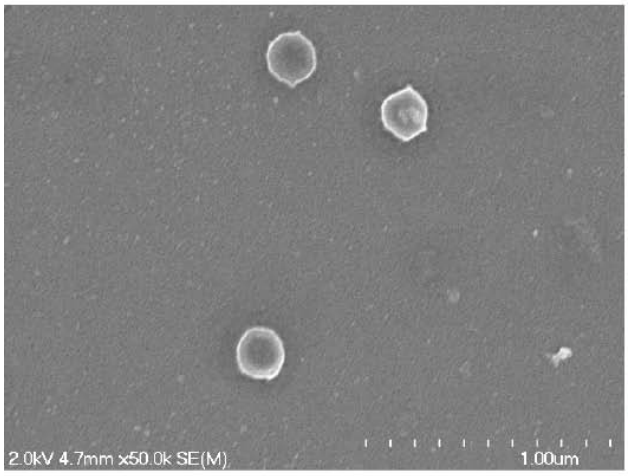
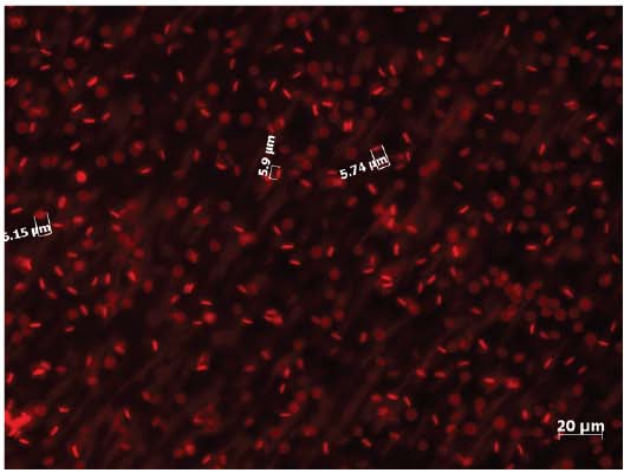
A

Particle	Size	PDI	Zeta (mV)
200nm PRINT cylinders	305.1nm	.066	-26.4
6µm PRINT microparticles	5.9µm	NA	-18.6
Qdots	32.1nm	.328	-37.8

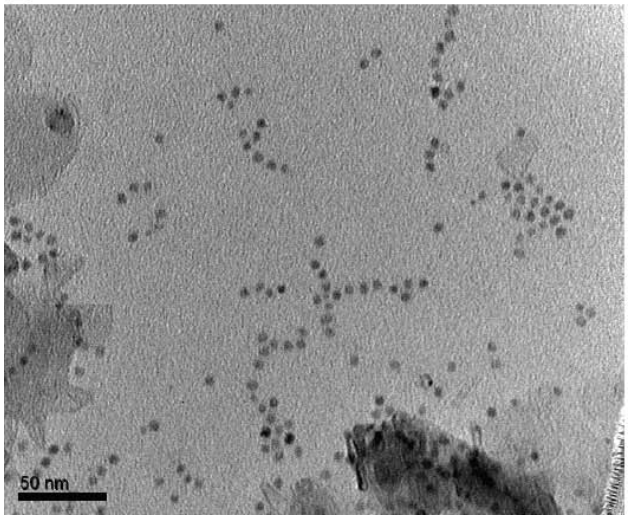
B



C

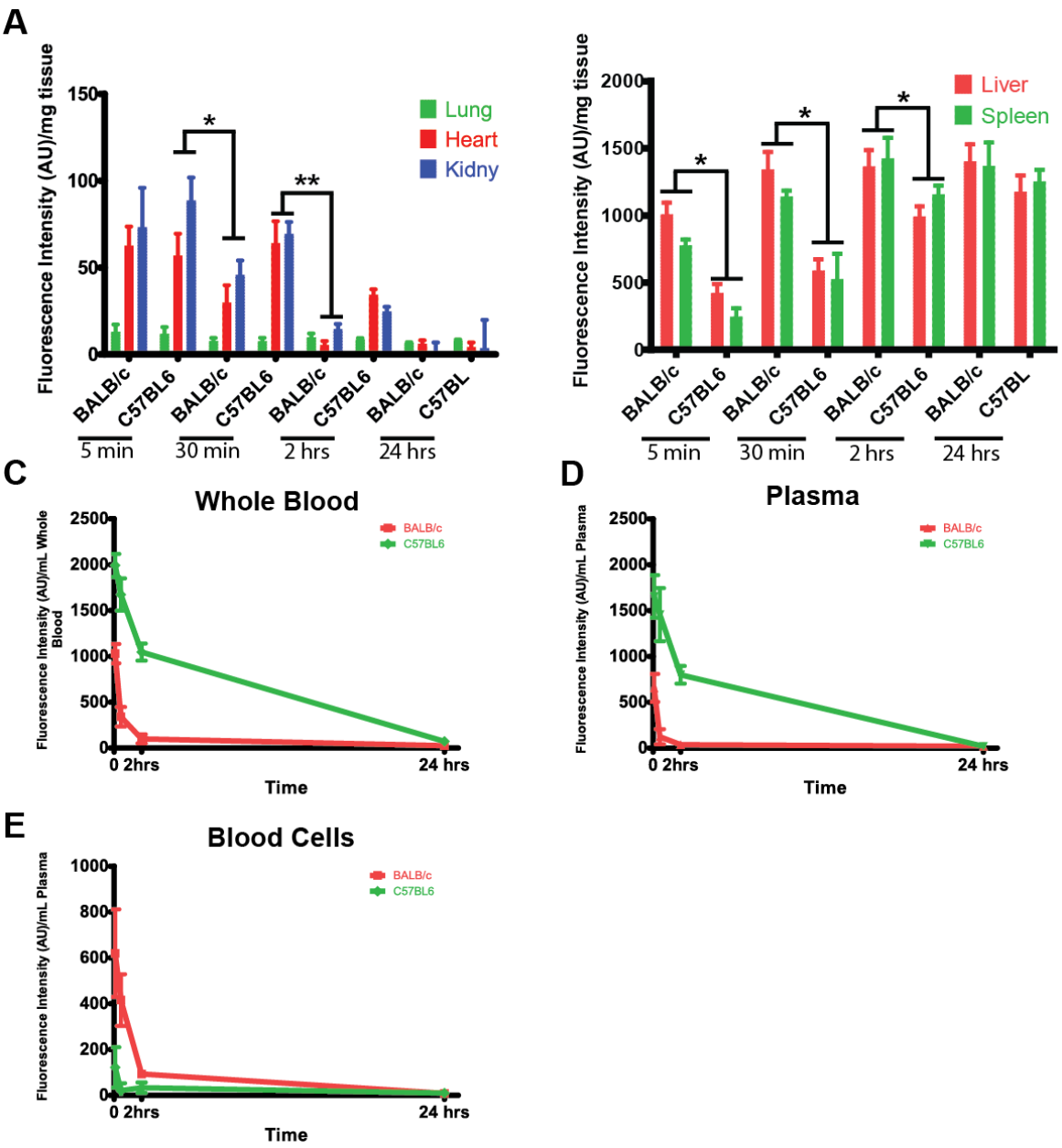


D



Supplemental Figure 1: Characterization of particles used in experiments.

- **(A) Size, PDI, and Zeta potential for all particles used. Size was determined by dynamic light scattering for 200nm PRINT particles and Qdots, fluorescence microscopy was used to determine size of microparticles.**
- **(B) Scanning electron micrograph of 200nm PRINT particles.**
- **(C) Fluorescence microscopy image of microparticles scale bar is 20µm.**
- **(D) TEM image of Qdots on glass scale bar is 50nm**



Supplemental Figure 2: Biodistribution of 300nm PRINT hydrogel particles in Balb/c and C57BL/6 mice.

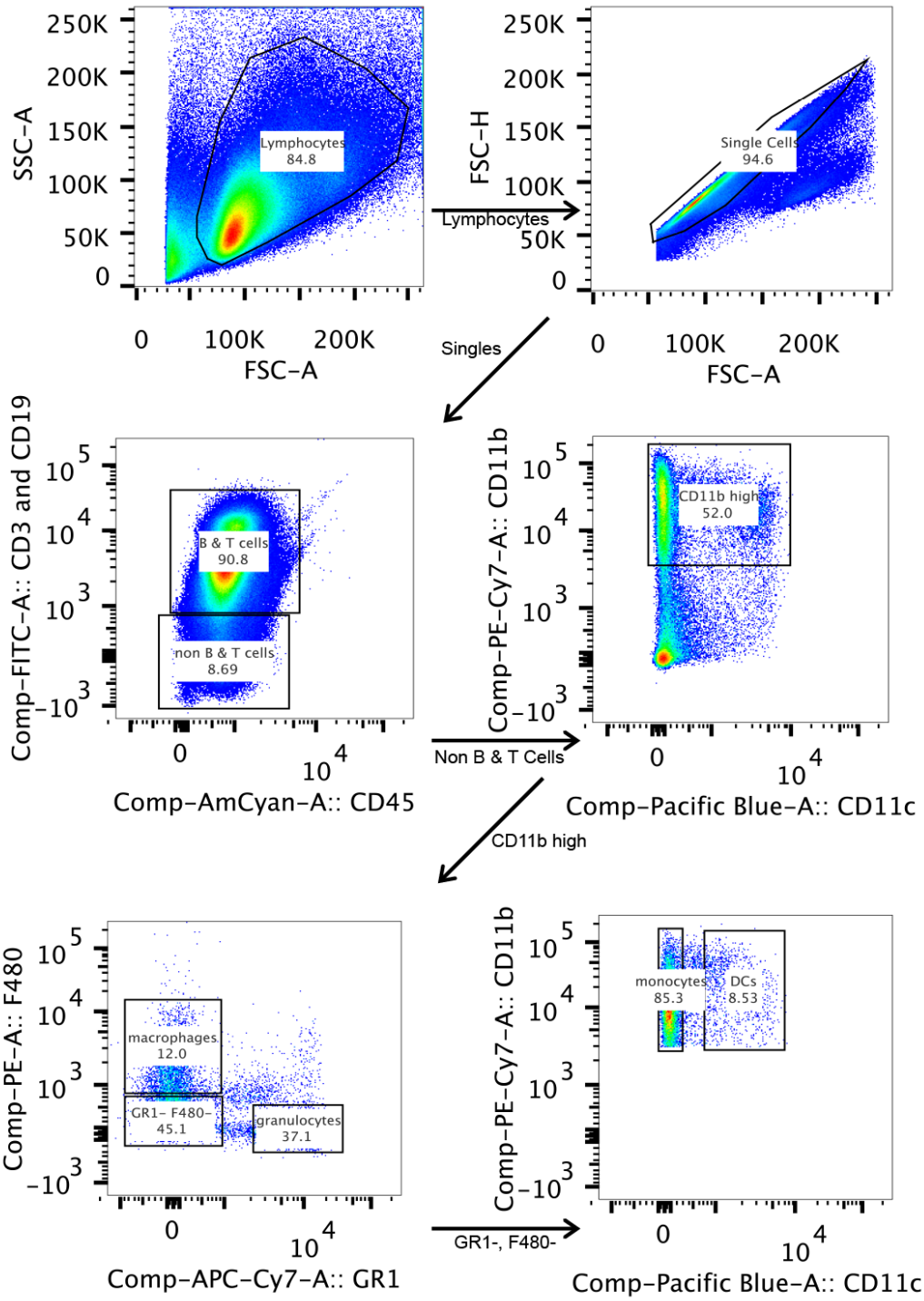
(A) Distribution of particles in the lungs, heart, and kidneys of Balb/c and C57BL/6 mice. C57BL/6 mice showed significantly higher amounts of particle in heart and kidneys then Balb/c mice at 30min ($P<.05$ t-test), and 2hrs ($P<.004$ t-test) (N=4).

(B) Distribution of particles in the liver and spleen of Balb/c and C57BL/6 mice. C57BL/6 mice showed significantly lower amounts of particle present in both liver and spleen compared to Balb/c mice at 5min, 30min and 2hrs ($P<.05$ t-test) (N=4).

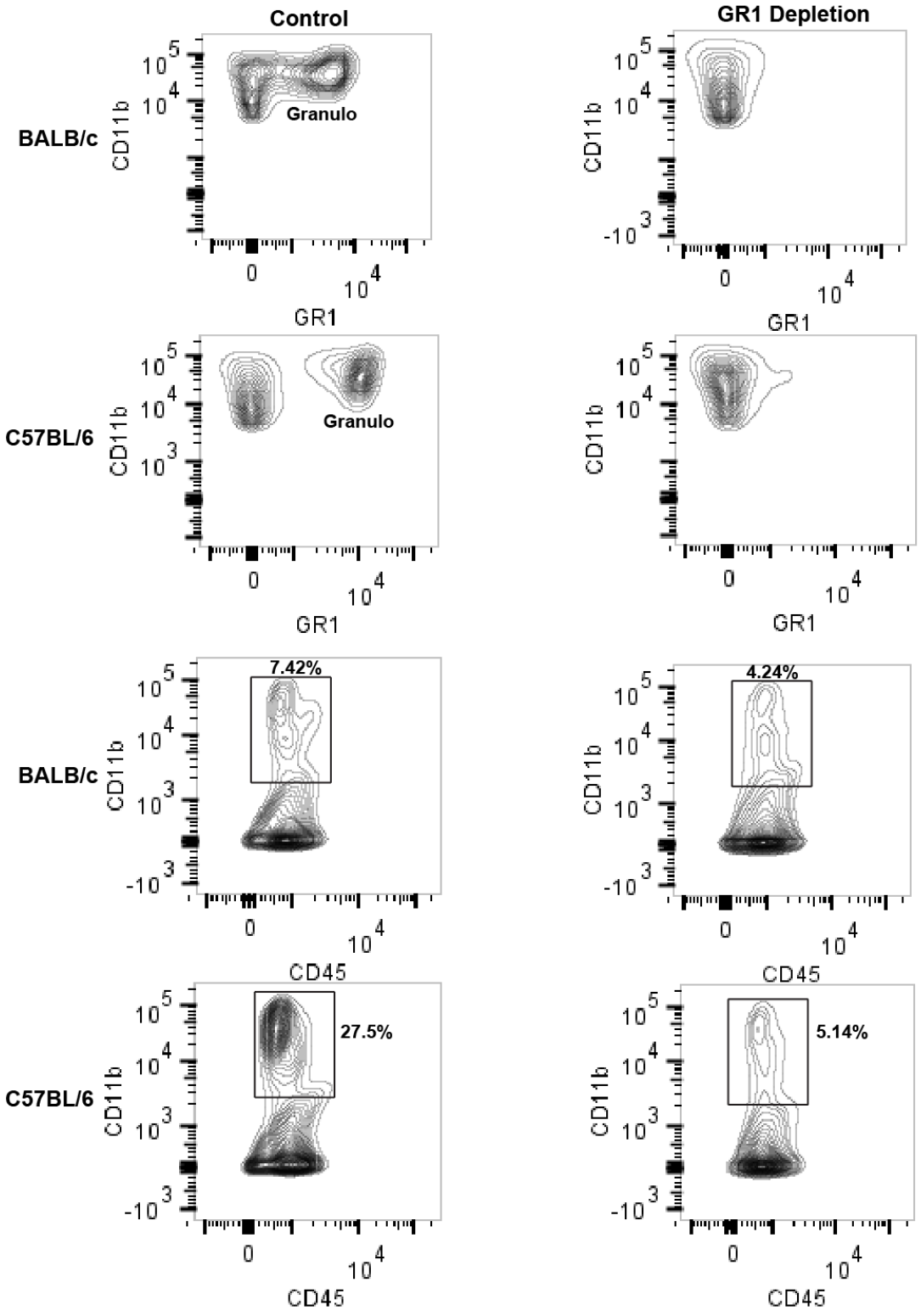
(C) Relative amounts of particle present in Balb/c and C57BL/6 whole blood, C57BL/6 had significantly higher levels of particles in blood at 5min, 30min and 2hrs compared to Balb/C mice ($P<.05$ t-test) (N=4).

(D) Relative amounts of particle present in Balb/c and C57BL/6 plasma, C57BL/6 had significantly higher levels of particles in blood at 5min, 30min and 2hrs compared to Balb/C mice ($P<.05$ t-test) (N=4).

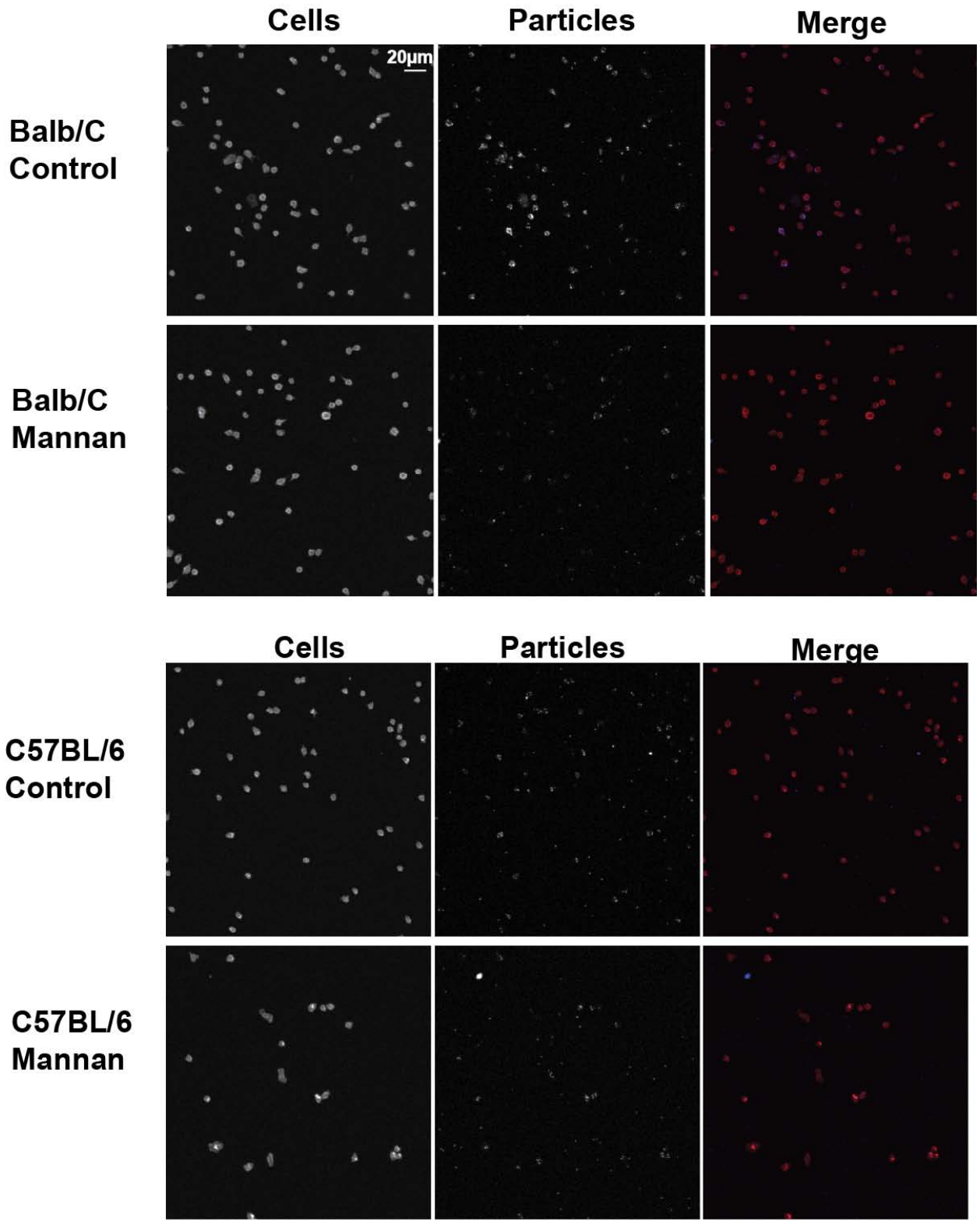
(E) Relative amounts of particle present in Balb/c and C57BL/6 blood cell fraction of whole blood, C57BL/6 had significantly higher lower of particles in blood at 5min, and 30min compared to Balb/C mice ($P<.05$ t-test) (N=4).



Supplemental Figure 3. Representative gating scheme from BALB/c mouse spleen. Single lymphocytes were gated by side scatter vs. forward scatter. Single cells were divided into B & T cells VS. non B & T cells by expression of CD3 and CD19. For Non B & T cells those expressing high CD11b were selected for further analysis. CD11b high were plotted for F480 and GR1, GR1 high F480- were categorized as granulocytes, F480high GR1- were categorized as macrophages. GRI- F480- cells were further divided into monocytes (CD11c low), and DC's (CD11c high).



Supplemental Figure 4: Depletion of granulocytes from mice using rat anti-mouse GR-1 antibodies.



Supplemental Figure 5: Confocal microscopy of Balb/c and C57BL/6 BMMs with and without mannan treatment.