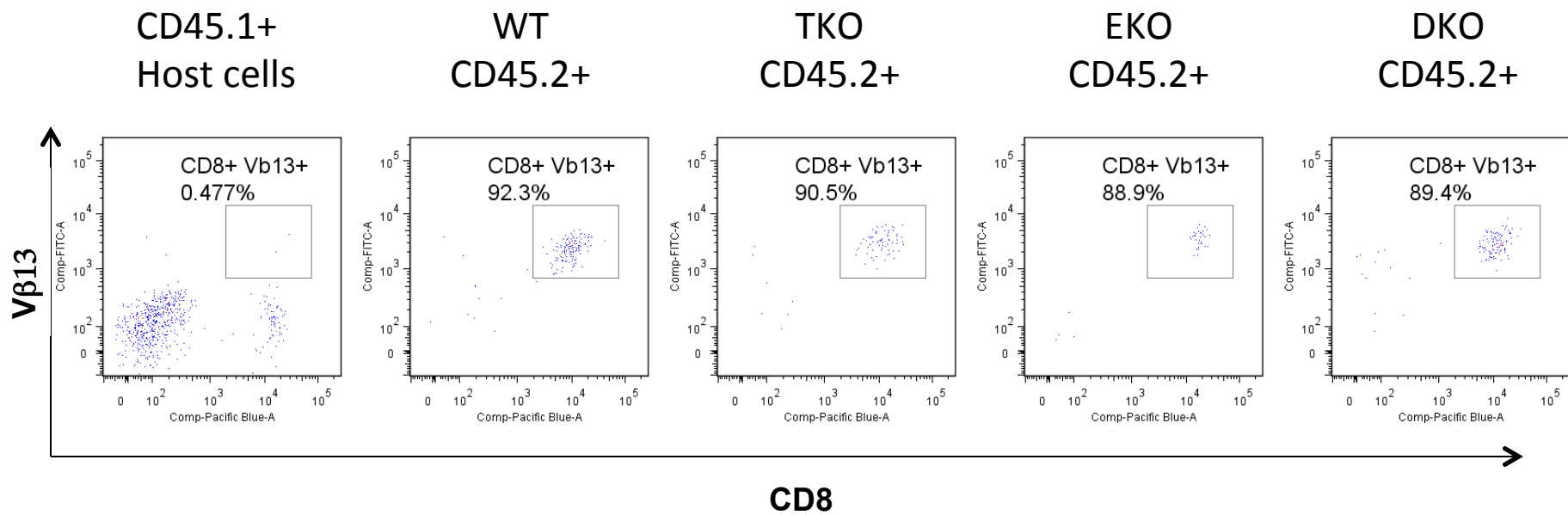


Figure S2. Pmel-1 T cells were the majority of the adoptively transferred cells. B6-LY5.2/Cr mice were challenged with 3×10^5 B16F0 cells *i.d.* 6 days later, mice were irradiated at 500 rad. On day 7, the mice were adoptively transferred with 5×10^5 WT, T-bet^{-/-}, Eomes^{-/-}, or T-bet/Eomes DKO pmel-1 T cells which had been cultured in Th1 conditions for 3 days. Mice were bled three days after T cell infusion. The percentage CD8⁺ Vβ13⁺ (pmel-1) T cells among the donor cells (CD45.2⁺) cells was determined by flow cytometry and the representative dot plots were shown.

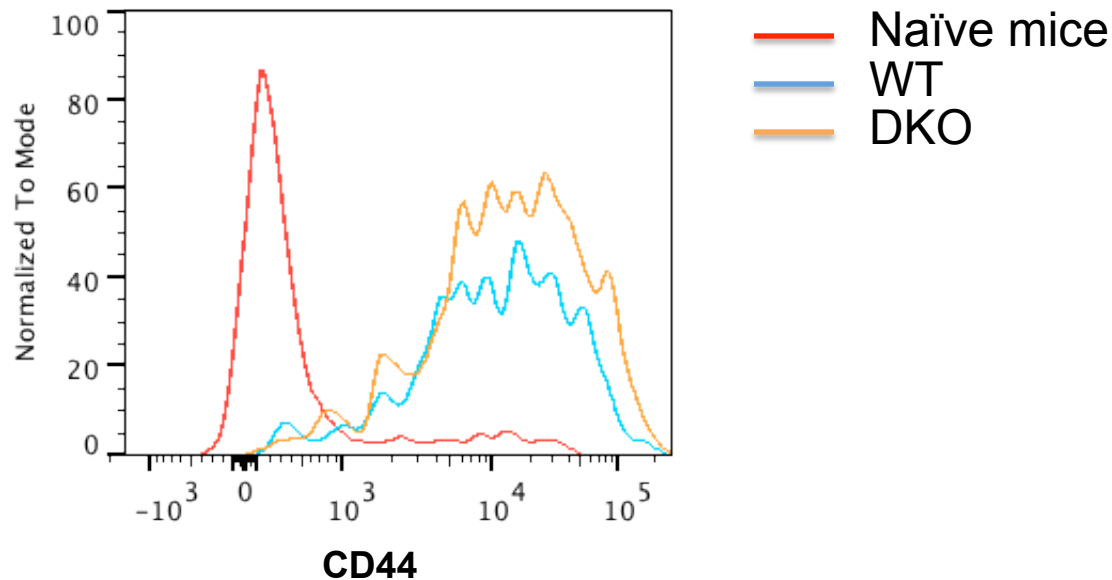


Figure S3. Up-regulation of CD44 on memory pmel-1 T cells after ex vivo re-stimulation. B6-LY5.2/Cr mice were irradiated at 500 rad, within 24 hours of irradiation, mice were adoptively transferred with 4×10^6 pmel-1 T cells (WT and DKO) which have been cultured in Th1 conditions for 4 days. Thirty days post T cell infusion, spleens were harvested. Splenocytes were stimulated with 0.01 mM gp100₂₅₋₃₃ peptide for 72 h hours. Then the donor cells (CD45.2⁺CD8⁺) were analyzed by flow cytometry for CD44 expression (WT denotes wild type donor cells, DKO denotes T-bet ^{-/-} Eomes ^{-/-} donor cells, and naïve mice denote CD8 T cells from unmanipulated wild type mice which were used as control).