SUPPLEMENTARY MATERIALS

Pulmonary CD103⁺ dendritic cells prime Th2 responses to inhaled allergens

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This file includes:

Legends for Supplementary Figures 1 through 11

Supplementary Figure Legends

Supplementary Figure 1 CD103⁺ DCs promote Th2 differentiation of naïve CD4⁺ T cells. DCs were purified from lungs of C57BL/6 mice 16 h post-OVA inhalation and cocultured with naïve CD4⁺ T cells from OT-II mice. (**a**) Proliferation of T cells as inferred from T cell recovery after 5 d of co-culture. (**b**) Cytokines in supernatants of 5 d cultures, as measured by ELISA.

Supplementary Figure 2 CD103⁺ DCs from BALB/c mice promote Th2 differentiation of naïve CD4⁺ T cells. DCs were prepared from lungs of BALB/c mice 16 h post-OVA inhalation and co-cultured with naïve CD4⁺ T cells from DO11.10 mice. (**a**) Proliferation of T cells as inferred from T cell recovery after 5 d of co-culture, and cytokines in supernatants of 5 d co-cultures. (**b**) Cytokines in supernatants of DO11.10 CD4⁺ T cells after 5 d of co-culture and subsequent incubation in anti-CD3 ϵ and anti-CD28-coated plates for 24 h. Cytokine levels assessed by ELISA.

Supplementary Figure 3 Effect of enzymatic digestion on lung resident DC function. DCs were prepared from lungs of OVA-treated mice, with or without digestion with Liberase TM, collagenase XI, Hyaluronidase and DNase. (**a**) CD11b^{hi} and CD103⁺ DCs recovered after flow cytometry-based sorting. (**b**) CD4 T cell proliferation in 5 d culture. (**c**) IL-4 production by T cells incubated for 24h with anti-CD3 ϵ and CD28 antibodies after 5 d of culture. Data shown are from one of two independent experiments giving similar results.

Supplementary Figure 4 Effect of adjuvants on T cell priming by CD103⁺ DCs. Lung CD103⁺ DCs were prepared from C57BL/6 mice that received OVA (100 μ g) together with either LPS (1 ng) or poly I:C (1 μ g). These DCs were cultured for 5 d with naïve CD4⁺ T cells from *Rag2^{-/-}* OT-II mice. IFN- γ and IL-4 production by T cells were assessed following 24 h incubation with anti-CD3 ϵ and -CD28 antibodies.

Supplementary Figure 5 Impaired allergic airway inflammation in mice lacking CD103⁺ DCs. WT B6C3F1 and CD103⁺ DC-deficient BXH2 mice were sensitized by oropharyngeal aspiration of OVA-LPS, and subsequently challenged by exposure to

2

aerosolized OVA. The number of total leukocytes and indicated subsets in BALF from unsensitized, challenged mice (open column) and from sensitized and challenged animals (filled column) are shown. *p*-value by Student's *t*-test is indicated (n=7).

Supplementary Figure 6 Notch ligand expression by lung CD11b^{hi} and CD103⁺ DCs. Lung DC subsets were purified from C57BL/6 mice 16 h after OVA-LPS instillation. mRNA levels of *Jagged 1*, *Jagged 2* and *Delta-like 4* in the indicated subsets were determined by quantitative real time PCR and normalized to GAPDH mRNA. Results of 3 independent experiments are shown.

Supplementary Figure 7 Cytokine production by lung CD11b^{hi} or CD103⁺ DCs. (**a**, **b**) Lung DC subsets were purified from naïve C57BL/6 mice, activated with PMA and ionomycin, and the culture supernatants analyzed for the indicated cytokines (**a**) and chemokines (**b**). (**c**) Intracellular IFN- γ staining of total non-autofluorescent CD11c^{hi} lung DCs. (**d**) Intracellular staining of CD11b^{hi} or CD103⁺ DCs with anti-IFN- γ antibodies (solid red line) or rat IgG₁ isotype control (dotted blue line) after stimulation with PMA and ionomycin. (**e**) Percentages of IFN- γ^+ cells among each DC subset with or without PMA and ionomycin stimulation.

Supplementary Figure 8 Effect of candidate gene disruption on DC-mediated T cell differentiation. DCs from the indicated mutant and WT mice were co-cultured with MHC-matched, OVA-specific, naïve CD4⁺ T cells for 5 d. Following subsequent 24 h incubation of the T cells in anti-CD3 ϵ -, CD28-coated plates, the indicated cytokines in supernatants were measured by ELISA. Mice bearing mutations in the following genes were tested; (a) IFN- γ (b) IL-4 (c) IL-2 (d) IL-9 (e) IL-10 (f) c-Kit.

Supplementary Figure 9 Cell surface molecule display on lung DC subsets. Lung DCs of naive C57BL/6 mice were stained with antibodies against the indicated molecules (solid line) or with isotype control antibodies (dotted line) and analyzed by flow cytometry. Display levels of costimulatory molecules (**a**) and myeloid markers (**b**) are shown. Similar results were obtained in two independent experiments.

Supplementary Figure 10 Effect of TLR ligands on lung DCs. C57BL/6 mice received OVA alone, or OVA together with the indicated amounts of microbial products by intratracheal instillation. Shown are individual flow plots (**a**) and histograms of compiled data (**b**) for cell surface levels of CD86 and MHC class II I-A^b and for OVA uptake (solid lines). Shaded regions represent staining with isotype control antibodies. *p*-value by Student's *t*-test (n=2).

Supplementary Figure 11 Naïve CD4⁺ T cell preparation. Flow cytometric analyses of T cells before and after sorting of naïve CD4⁺ T cells. Non-naïve CD4⁺ T cells were depleted from cells of pooled LNs and spleen by magnetic activated sorter (MACS) and antibodies against CD8 α , CD8 β , CD11b, CD11c, CD16/32, CD19, CD25, CD44, B220, CD49b, I-A, Ly6C/G.





 $\boldsymbol{a}\,$ BALB/c lung DC, T cell reponses in primary culture

















