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# **Supplemental Information**

# **Crystal Structures of Malonyl-Coenzyme A**

**Decarboxylase Provide Insights into Its Catalytic** 

# **Mechanism and Disease-Causing Mutations**

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**Inventory of Supplemental Information**

**Fig. S1, related to Fig. 2 Fig. S2, related to Fig. 3 Fig. S3, related to Fig. 5 Table S1, related to Fig. 6 Text References** 









## **Fig. S1, related to Fig. 2.**

- **A.** (*Top*) Plot of real-space correlation coefficient (CC) for the two chains of HsMCD structure. The majority of the residues has CC greater than 0.9. Regions of poorer fit correspond to disordered/missing surface loops or alternate conformations. (*Bottom*) Quality of the HsMCD electron density map.  $2F_0-F_c$  map contoured at 1.2  $\sigma$  is shown in blue mesh for four representative regions of the structure.
- **B.** Schematic drawings of the structures of the N-terminal helical domain of MCD. The structures are colored from blue at the N-terminus to red at the C-terminus.
- **C.** Overlay of MCD monomer structures. The overlay is based on the catalytic domain. One monomer is shown in color, and the other in gray. The catalytic domain of AvMCD is most similar to that of HsMCD.











## **Fig. S2, related to Fig. 3.**

- **A** (*Top left*) Gel filtration chromatography of HsMCD in the apo form (blue) and supplemented with malonyl-CoA (mCoA, red) reveals a tetramer. Arrows indicate retention volumes of molecular weight standards. (*Top right*) Analytical ultracentrifugation shows HsMCD exhibits a narrow sedimentation coefficient distribution estimated to be a tetramer of ~200 kD. (*Bottom*) Left Panel: Representative electron microscopy image of HsMCD. The scale bar shown is 100 nm. Right Panel: Representative examples of 2D classifications (C) and reprojections (R) from the electron micrograph. Box size is 410 Å.
- **B** (*Top*) Differences in the relative orientations of the dimers in the tetramers of HsMCD, RpMCD and AvMCD. The plane of the dimer in the back (cyan and magenta) is along the horizontal direction (indicated by the horizontal black line). The plane of the second dimer (green and orange, in the front) makes different angles in the three tetramers. (*Bottom*) Overlay of the structures of HsMCD (in color) and RpMCD (in gray) dimers. The overlay is based on the catalytic domain of monomer A.
- **C** Overlay of the structures of HsMCD monomer (*left*) and tetramer (*right*), reported here (in color) and by (Aparicio et al., 2013) (in gray).





## **Fig. S3, related to Fig. 5.**

- **A** (*Top*) Overlay of the active site region of HsMCD (in cyan) and CurA (gray). Residues in HsMCD are labeled. (*Bottom*) Overlay of the active site region of RpMCD (in cyan) and CurA (gray). Both panels are in stereo view.
- **B** Structural model of HsMCD docked *in silico* with malonyl-CoA (mCoA). (*Left*) Surface representation of the substrate channel showing in sticks the docked mCoA as well as acetyl CoA (AcCoA) from the CurA structure. Surface is coloured on the basis of sequence conservation among MCD homologues (magenta, most conserved; cyan, least conserved). (*Right*) Active site showing residues (in sticks) that can interact with the docked mCoA (hydrogen bonds, red dashed lines; distance in angstrom, black line).

No.	<b>Mutation</b>	<b>Conservation</b>	<b>Location/proposed consequence</b>
1	G3D	variable	Affect protein targeting?
			N-terminal methionine for the
$\overline{2}$	<b>M40T</b>	conserved	mitochondrial isoform; Affect protein
			targeting?
3	A69V	variable	N-domain helix $\alpha$ B
$\overline{4}$	L161P	conserved	N-domain helix $\alpha G$
5	<b>S290F</b>	conserved	GNAT core, strand $\beta$ 2; in active site
			GNAT core, linker between $\beta$ 4 and
6	G300V	conserved	following helix; close to entrance of
			substrate channel
7	L307R	conserved	GNAT core, helix following $\beta$ 4; close to
			active site
8	W384C	variable	GNAT core, in a $3_{10}$ helix
9	S440I	conserved	<b>GNAT</b> core
10	Y456S	invariant	GNAT core, strand $\beta$ 7; hydrogen bond to
			catalytic residue His423
11	<b>S477F</b>	conserved	<b>GNAT</b> core

**Table S1, related to Fig. 6. List of missense pathogenic mutations of HsMCD** 

### **Supplementary text**

## **An N-terminal helix in the central cavity of RpMCD tetramer**

The expression construct for RpMCD covered residues 8-451 of this enzyme. Residues 10-29 form a helix  $(\alpha A^{\prime}, Fig. 2B)$  and are located in the center of the tetramer (Fig. 3C). However, only two of the four helices of the tetramer are observed, as there is not sufficient space in the center of the tetramer to accommodate all four helices. In fact, the helix is situated on a two-fold axis of the tetramer (Fig. 3C). In an attempt to assess the importance of these residues for the tetramerization of RpMCD, we deleted the first 39 residues, but the resulting protein was not expressed in *E. coli*.

At the C-terminus, the expression construct of RpMCD lacked the last 25 residues of the protein. They are hydrophilic and poorly conserved among the MCDs (Fig. 1), suggesting that they are likely to be flexible. This C-terminal segment for HsMCD is much shorter, and it also contains the peroxisome targeting signal (SKL).

#### *In silico* **substrate docking of HsMCD**

The refined HsMCD structure was used as receptor for *in silico* docking with the substrate malonyl-CoA. Docking was performed with fully flexible ligands and potential grid maps representing five different features of the receptor, as implemented in the ICM software (Totrov et al., 1997). No restraints were introduced to receptor or ligand to allow full exploration of the active site, with the exception of thioester carbonyl oxygen atom. This atom was tethered to the oxygen atom of the β-alanine carbonyl of a bound acetyl-CoA (product of malonyl-CoA decarboxylation) in the CurA co-crystal structure (Gu et al., 2007), which has been superimposed to the crystal structure of HsMCD. The best poses were visually screened and selected if distance requirements for catalysis were found to be acceptable for atoms involved in the proposed catalytic mechanism. The docking result is shown in Fig. S6. The docked malonyl-CoA binds in a similar manner to that of acetyl-CoA in the CurA structure.

### **References**

Aparicio, D., Perez-Luque, R., Carpena, X., Diaz, M., Ferrer, J.C., Loewen, P.C., and Fita, I. (2013). Structural asymmetry and disulfide bridges among subunits modulate the activity of human malonyl-CoA decarboxylase. J Biol Chem *288*, 11907-11919.

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Totrov, M. & Abagyan, R. Flexible protein-ligand docking by global energy optimization in internal coordinates. *Proteins* Suppl 1, 215-20 (1997).