



Fig. S1. An ELISA procedure was used to assess the ability of fibronectin (Fn-left panel) and fibrinogen (Fg-right panel) to bind to various purified bacterial surface proteins. Increasing concentration of either Fn or Fg were incubated with target proteins that were fixed to micro-titer plates. Binding of Fn or Fg to the target proteins was detected using Fn or Fg specific antibodies. Values are the mean of three independent experiments ( $\pm$ SD). FnbB and ClfA are *S. aureus* fibronectin binding protein B and clumping factor A, respectively.

TABLE S1. Primers used in this study

Primer	Amplified region	Approx. Fragment Size (bp)	Sequence <sup>a</sup>
<b><u>Deletion mutants</u></b>			
chtAUP1 chtAUP2	Upstream of <i>chtA</i>	720	5'-GCGGGATCCTGCCACTGTTGTGGTGA-3' 5'-GCGCCC <del>GGG</del> ACACAAACCGTGAAAGG-3'
chtADN1 chtADN2	Downstream of <i>chtA</i>	2060	5'-GCGGGATCCGTCCATCACTTCTTTGG-3' 5'-GCGAAGCTTCCC <del>GGG</del> TTGCTCGCGCATGAGCC-3'
chtBUP1 chtBUP2	Upstream of <i>chtB</i>	1120	5'-GCGGGATCCGGAACGGTTACATCACATAACC-3' 5'-GCGCCC <del>GGG</del> TGTAGACGCCAACAAAGG-3'
chtABDN3 chtABDN4	Downstream of <i>chtB</i>	1600	5'-GCGGGATCCGCACCAAGCGCTAGAGC-3' 5'-GCGAAGCTTCCC <del>GGG</del> CTATATCGATGCTCAGG-3'
chtC-UP1 chtC-UP2	Upstream <i>chtC</i>	690	5'-GCGGGATCCCTTGCCTGAAGTGGCGA-3' 5-GCGAAGCTTAGG <del>CCT</del> CCTCCAAGTCTGAAGAAGC-3'
chtC-DN1 chtC-DN2	Downstream <i>chtC</i>	900	5'-GCGGGATCCGGCAAGCTGTCCACAGTG-3' 5'-GCGAGG <del>CCT</del> CACCAAGGACGACGCCG-3'
<b><u>Promoter fusions</u></b>			
chtBPo-SalI chtBPo-BamHI	<i>chtB</i> promoter	390	5'-GCGGTCGACGTGGAAGATTATTCTTGG-3' 5'-GCGGGATCCC <del>GGC</del> CAGCACCGACTACCG-3'
7.40B 7.40A	<i>chtA</i> promoter	211	5'-GCGGGATCCCAAAGACGTAAGTGTTC-3' 5'-GCGGTCGACGAGTTGTGGTCTAGGTGG-3'
PO1UP PO1DN PO2UP PO3DN	<i>cirA</i> promoter	269 166 114	5'-GCGGTCGACCATCAGAACTAACCACC-3' 5'-GCGGGATCCAGACTTGAGTGGTAG-3' 5'-GCGGTCGACGCACACAATAAAAGAAG-3' 5'-GCGGGATCCTTATTGTGTGCTTGTGGGG-3'
<b><u>Protein expression GST-fusions</u></b>			
FGex522RI RXho522-Tm	<i>chtC</i> for pGEX-6P-1 without transmembrane	1800	5'-GCGGAATTCGAGGACGTCGAAAAGCCT-3' 5'-GCGCTCGAGCTAAAGCCACCTGTGGATGAT-3'
FGex523RI RXho523-Tm	<i>cirA</i> for pGEX-6P-1 without transmembrane	840	5'-GCGGAATTCTCTGAGCCCCTGCCGAC-3' 5'-GCGCTCGAGCTACTTGAAAAAGTAACTAT-3'
FGexChtBRI RXhoChtB-Tm	<i>chtB</i> for pGEX-6P-1 without transmembrane	720	5'-GCGGAATTCGAAAACGTGGCTGCTACC-3' 5'-GCGCTCGAGCTATTTAGGGAGACCGCCAAA-3'
FGexChtARI RXhoChtA-Tm	<i>chtA</i> for pGEX-6P-1 without transmembrane	2100	5'-GCGGAATTCGAGCACGTAGTGGAGGAT-3' 5'-GCGCTCGAGCTACTTCCACGGCTTAGAAAG-3'
<b><u>Strep-tag fusions</u></b>			
FstrepChtA RstrepChtA	<i>chtA</i> strep tag	2050	5'-GCGCATATGGCAAGCTGGAGCCACCCGCAGTTC GAAAAGGGTGCAGAGCACGTAGTGGAGGAT-3' 5'-GCGGAATTCCTACTTCCACGGCTTAGAAAG-3'

RstrepChta-CR	<i>chtA</i> -CR	700	5'-GCG <u>GAAATTC</u> CTATGATTTGTCAAAACTAAA-3'
FstrepChtA-CT	<i>chtA</i> C-terminal	1320	5'-GCGCATATGGCAAGCT <b>TGGAGCCACCCGCAGTTC</b> GAAAAGGGTGCATCACAACCTGGCGATAAT-3'
FstrepChtB	<i>chtB</i> strep tag	800	5'-GCGCATATGGCAAGCT <b>TGGAGCCACCCGCAGTTC</b> GAAAAGGGTGCAGAAAACGTGGCTGCTACC-3'
RstrepChtB			5'-GCG <u>GAAATTC</u> CTAGATGCCCCAGATTTTAGG-3'
FstrepChtC	<i>chtC</i> strep tag	1850	5'-GCGCATATGGCAAGCT <b>TGGAGCCACCCGCAGTTC</b> GAAAAGGGTGCAGAGGACGTCGAAAAGCCT-3'
RstrepChtC			5'-GCG <u>GAAATTC</u> CTACTTGCCCCAAAACGCTCGA-3'

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<sup>a</sup>Restriction sites are underlined and bold indicates strep tag sequence.