

Supporting Information

Chlorizidine, A Cytotoxic 5*H*-Pyrrolo[2,1-*a*]isoindol-5-one-Containing Alkaloid from a Marine *Streptomyces* sp.

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General. Pyridine was distilled from calcium hydride prior to use, and dimethylformamide (DMF) was dried prior to use by eluting through a column of activated alumina. All other reagents and solvents were purchased commercially and were used without further purification. Organic extracts were dried with sodium sulfate. Reactions were analyzed by TLC (Merck® silica gel 60 F₂₅₄) or, when appropriate, an analytical 1090 Series HP system (0.7 mL min⁻¹) with UV detection (254 nm) using a C18(2) Phenomenex® Luna column (5 µm, 100 x 4.6 mm). Semi-preparative RP-HPLC purification was performed on a C8(2) Phenomenex® Luna (5 µm, 250 x 10 mm), C18(2) Phenomenex® Luna (5 µm, 250 x 10 mm) or Kromasil® Diol (5 µm, 250 x 10 mm) column with UV detection (254 nm). ¹H NMR spectra were recorded at 500 MHz or 600 MHz in DMSO-*d*₆ (residual solvent referenced to 2.50 ppm), CD₃CN (residual solvent referenced to 1.94 ppm), or benzene-*d*₆ (residual solvent referenced to 7.16 ppm) on a Varian Inova 500 MHz or Bruker 600 MHz NMR spectrometer. ¹³C NMR spectra were recorded at 125 MHz in DMSO-*d*₆ (referenced to 39.5 ppm), CD₃CN (referenced to 1.2 ppm) or benzene-*d*₆ (referenced to 128.0 ppm) on a Varian Inova 500 MHz NMR spectrometer.

Phylogeny of strain CNH-287. The identity of strain CNH-287 was determined by phylogenetic analysis of the 16S rRNA gene sequence. The sequence from this strain was used as a query in a BLASTn search at the NCBI database. The top hits were all members of the *Streptomyces* genus. Twenty-two of the top BLAST hits were from strains that shared 99% or greater identity with the CNH-287 sequence. Of these twenty-two strains, all that had a specified location were cultured from the marine environment, suggesting that strain CNH-287 may be part of a previously undescribed marine *Streptomyces* lineage.

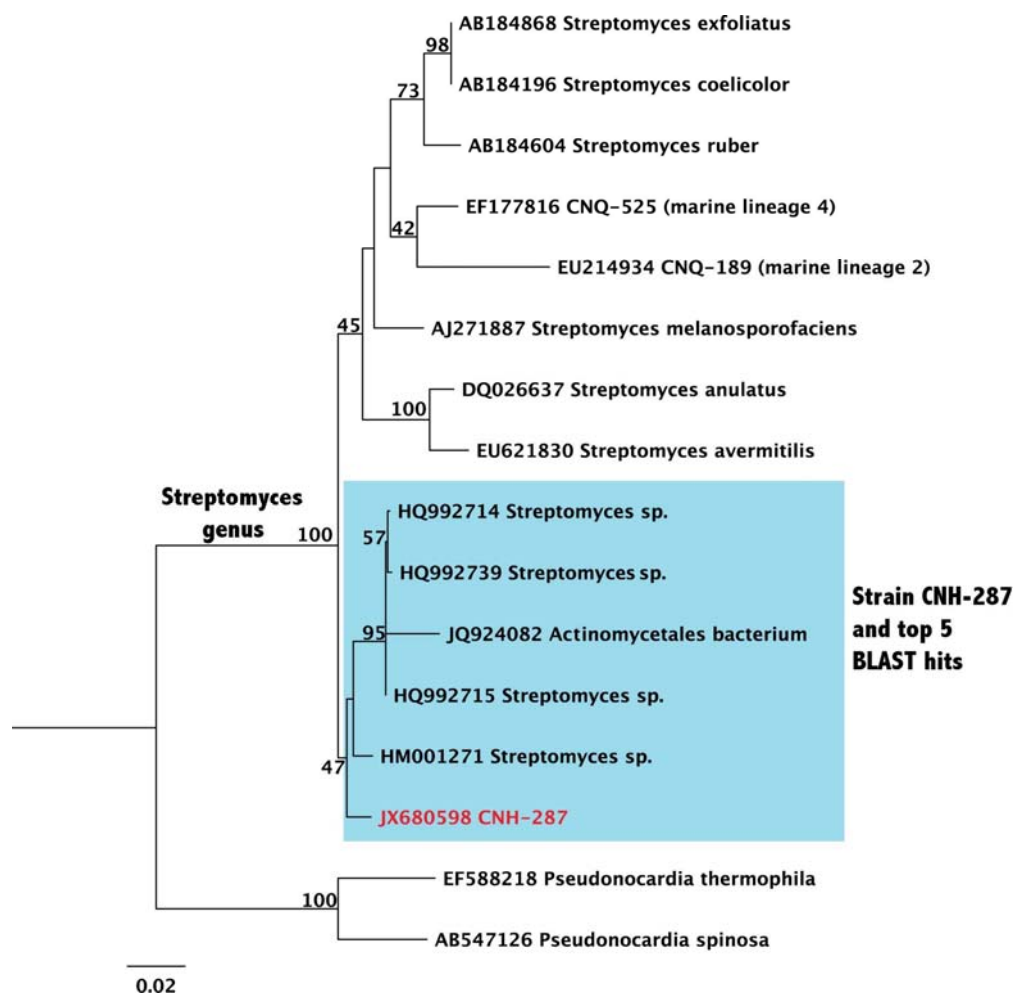
A phylogeny of strain CNH-287 was built using the top five BLAST hits for this strain and a number of diverse *Streptomyces* sequences, including two from described marine *Streptomyces* lineages. Two outgroups were used that fell into the genus *Pseudonocardia*, members of the Pseudonocardiaceae which clade closely with streptomycetes in overall actinobacterial phylogenies.¹ Sequences were aligned using MUSCLE,² and a maximum likelihood phylogeny built using raxmlGUI.³ GTR+G was used as a substitution model with 100 thorough bootstraps.

(1) Stackebrandt, E.; Rainey, F. A.; Ward-Rainey, N. L. *Int. J. Syst. Bacteriol.* **1997**, *47*, 479-491.

(2) Edgar, R. C. *Nucleic Acids Res.* **2004**, *32*, 1792-1797.

(3) Silvestro, D.; Michalak, I. *Org. Divers. Evol.* **2011**.

Figure S1. Phylogenetic tree for CNH-287.



Cultivation of strain CNH-287. The strain was cultured in 2.8 L Fernbach flasks (20 x 1 L) in a seawater-based medium (per 1 L seawater: 10 g of starch, 4 g of yeast extract, 2 g of peptone, 1 g of CaCO₃, 40 mg of Fe₂(SO₄)₃•4H₂O, 100 of mg KBr) and shaken at 230 rpm at 27 °C.

Isolation and purification of chlorizidines A (1) and B (7). After one day of a 20 L cultivation, sterilized Amberlite XAD-18 resin (20 g L⁻¹) was added. The culture and resin were shaken for six days. The resin was filtered through cheesecloth, washed with deionized water, and eluted with acetone. The acetone was removed under reduced pressure, and the resulting aqueous layer was extracted with EtOAc (3 x 400 mL). The combined extracts were concentrated to yield 1.7 g of crude extract. The extract was fractionated by column chromatography on silica gel (20 g), eluting with a step gradient of isooctane, EtOAc, and CH₃OH. The 1:1 isooctane/EtOAc fraction contained chlorizidine A (1). This sample was

further fractionated on C18 column (3 g) under vacuum, eluting with a step gradient of CH₃CN and water. Fractions 4:1 and 9:1 CH₃CN/water were purified by reversed-phase HPLC (80% CH₃CN in water, t_R = 16 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to afford chlorizidine A (**1**) (50.0 mg) as a yellow film. In a separate 20 L culture, resin was instead added at the end of the cultivation (day seven). The crude extract (2.0 g) was fractionated by column chromatography on silica gel (20 g), again eluting with a step gradient of isooctane, EtOAc, and CH₃OH. The 3:2 isooctane/EtOAc fraction contained chlorizidine A (**1**) and chlorizidine B (**7**). Chlorizidine A (**1**) was purified as before to yield 10.0 mg. Chlorizidine B (**7**) was purified by reversed-phase HPLC (70% CH₃CN in water, t_R = 16.5 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to give 16.0 mg as a rapidly decomposing white solid.

Chlorizidine A (1): UV/vis (CH₃CN) λ_{\max} = 212 (31800), 261 (47400), 340 (7300), 356 (8500), 409 (2300); $[\alpha]_D$ -35 (*c* 0.50, CH₃CN); IR (film) $\tilde{\nu}$ = 3119, 1721, 1626 cm⁻¹; ¹H NMR see Table 1; ¹³C NMR (CD₃CN) δ 163.9, 163.0, 157.5, 136.8, 135.5, 132.7, 116.9, 113.3, 113.0, 109.7, 108.2, 106.0, 105.9, 101.9, 99.3, 53.0, 32.5, 25.4; HRESI-FT-MS (Orbit-Trap-MS): m/z (M+H)⁺ calcd for C₁₈H₁₁³⁵Cl₄N₂O₃ 442.9524, found 442.9511.

Chlorizidine B (7): UV/vis (CH₃CN) λ_{\max} = 299; ¹H NMR see Table S1; HRESI-FT-MS (Orbit-Trap-MS): m/z (M+H)⁺ calcd for C₁₇H₁₃³⁵Cl₄N₂O₂ 416.9731, found 416.9724.

Diacetyl chlorizidine (2): To a solution of chlorizidine A (**1**) (9.0 mg, 0.020 mmol) in dry CH₃CN (2.0 mL) at room temperature was added dry Et₃N (85 μ L, 0.61 mmol) followed by acetic anhydride (30 μ L, 0.30 mmol). The mixture was stirred at rt for 3 h, then diluted with water (10 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were dried, filtered, and concentrated. The crude material was filtered on a short plug of silica gel with EtOAc. The organic layer was concentrated, and the product was purified by reversed-phase HPLC (85% CH₃CN in water, t_R = 11 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to yield 5.7 mg (54%) of **2** as a yellow film: UV/vis (CH₃CN) λ_{\max} = 205 (26000), 267 (38400), 303 (2200), 316 (2200), 330 (1400), 414 (1600); $[\alpha]_D$ -140 (*c* 0.10, CH₃CN); IR (film) $\tilde{\nu}$ = 3449, 1767, 1628 cm⁻¹; ¹H NMR see Table S2; ¹³C NMR (CD₃CN) δ 168.7, 168.0, 167.2, 165.7, 157.9, 155.3, 154.9, 148.1, 147.5, 135.5, 134.9, 131.4, 125.8, 125.2, 117.7, 117.3, 114.6, 113.5, 113.0, 109.3, 109.2, 106.2, 100.1, 100.0, 53.0, 33.8, 33.6, 24.9, 24.8, 20.7, 20.3, 20.0, 19.6; HRESI-FT-MS (Orbit-Trap-MS): m/z (M+Na)⁺ calcd for C₂₂H₁₄³⁵Cl₄N₂O₅Na 548.9555, found 548.9556.

Dimethyl chlorizidine (3): To a solution of chlorizidine A (**1**) (8.0 mg, 0.018 mmol) in dry acetone (3.0 mL) at room temperature was added potassium carbonate (50 mg, 0.36 mmol) followed by dimethyl sulfate (21 μ L, 0.18 mmol). The mixture was stirred at rt overnight, then diluted with water (10 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were dried, filtered, and concentrated. The

crude material was filtered through a short plug of silica gel with EtOAc. The organic layer was concentrated, and the product was purified by reversed-phase HPLC (95% CH₃CN in water, t_R = 14 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to give 7.5 mg (87%) of **3** as a yellow film: UV/vis (CH₃CN) λ_{max} = 214 (32700), 267 (56900), 330 (4900), 343 (5200), 408 (2100); [α]_D -77 (*c* 0.10, CH₃CN); IR (film) $\tilde{\nu}$ = 3421, 2945, 2854, 1750, 1615, 1581 cm⁻¹; ¹H NMR see Table S2; ¹³C NMR (DMSO-*d*₆) δ 165.2, 164.1, 159.8, 158.7, 158.5, 137.5, 136.1, 132.1, 120.0, 116.2, 112.6, 107.4, 105.0, 100.9, 100.1, 99.2, 62.8, 61.8, 57.1, 52.7, 33.0, 25.2; HRESI-FT-MS (Orbit-Trap-MS) m/z (M+H)⁺ calcd for C₂₀H₁₅³⁵Cl₄N₂O₃ 470.9837, found 470.9833.

Diisobutryl chlorizidine (4): To a solution of chlorizidine A (**1**) (4.5 mg, 0.010 mmol) in dry CH₃CN (1.5 mL) at room temperature was added dry Et₃N (27 μ L, 0.20 mmol) followed by isobutryl chloride (10 μ L, 0.10 mmol). The mixture was stirred at rt for 3 h, then diluted with water (10 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were dried, filtered, and concentrated. The crude material was filtered on a short plug of silica gel with EtOAc. The organic layer was concentrated, and the product was purified by reversed-phase HPLC (95% CH₃CN in water, t_R = 12 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to give 4.2 mg (72%) of **4** as a yellow film: UV/vis (CH₃CN) λ_{max} = 205 (34000), 267 (53500), 303 (3700), 316 (3700), 330 (3300), 414 (2300); [α]_D -100 (*c* 0.10, CH₃CN); IR (film) $\tilde{\nu}$ = 2979, 2938, 1627 cm⁻¹; ¹H NMR Table S2; ¹³C NMR (DMSO-*d*₆) δ 174.3, 174.2, 173.7, 173.2, 171.5, 157.9, 157.8, 155.7, 155.1, 155.0, 148.6, 148.3, 147.6, 135.4, 135.1, 135.0, 134.9, 131.5, 131.4, 131.3, 125.4, 125.2, 125.0, 119.0, 118.4, 117.7, 117.2, 114.4, 113.6, 113.5, 113.4, 113.2, 112.9, 109.5, 109.3, 109.2, 109.0, 106.5, 106.4, 100.3, 100.2, 100.1, 54.4, 53.5, 53.3, 33.8, 33.7, 33.4, 33.3, 33.2, 32.9, 32.7, 24.9, 24.8, 24.4, 19.0, 18.6, 18.5, 18.3, 17.8, 17.7, 17.5; HRESI-FT-MS (Orbit-Trap-MS) m/z (M+Na)⁺ calcd for C₂₆H₂₂³⁵Cl₄N₂O₅Na 605.0181, found 605.0174.

Dipivaloyl chlorizidine (5): To a solution of chlorizidine A (**1**) (4.5 mg, 0.010 mmol) in dry CH₃CN (1.5 mL) at room temperature was added dry Et₃N (27 μ L, 0.20 mmol) followed by pivaloyl chloride (12 μ L, 0.10 mmol). The mixture was stirred at rt for 3 h, then diluted with water (10 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were dried, filtered, and concentrated. The crude material was filtered on a short plug of silica gel with EtOAc. The organic layer was concentrated, and the product was purified by reversed-phase HPLC (97% CH₃CN in water, t_R = 13 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to give 4.7 mg (77%) of **5** as a yellow film: UV/vis (CH₃CN) λ_{max} = 205 (36000), 267 (56700), 303 (4300), 316 (4300), 330 (2400), 414 (2400); [α]_D -57 (*c* 0.10, CH₃CN); IR (film) $\tilde{\nu}$ = 3401, 2976, 1761, 1627 cm⁻¹; ¹H NMR Table S2; ¹³C NMR (DMSO-*d*₆) δ 175.7, 175.4, 173.6, 158.0, 157.9, 156.4, 155.7, 155.4, 149.3, 149.0, 147.9, 135.4, 135.3, 135.2, 134.8, 131.4, 131.3, 124.2, 124.1, 118.7,

118.1, 117.8, 117.3, 117.2, 114.1 113.7, 113.6, 113.5, 113.2, 113.0, 109.7, 109.6, 109.3, 109.2, 109.1, 107.1, 106.7, 106.6, 100.6, 100.5, 55.0, 54.3, 53.8, 26.8, 26.7, 26.5, 26.4, 26.3, 24.6; HRESI-FT-MS (Orbit-Trap-MS) m/z (M+Na)⁺ calcd for C₂₈H₂₆³⁵Cl₄N₂O₅Na 633.0494, found 633.0487.

Dibenzoyl chlorizidine (6): To a solution of chlorizidine A (**1**) (8.0 mg, 0.018 mmol) in dry CH₃CN (3 mL) at room temperature was added dry Et₃N (50 μ L, 0.36 mmol) followed by benzoyl chloride (21 μ L, 0.18 mmol). The mixture was stirred at rt for 3 h, then diluted with water (10 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were dried, filtered, and concentrated. The crude material was filtered on a short plug of silica gel with EtOAc. The organic layer was concentrated, and the product was purified by reversed-phase HPLC (90% CH₃CN in water, t_R = 14 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) to give 5.0 mg (42%) of **6** as a yellow film: UV/vis (CH₃CN) λ_{max} = 233 (50000), 267 (63700), 303 (4200), 316 (3900), 330 (2300), 414 (2300); [α]_D -74 (*c* 0.050, CH₃CN); IR (film) $\tilde{\nu}$ = 3485, 3058, 2963, 1754, 1629 cm⁻¹; ¹H NMR Table S2; ¹³C NMR (benzene-*d*₆) δ 164.4, 164.1, 163.9, 163.4, 162.1, 157.9, 157.7, 157.5, 155.9, 155.1, 155.0, 149.9, 149.5, 148.8, 136.0, 135.9, 135.0, 134.7, 134.6, 134.4, 134.3, 134.2, 133.5, 131.3, 131.2, 131.1, 131.0, 130.9, 130.7, 130.6, 130.5, 130.4, 129.3, 129.2, 129.0, 128.8, 128.7, 128.6, 128.5, 128.4, 128.2, 128.1, 128.0, 127.1, 126.6, 125.9, 120.6, 119.9, 118.9, 118.2, 118.1, 114.8, 114.1, 112.1, 112.0, 111.8, 111.7, 108.3, 108.2, 108.1, 107.7, 107.5, 101.2, 54.6, 54.1, 53.8, 34.4, 34.0, 33.8, 25.1, 24.7; HRESI-FT-MS (Orbit-Trap-MS) m/z (M+H)⁺ calcd for C₃₂H₁₉³⁵Cl₄N₂O₅ 651.0048, found 651.0033.

Diacetyl chlorizidine B (8): To a solution of chlorizidine B (**7**) (16 mg, 0.039 mmol) in dry CH₃CN (5.0 mL) at room temperature was added dry Et₃N (85 μ L, 0.78 mmol) followed by acetic anhydride (36 μ L, 0.39 mmol). The mixture was stirred at rt for 3 h, then diluted with water (10 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were dried, filtered, and concentrated. The crude material was filtered on a short plug of silica gel with EtOAc. The organic layer was concentrated, and the product was first purified by reversed-phase HPLC (80% CH₃CN in water, t_R = 12 min, C18(2) Phenomenex® Luna, 2.5 mL min⁻¹) and then purified by normal phase HPLC (85% isooctane in isopropanol, t_R = 15 min, Kromasil® 60-5-Diol, 2.5 mL min⁻¹) to furnish 6.0 mg (31%) of **8**: UV/vis (CH₃CN) λ_{max} = 300 (23500); [α]_D -75 (*c* 0.10, CH₃CN); IR (film) $\tilde{\nu}$ 3131, 2989, 1767, 1625 cm⁻¹; ¹H NMR Table S1; ¹³C NMR (DMSO-*d*₆) δ 169.3, 167.7, 149.5, 149.4, 135.3, 131.8, 128.2, 122.7, 116.9, 114.9, 113.0, 109.1, 109.0, 107.3, 106.4, 99.9, 53.9, 33.8, 24.7, 20.6, 20.0; HRESI-FT-MS (Orbit-Trap-MS): m/z (M+H)⁺ calcd for C₂₁H₁₇³⁵Cl₄N₂O₄ 500.9942, found 500.9935.

Thioester 9: To a solution of chlorizidine A (**1**) (15.4 mg, 0.0347 mmol) and potassium carbonate (40 mg, 0.29 mmol) in dry DMF (2.0 mL) at room temperature was added *N*-acetylcysteamine (20 μ L, 0.19 mmol) via syringe. The mixture was stirred at 60 °C for 1 h, then diluted with water (5 mL) and a saturated aq. NH₄Cl solution (5 mL) and extracted with EtOAc (3 x 5 mL). The combined extracts were washed with brine (4 mL), dried, filtered, and concentrated. The product was purified by reversed-phase HPLC (55% CH₃CN in water, 0.2% TFA, t_R = 27 min, C8(2) Phenomenex® Luna, 3 mL min⁻¹) to give 10.2 mg (52%) of **9**: UV/vis (CH₃CN/water) λ_{max} = 223, 266, 360, 417; $[\alpha]_D$ -19 (*c* 0.38, CH₃CN); IR (film) $\tilde{\nu}$ = 3352, 1728, 1626 cm⁻¹; ¹H and 2D NMR Table S3; HRMS (ESI-TOF) *m/z* (M+H)⁺ calcd for C₂₂H₂₀³⁵Cl₄N₃O₄S 561.9929, found 561.9925.

Amide 10: To a solution of chlorizidine A (**1**) (5.3 mg, 0.012 mmol) in dry CH₃CN (2.0 mL) at room temperature was added benzylamine (30 μ L, 0.27 mmol) via syringe. The mixture was stirred at rt for 3 h and then concentrated. The product was purified by reversed-phase HPLC (70% CH₃CN in water, 0.2% TFA, t_R = 20 min, C8(2) Phenomenex® Luna, 3 mL min⁻¹), collecting the eluent in a vial containing an aq. 0.2 M NaHCO₃ solution. After concentration, the white solid was triturated with CH₃CN. The solution concentrated onto C18 silica (100 mg) and fractionated on a C18 SPE cartridge (100 mg), eluting with water and then with CH₃CN. The organic fraction was concentrated to give 2.2 mg (33%) of **10**: UV/vis (CH₃CN/water) λ_{max} = 239, 307; $[\alpha]_D$ +2 (*c* 0.34, CH₃CN); IR (film) $\tilde{\nu}$ = 3237, 1613 cm⁻¹; ¹H and 2D NMR Table S4; HRMS (ESI-TOF) *m/z* (M+H)⁺ calcd for C₂₅H₂₀³⁵Cl₄N₃O₃ 550.0259, found 550.0264.

Ester 11: To chlorizidine A (**1**) (1.3 mg, 0.0029 mmol) and K₂CO₃ (10 mg) was added dry MeOH (1.0 mL) at room temperature. The solution was stirred at rt overnight, then diluted with a saturated aq. NH₄Cl solution (4 mL) and extracted with EtOAc (2 x 2 mL). The combined extracts were washed with brine (1 mL), dried, filtered, and concentrated. The product was purified by reversed-phase HPLC (65% CH₃CN, 0.2% TFA, t_R = 30 min, C8(2) Phenomenex® Luna, 3 mL min⁻¹) to give 0.4 mg (29%) of **11**: HRMS (ESI-TOF) *m/z* (M-H)⁻ calcd for C₁₉H₁₃³⁵Cl₄N₂O₄ 472.9629, found 472.9646. The complete characterization of **11** was not possible due to its instability.

Diacetylated thioester 12: To a solution of thioester **9** (3.0 mg, 0.0053 mmol) in dry pyridine (1.0 mL) at room temperature was added acetic anhydride (0.10 mL). The mixture was stirred at rt for 4 h, CH₃OH (1 mL) was added, and the solution was concentrated. The product was purified by reversed-phase HPLC (70% CH₃CN, 0.2% TFA, t_R = 16 min, C8(2) Phenomenex® Luna, 3 mL min⁻¹) to give 1.8 mg (52%) of **12**: UV/vis (CH₃CN/water) λ_{max} = 219, 267, 321, 417; $[\alpha]_D$ -4 (*c* 0.17, CH₃CN); IR (film) $\tilde{\nu}$ = 1761, 1684, 1626 cm⁻¹; ¹H NMR (600 MHz, CD₃CN) δ 10.02 (br s), 9.95 (br s), 7.12 (s), 7.09 (s), 6.46 (s), 5.86 (s),

5.83 (s), 4.04 (m), 3.35 (m), 3.08 (m), 2.58 (m), 2.42 (m), 2.36 (s), 2.32 (s), 2.29 (s), 2.22 (s), 1.87 (s), 1.85 (s); HRMS (ESI-TOF) m/z (M+H)⁺ calcd for C₂₆H₂₄³⁵Cl₄N₃O₆S 646.0140, found 646.0130.

Triacetylated amide 13: To a solution of chlorizidine A (**1**) (5.0 mg, 0.011 mmol) in dry CH₃CN (2.0 mL) at room temperature was added benzylamine (30 μL, 0.27 mmol). The mixture was stirred at rt for 5 h and then concentrated. To a solution of the crude product in dry pyridine (1 mL) was added acetic anhydride (0.20 mL). The mixture was stirred at rt overnight, CH₃OH (1 mL) was added, and the solution was concentrated. The product was purified by reversed-phase HPLC (70% CH₃CN, 0.2% TFA, t_R = 24 min, C8(2) Phenomenex® Luna, 3 mL min⁻¹) to give 3.4 mg (45%) of **13**: UV/vis (CH₃CN/water) λ_{max} = 235, 282; [α]_D -17 (c 0.29, CH₃CN); IR (film) $\tilde{\nu}$ = 1773, 1684 cm⁻¹; ¹H NMR (600 MHz, CD₃CN) δ 7.34-7.22 (m), 7.14-7.03 (m), 7.08 (s), 6.32 (s), 6.30 (s), 5.88 (s), 5.85 (s), 5.64 (dd, J = 8.8, 5.5 Hz), 5.49 (dd, J = 8.8, 6.4 Hz), 4.39 (d, J = 5.8 Hz), 4.36 (d, J = 6.4 Hz), 4.33 (d, J = 5.8 Hz), 4.30 (d, J = 5.8 Hz), 4.26 (d, J = 6.2 Hz), 3.03 (m), 2.92 (m), 2.44 (m), 2.33 (s), 2.27 (s), 2.15 (s), 1.71 (s); HRMS (ESI-TOF) m/z (M+H)⁺ calcd for C₃₁H₂₆³⁵Cl₄N₃O₆ 676.0576, found 676.0595.

Triacetylated ester 14: To a solution of diacetyl chlorizidine A (**2**) (7.9 mg, 0.015 mmol) in dry CH₃OH (2.0 mL) at room temperature was added an aqueous solution of NaOH (0.10 mL, 0.05 M). The mixture was stirred at rt overnight, then diluted with a saturated aq. NH₄Cl solution (4 mL) and extracted with EtOAc (3 x 2 mL). The combined extracts were washed with brine (2 x 1 mL), dried, filtered, and concentrated. To a solution of the crude product in dry pyridine (1 mL) was added acetic anhydride (0.20 mL). The mixture was stirred at rt overnight, CH₃OH (1 mL) was added, and the solution was concentrated. The product was purified by reversed-phase HPLC (70% CH₃CN, 0.2% TFA, t_R = 25 min, C8(2) Phenomenex® Luna, 3 mL min⁻¹) to give 3.4 mg (38%) of **14**: UV/vis (CH₃CN/water) λ_{max} = 238, 287; [α]_D -21 (c 0.28, CH₃CN); IR (film) $\tilde{\nu}$ = 1773, 1684 cm⁻¹; ¹H NMR (600 MHz, CD₃CN) δ 7.18 (s), 7.09 (s), 6.29 (s), 5.89 (s), 5.68 (dd, J = 9.4, 5.5 Hz), 5.57 (dd, J = 8.8, 5.8 Hz), 3.71 (s), 3.59 (s), 3.06 (m), 2.94 (m), 2.46 (m), 2.34 (s), 2.32 (s), 2.31 (s), 2.29 (s); HRMS (ESI-TOF) m/z (M+H)⁺ calcd for C₂₅H₂₁³⁵Cl₄N₂O₇ 601.0103, found 601.0091.

Figure S2. HMBC correlations for chlorizidine A (**1**)

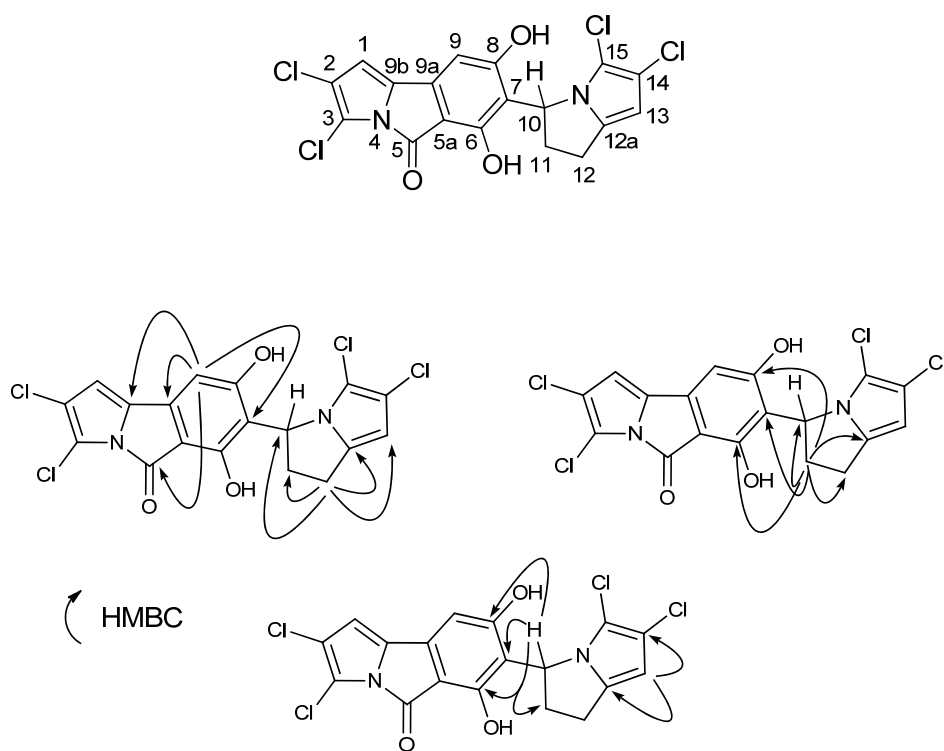
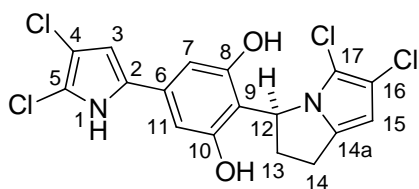
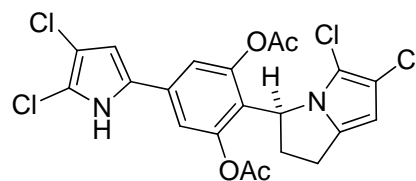


Table S1. Selected NMR spectral data for **7** and **8**



7

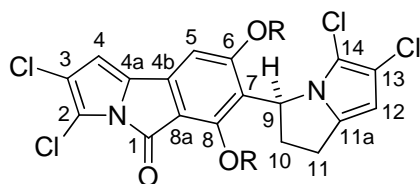


8

position	δ_{H} (7), mult. (<i>J</i> , Hz) ^[a] CD ₃ CN	δ_{H} , (8), mult. (<i>J</i> , Hz) ^[a] DMSO- <i>d</i> ₆
1	10.06, br s	12.42, br s
3	6.45, d (3.0)	6.79, d (3.0)
7	6.51, br s ^[b]	7.35, s ^[b]
11	6.51, br s ^[b]	7.40, s ^[b]
12	5.84, dd (9.5, 6.5)	5.59, dd (6.5, 5.5)
13	2.80, ddd (17.5, 14.0, 5.0)	2.84, ddd (18.5, 10.0, 4.5)
	2.55, ddd (17.5, 10.0, 6.0)	2.25, dd (12.0, 6.0)
14	3.06, ddd (14.5, 10.0, 5.0)	2.99, ddd (19.0, 14.5, 6.0)
	2.88, ddd (14.5, 10.0, 6.0)	2.88, ddd (17.5, 9.5, 5.5)
15	5.79, s	5.95, s
Ac		2.39, s
		1.94, s

^[a] 500 MHz. ^[b] H-7 and H-11 are not distinguishable due to the pseudo-symmetry of **7** and **8**.

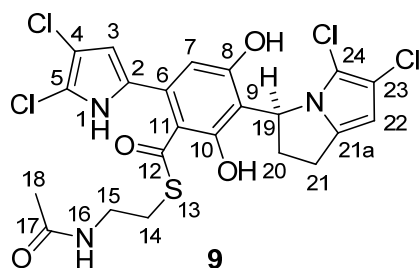
Table S2. Selected NMR spectral data for **2-6**



2, R = Ac
3, R = Me
4, R = CO*i*-Pr
5, R = Piv
6, R = Bz

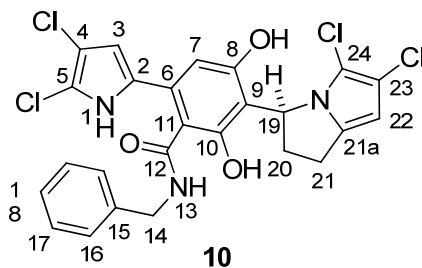
position	δ_{H} (2) mult. (<i>J</i> , Hz) ^[a] DMSO- <i>d</i> ₆	δ_{H} (3) mult. (<i>J</i> , Hz) ^[a] DMSO- <i>d</i> ₆	δ_{H} (4) mult. (<i>J</i> , Hz) ^[a] DMSO- <i>d</i> ₆	δ_{H} (5) mult. (<i>J</i> , Hz) ^[a] DMSO- <i>d</i> ₆	δ_{H} (6) mult. (<i>J</i> , Hz) ^[a] benzene- <i>d</i> ₆
1	6.80, s 6.79, s	7.17, s 7.10, s	6.80, s 6.78, s	6.80, s 6.79, s	5.48, s 5.43, s 5.40, s
9	7.42, s 7.36, s	6.67, s	7.45, s 7.42, s 7.33, s	7.44, s 7.43, s 7.30, s	6.43, s 6.38, s 6.35, s
10	5.70, br s 5.75, dd (6.0, 5.0)	5.82, s	5.63, t (8.5) 5.61, t (8.5) 5.58, t (6.0)	5.48, br s 5.46, t (8.5) 5.40, t (8.5)	5.17, dd (9.0, 5.0) 5.10, dd (9.5, 4.5)
11	2.88, m 2.36, br s 2.25, ddd (18.0, 14.0, 5.5)	2.86, m 2.36, m	2.81, m 2.40, m 2.29, ddd (17.5, 14.0, 5.5) 2.03, m	2.75-2.57, m 2.30, br s 2.01, ddd (19.0, 10.0, 10.0)	2.36, m 2.30, m 2.11, m
12	3.10, t (10.0) 3.01, dd (15.0, 12.0) 2.91, m	3.00, m 2.88, m	2.97-2.76, m	2.94-2.77, m	2.55, ddd (15.0, 5.0, 5.0) 2.27, m 2.11, m 2.02, dd (20.0, 10.0)
13	5.96, s 5.94, s	5.90, s 5.86, s	5.98, s 5.95, s 5.92, s	6.00, s 5.96, s 5.95, s	5.58, s 5.40, s 5.26, s
R	2.43, s 2.41, s 2.00, s 1.95, s	4.04, s 4.00, s 3.68, s 3.48, s	3.03, 2.53, 2.44 2.40, sep (9.0) 1.33, 1.32, 1.29, 1.28, 1.24, 1.13, 1.12, 1.05, 1.05, 0.98, 0.95, d (9.0)	1.39, s 1.36, s 1.35, s 1.11, s 1.10, s 1.08, s	8.34, br s 8.25, 8.20, 8.08, 8.06, 7.91, d (7.5) 7.21-7.04, m

^[a] 500 MHz.

Table S3. Selected NMR spectral data for **9** (CD₃CN)

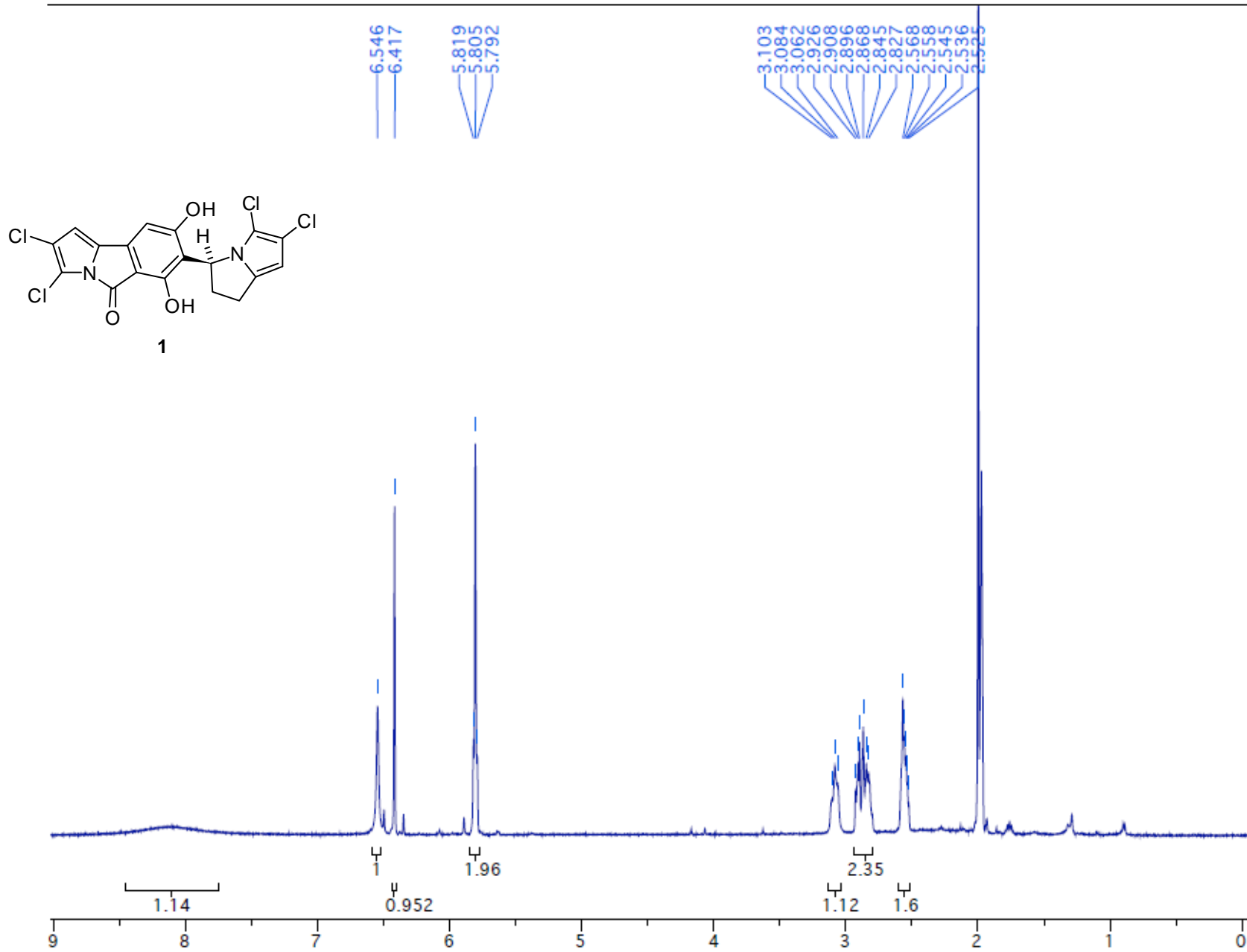
position	δ_C ^[a]	δ_H , mult. (J, Hz) ^[b]	HMBC
1		9.66, br s	3
3	107.7	6.24, s	
7	102.7	6.46, s	8,9,11,19
8	164.4		
9	114.9		
10	158.1		
11	106.0		
12	--		
14	31.5	2.55, m	15
15	39.9	3.32, m	14,17
16		6.84, br s	
17	171.8		
18	22.6	1.87, s	17
19	38.6	4.43, m	8,10,20,21
20	33.6	2.28, m	9,19,21,21a
		2.17, m	
21	26.5	2.51, m	19,20,21a,22
21a	131.6		
22	106.4	5.81, d (3.0)	21,21a

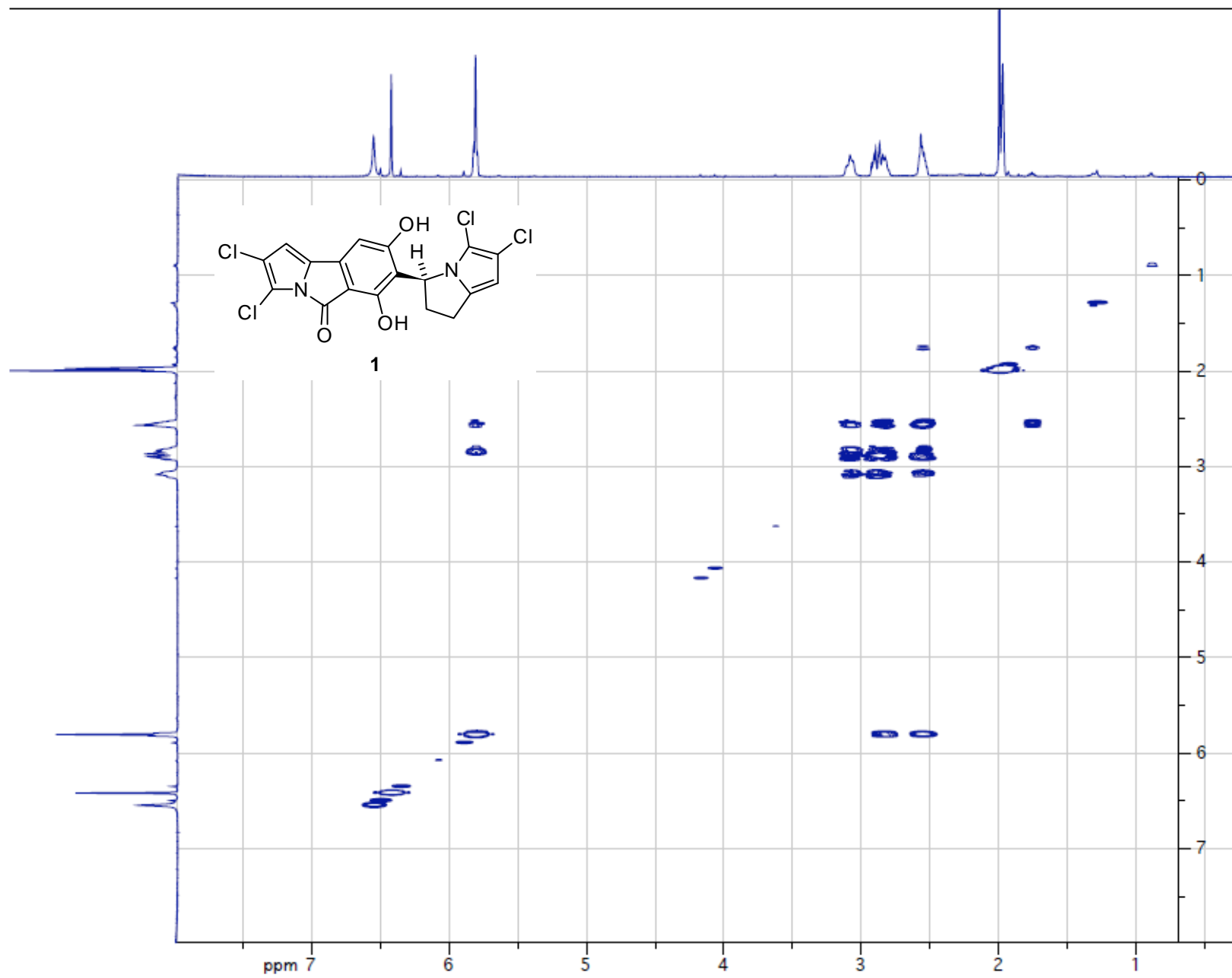
^[a] Carbon assignments were based solely on HSQC and HMBC data collected at 600 MHz. Carbons 2, 4, 5, 6, 12, 23, and 24 could not be assigned. ^[b] 600 MHz.

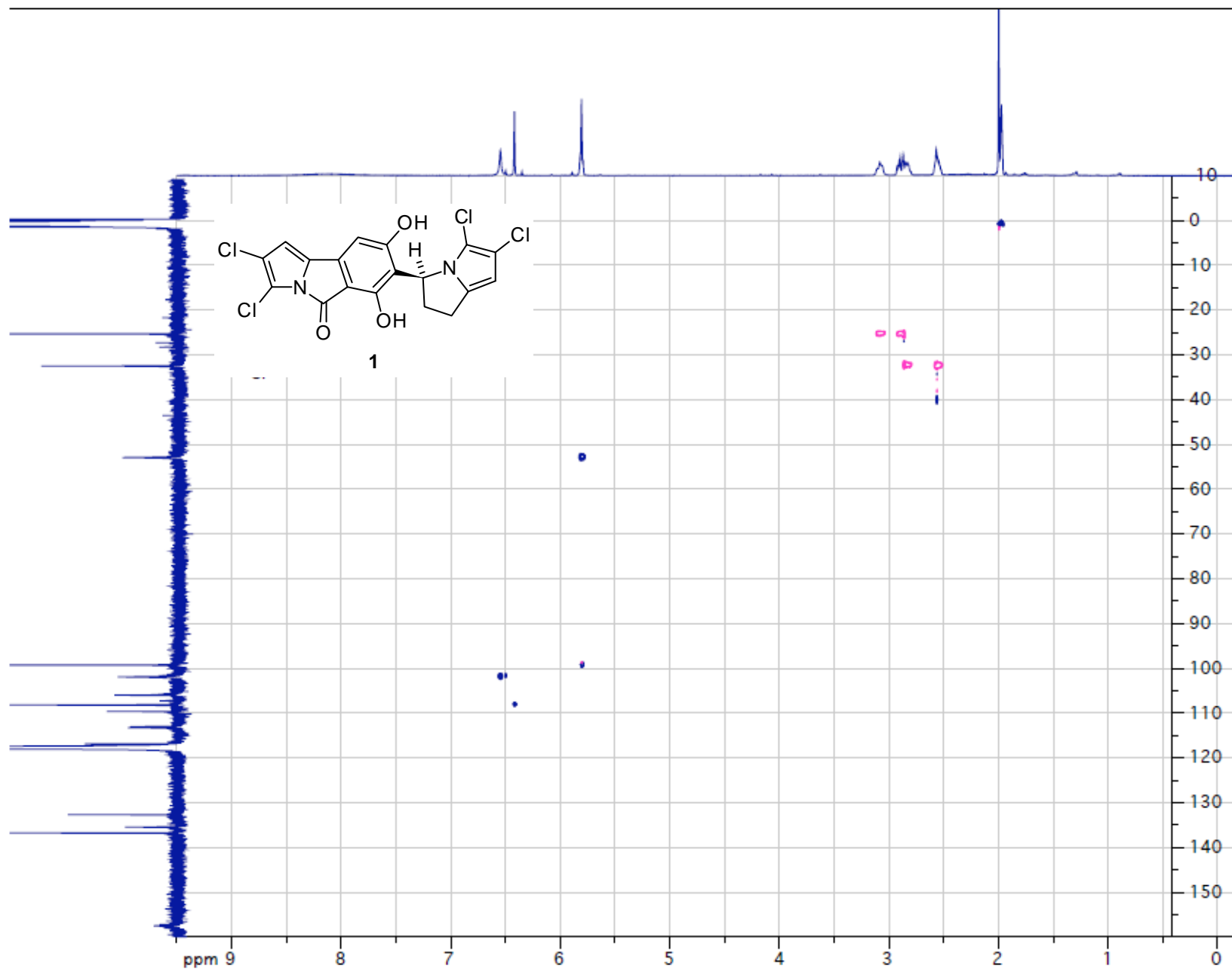
Table S4. Selected NMR spectral data for **10** (CD₃CN)

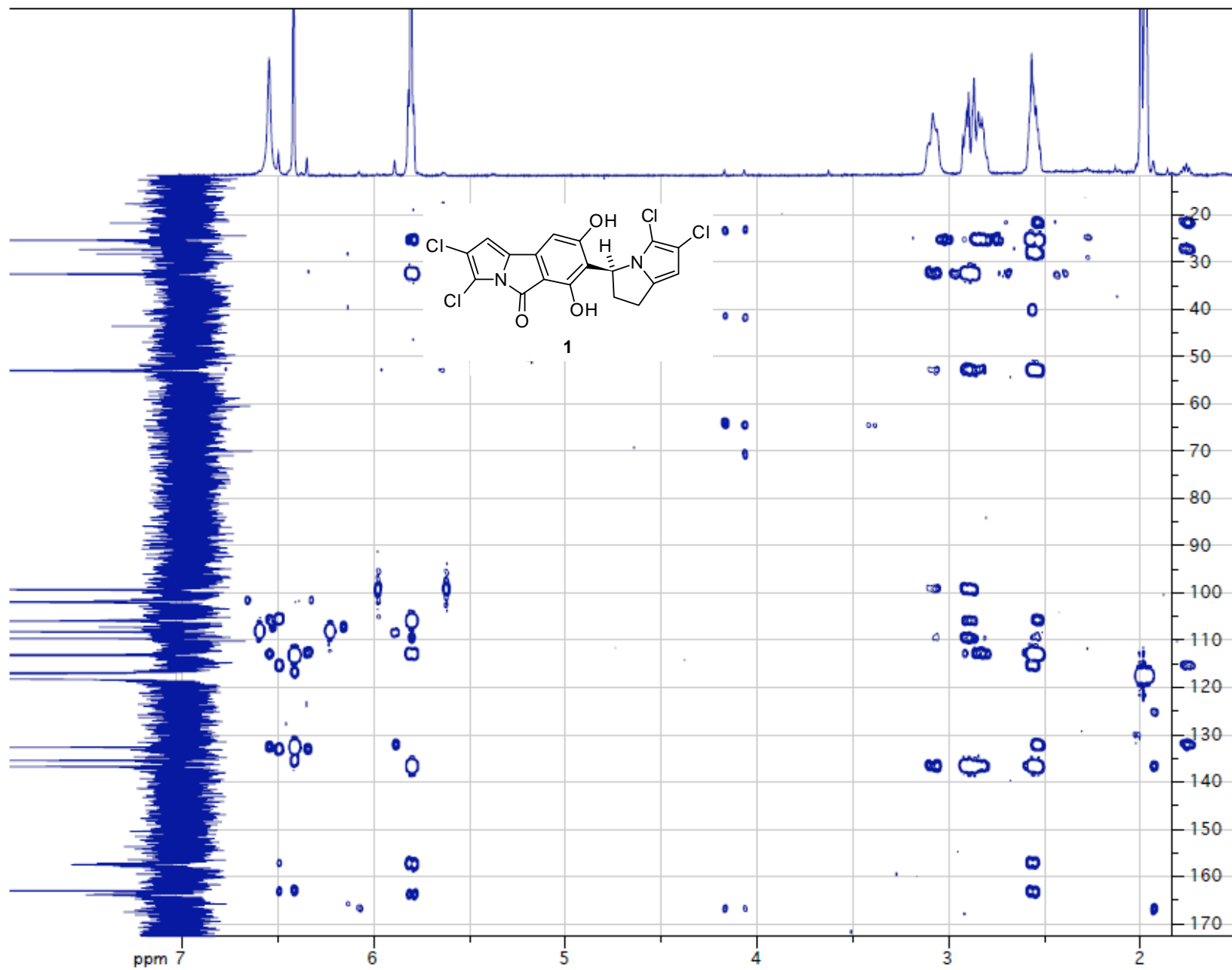
position	δ_C [a]	δ_H , mult. (J, Hz) [b]	HMBC
1		10.03, br s	
3	110.0	6.21, s	
7	110.2	6.32, br s	
9	114.3		
11	106.2		
12	170.3		
13		6.46, br s	
14	44.1	4.34, br s	12,15,16
15	138.3		
16	128.3	7.15, m	14
17	129.1	7.32, m	14,15
18	127.8	7.26, m	15,16
19	53.9	5.89, br s	
20	32.8	2.81, m	
		2.53, br s	9,19,21,21a
21	26.0	3.07, br s	
		2.87, m	19,20,21a,22
21a	137.5		
22	99.5	5.77, s	19,21,21a

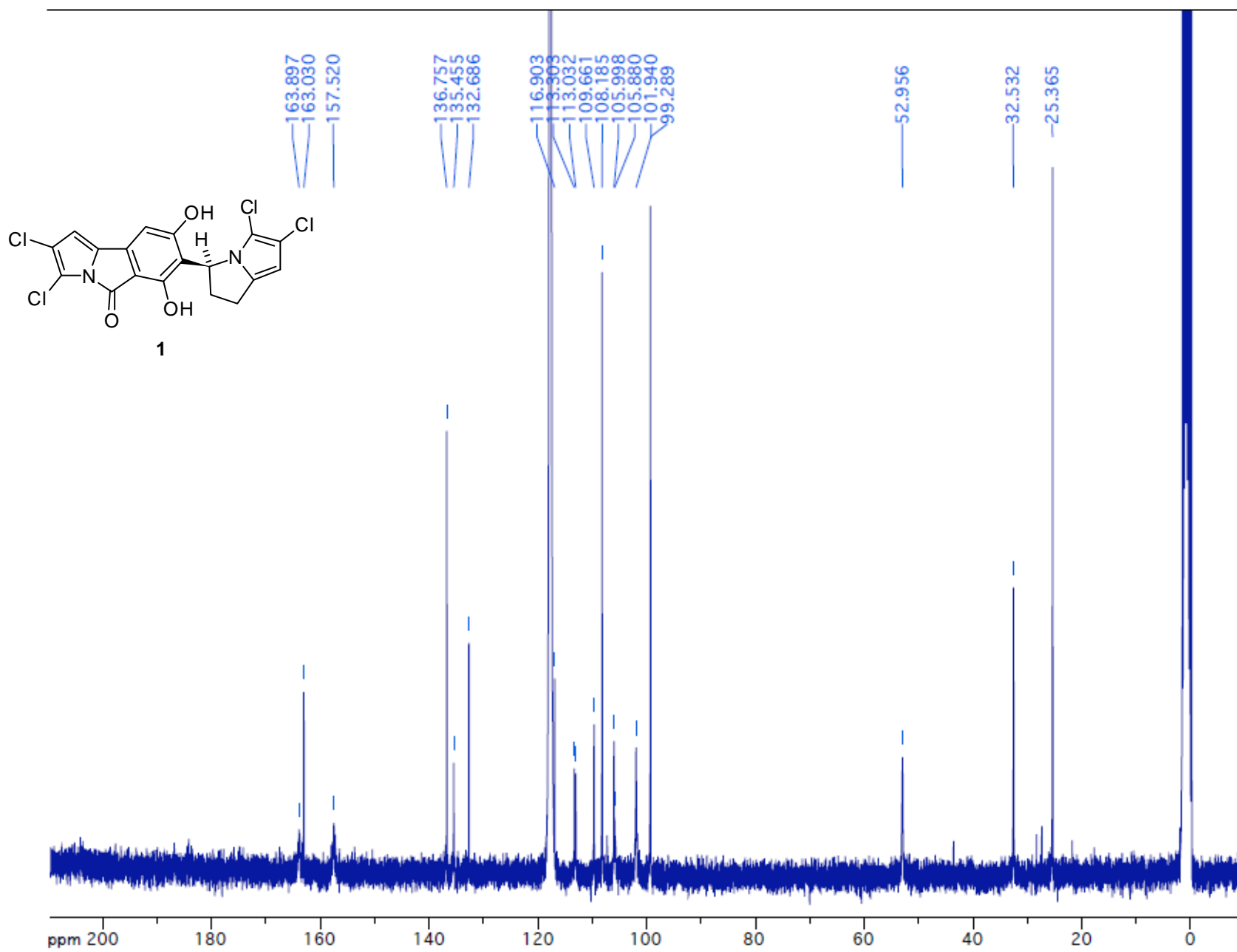
[a] Carbon assignments were based solely on HSQC and HMBC data collected at 600 MHz. Carbons 2, 4, 5, 6, 8, 10, 23, and 24 could not be assigned. [b] 600 MHz.

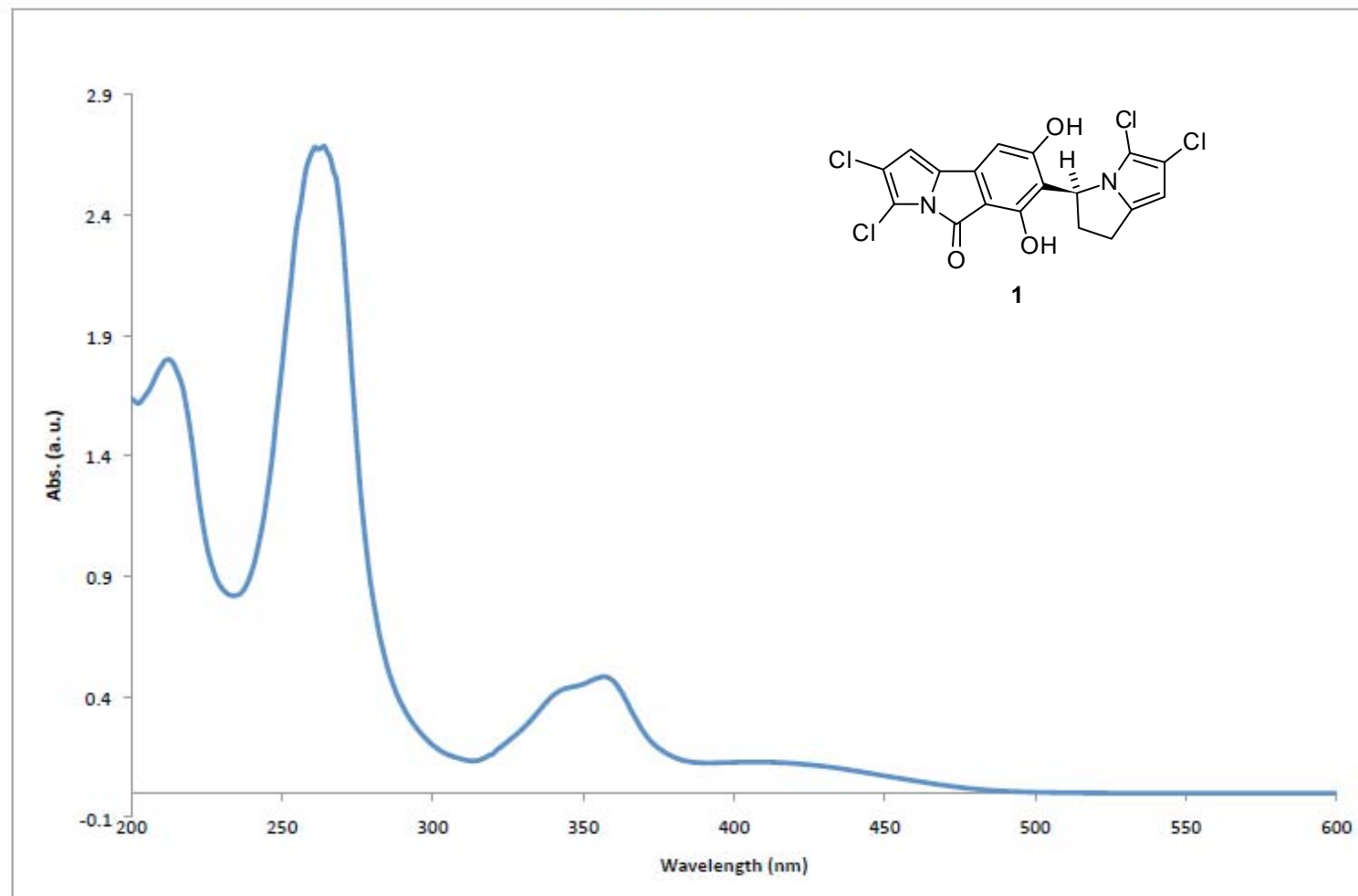




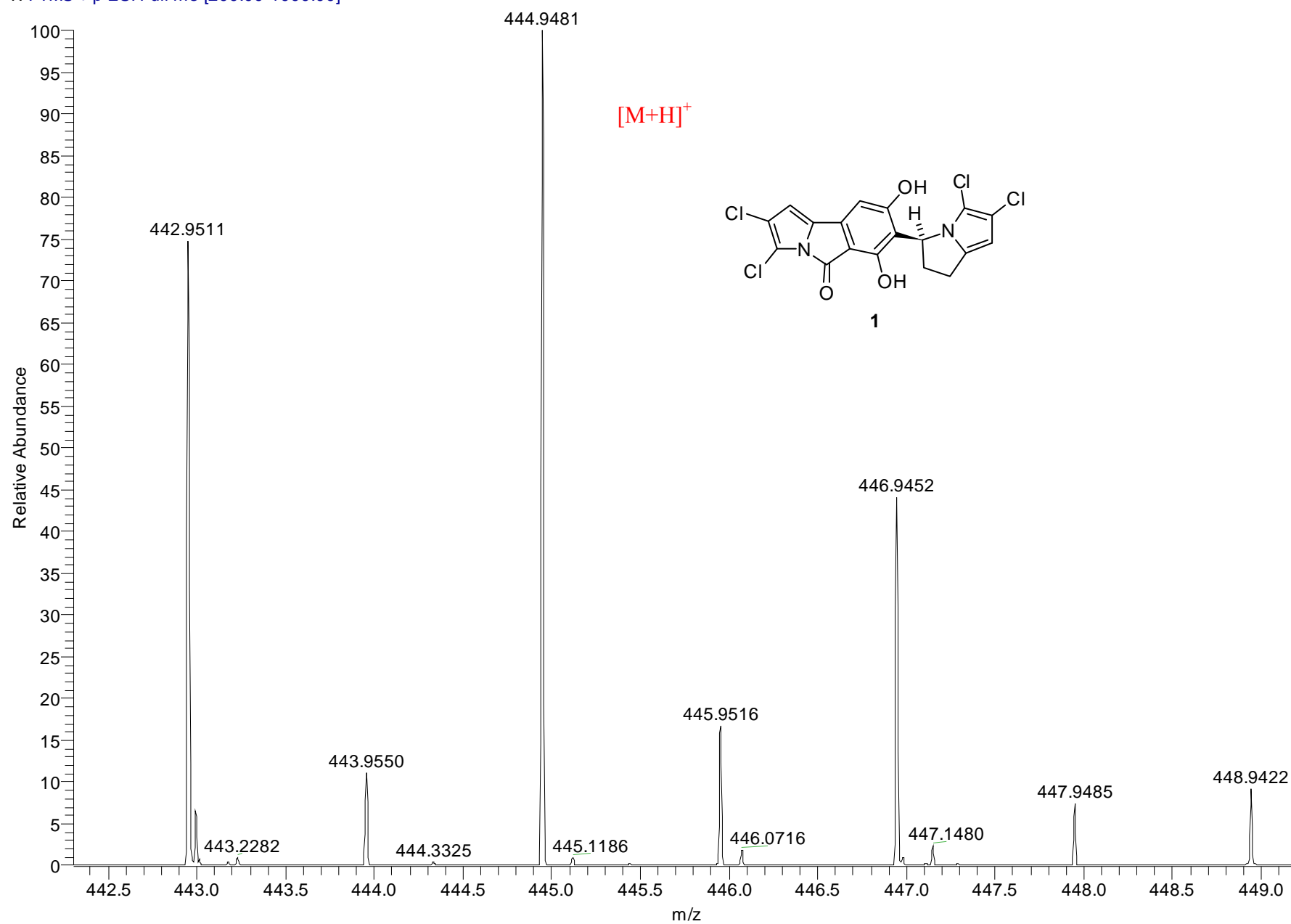


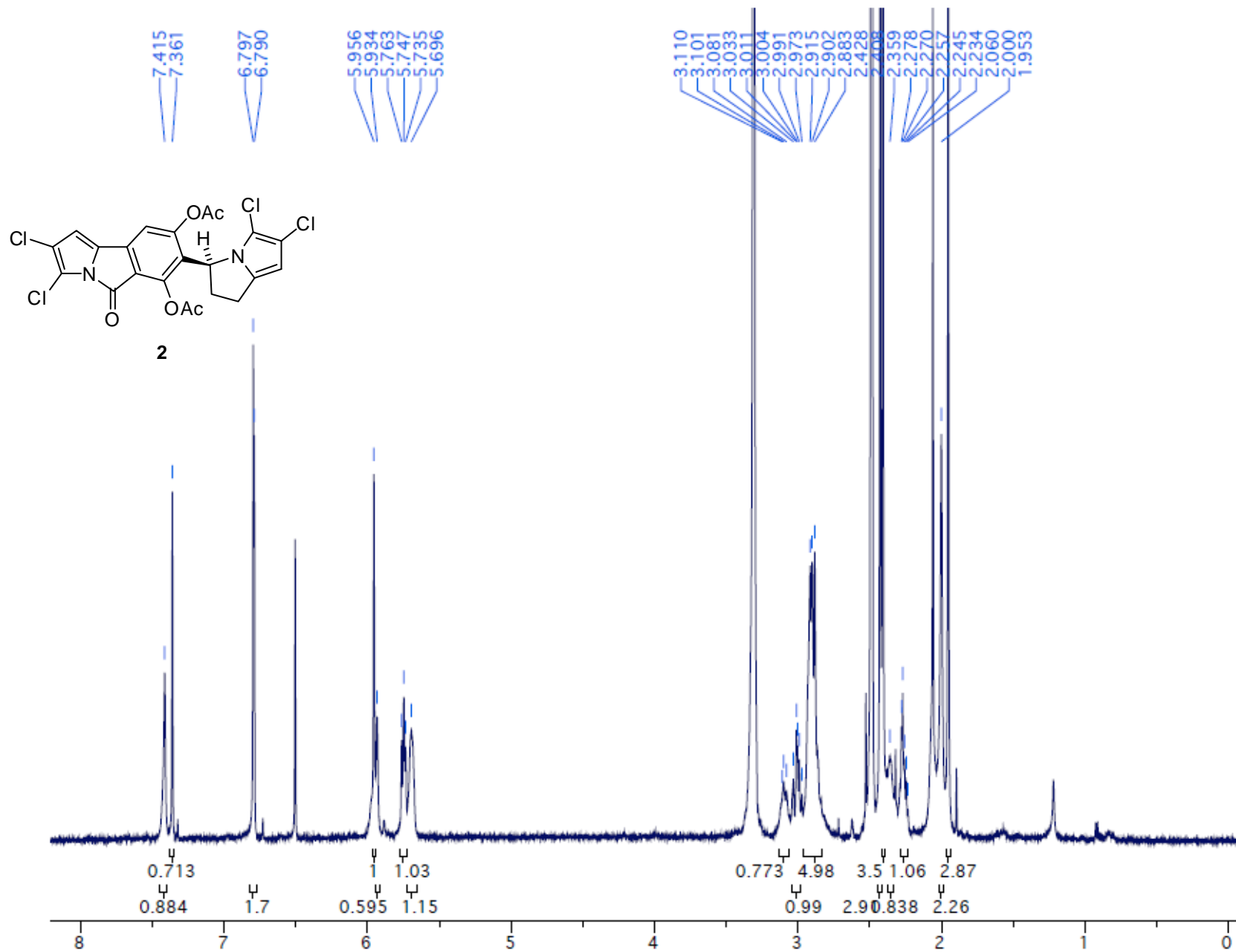


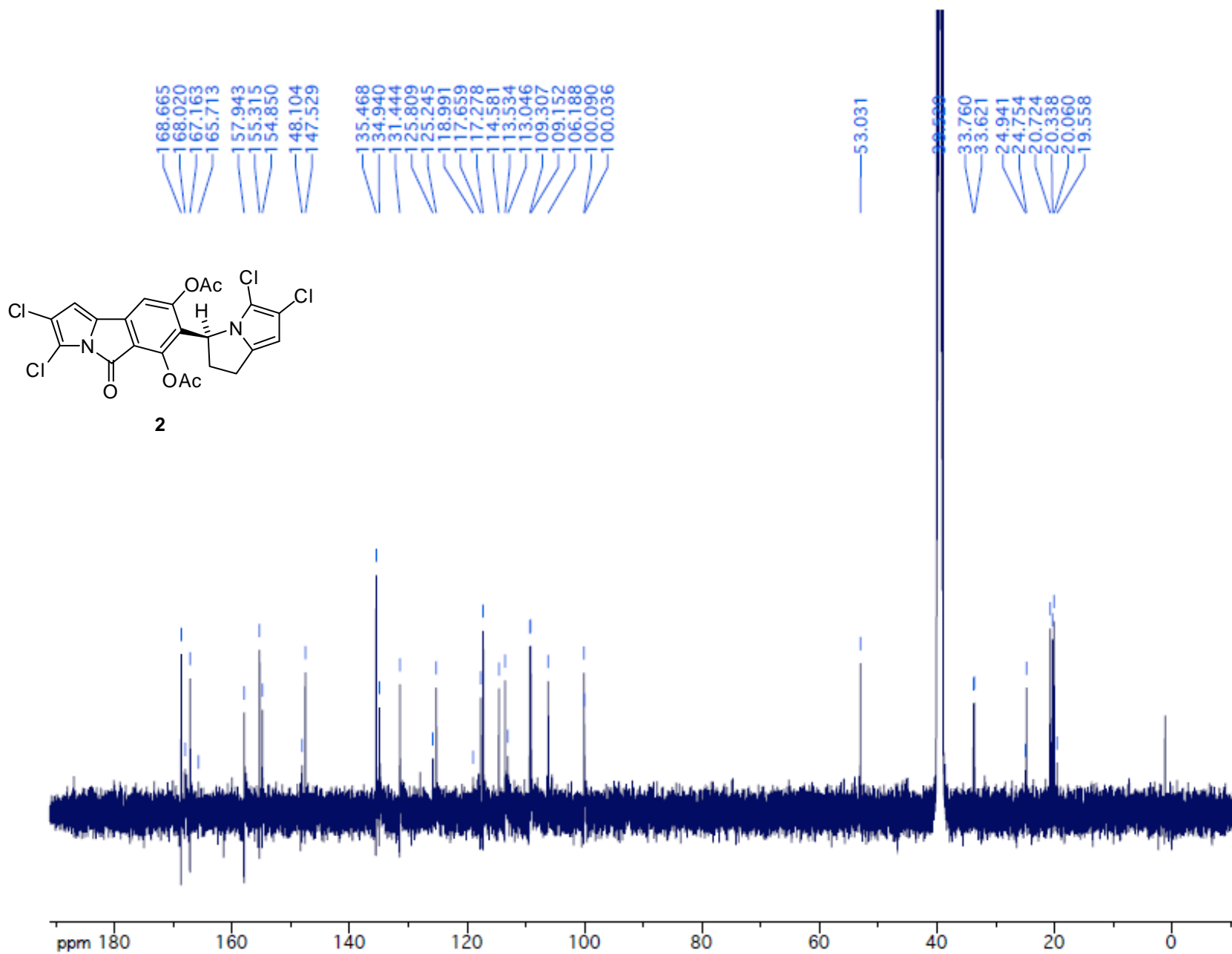


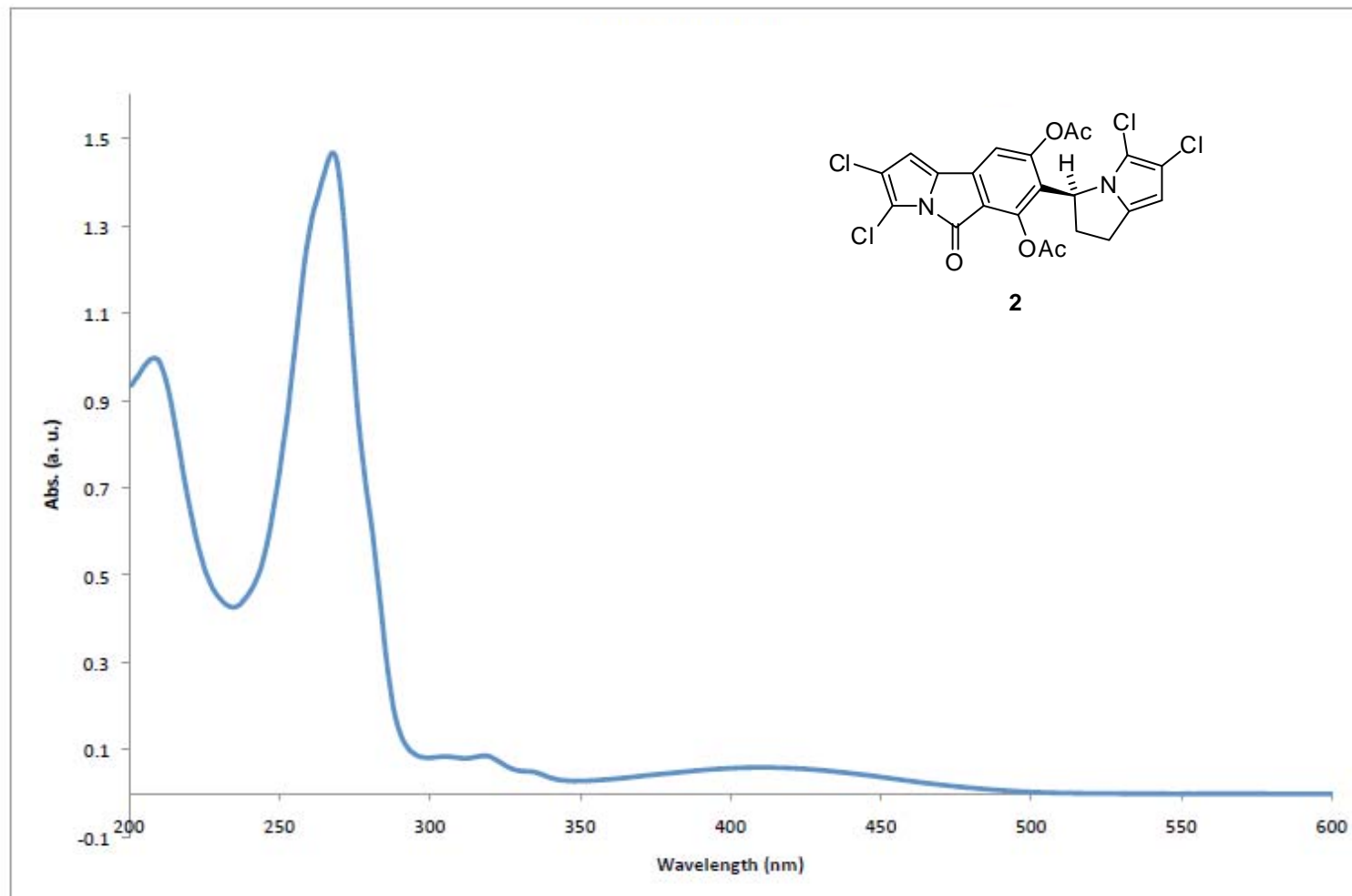


CNH287442 #110-119 RT: 1.47-1.55 AV: 10 NL: 1.85E5
T: FTMS + p ESI Full ms [200.00-1000.00]



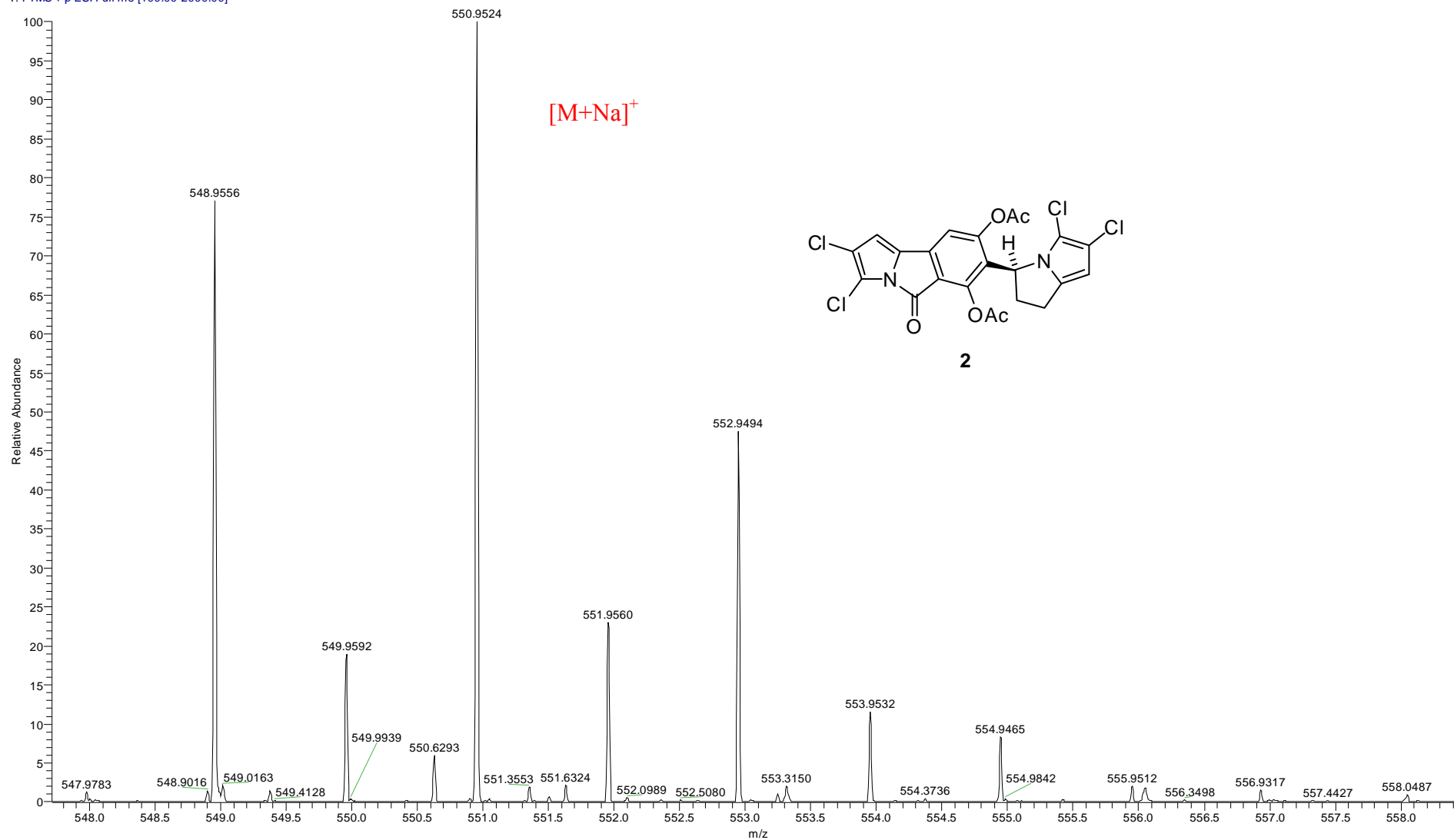


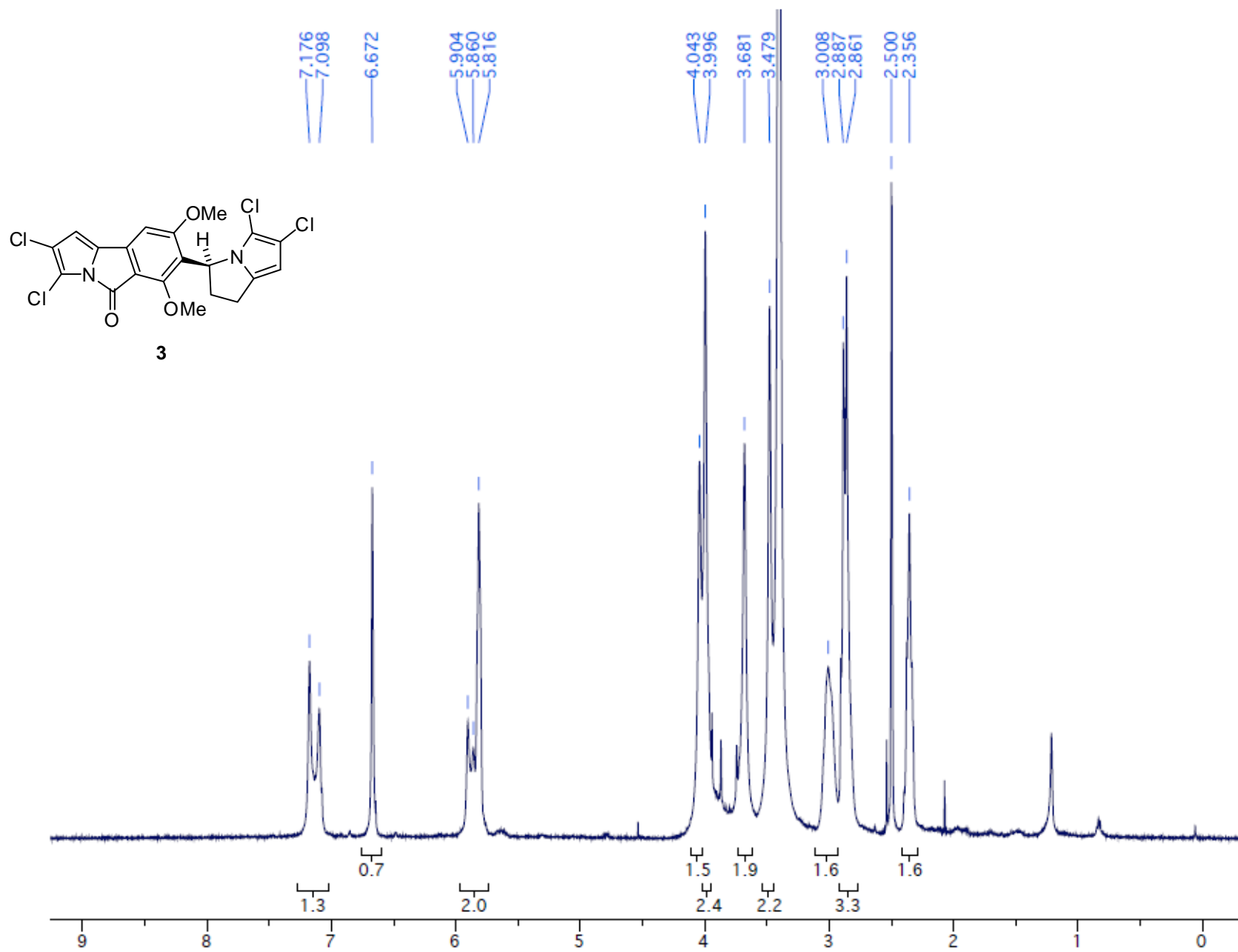


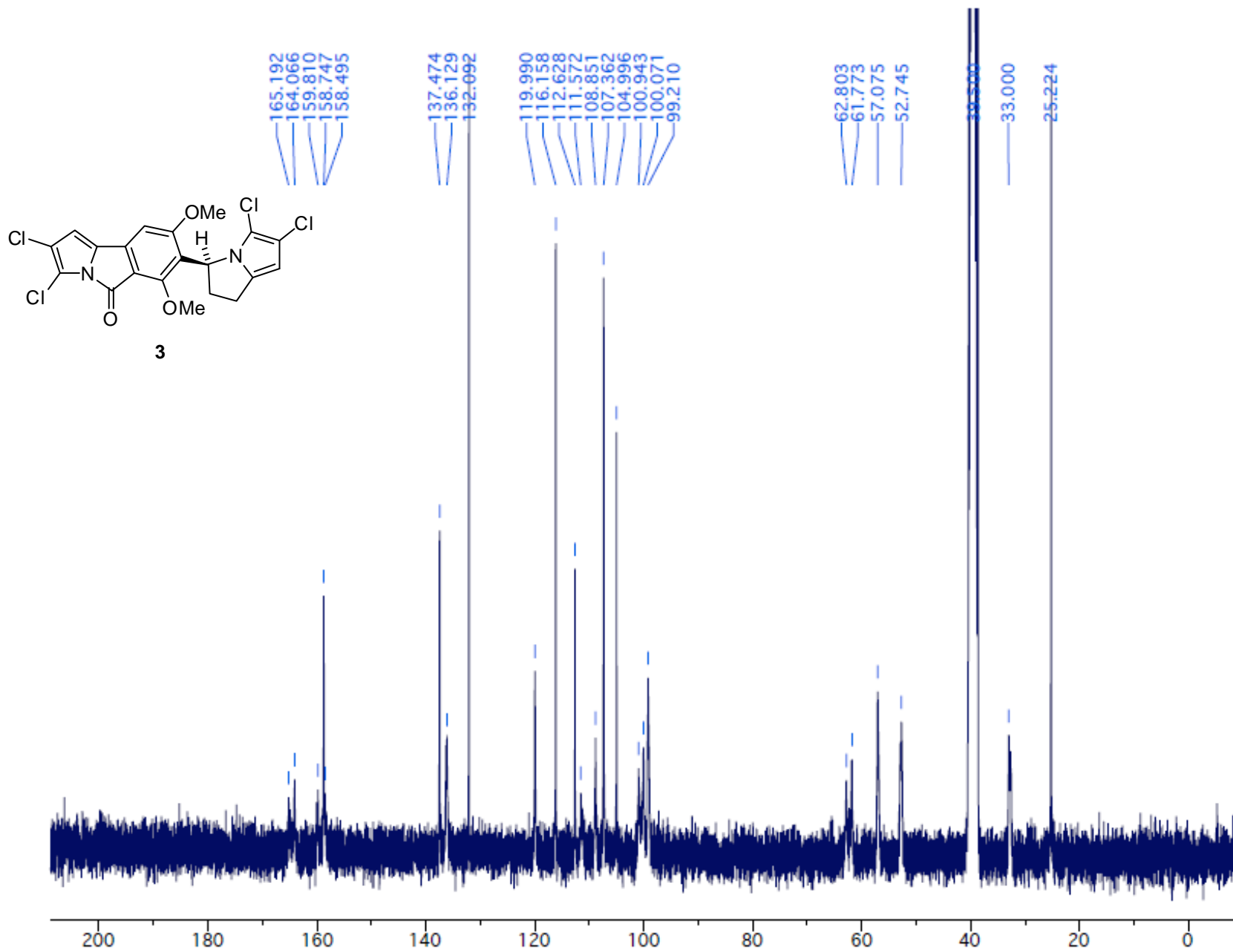


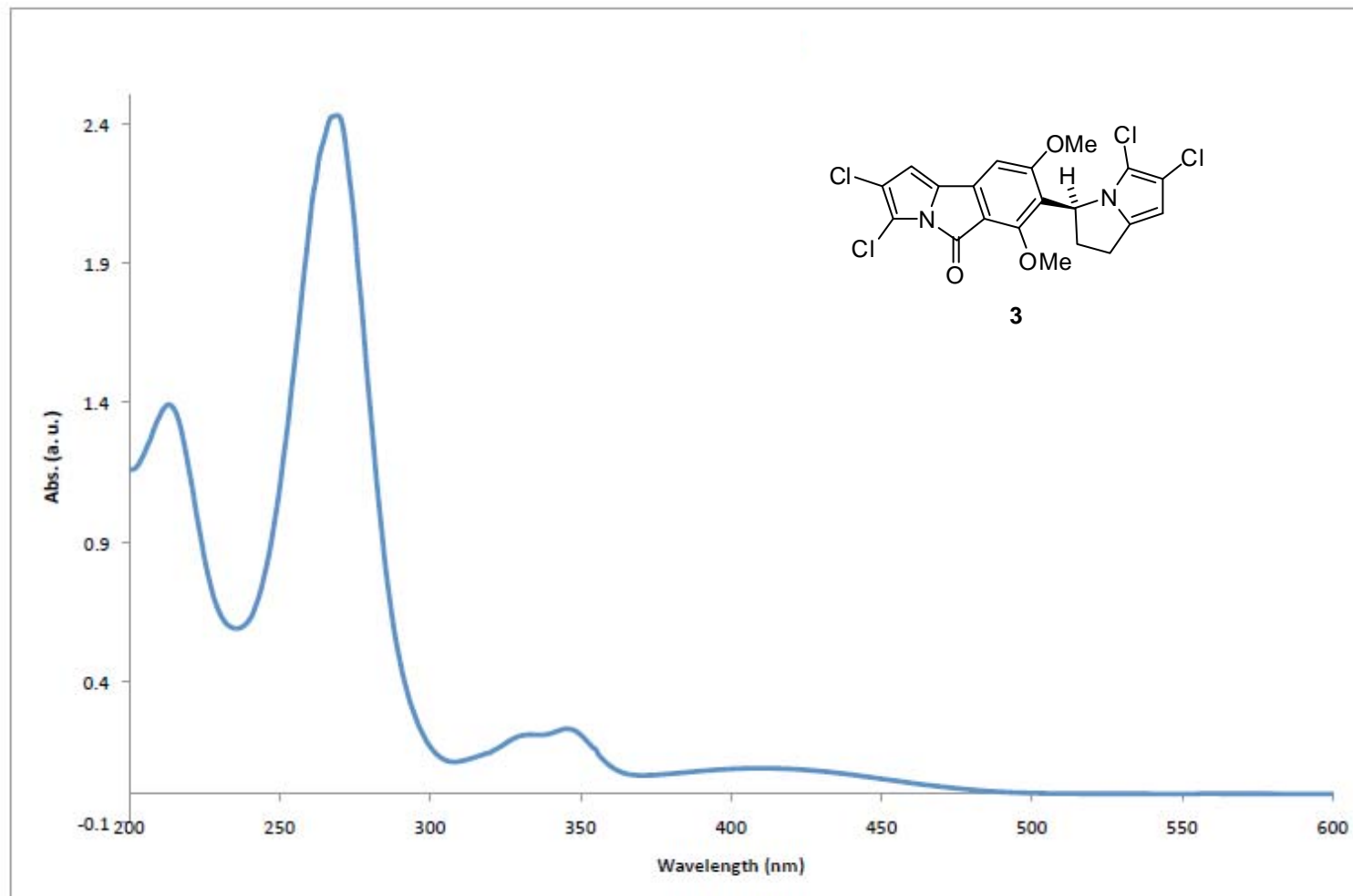
HRESI-FT-MS (Orbit-Trap-MS) of diacetyl chlorizidine A (2)

H287-Ac #114-117 RT: 0.99-1.02 AV: 4 NL: 1.31E6
 T: FTMS + p ESI Full ms [100.00-2000.00]



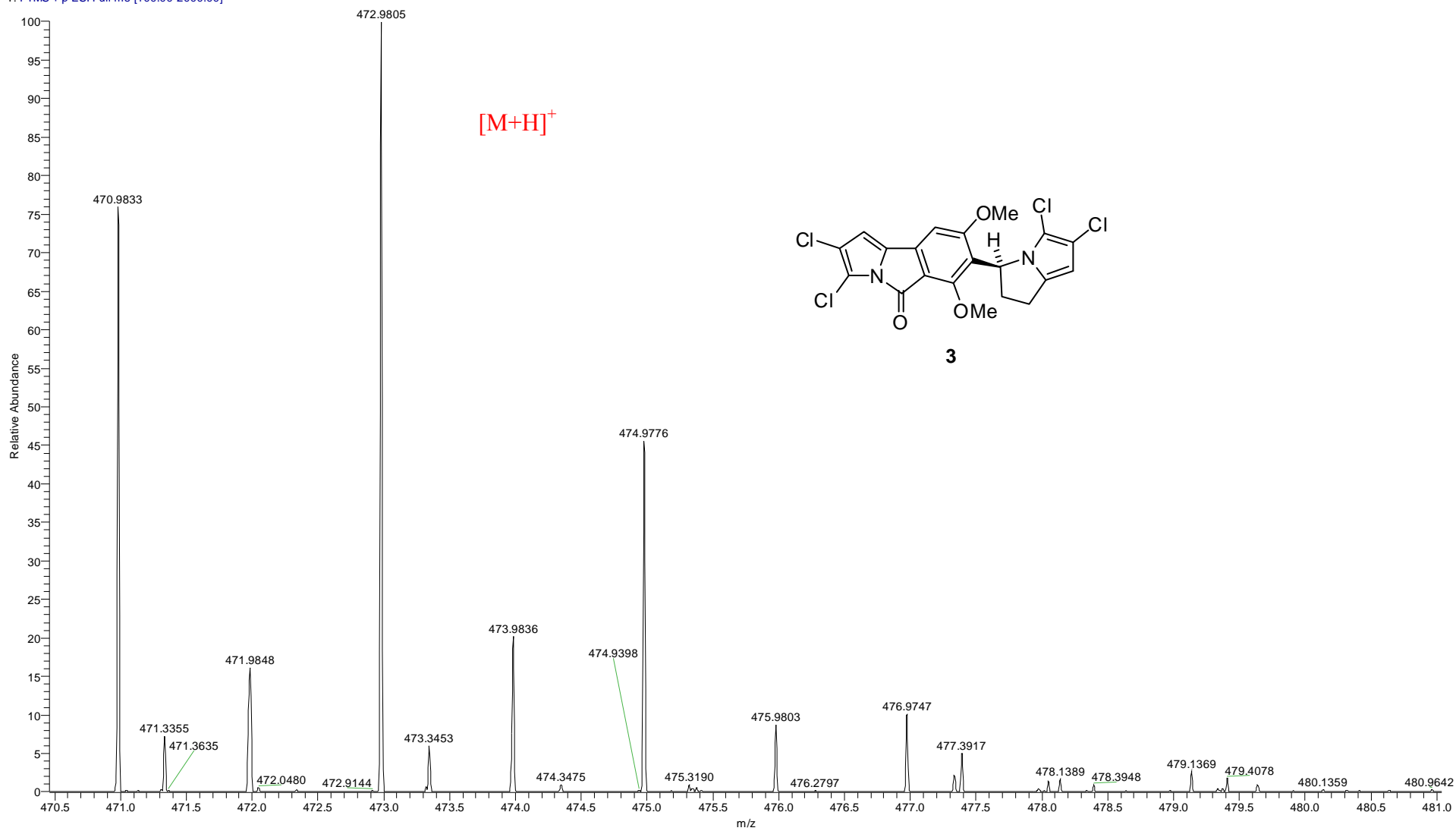


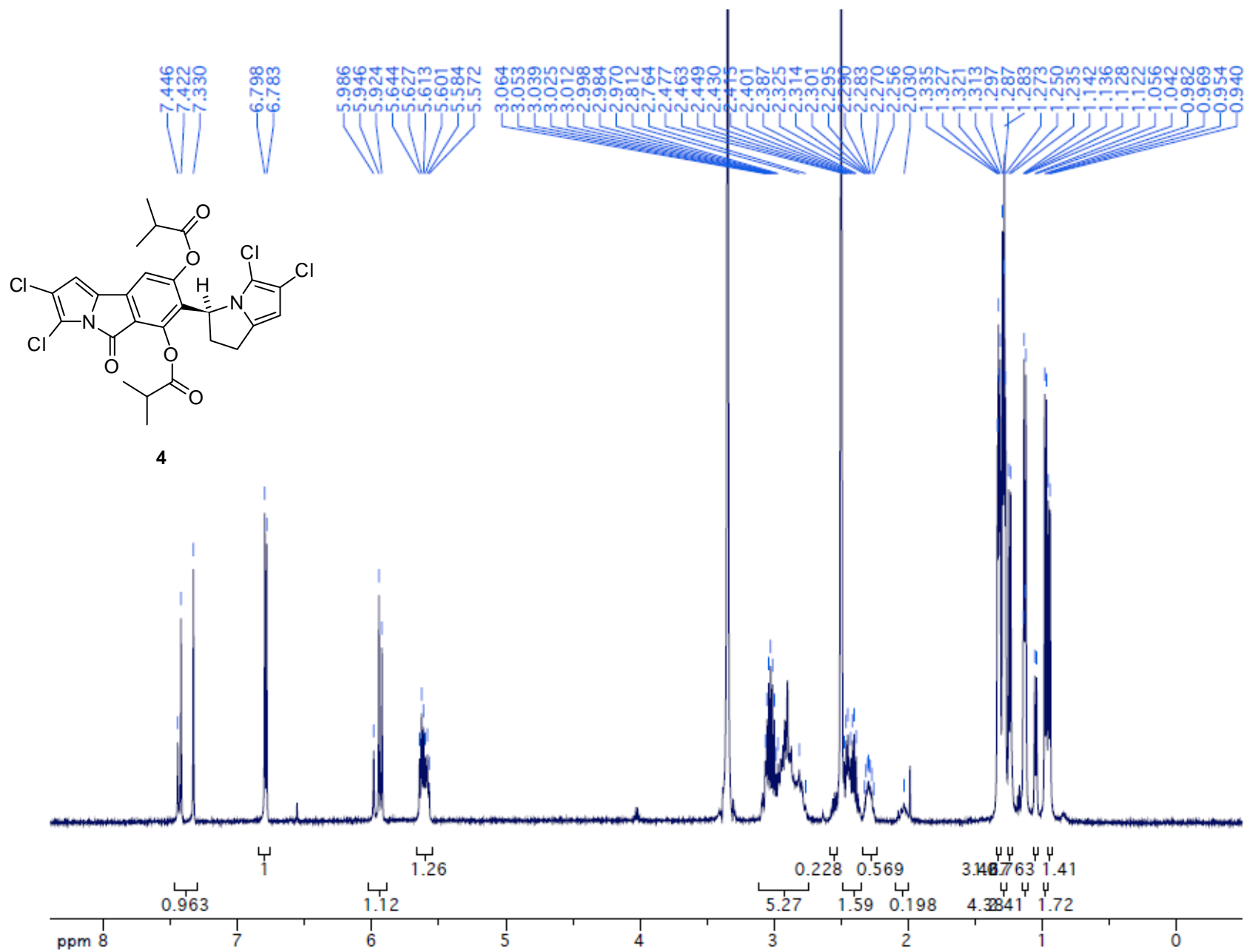


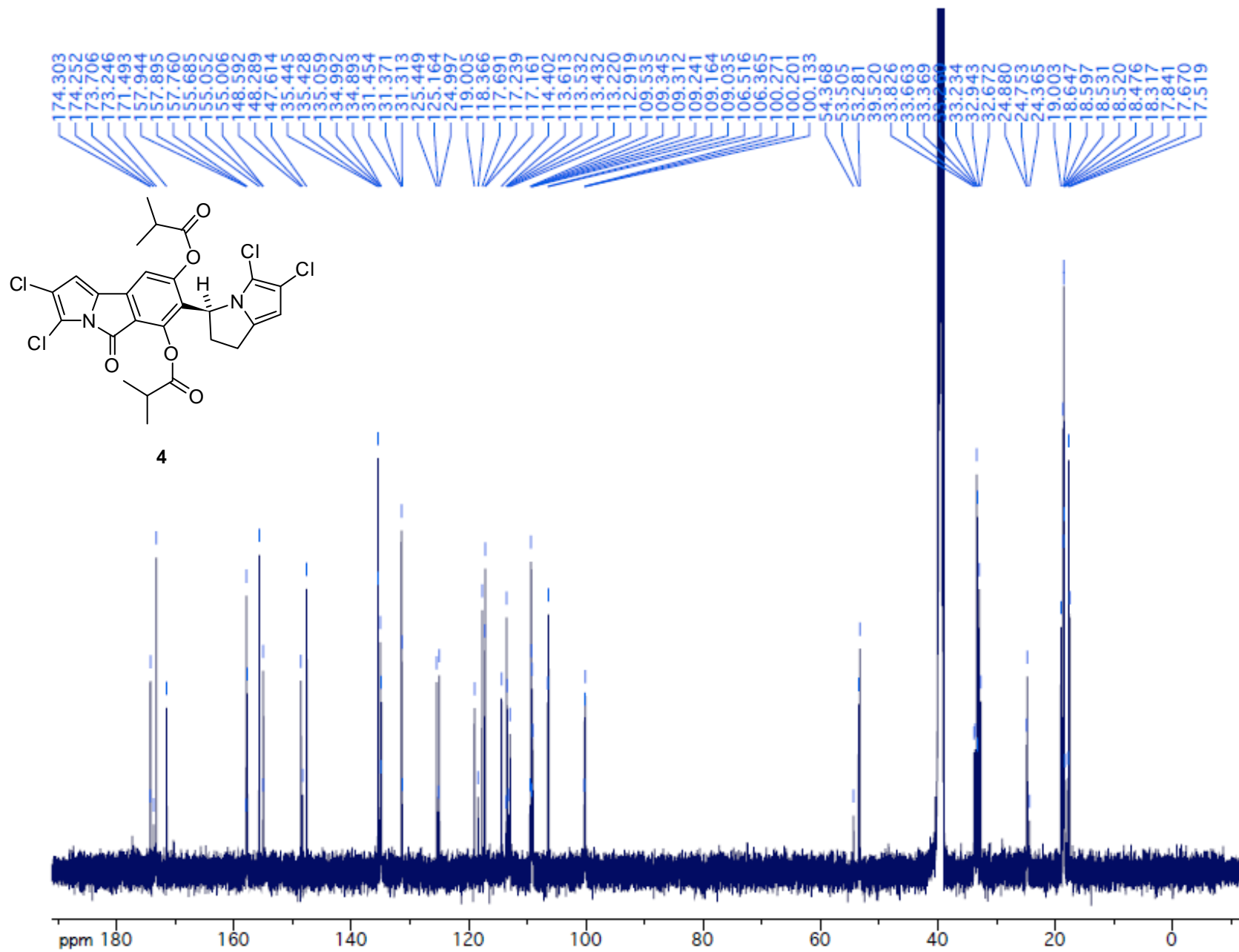


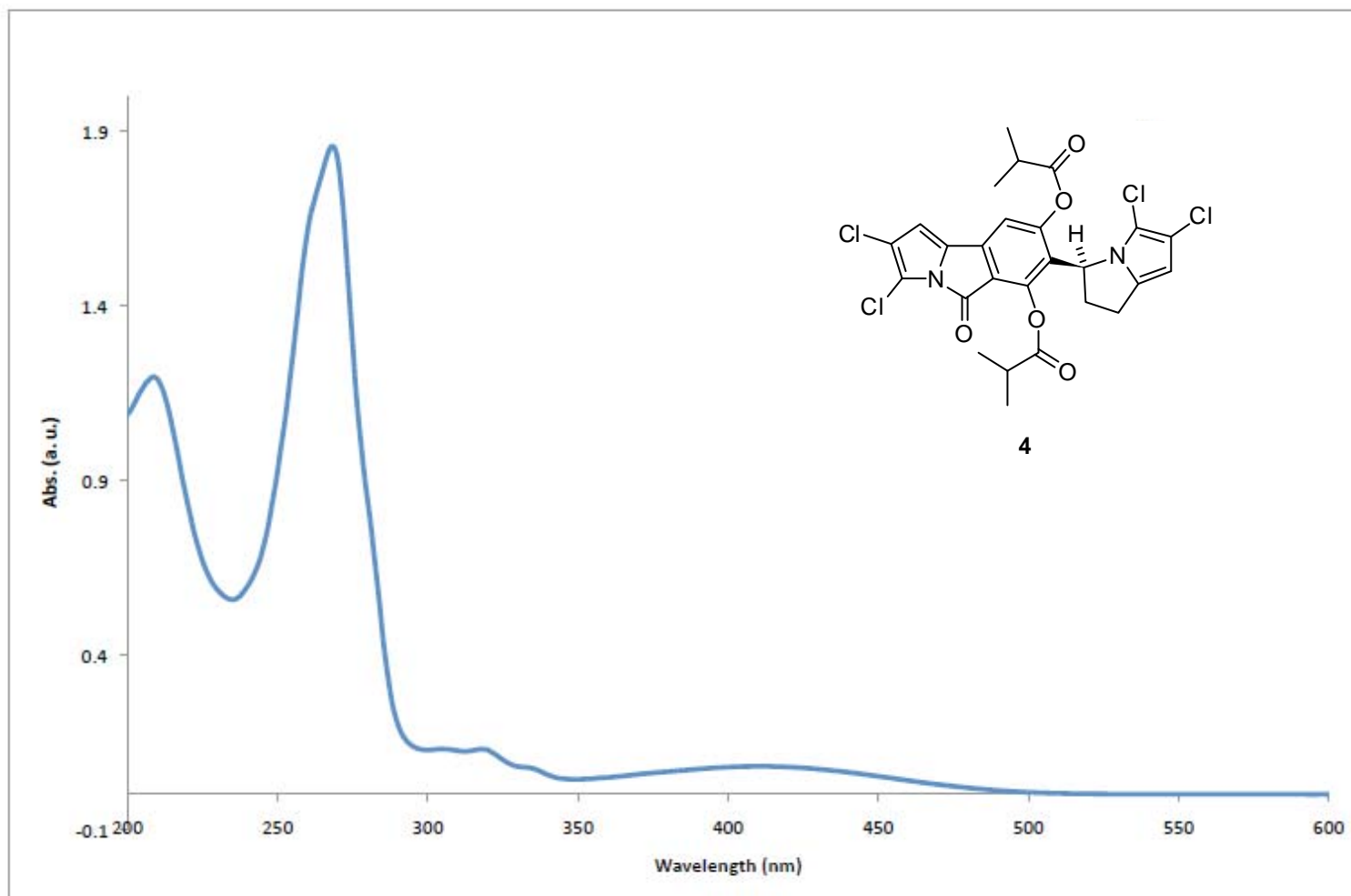
HRESI-FT-MS (Orbit-Trap-MS) of dimethyl chlorizidine A (3)

H287-Me #554-564 RT: 5.07-5.16 AV: 11 NL: 1.41E6
T: FTMS + p ESI Full ms [100.00-2000.00]

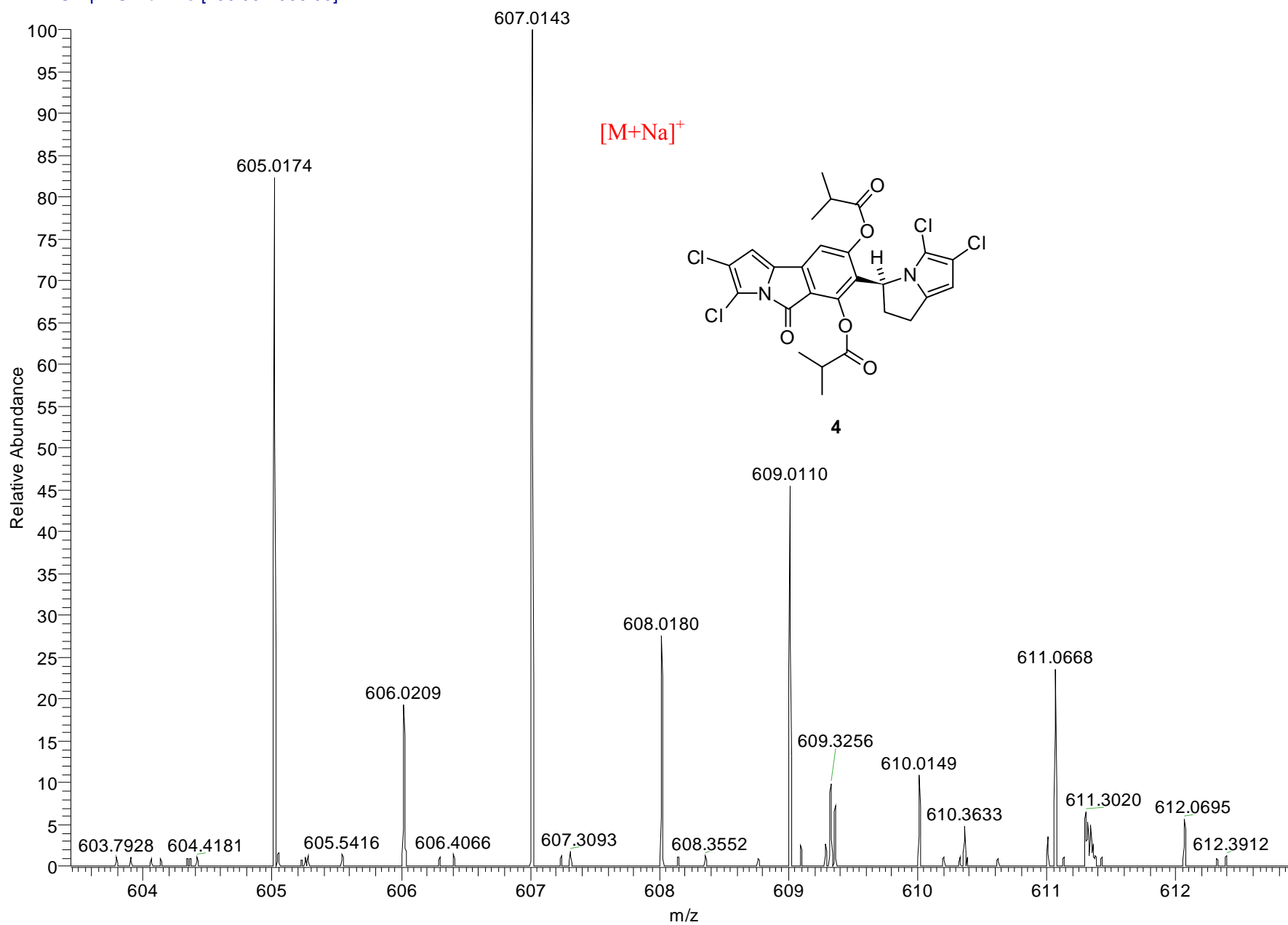


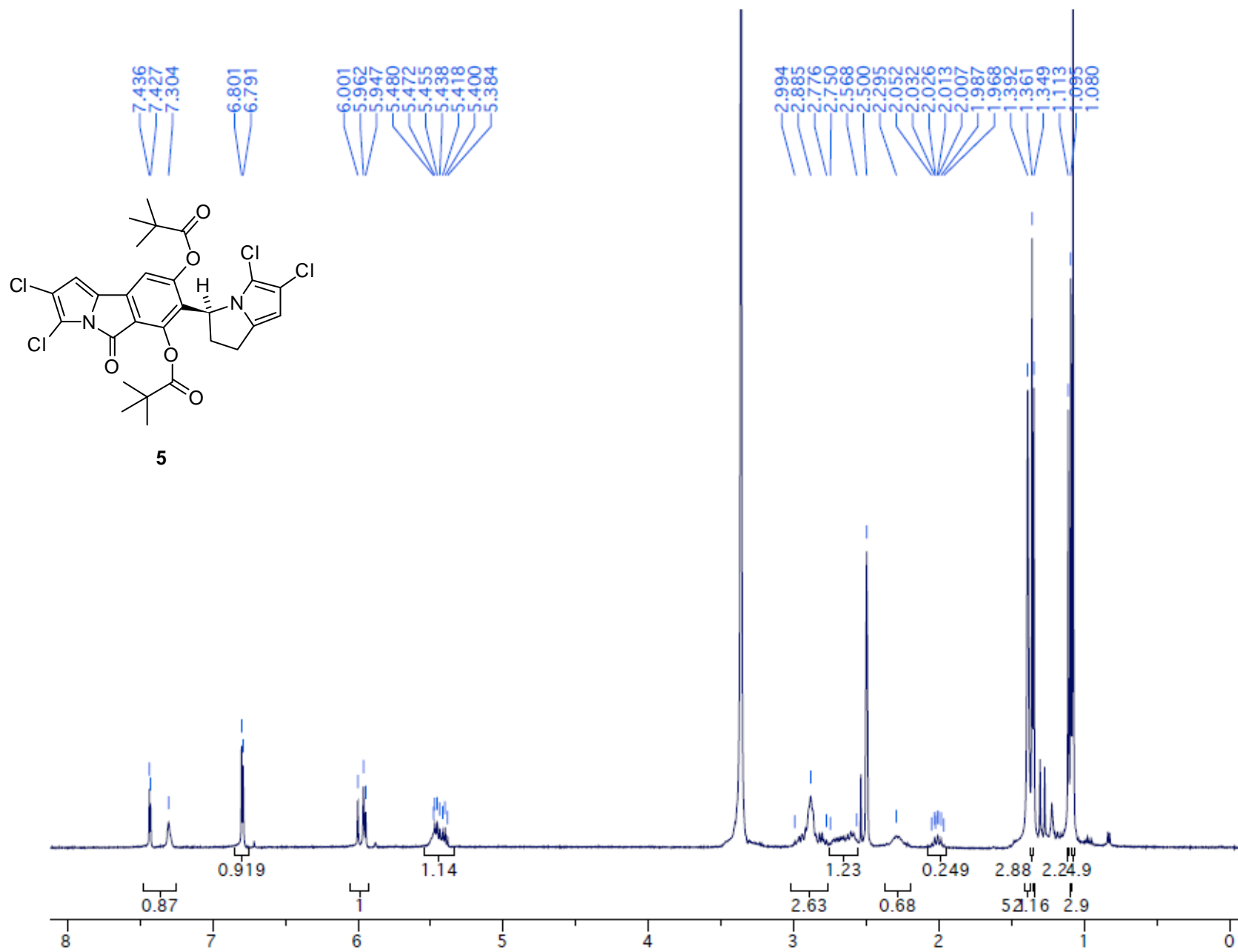


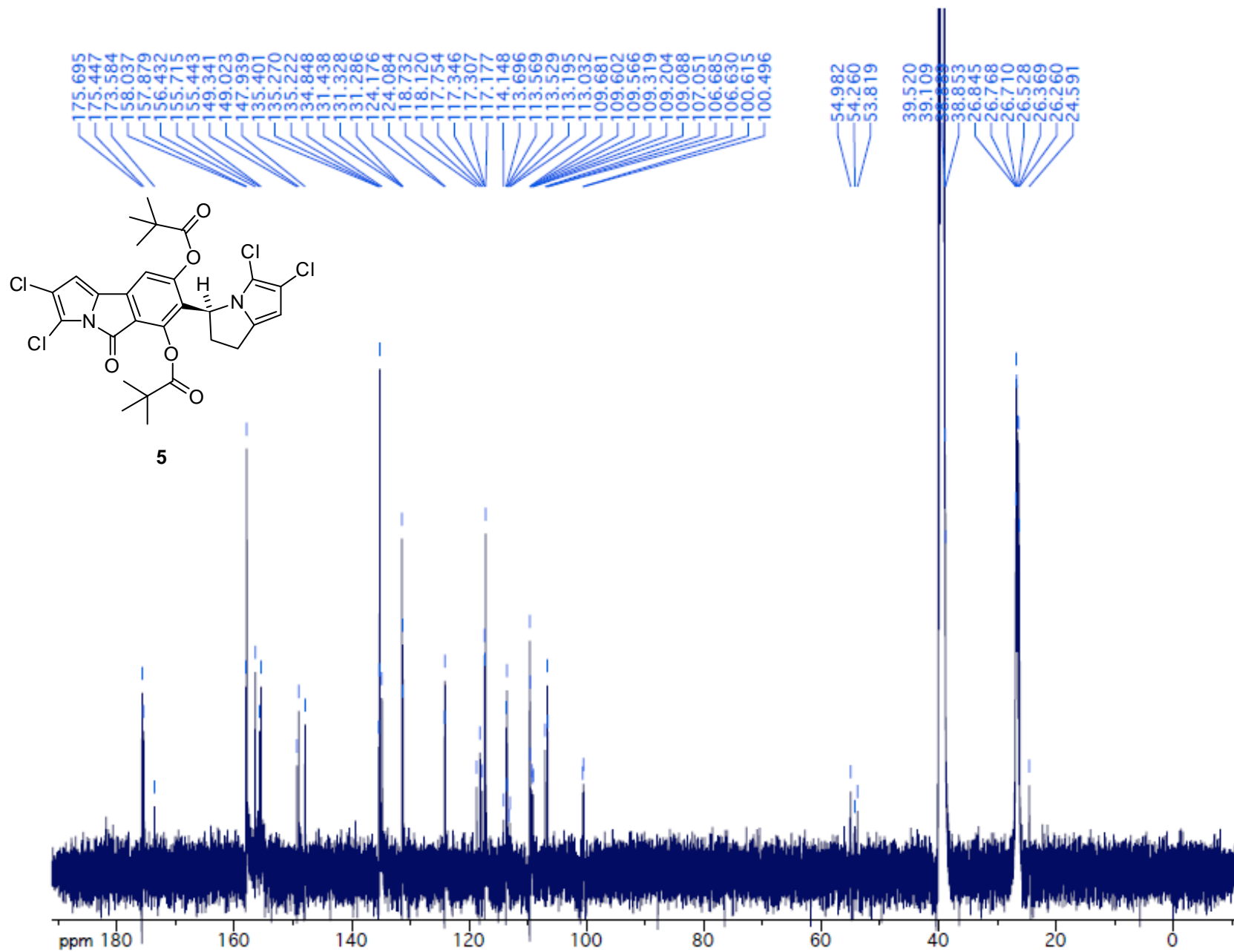


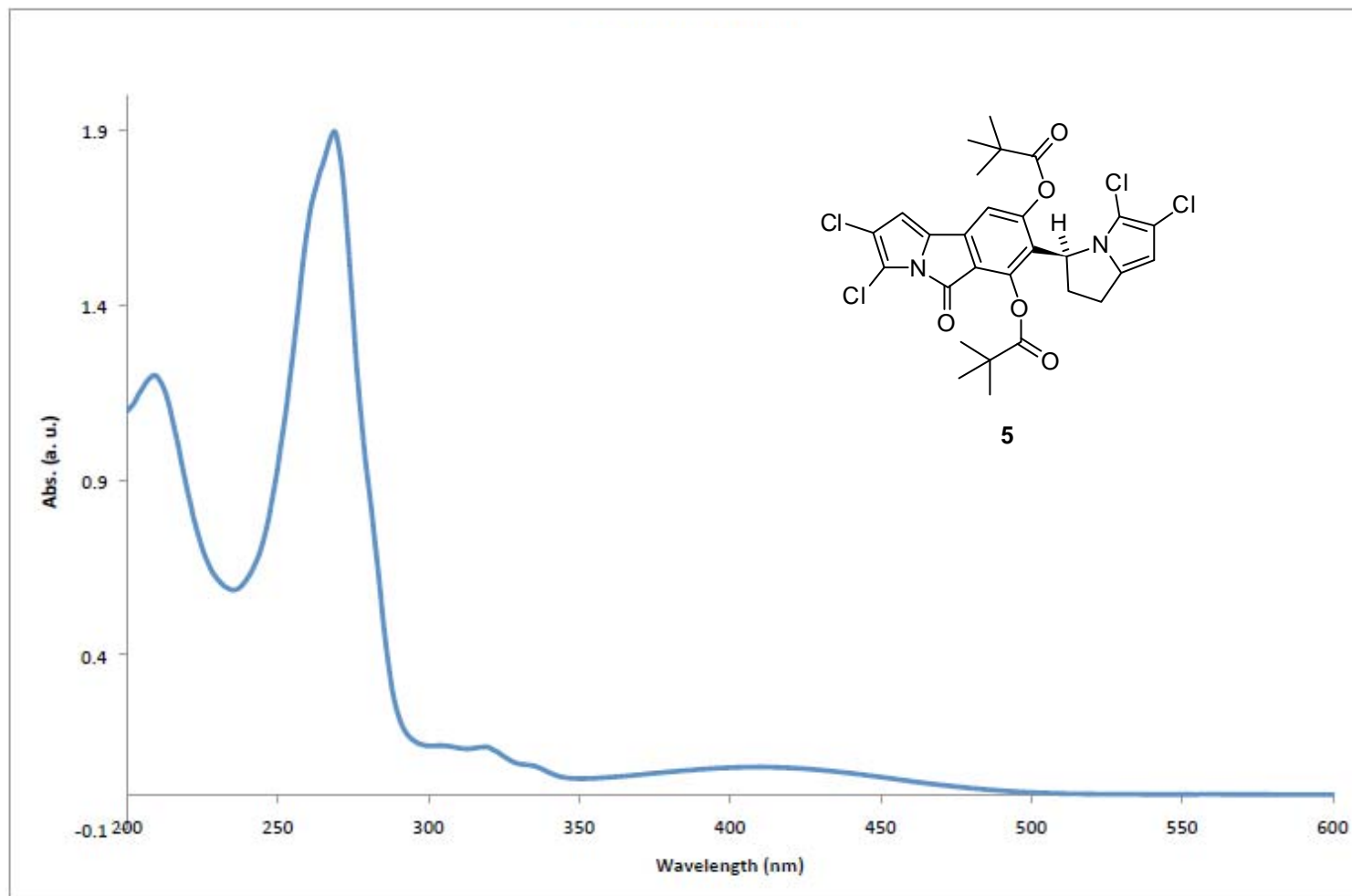


Zsobuty-a #130-135 RT: 2.36-2.47 AV: 6 NL: 8.49E3
T: FTMS + p ESI Full ms [100.00-2000.00]

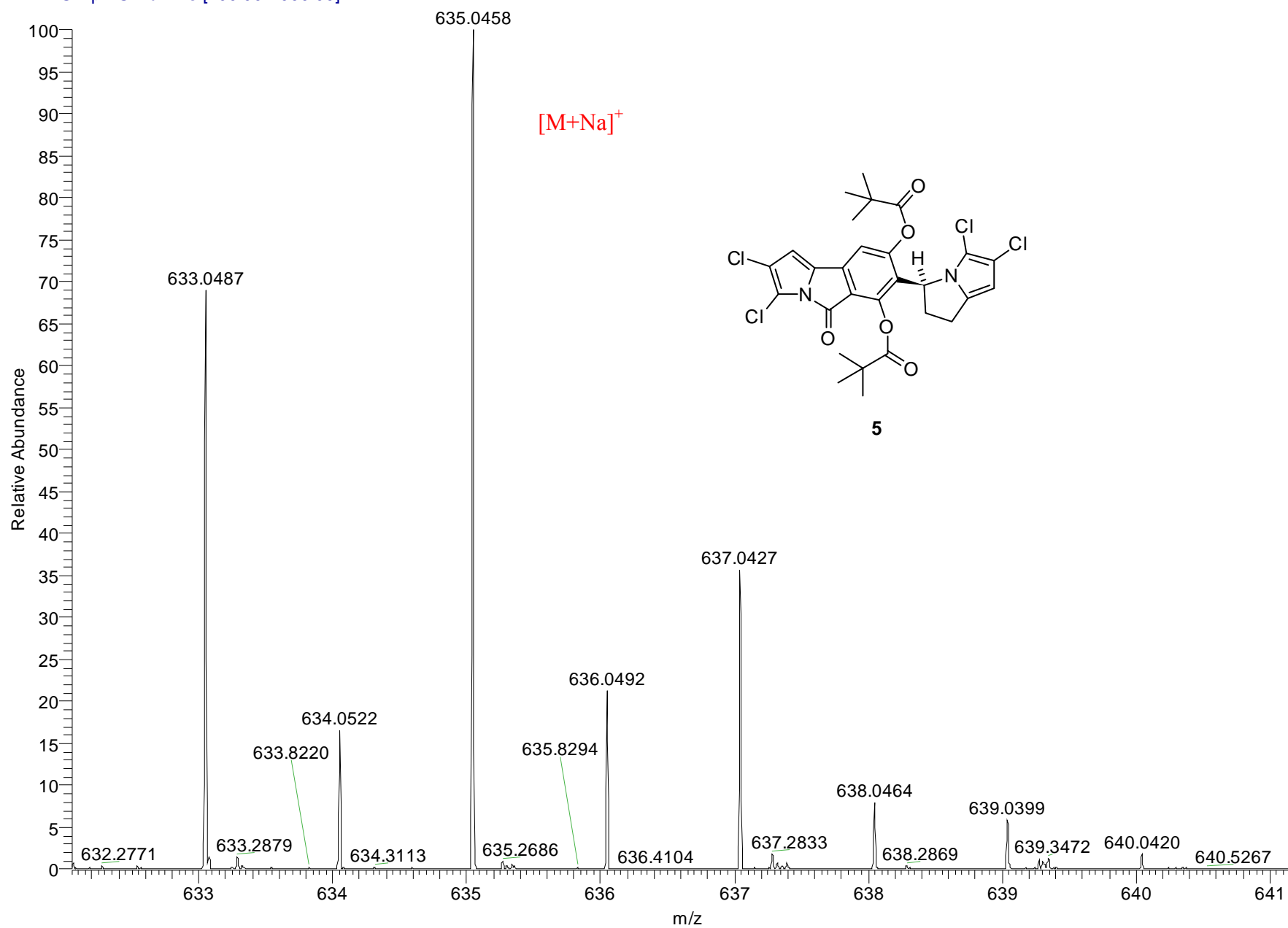


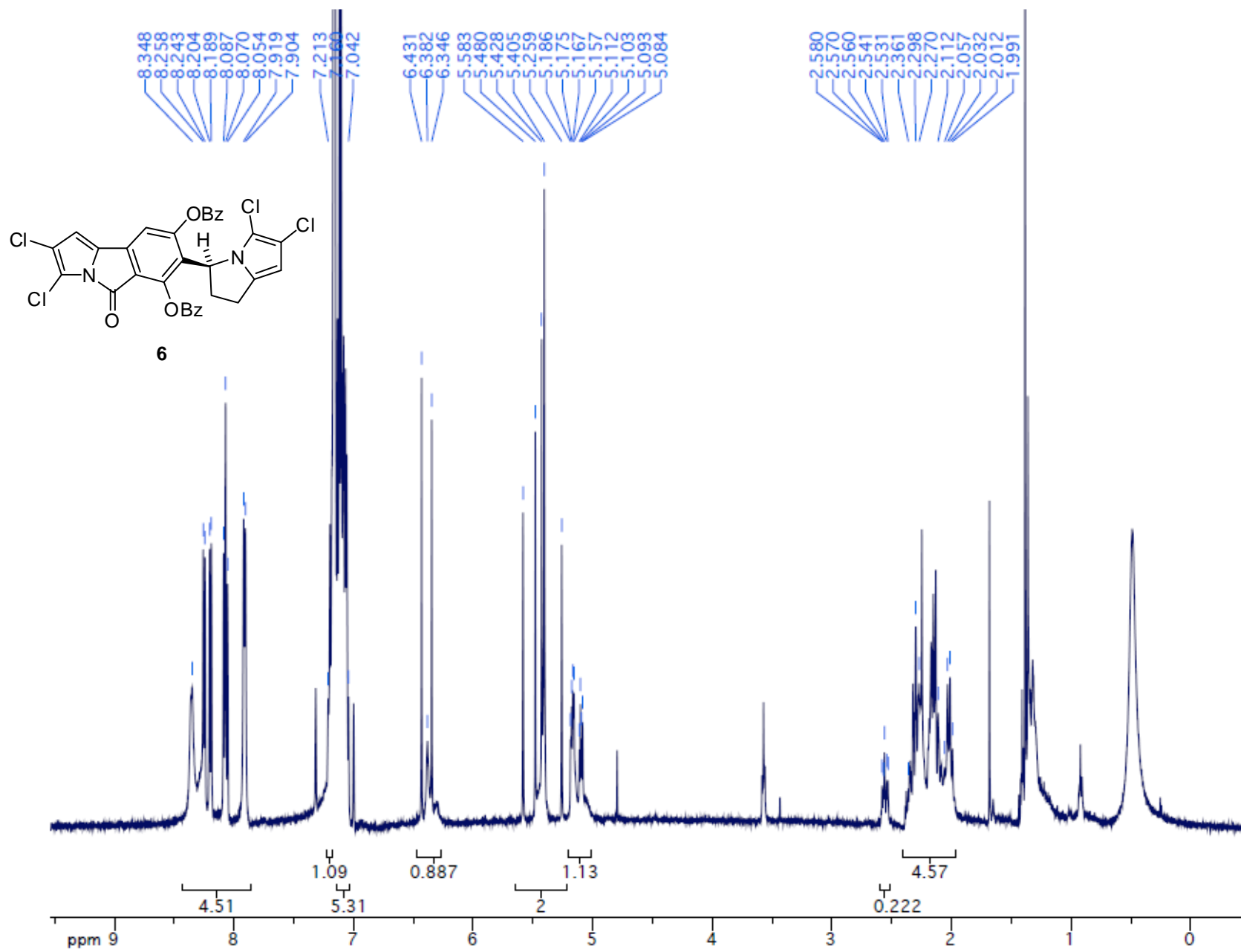


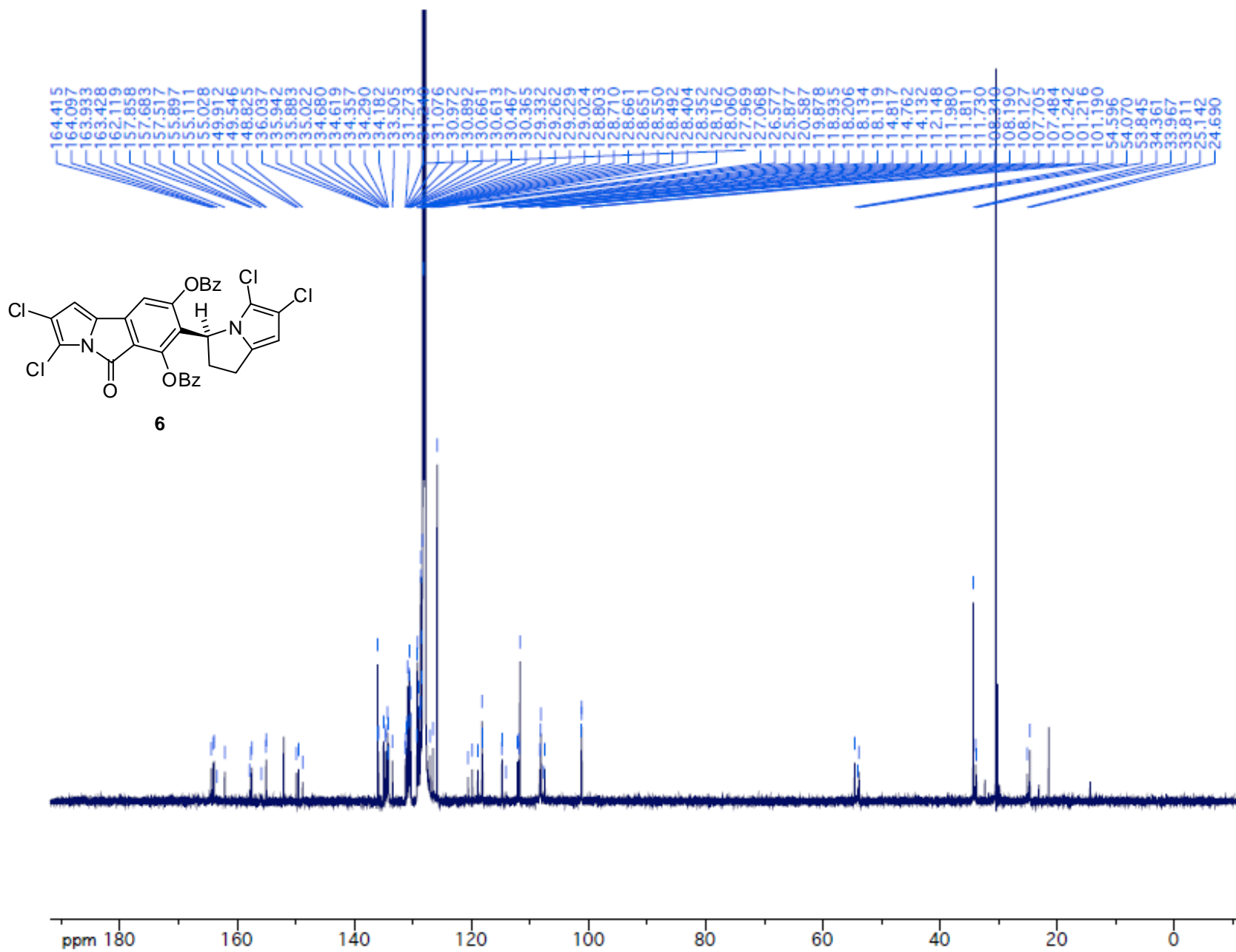


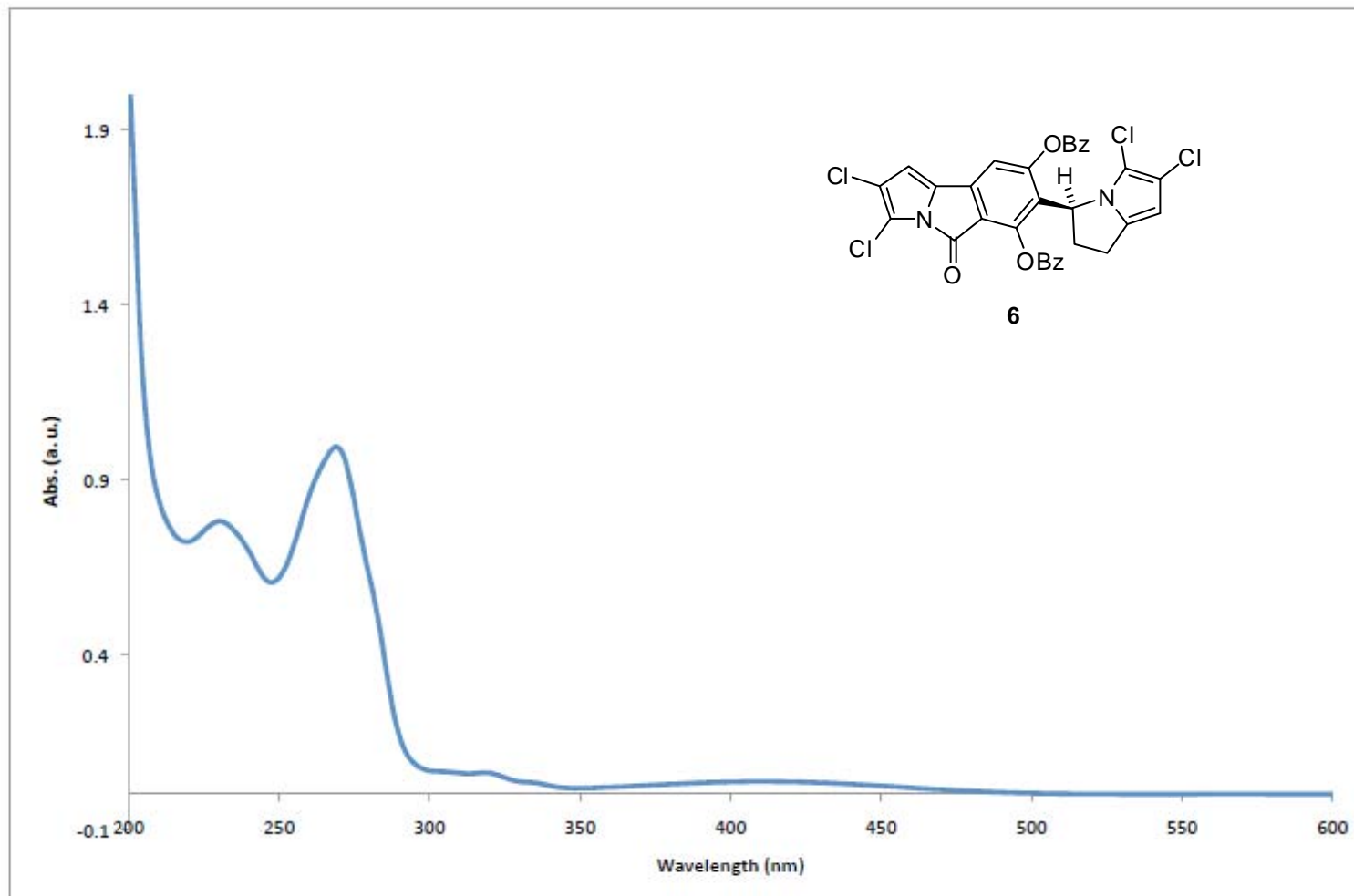


Pivaloyl-a #168-175 RT: 2.91-3.04 AV: 8 NL: 9.66E4
T: FTMS + p ESI Full ms [100.00-2000.00]

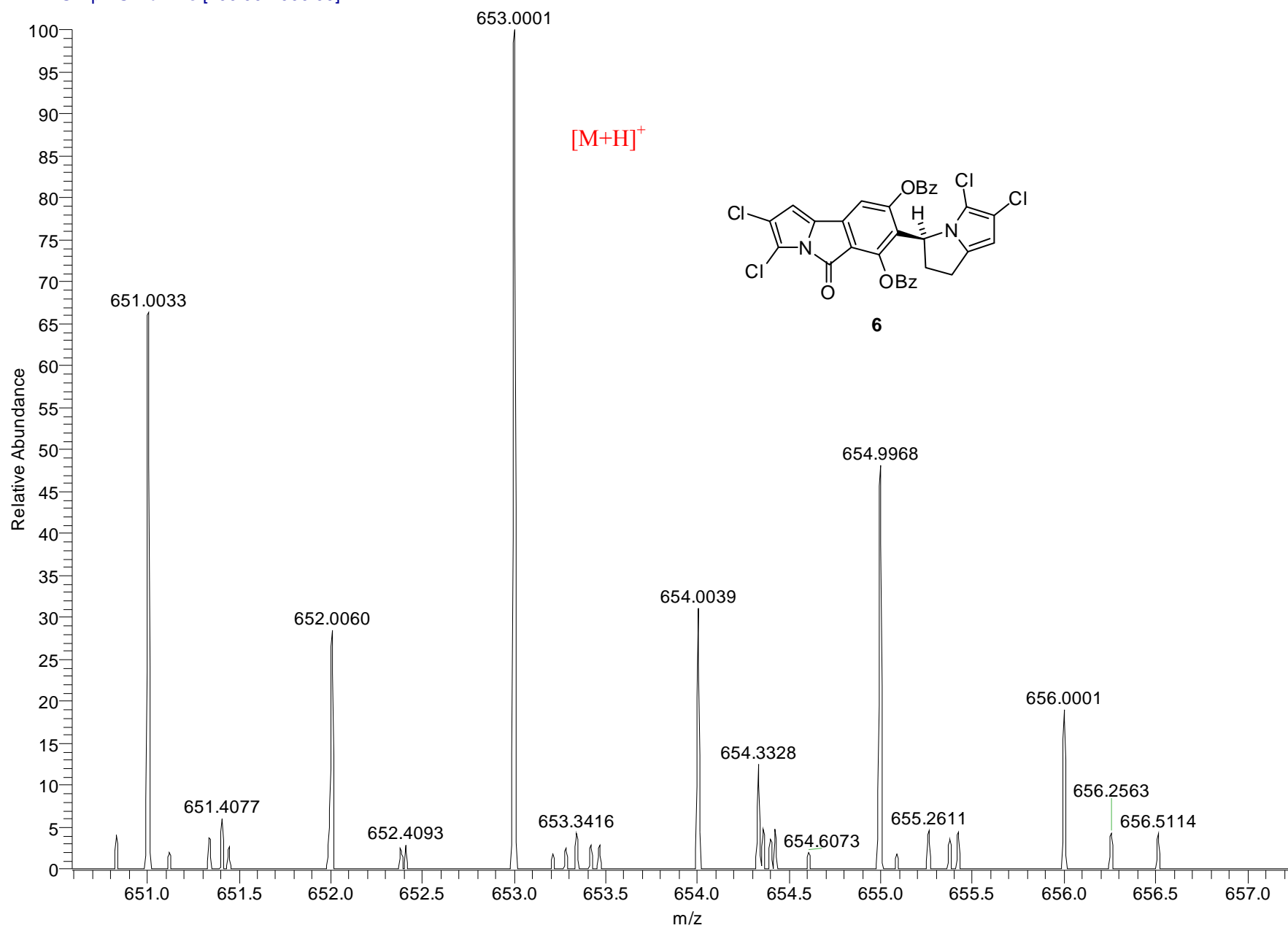


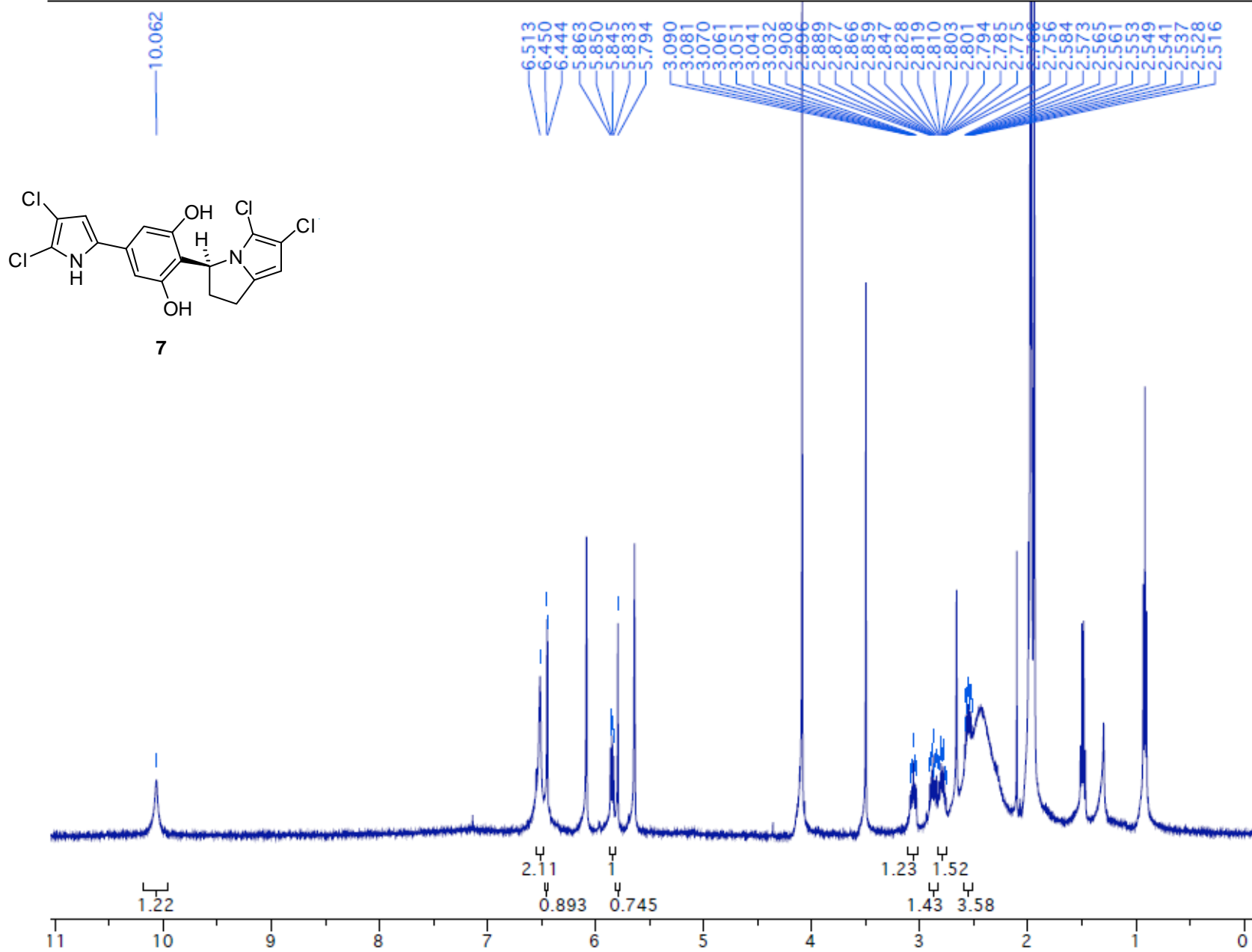






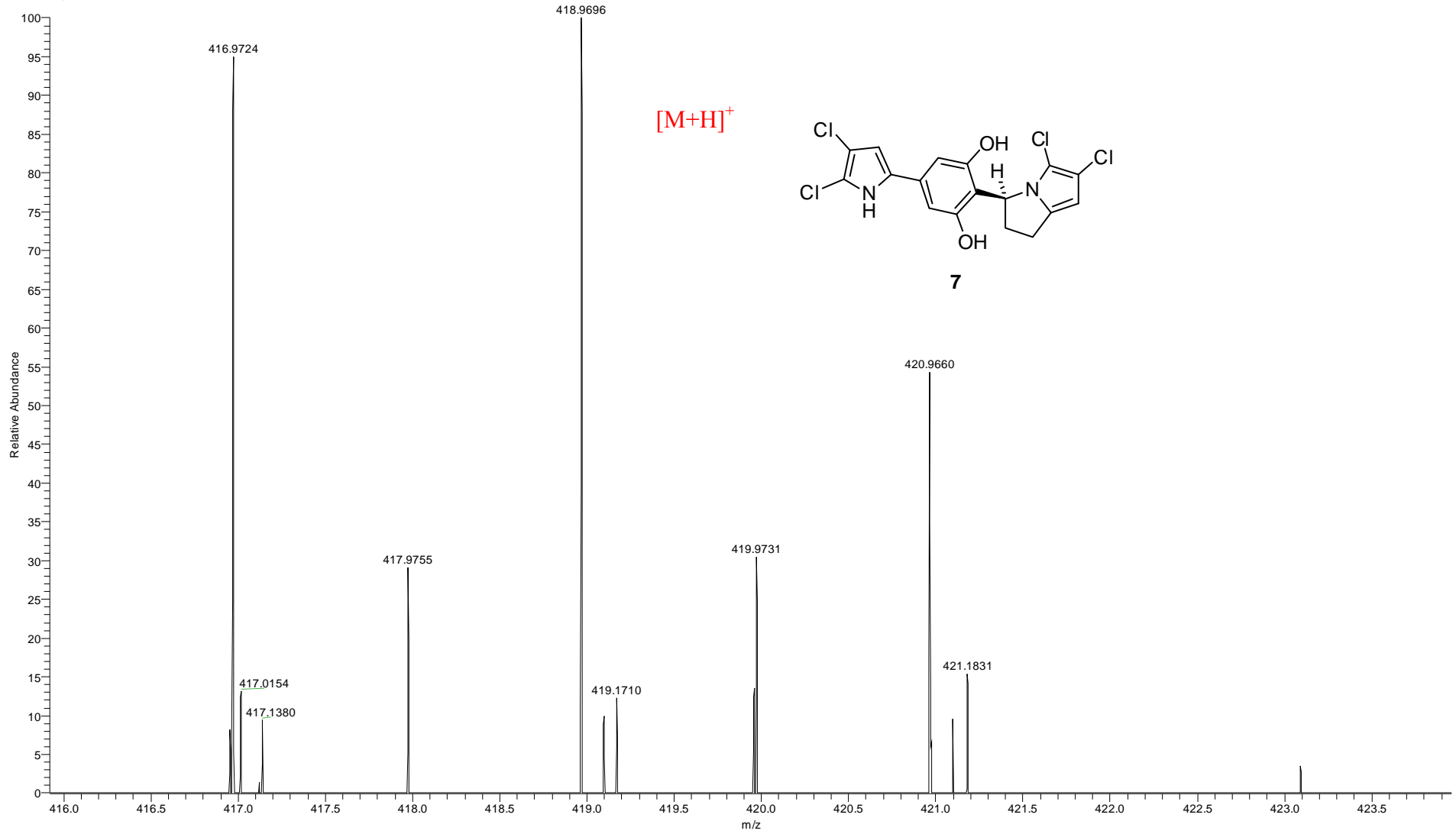
650-b #33-35 RT: 0.53-0.56 AV: 3 NL: 3.68E4
T: FTMS + p ESI Full ms [100.00-2000.00]

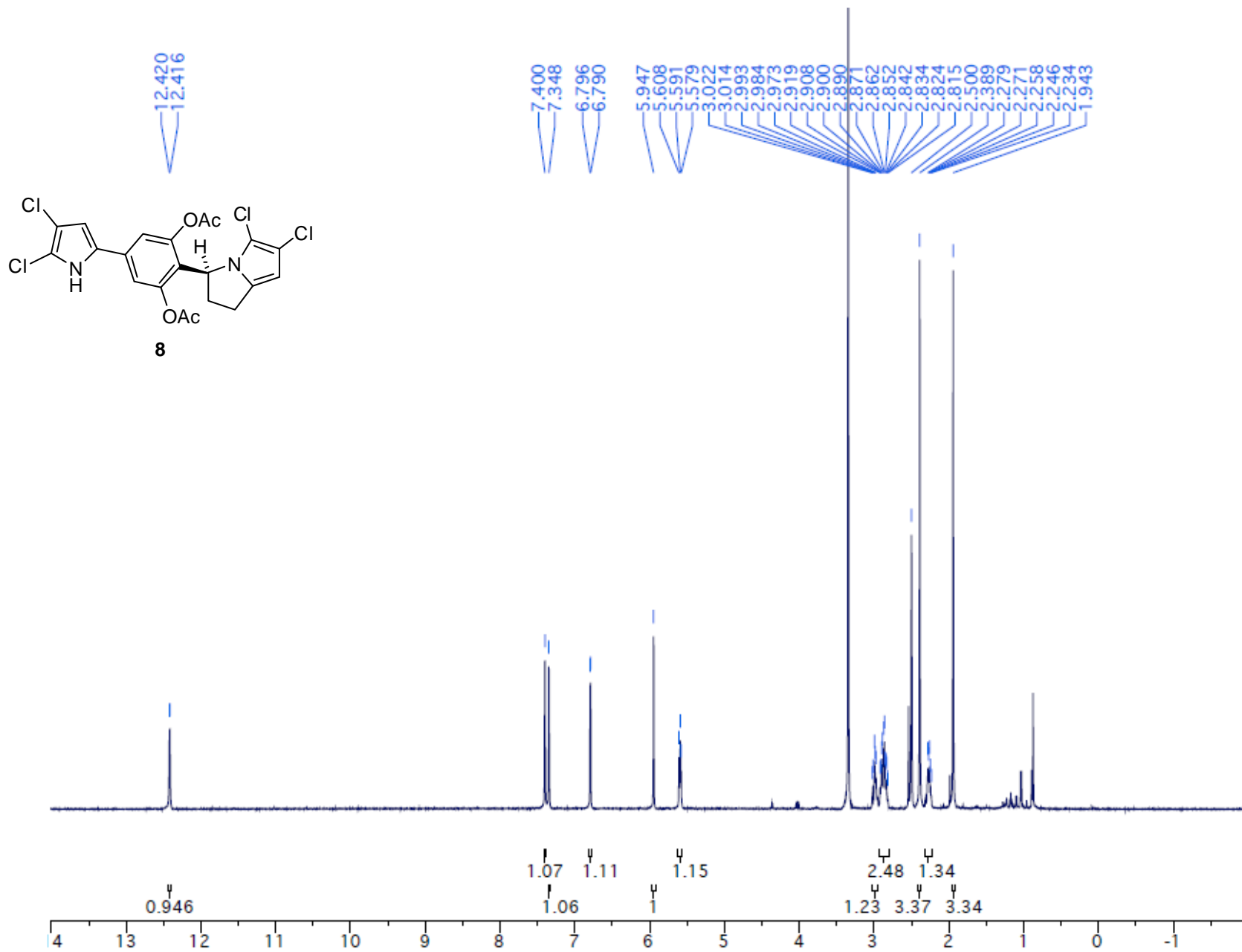


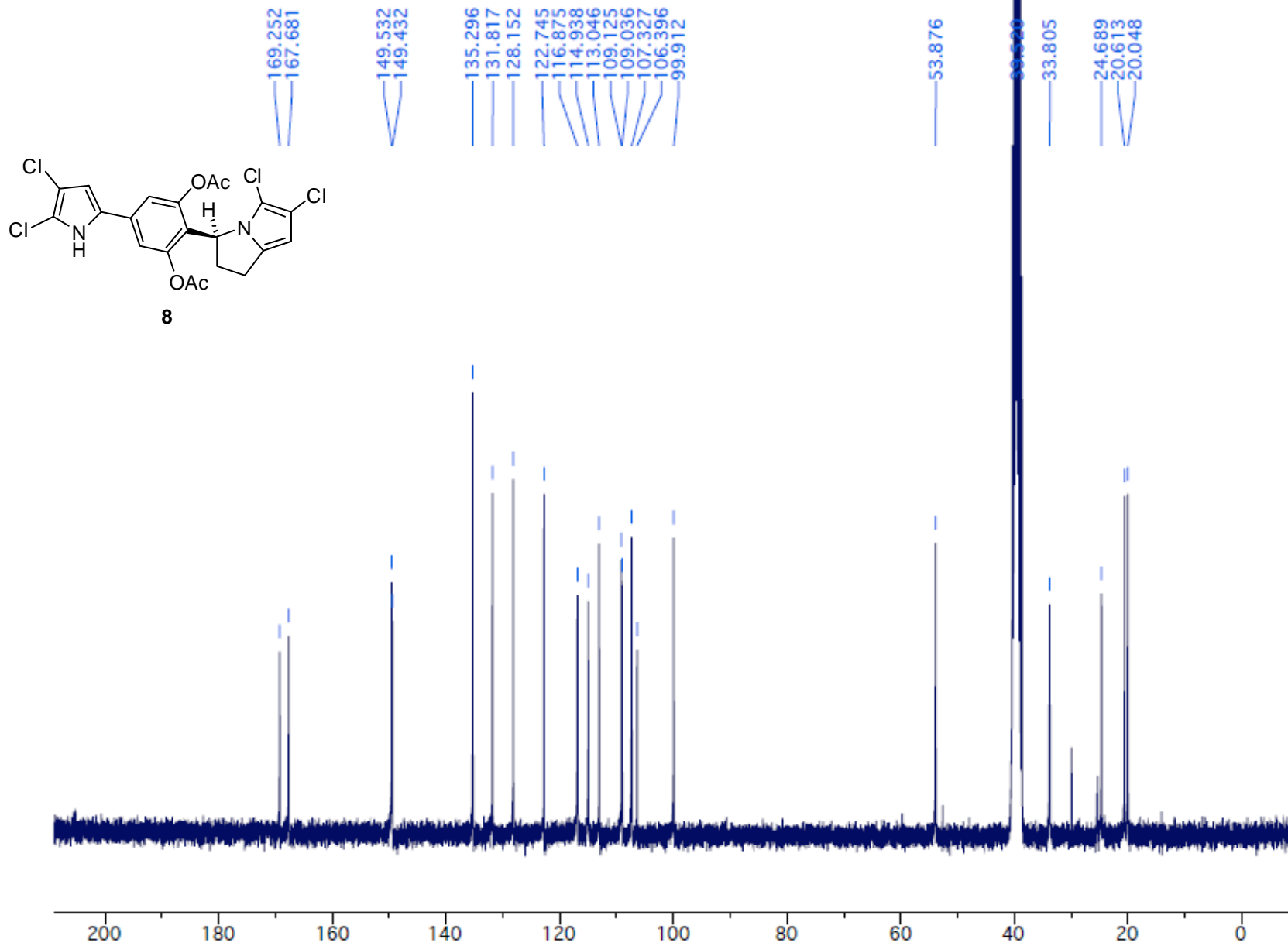


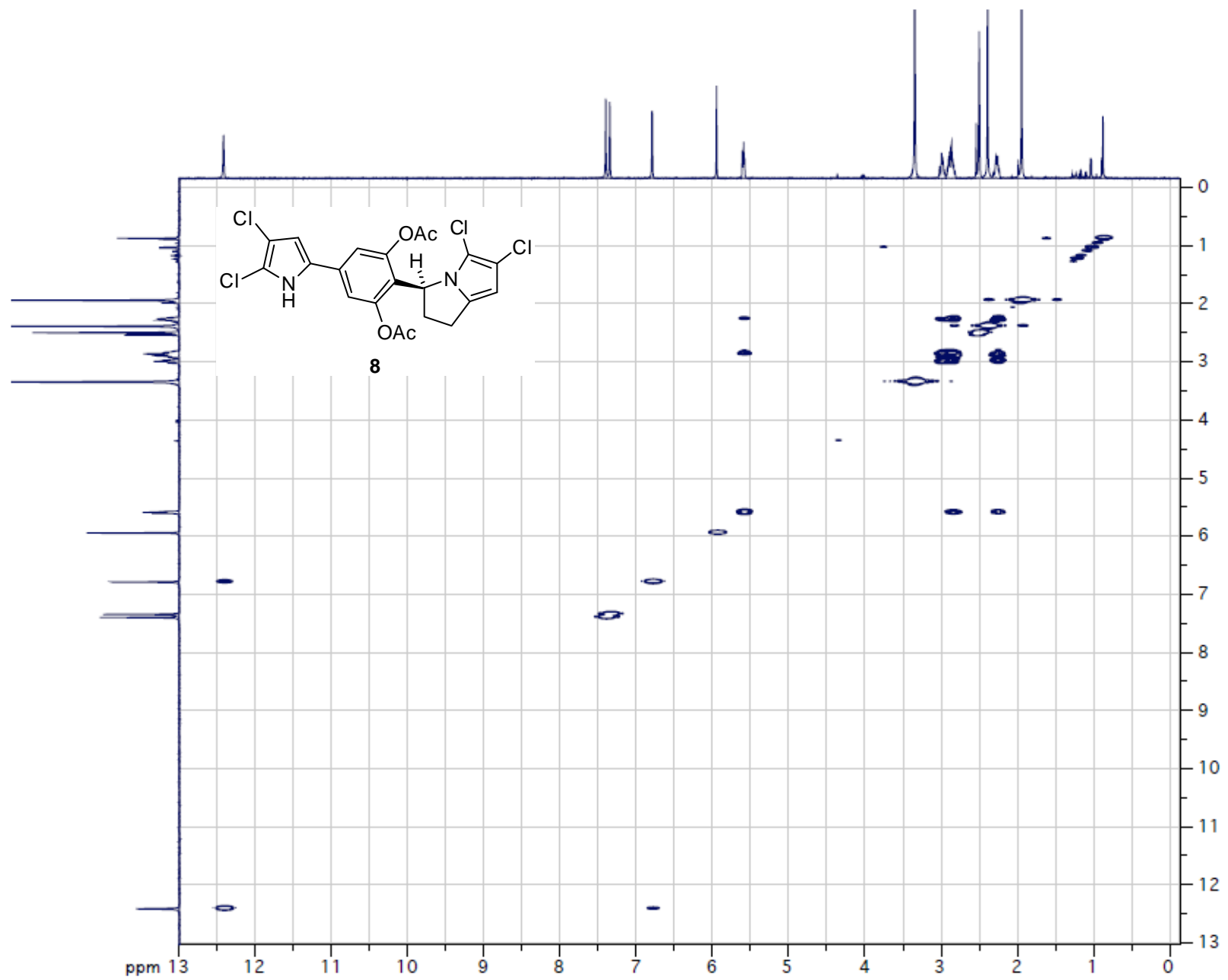
HRESI-FT-MS (Orbit-Trap-MS) of chlorizidine B (7)

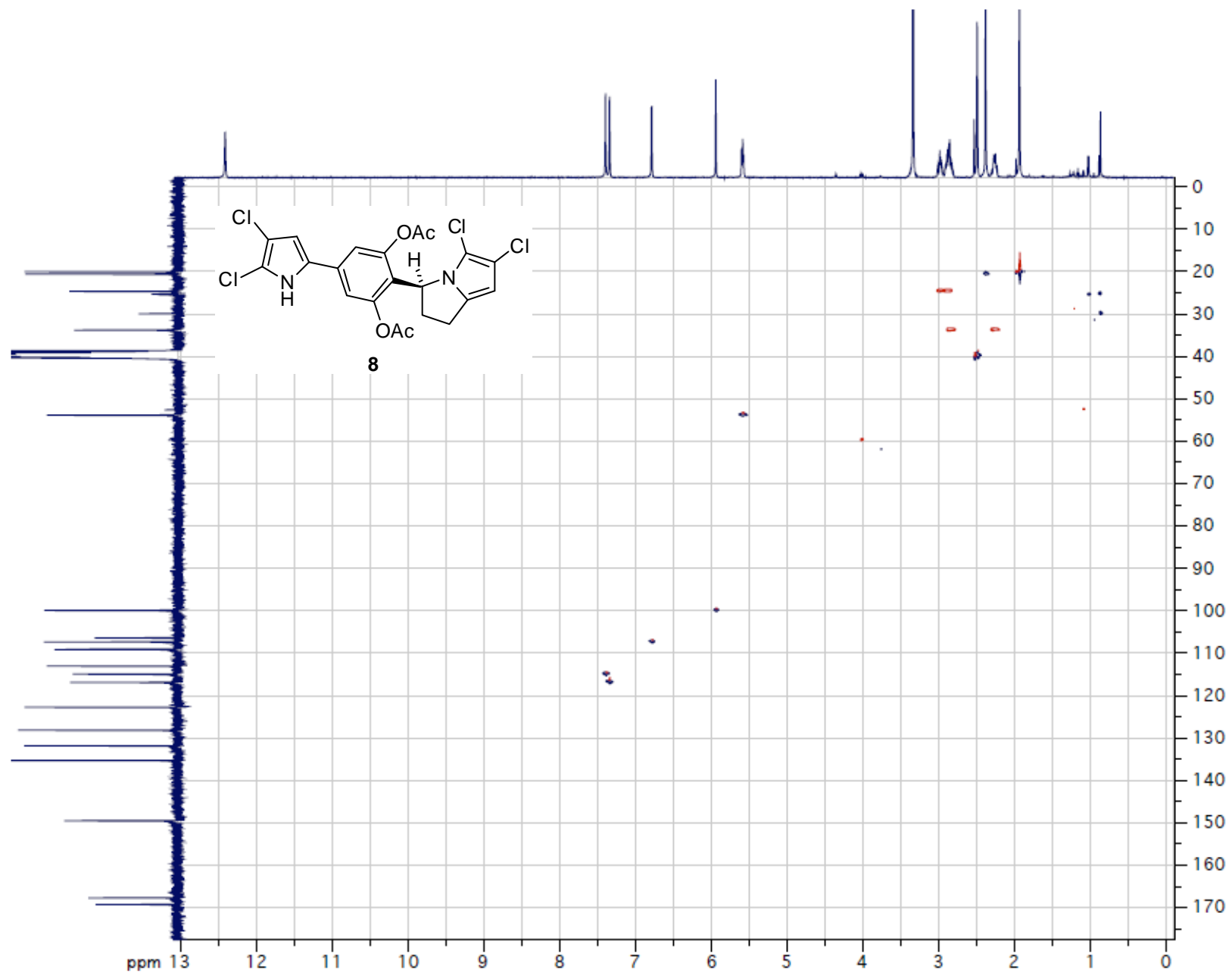
H287416 #316 RT: 5.61 AV: 1 SB: 4 3.37-3.41 NL: 4.08E4
T: FTMS + p ESI Full ms [100.00-2000.00]

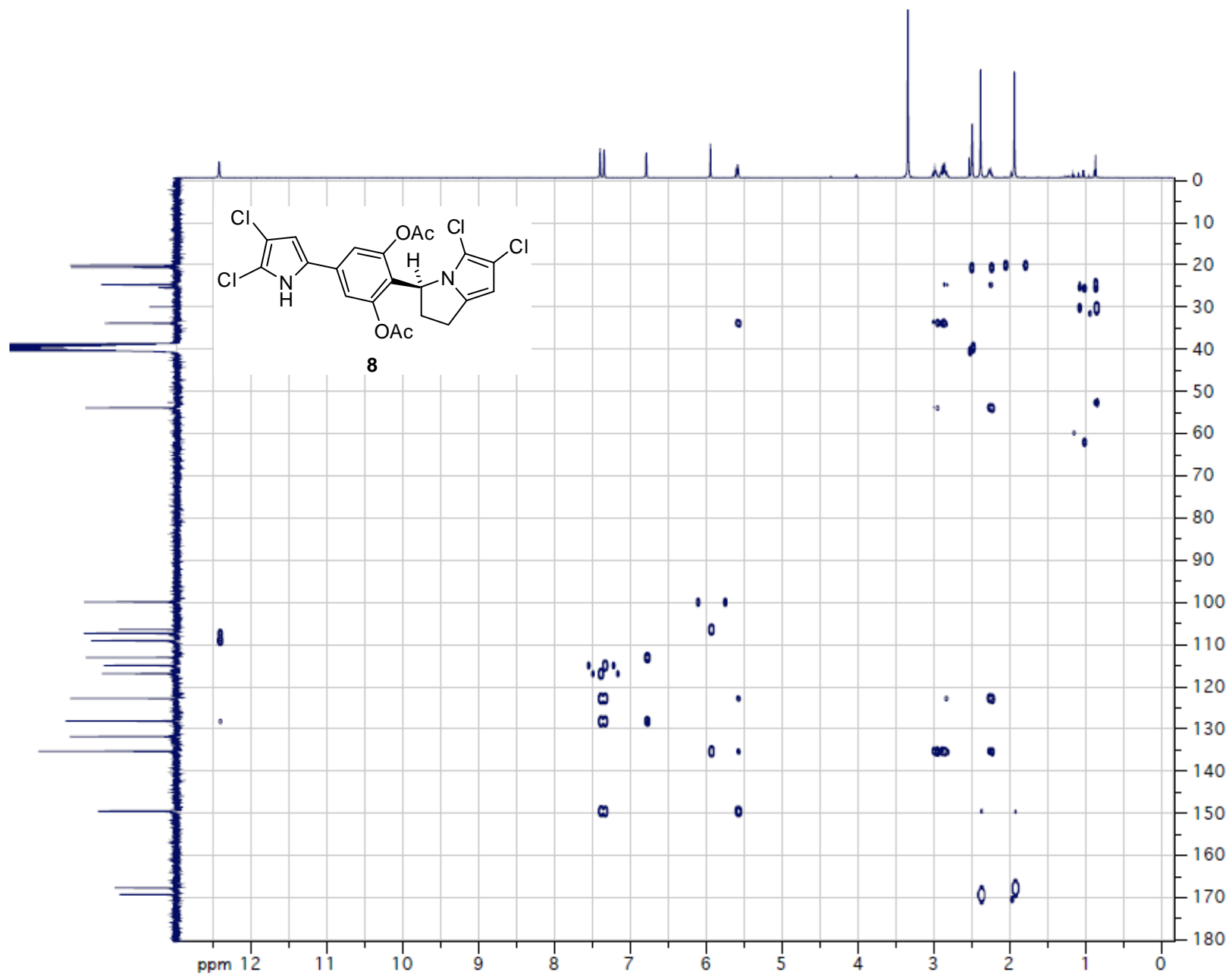


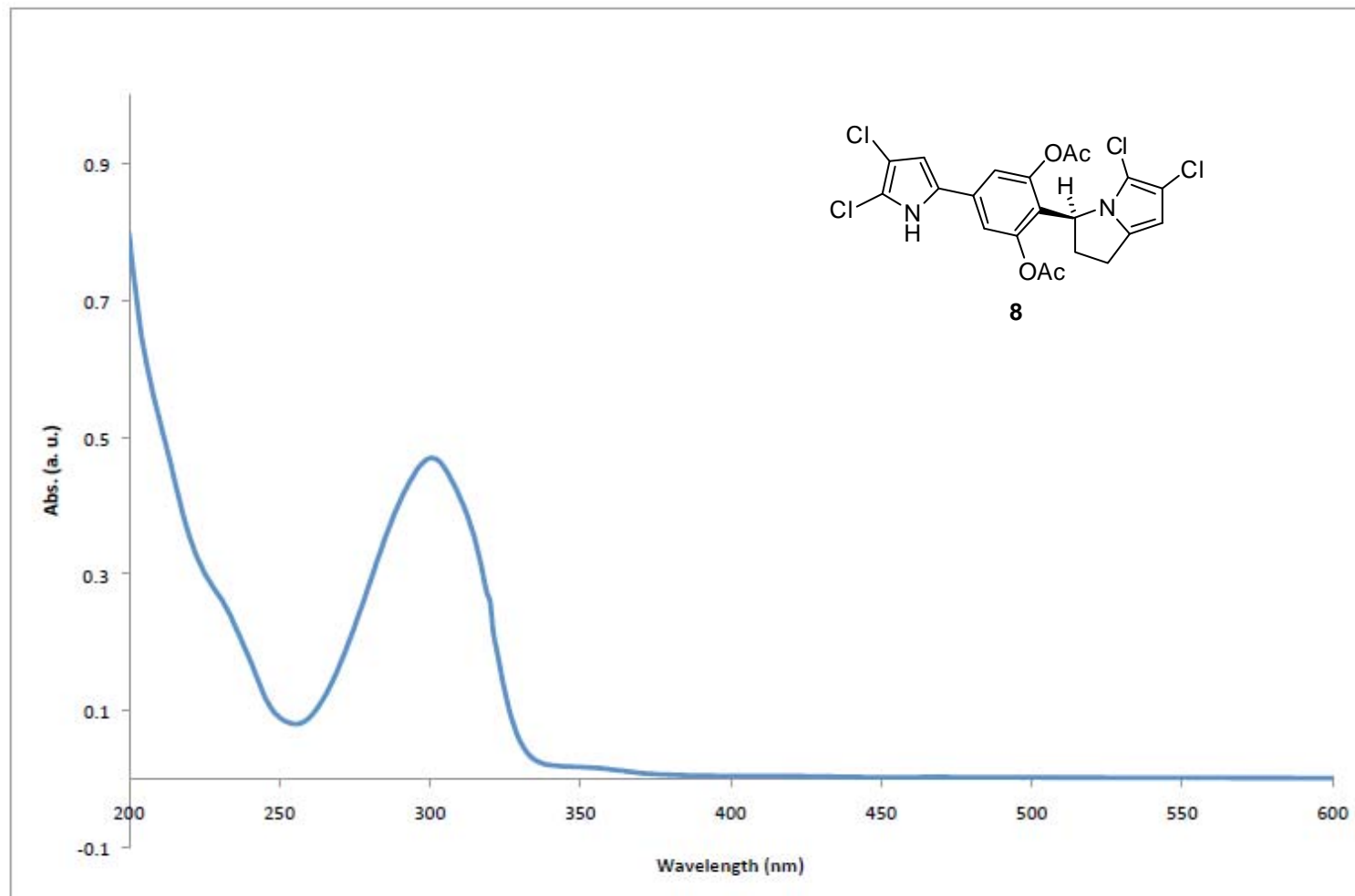




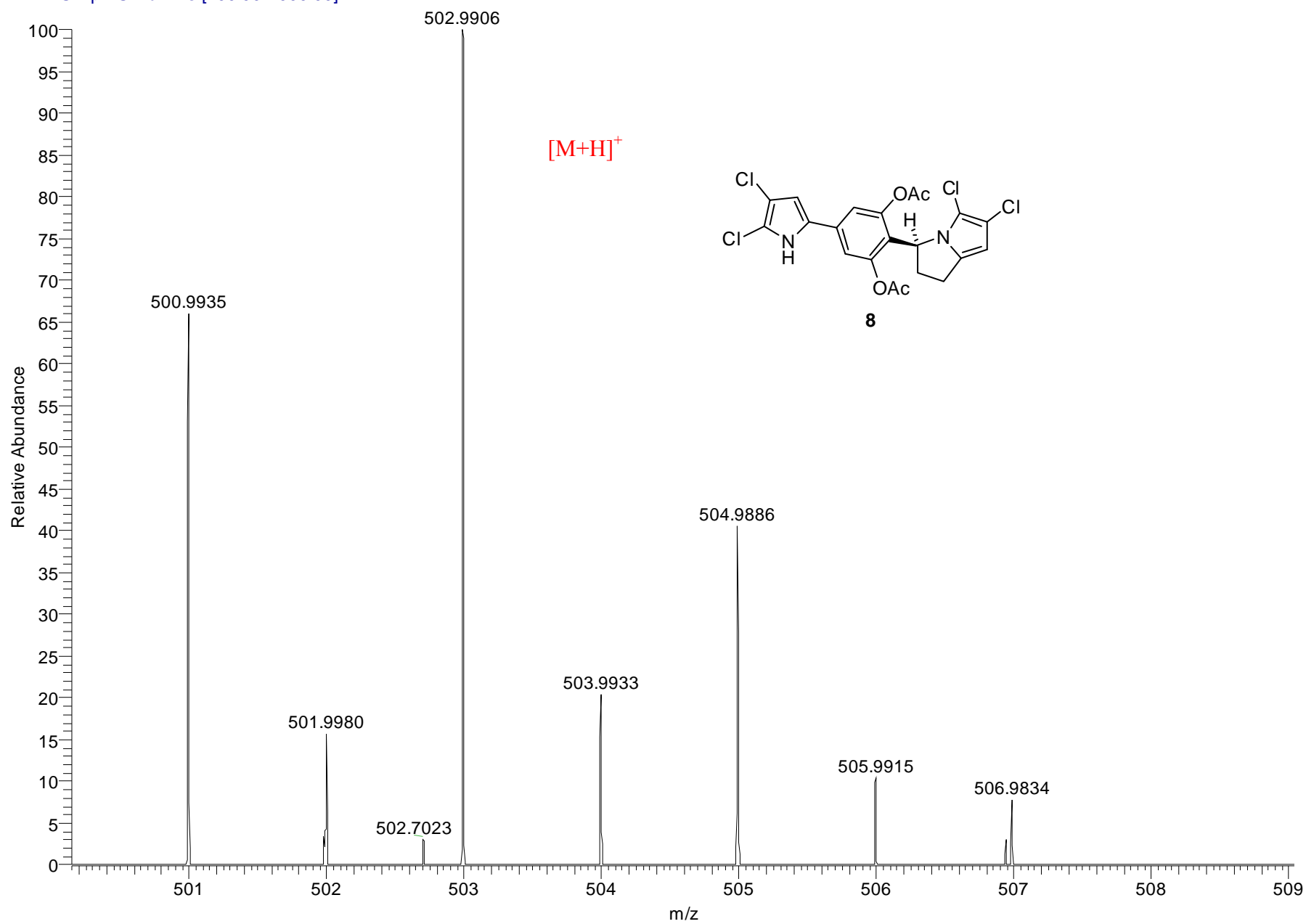




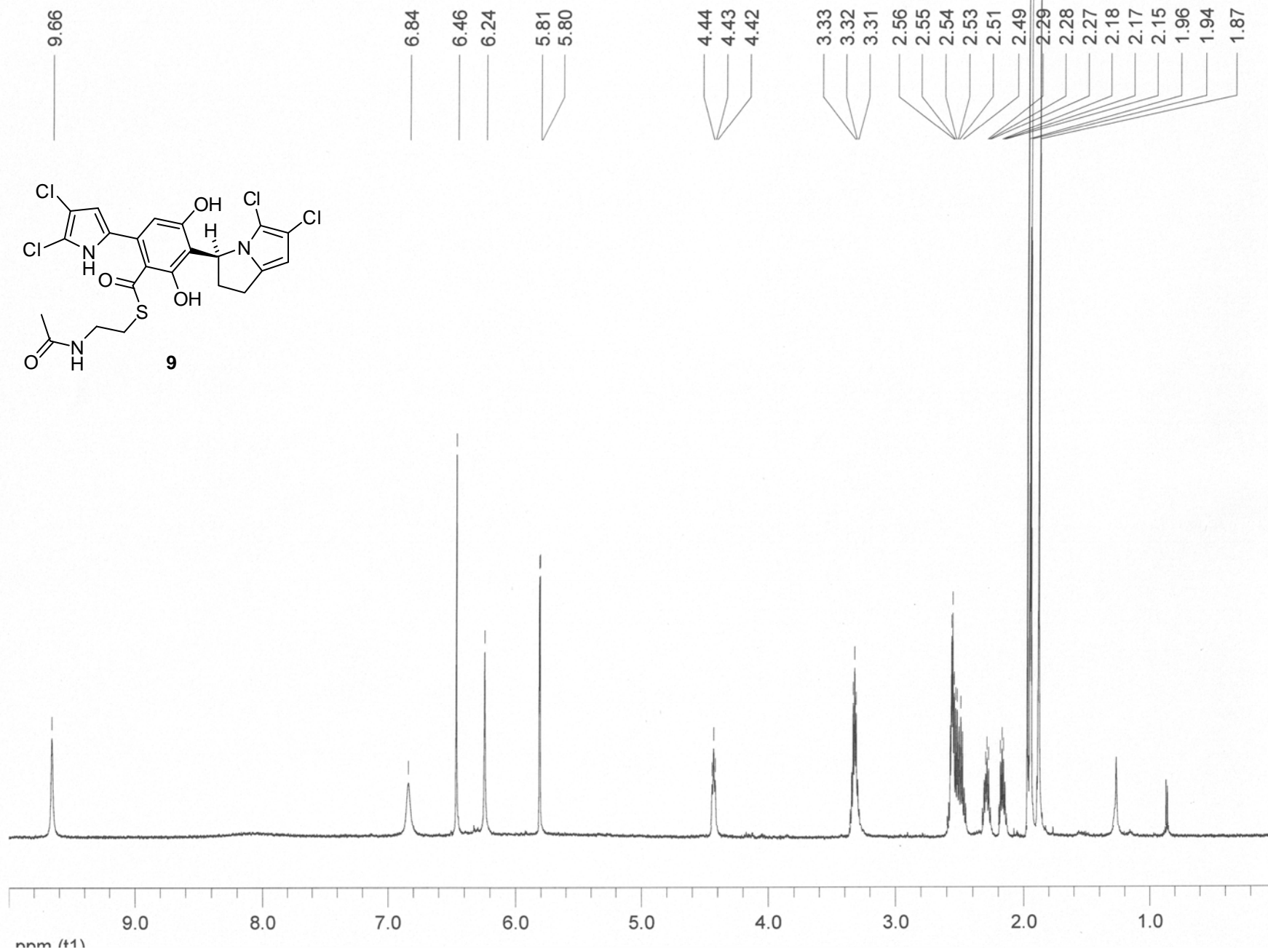




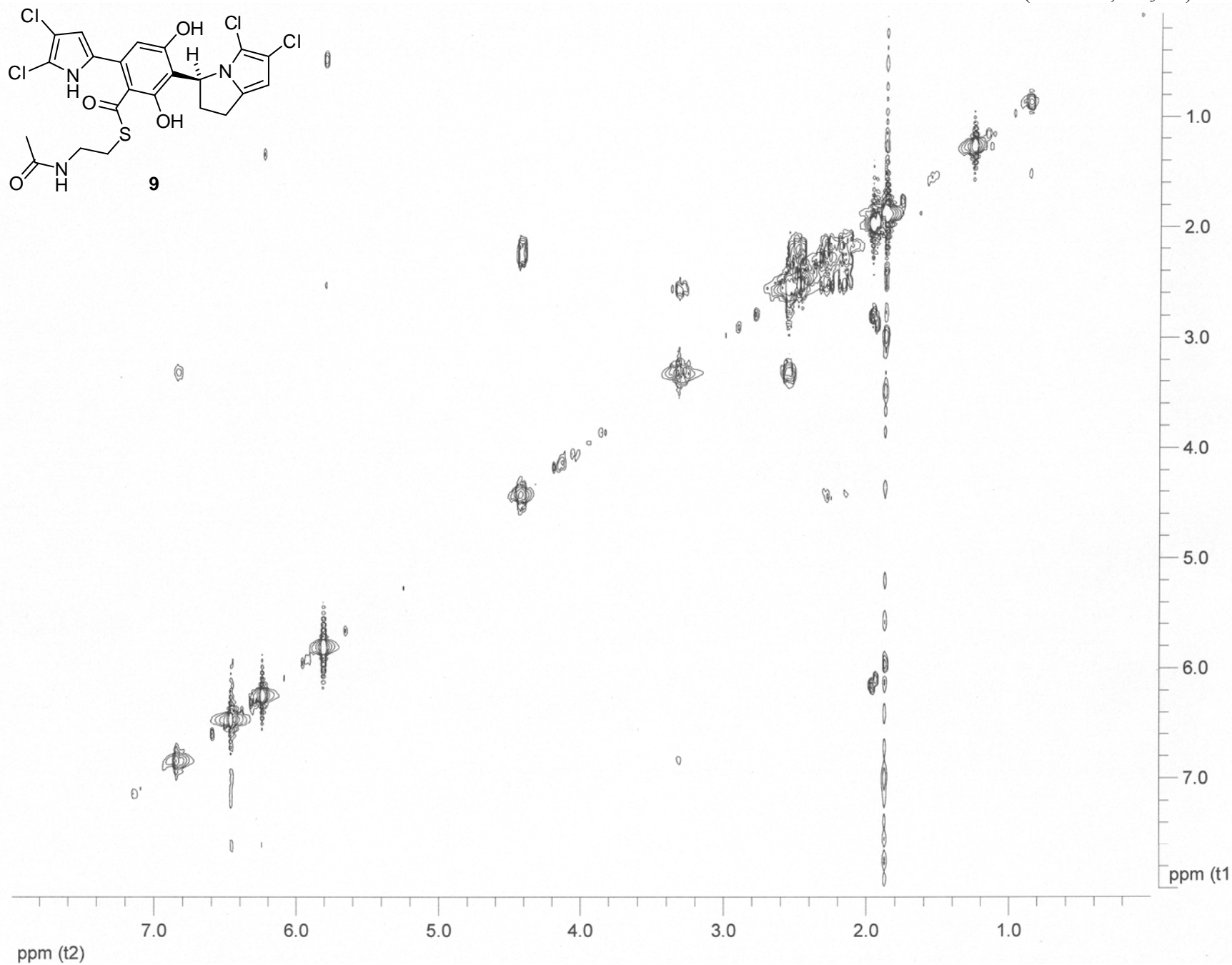
500 #98 RT: 1.57 AV: 1 NL: 3.62E5
T: FTMS + p ESI Full ms [100.00-2000.00]



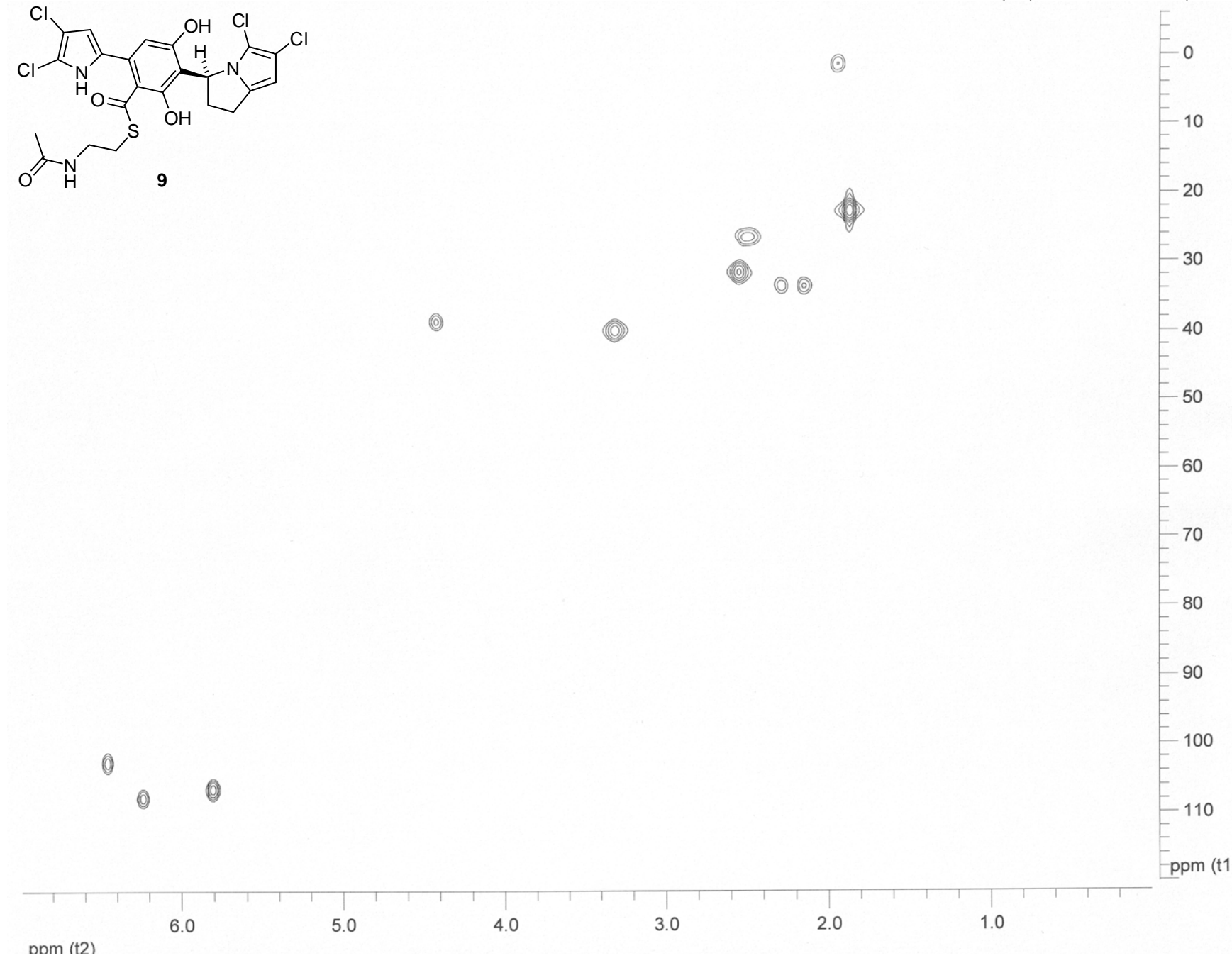
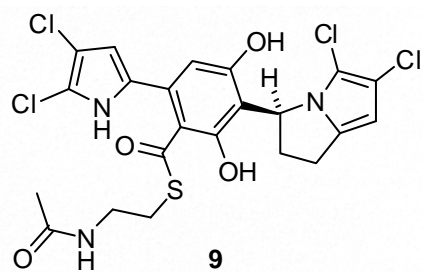
¹H NMR (600 MHz, CD₃CN) of thioester **9**



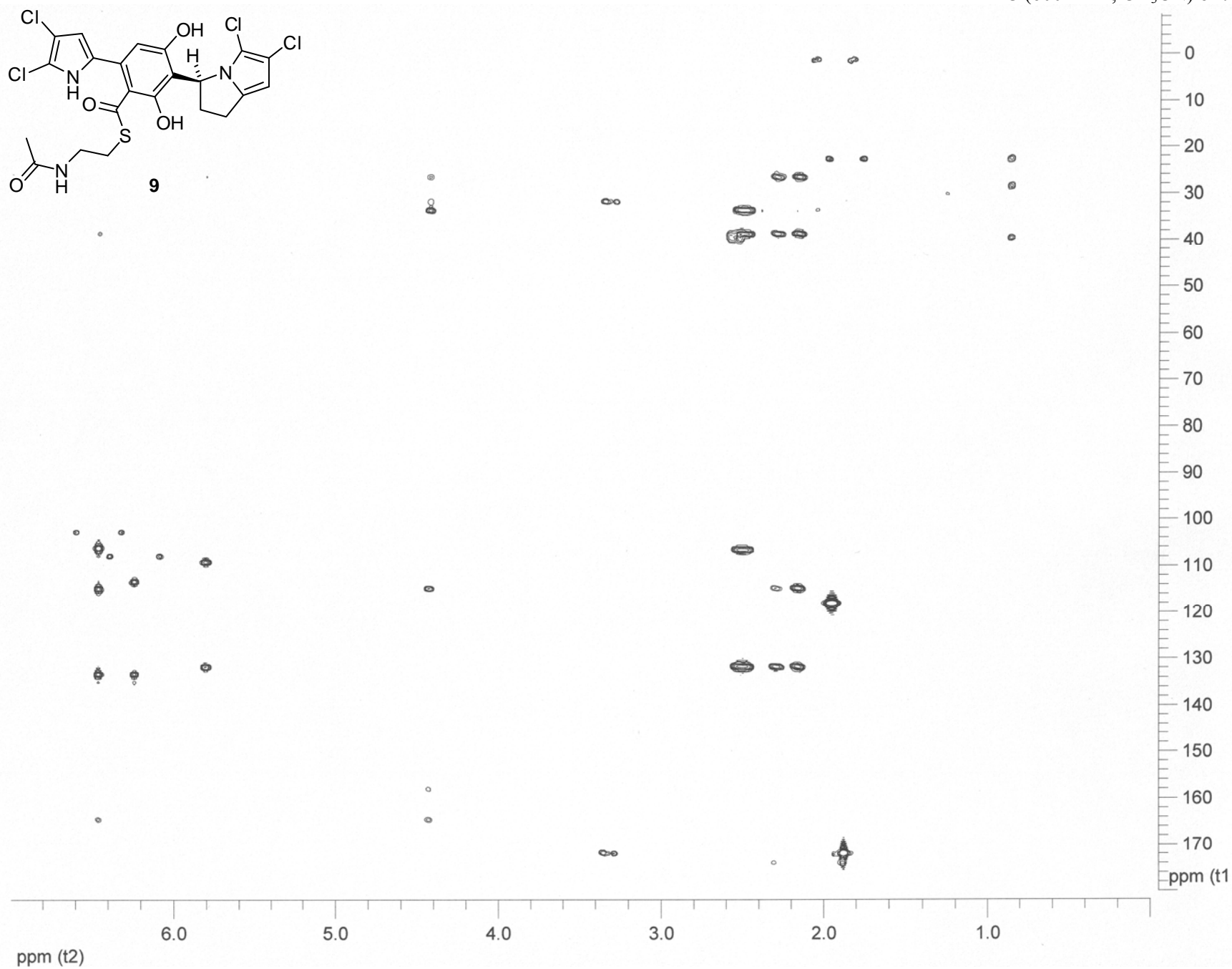
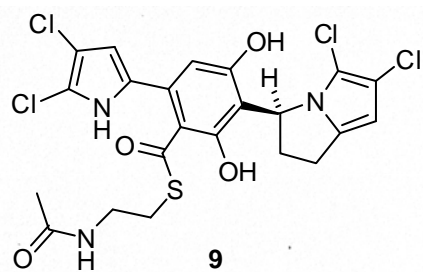
COSY (600 MHz, CD₃CN) of thioester **9**



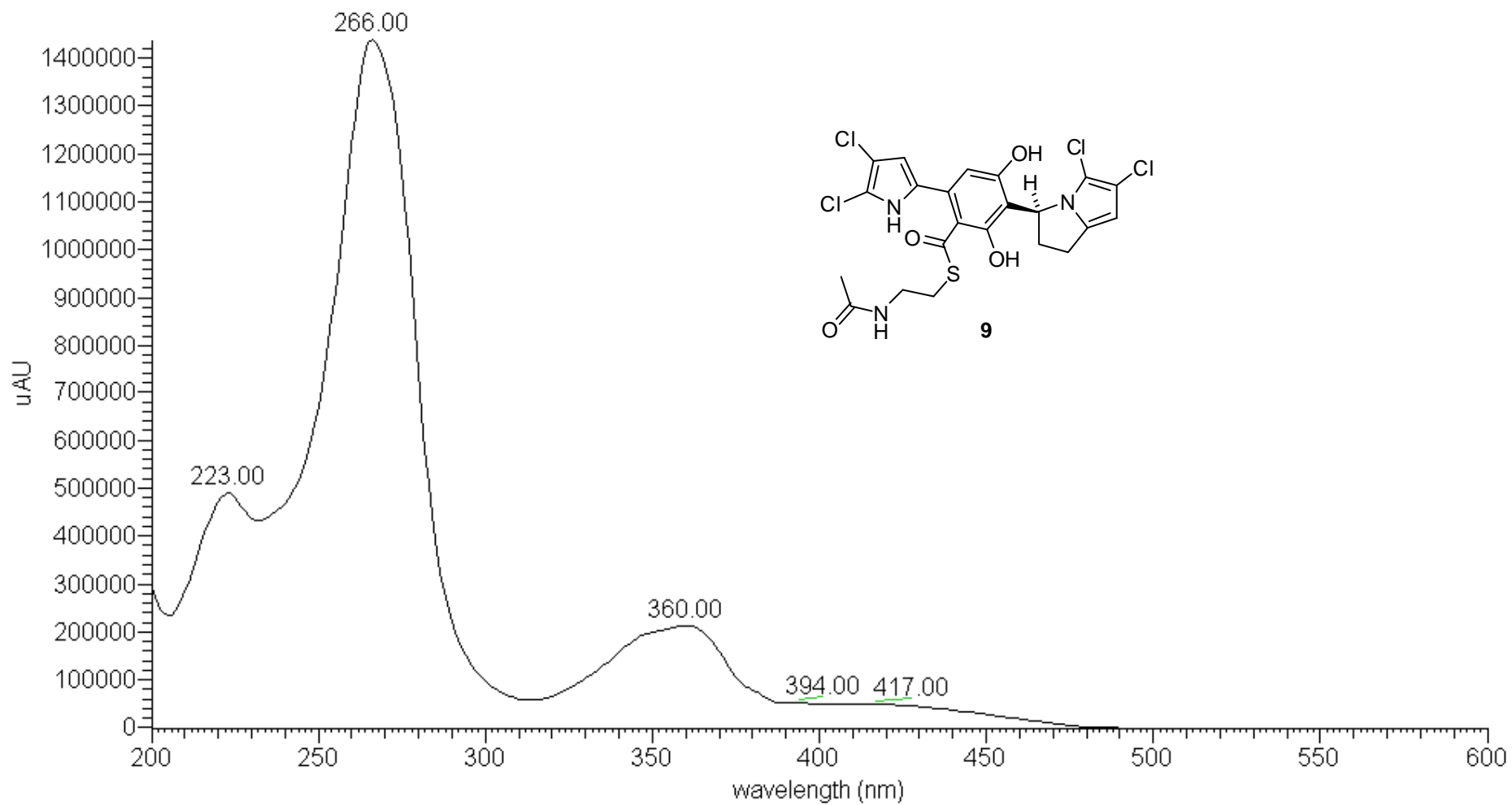
HSQC (600 MHz, CD₃CN) of thioester **9**



HMBC (600 MHz, CD₃CN) of thioester **9**

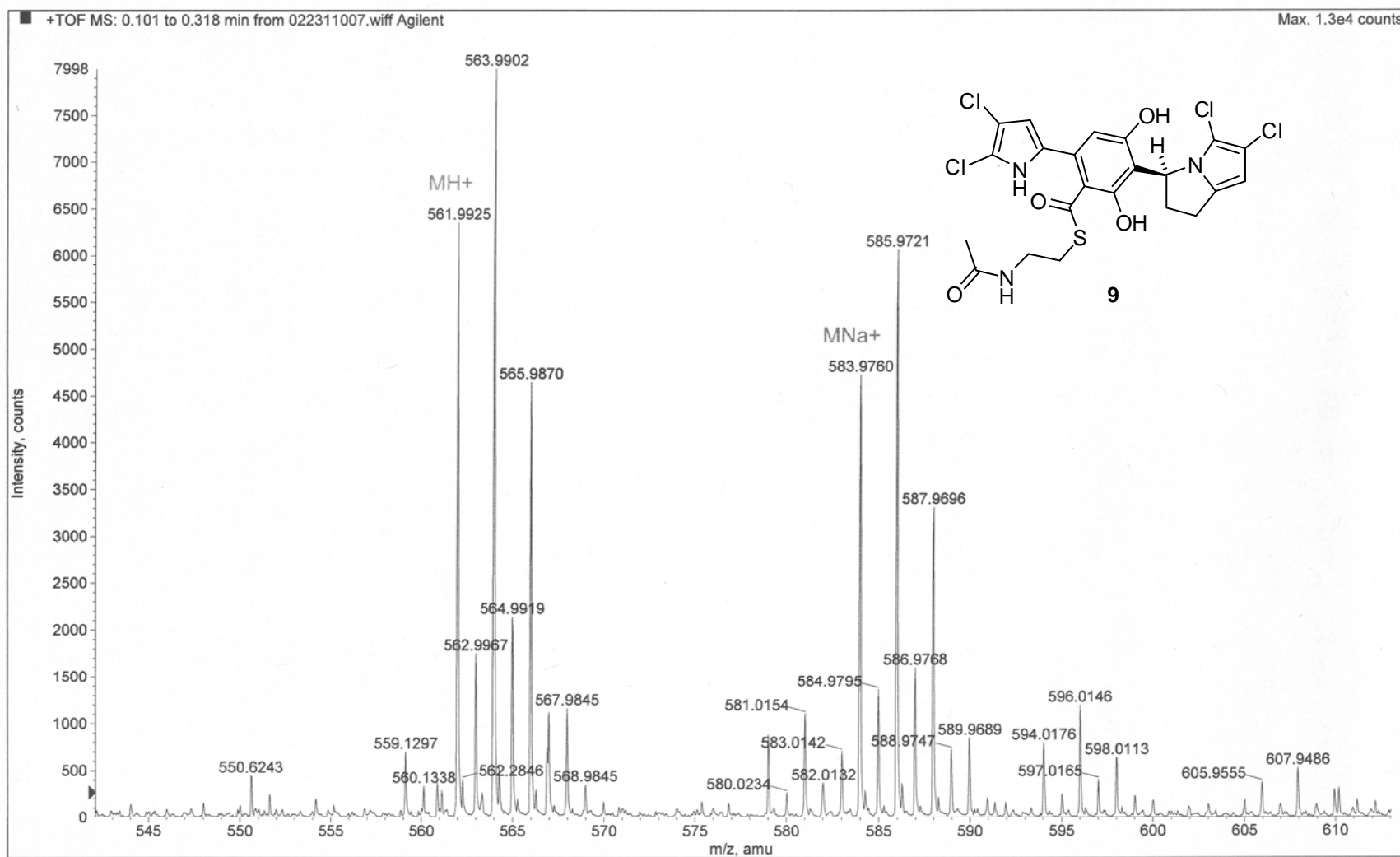


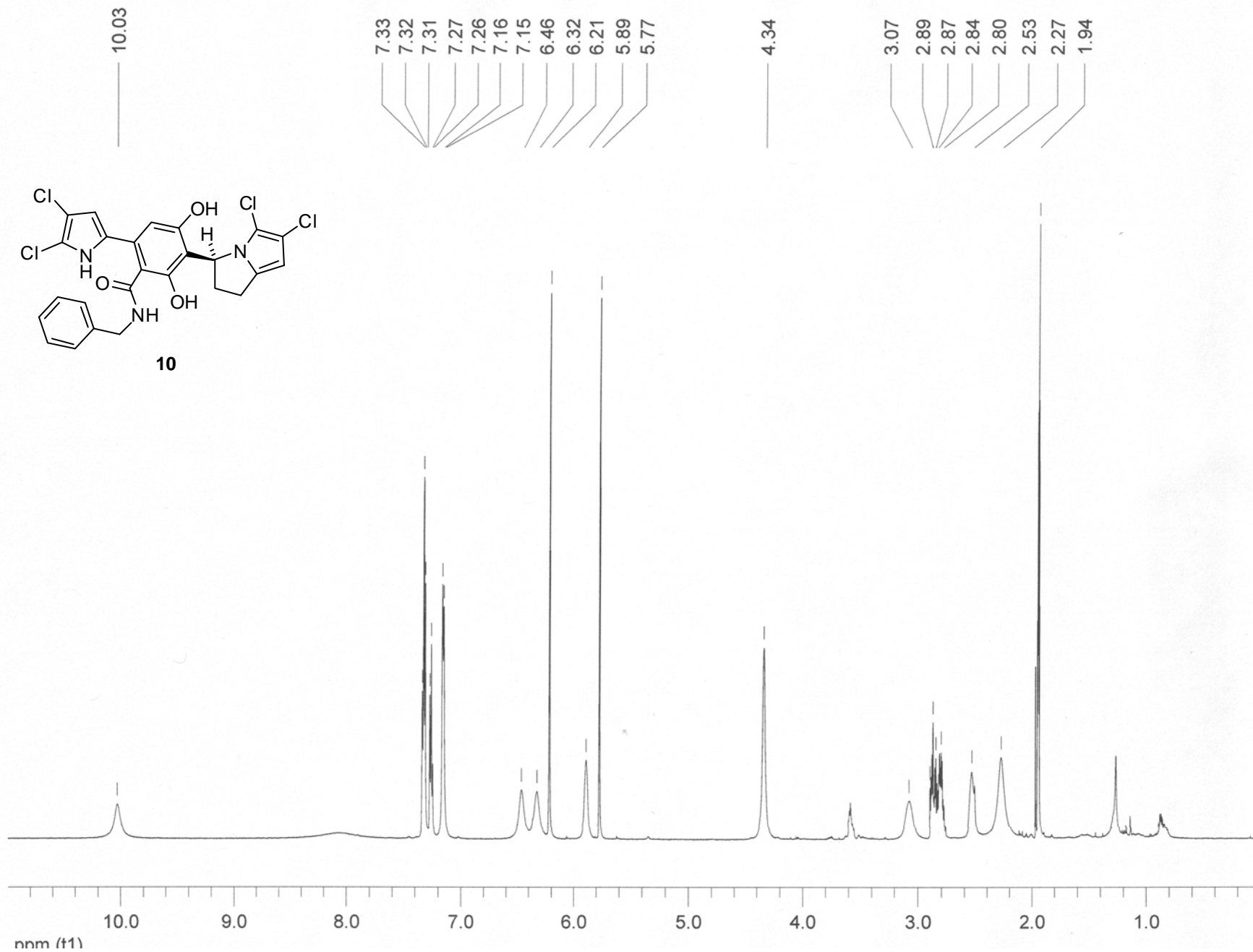
thiolsemip1 #4764 RT: 15.88 AV: 1 NL: 1.44E6 microAU

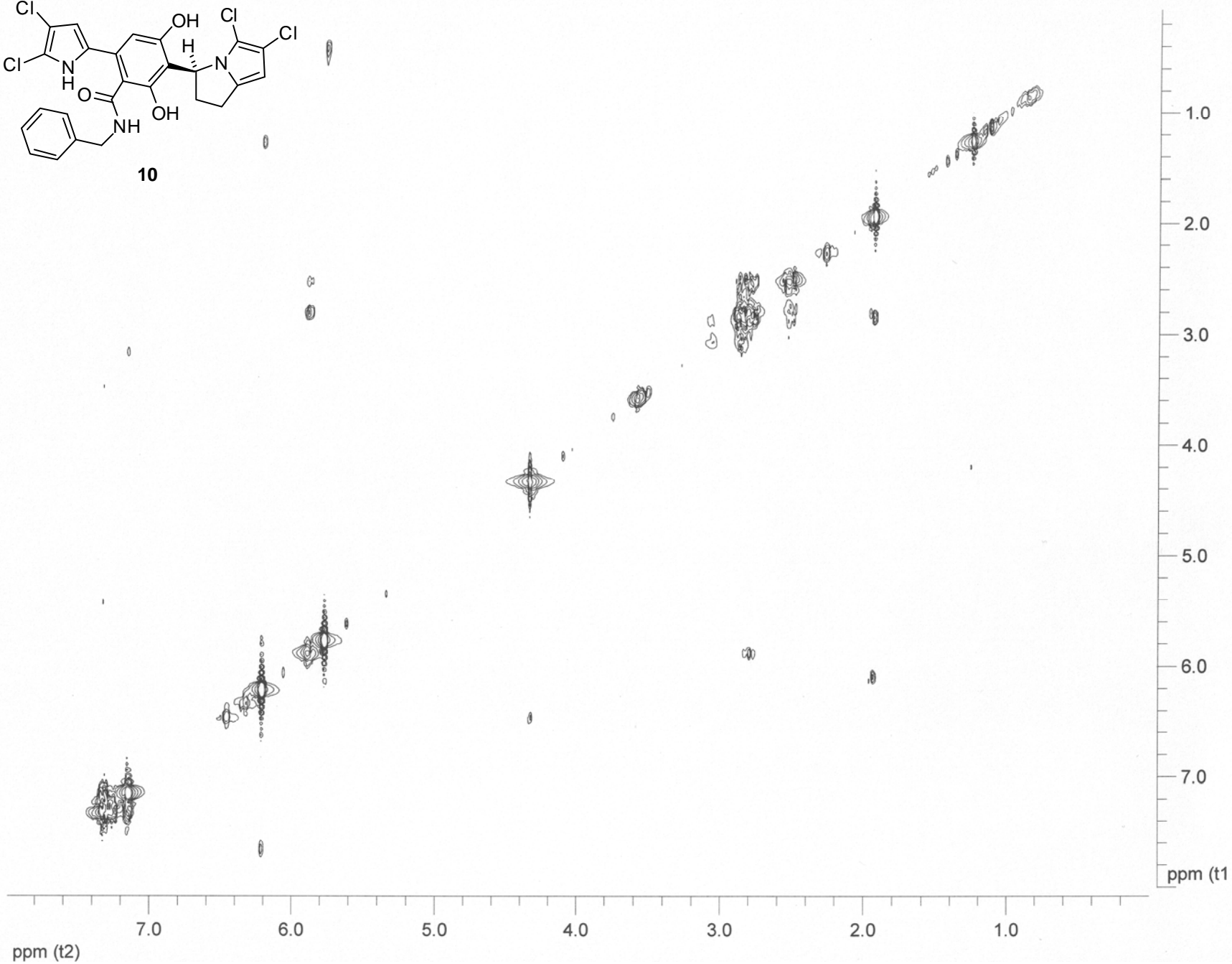
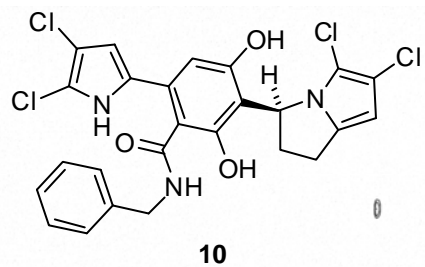


Polarity/Scan Type: Positive

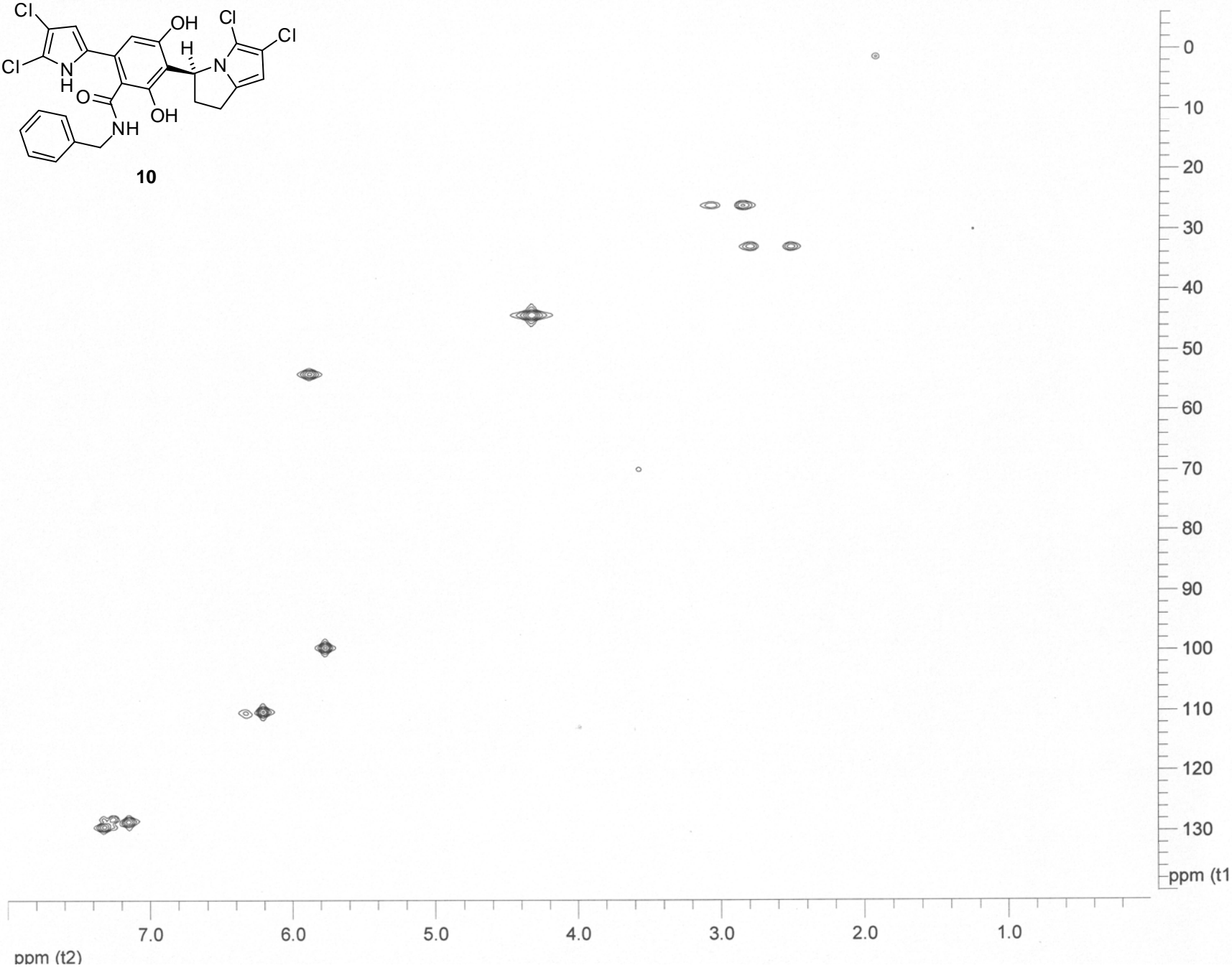
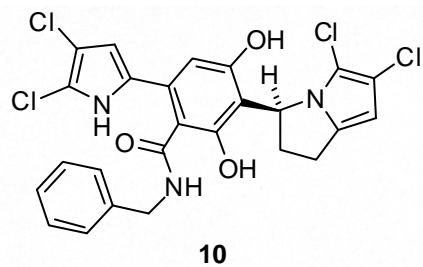
Sample Name: flch8376

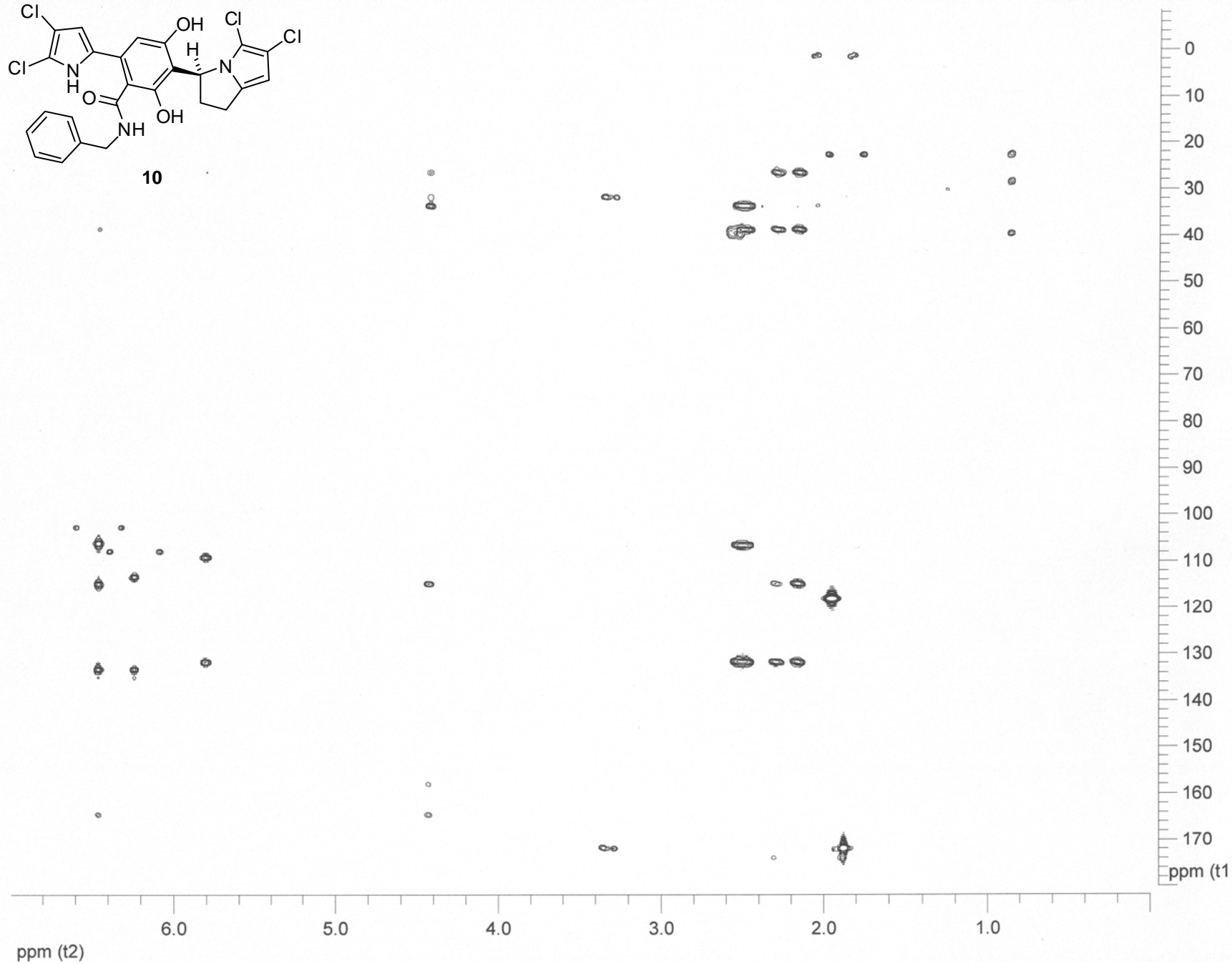
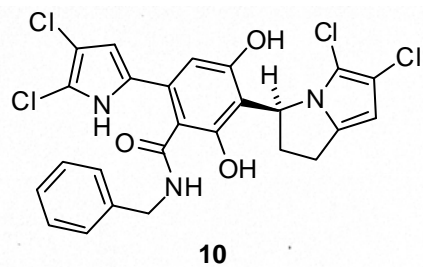
Acq. File: 022311007.wiff
Acq. Date: Wednesday, February 23



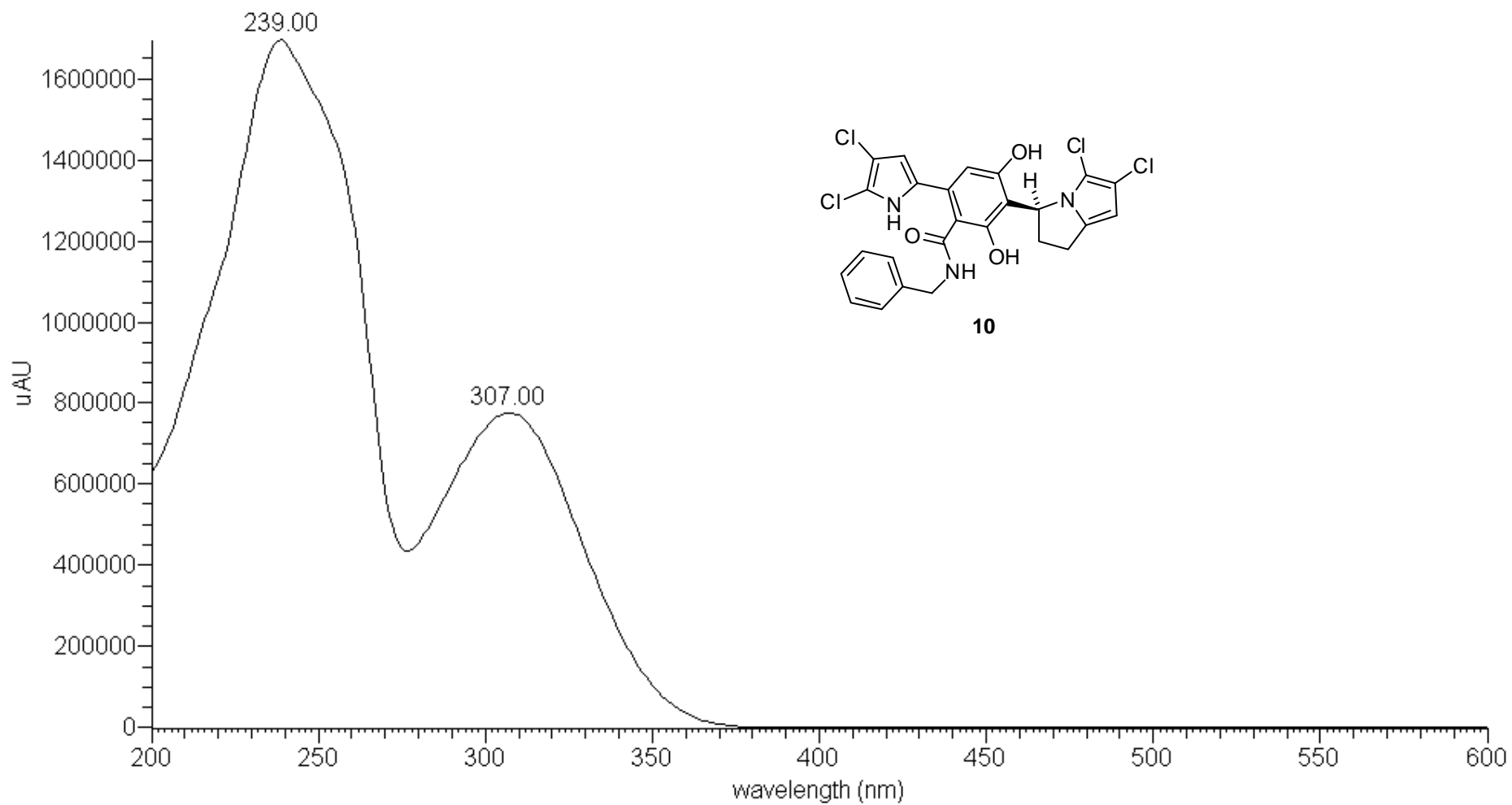


HSQC (600 MHz, CD₃CN) of amide **10**



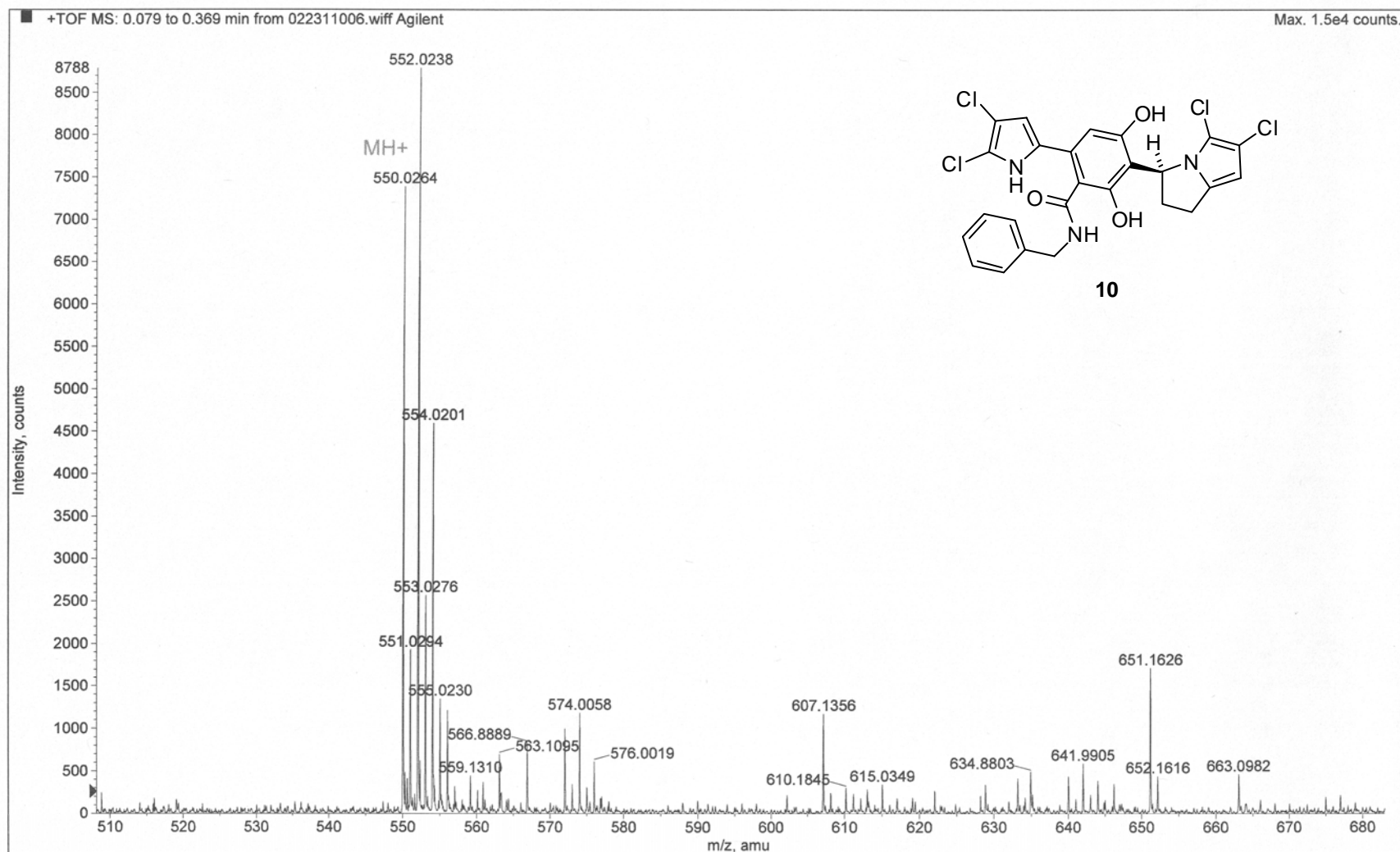


may3benzylaminebig1 #5605 RT: 18.68 AV: 1 NL: 1.69E6 microAU

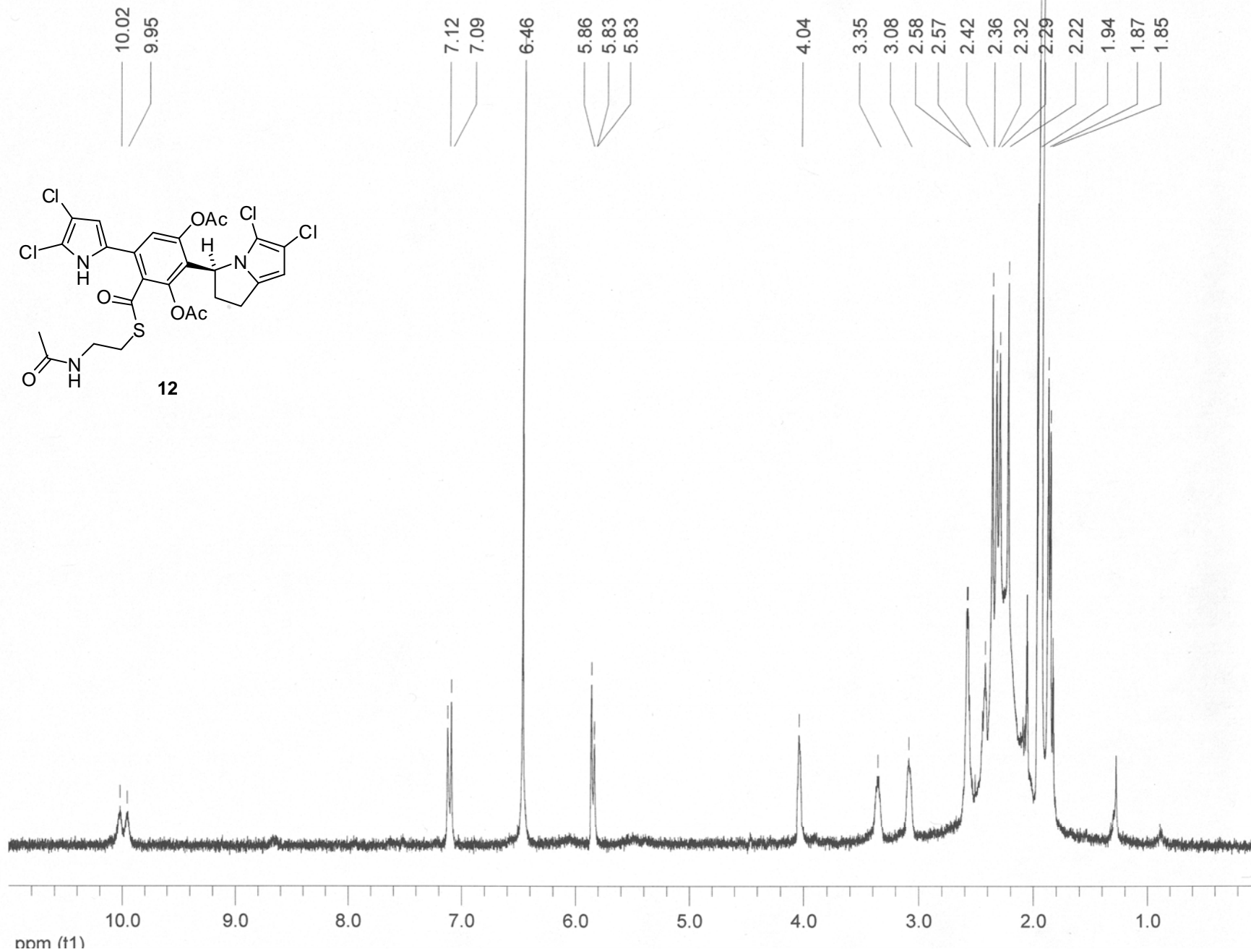


Polarity/Scan Type: Positive

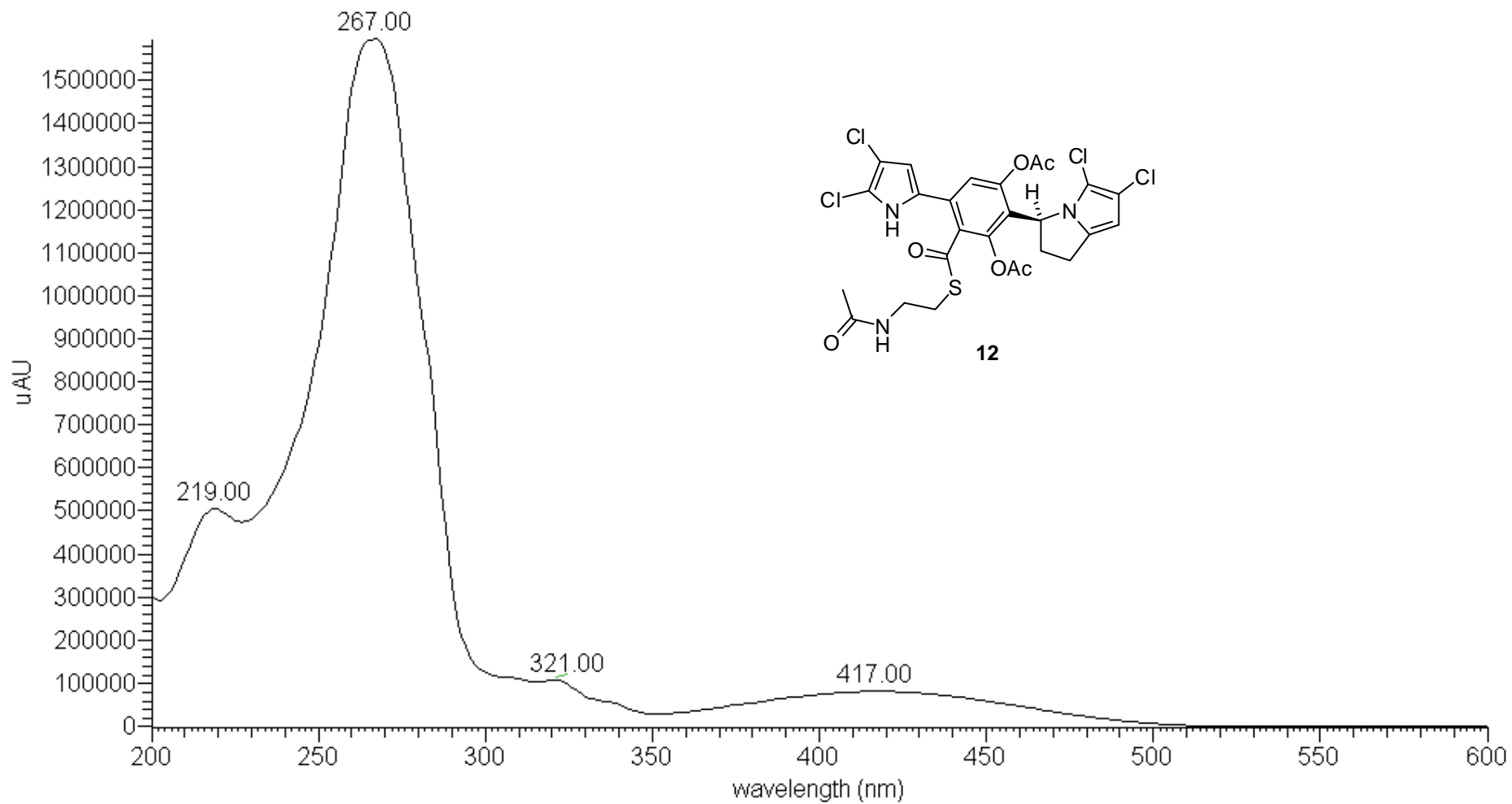
Sample Name: fch8375

Acq. File: 022311006.wiff
Acq. Date: Wednesday, February 23

¹H NMR (600 MHz, CD₃CN) of diacetylated thioester **12**

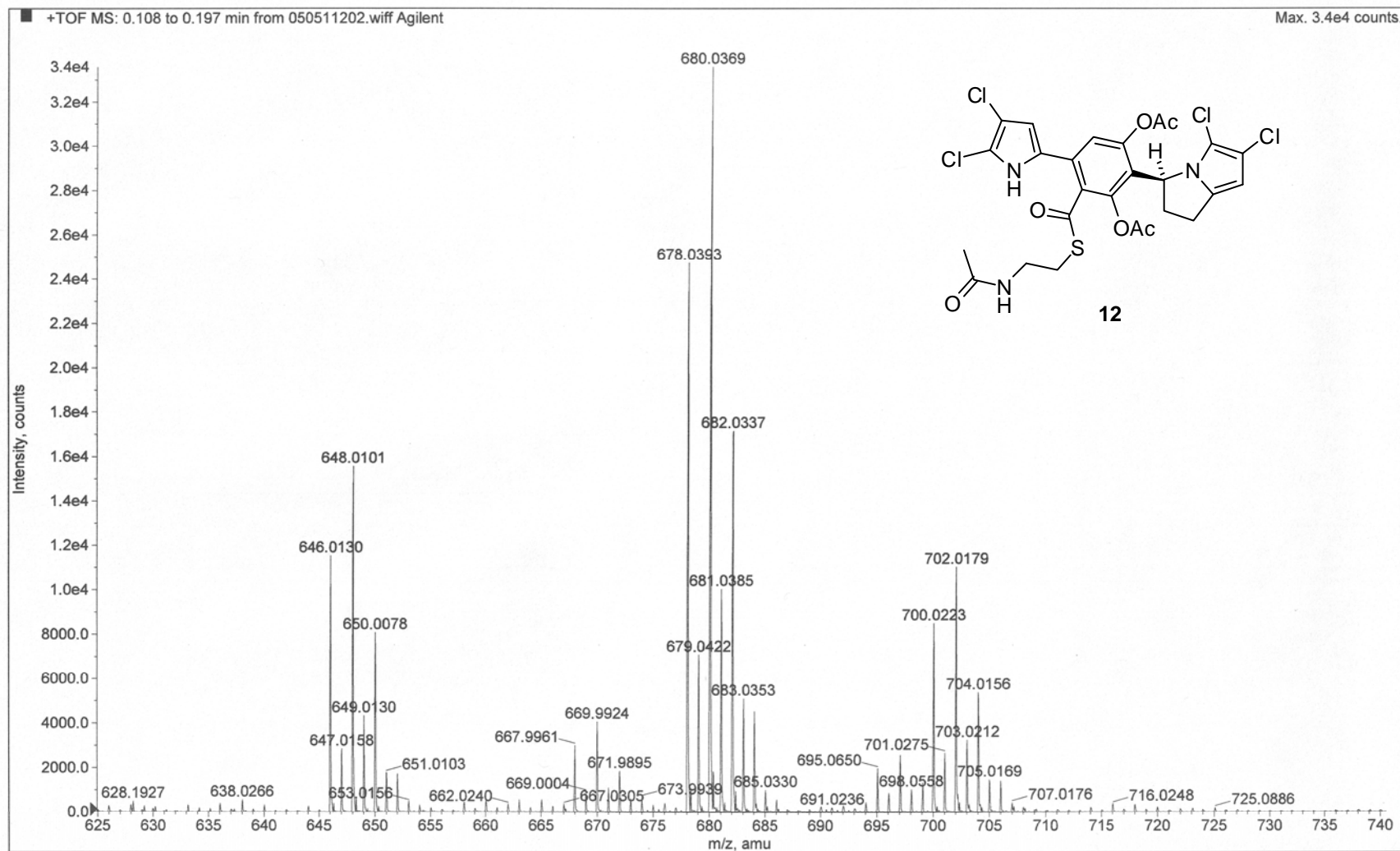


may4thioesterAc201P #5285 RT: 17.61 AV: 1 NL: 1.59E6 microAU

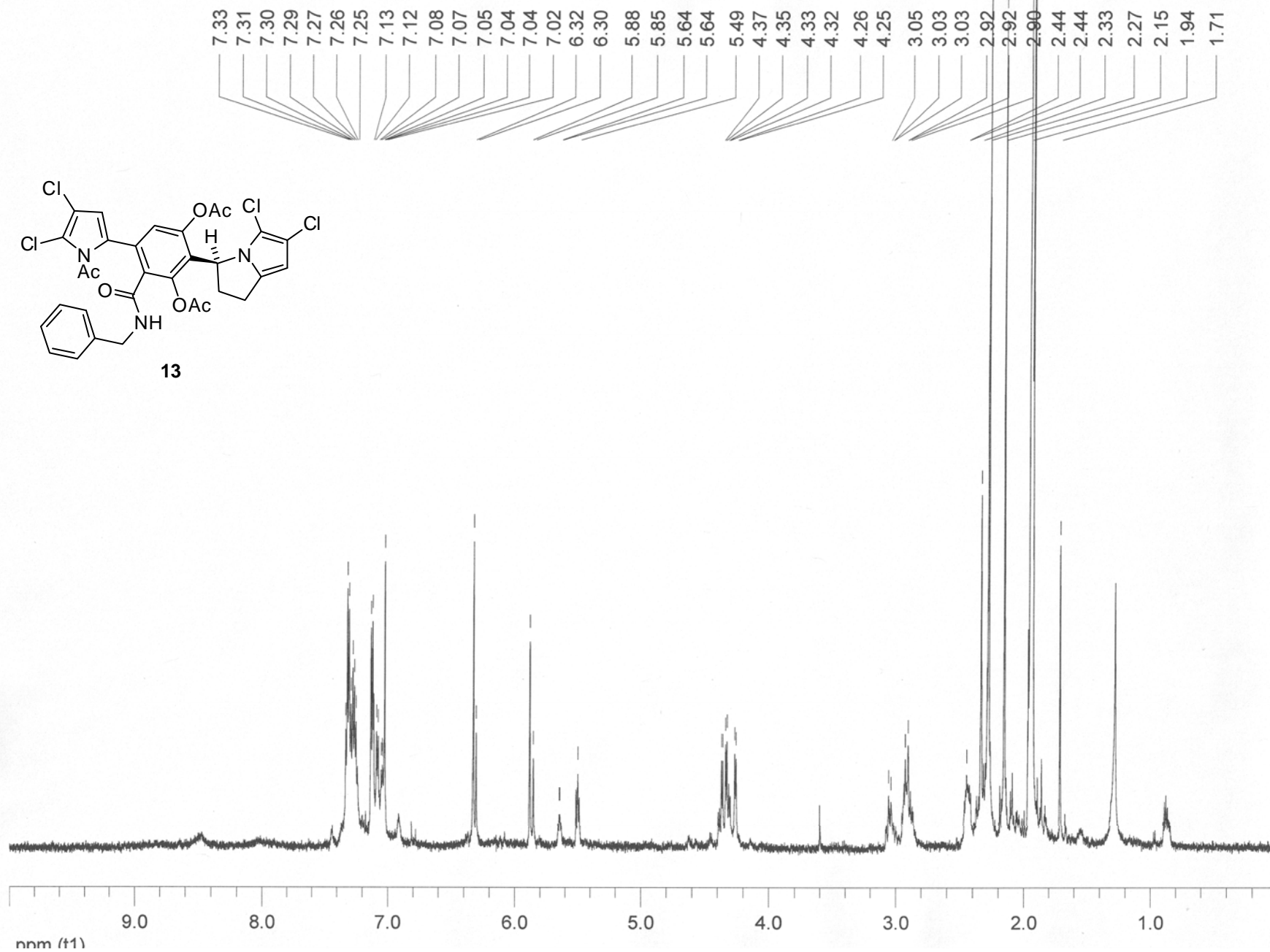
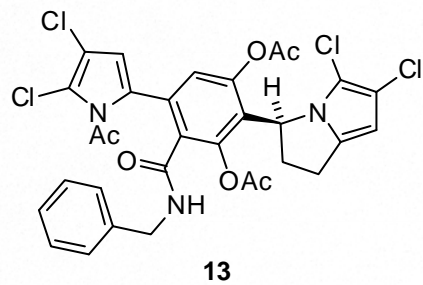


Polarity/Scan Type: Positive

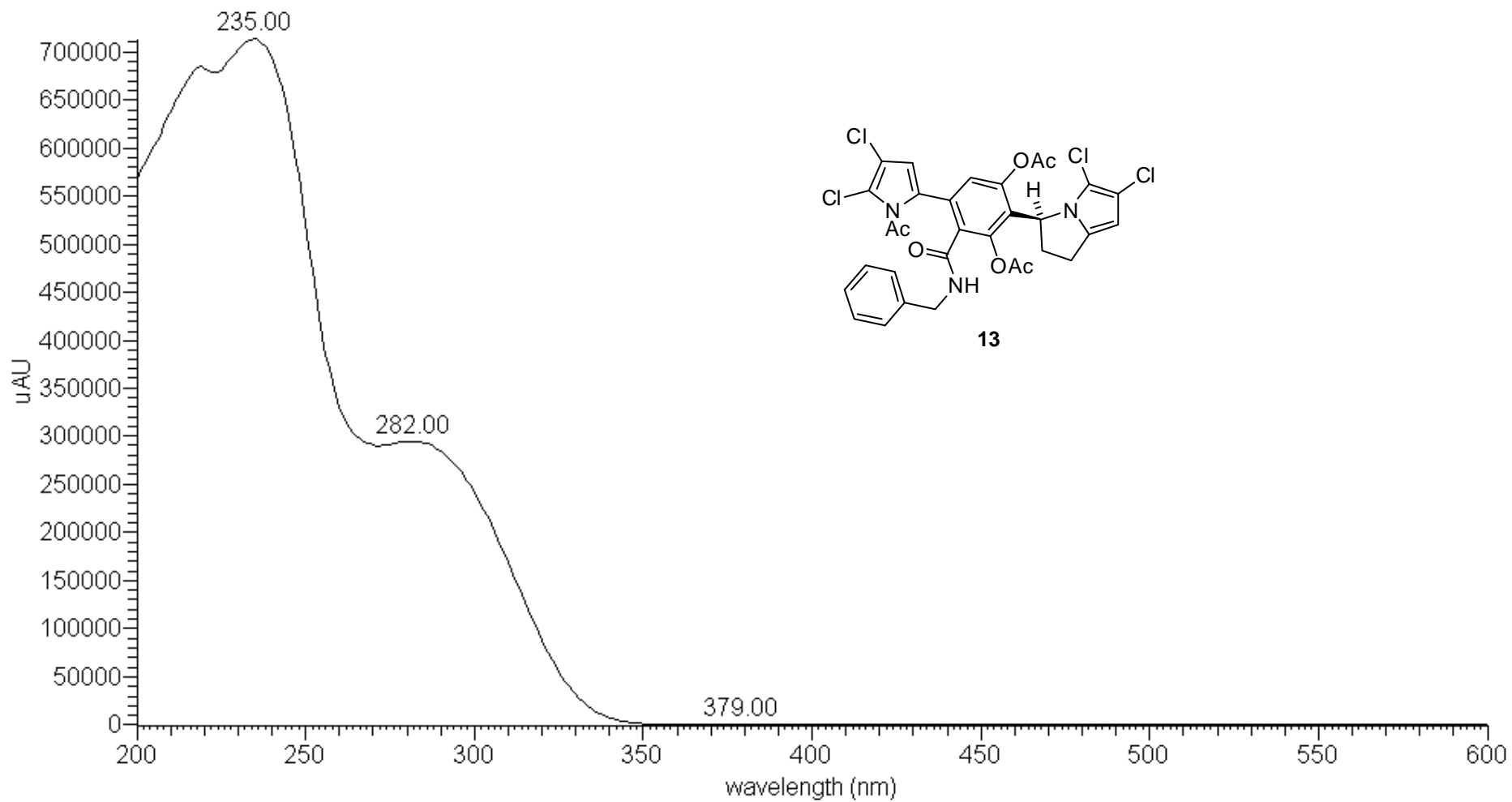
Sample Name: fch1277

Acq. File: 050511202.wiff
Acq. Date: Thursday, May 05, 2011

¹H NMR (600 MHz, CD₃CN) of triacetylated amide **13**

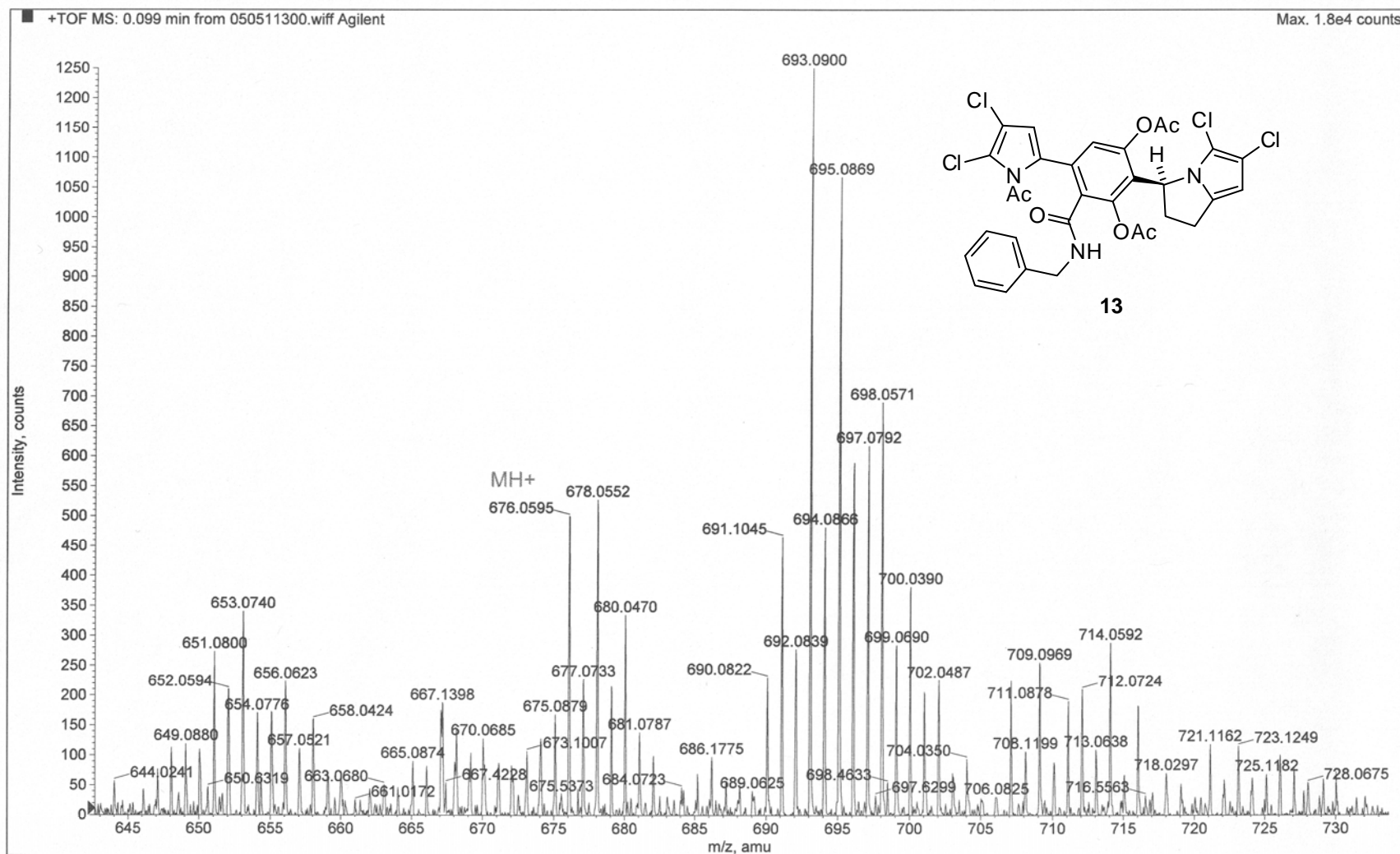


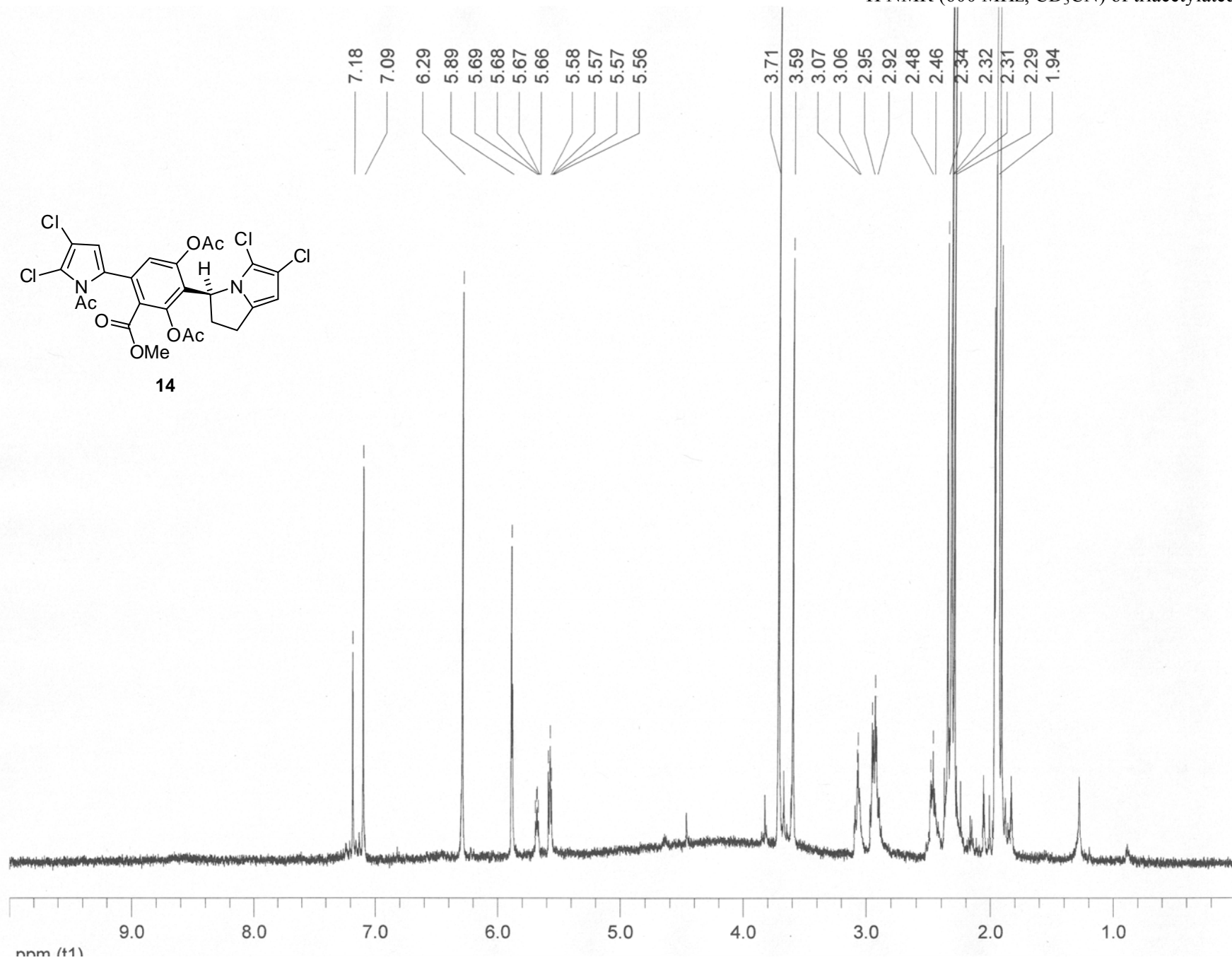
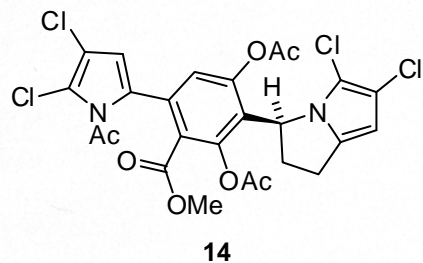
may3amideAc201 #5523 RT: 18.41 AV: 1 NL: 7.13E5 microAU



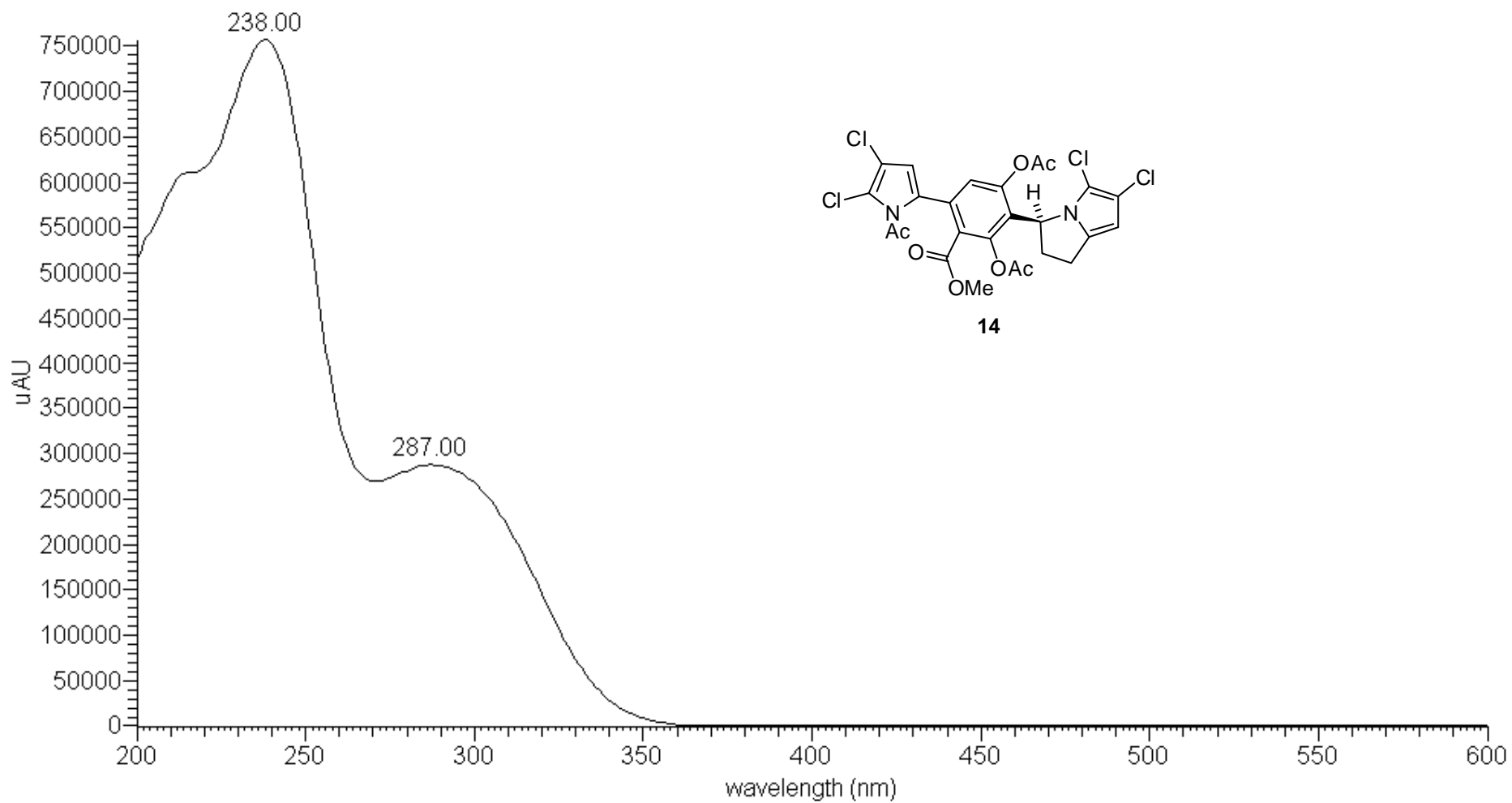
Polarity/Scan Type: Positive

Sample Name: fch1280

Acq. File: 050511300.wiff
Acq. Date: Thursday, May 05, 2011



may3esterAc201 #5523 RT: 18.41 AV: 1 NL: 7.56E5 microAU



Polarity/Scan Type: Positive

Sample Name: fch1279

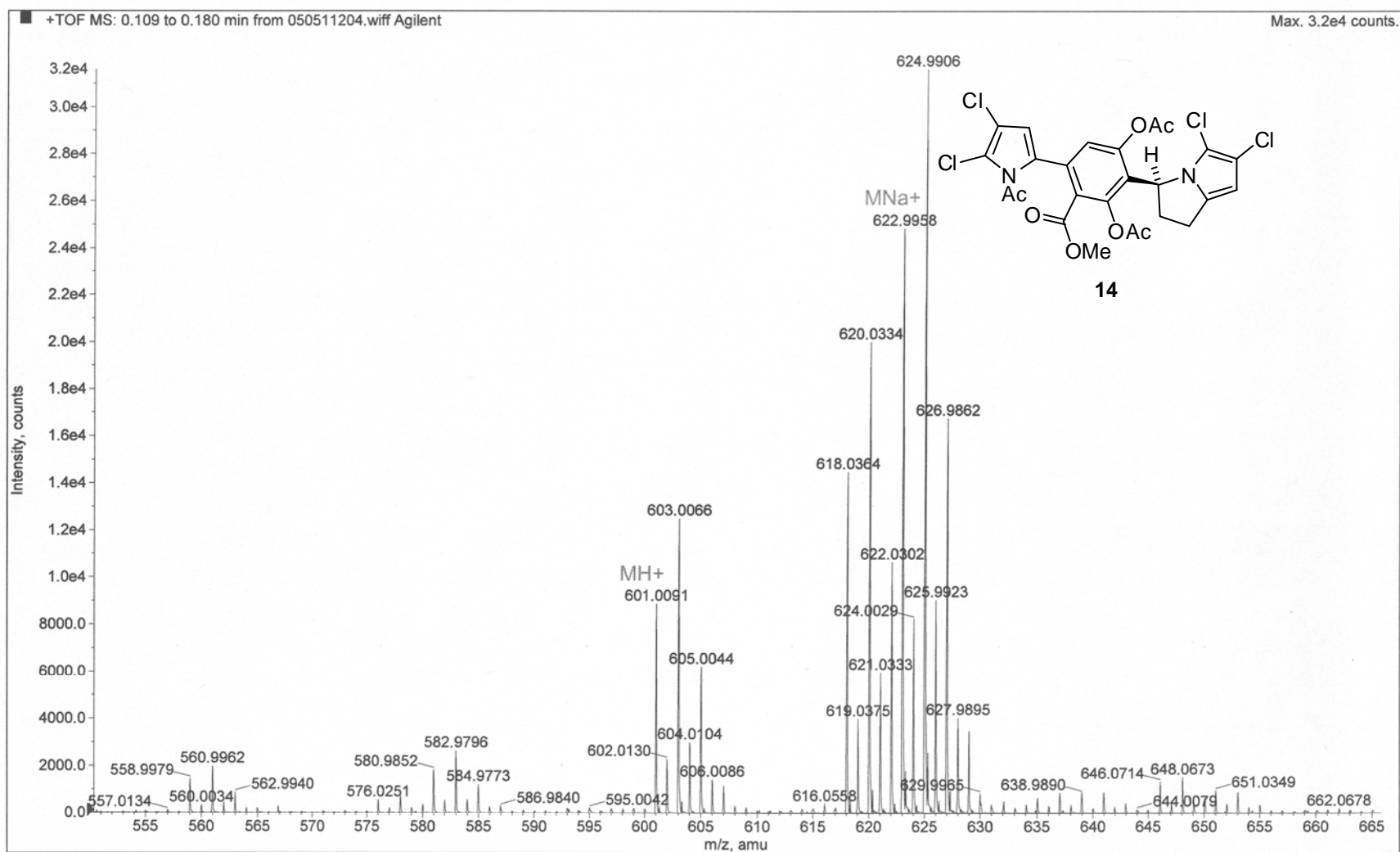
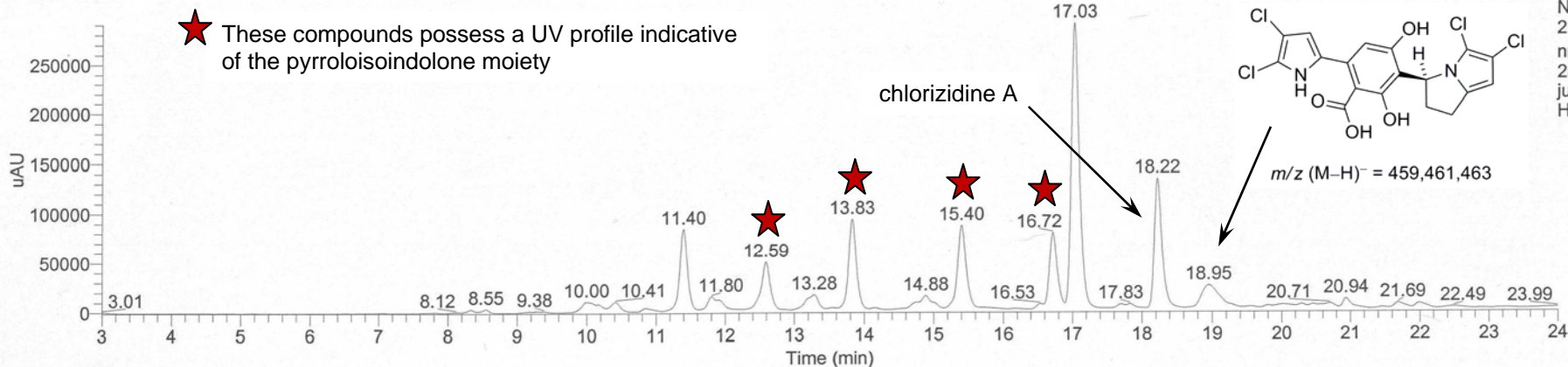
Acq. File: 050511204.wiff
Acq. Date: Thursday, May 05, 2011

Figure S3. LC/(-)-LRESI-MS chromatogram of chlorizidine (**1**) in pH 10 buffer

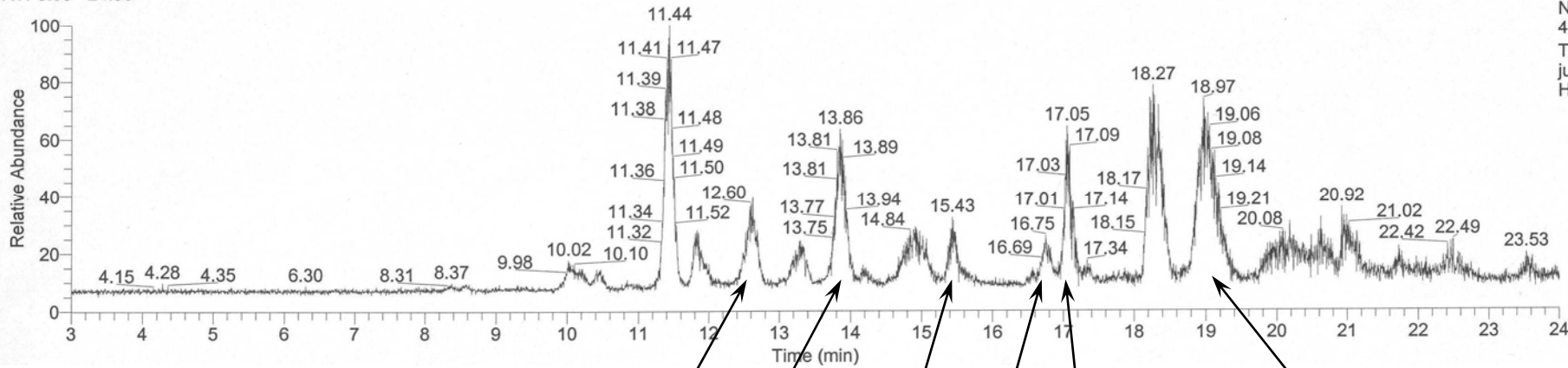
C:\Xcalibur\...july5pH10NH4CIN1

7/5/2011 1:55:47 PM

RT: 3.00 - 24.00



RT: 3.00 - 24.00



$m/z = 405,407,409; 423,425,427$

$m/z = 403,405,407; 439,441,443$

$m/z = 387,388$

$m/z = 373,375,377; 437,439,441$

$m/z = 420,423,425$

$m/z = 459,461,463$

Figure S4. X-Ray structure of chlorizidine A (**1**)

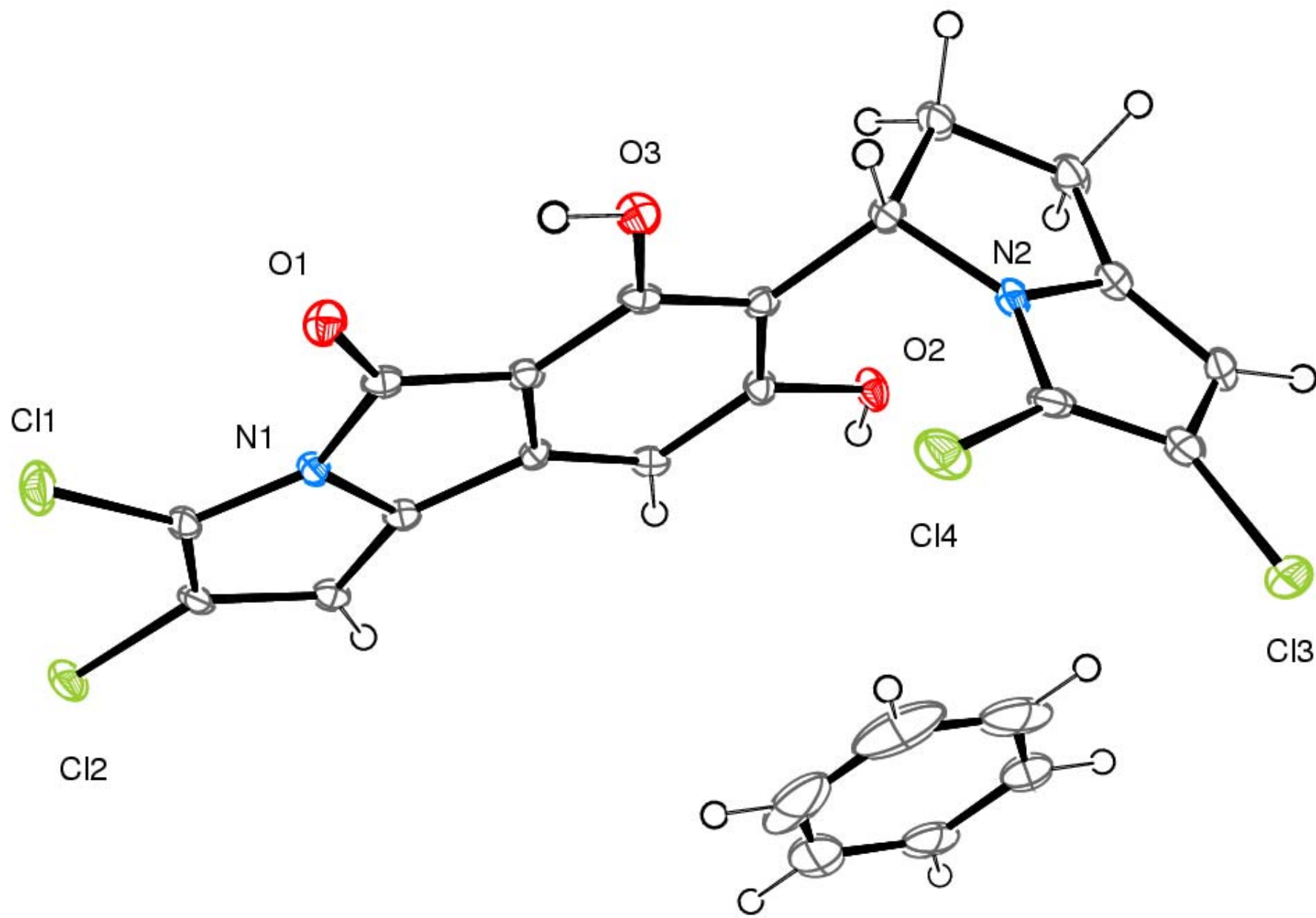
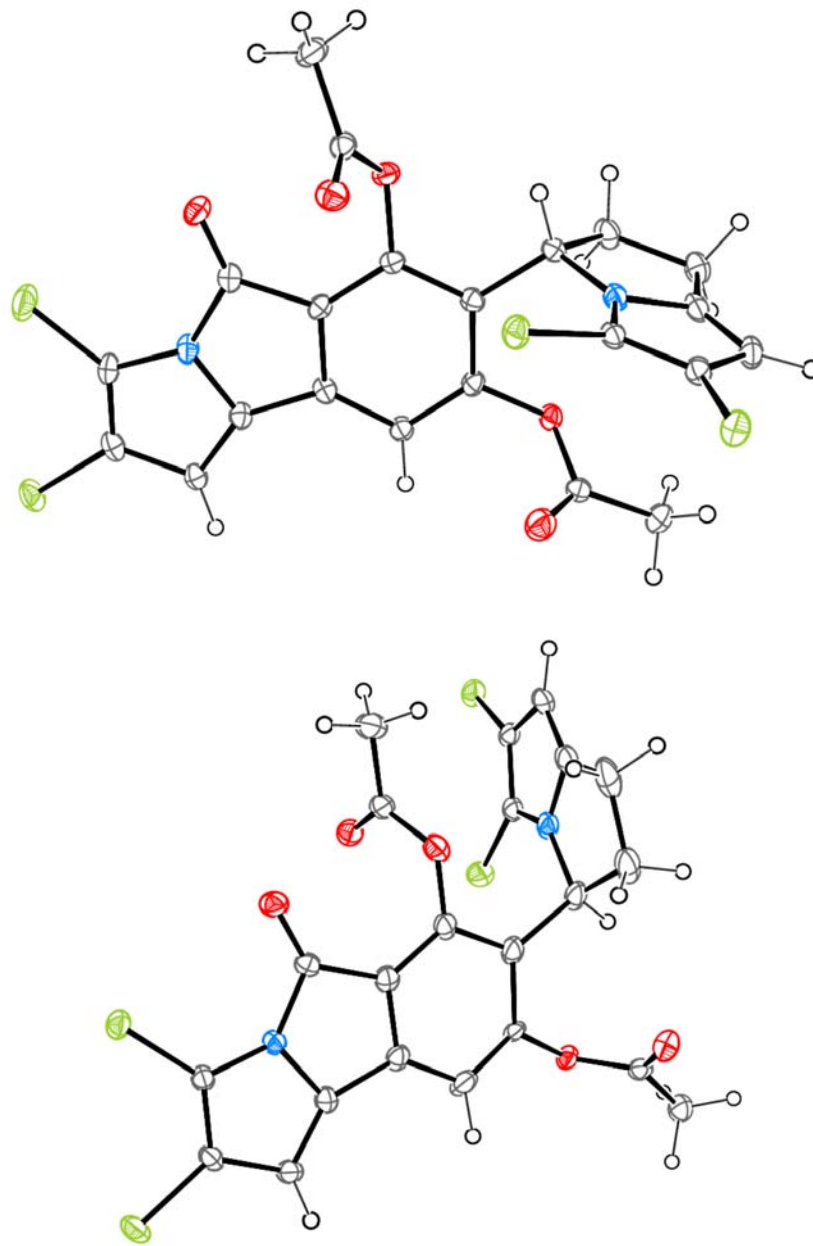


Figure S5. X-Ray structure of diacetyl chlorizidine A (2)



Developmental Therapeutics Program

NSC: D-754768 / 1

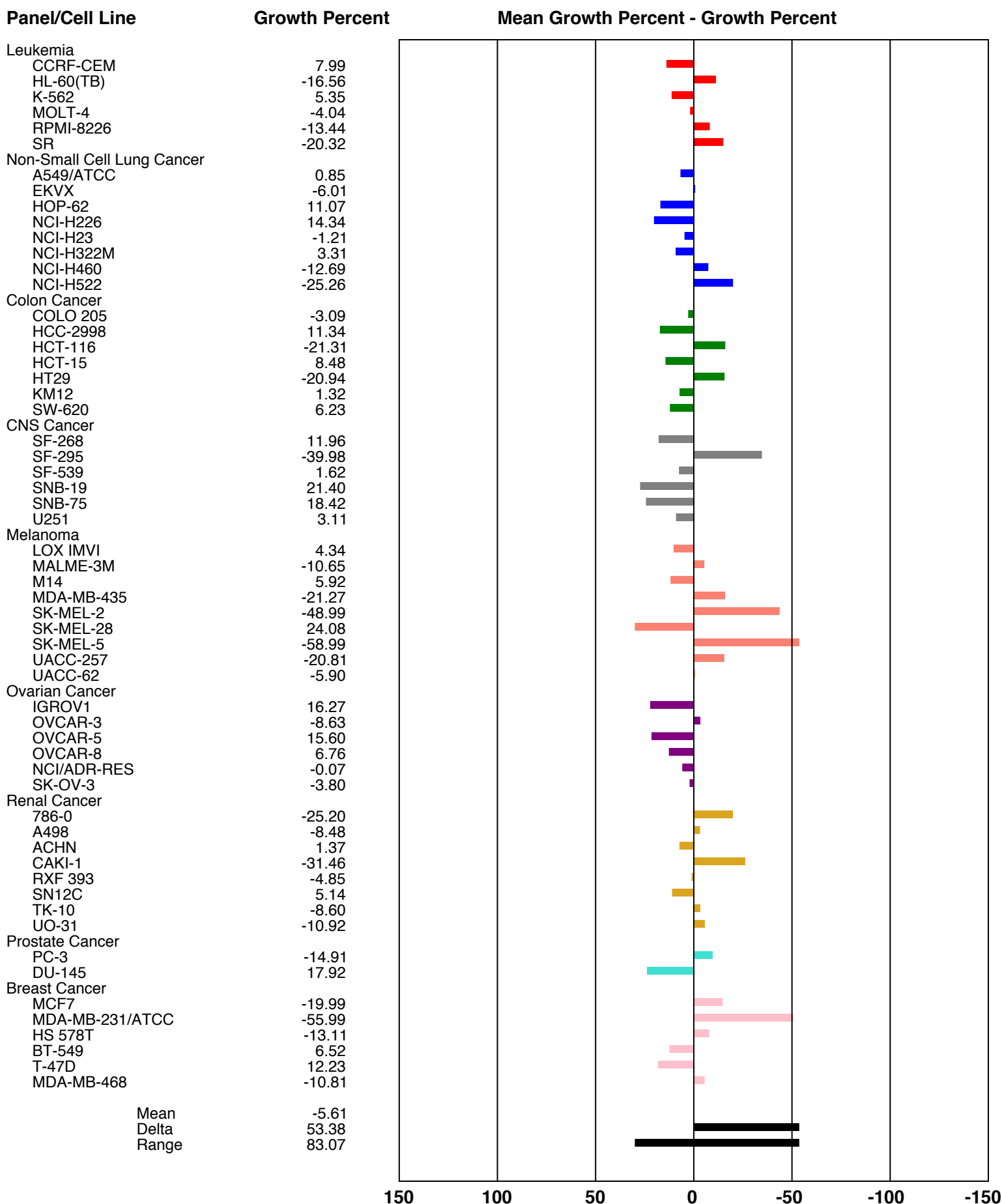
Conc: 1.00E-5 Molar

Test Date: Nov 01, 2010

One Dose Mean Graph

Experiment ID: 1011OS45

Report Date: Dec 03, 2010



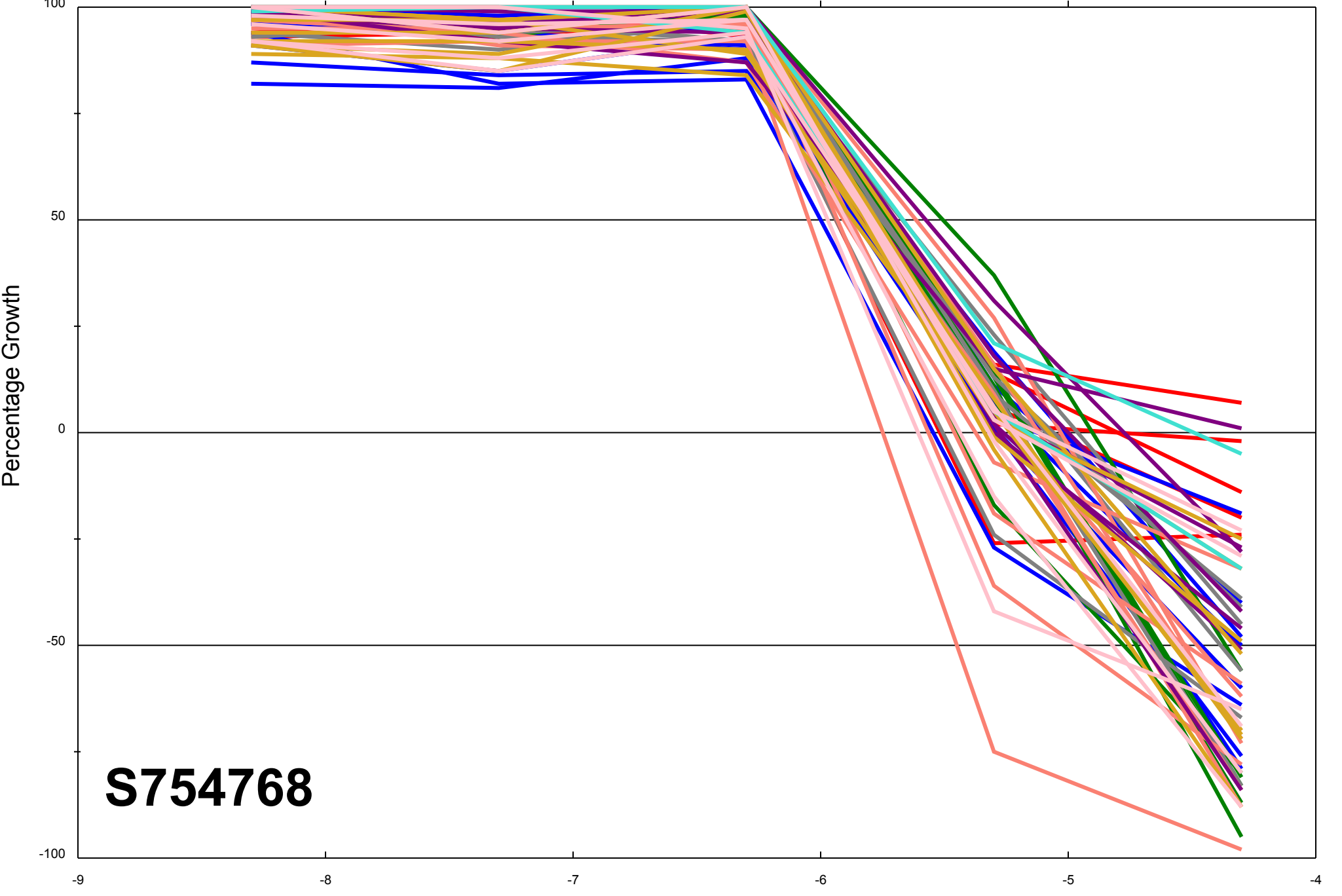
150 100 50 0 -50 -100 -150

Dose Response Curves

Report Date:February 01, 2011

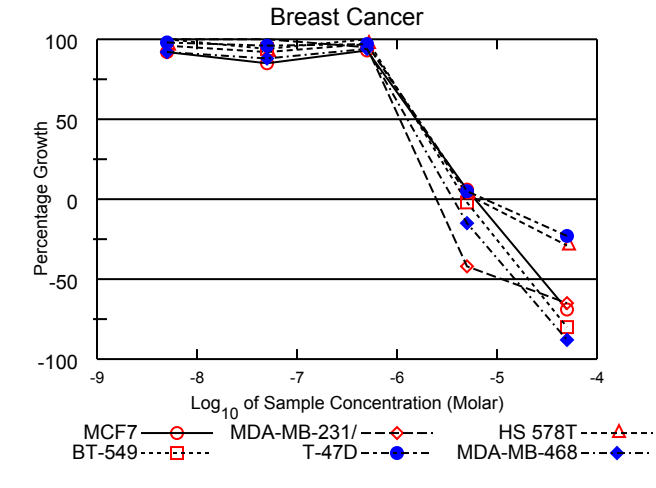
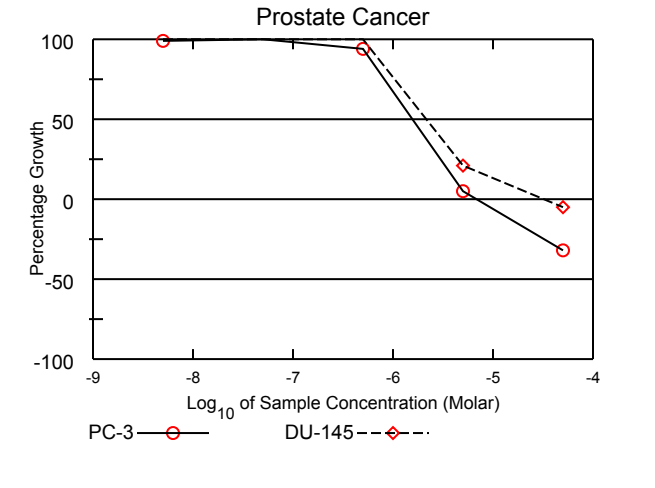
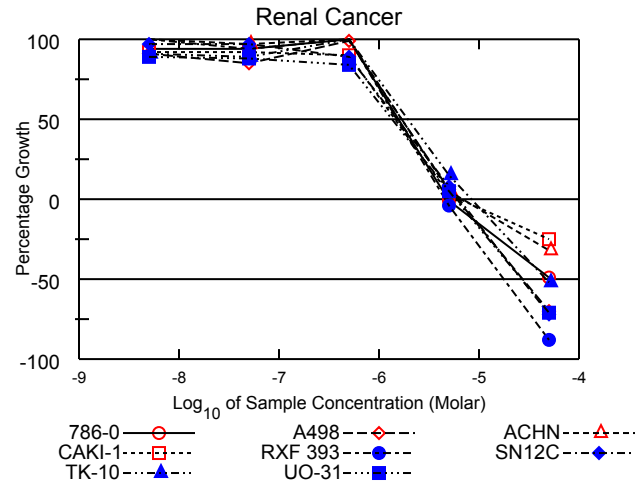
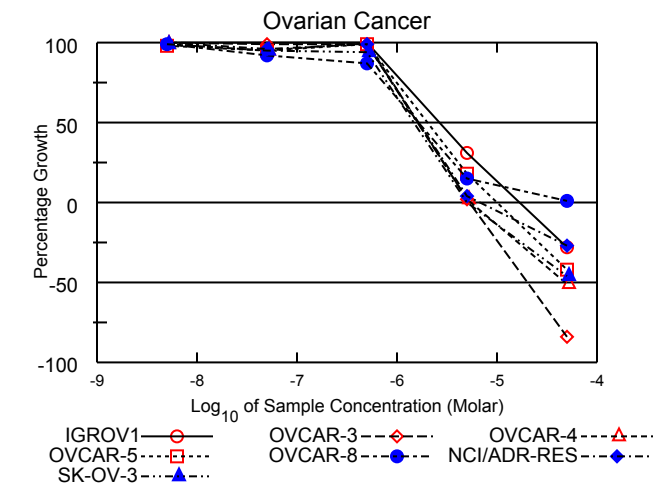
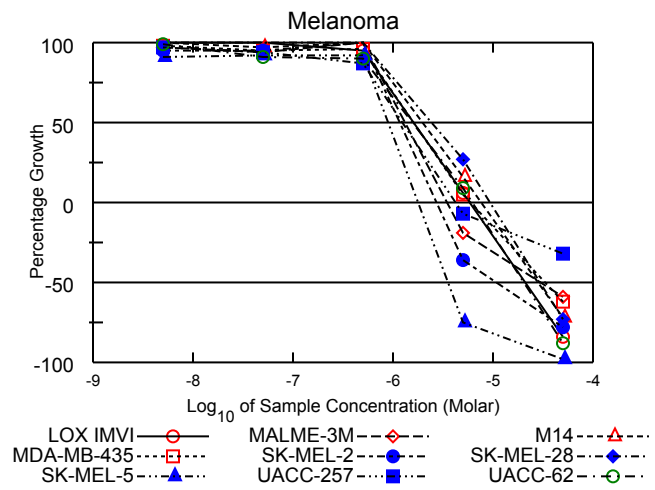
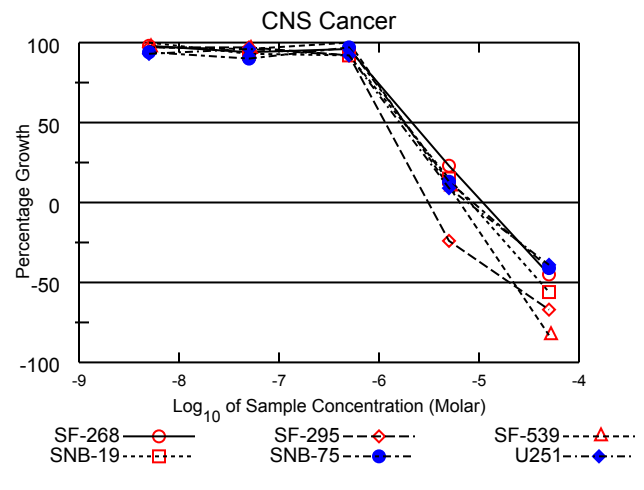
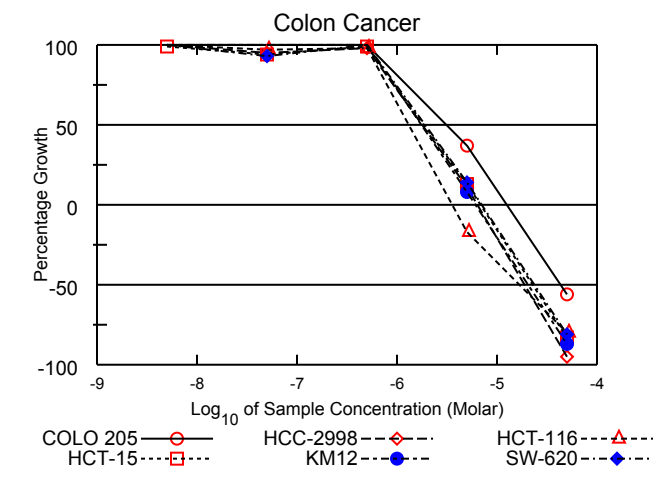
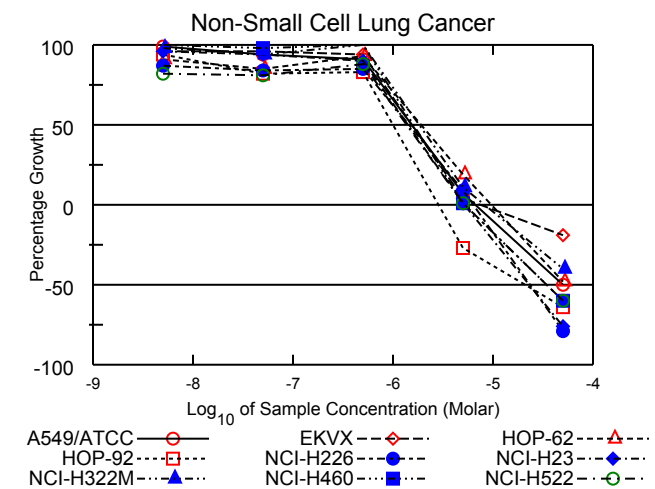
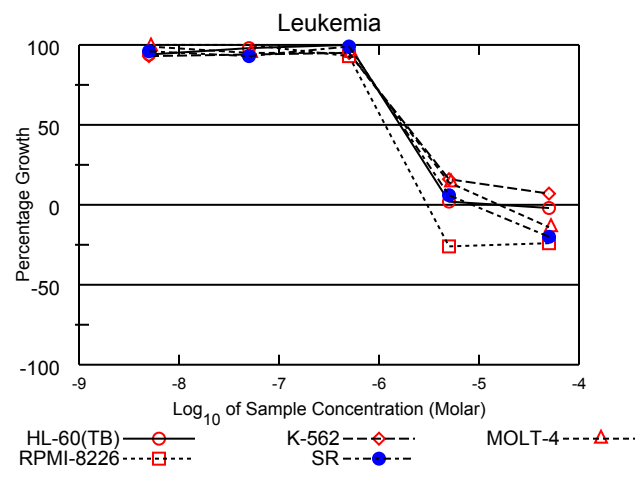
Test Date:December 06, 2010

All Cell Lines



S754768

Log₁₀ of Sample Concentration (Molar)



National Cancer Institute Developmental Therapeutics Program In-Vitro Testing Results

NSC : D - 754768 / 1	Experiment ID : 1012NS49	Test Type : 08	Units : Molar
Report Date : February 01, 2011	Test Date : December 06, 2010	QNS :	MC :
COMI : CNH287.528_SS_XM (100117)	Stain Reagent : SRB Dual-Pass Related	SSPL : 075T	

Panel/Cell Line	Log10 Concentration												GI50	TGI	LC50
	Time Zero	Ctrl	-8.3	-7.3	-6.3	-5.3	-4.3	-8.3	-7.3	-6.3	-5.3	-4.3			
Leukemia															
HL-60(TB)	0.986	2.925	2.818	2.883	2.971	1.031	0.965	94	98	102	2	-2	1.67E-6	1.65E-5	> 5.00E-5
K-562	0.187	1.718	1.603	1.620	1.648	0.437	0.291	93	94	95	16	7	1.88E-6	> 5.00E-5	> 5.00E-5
MOLT-4	0.707	2.625	2.613	2.528	2.534	0.972	0.607	99	95	95	14	-14	1.80E-6	1.56E-5	> 5.00E-5
RPMI-8226	0.773	2.231	2.265	2.225	2.125	0.571	0.590	102	100	93	-26	-24	1.14E-6	3.01E-6	> 5.00E-5
SR	0.593	2.320	2.252	2.200	2.296	0.693	0.474	96	93	99	6	-20	1.67E-6	8.36E-6	> 5.00E-5
Non-Small Cell Lung Cancer															
A549/ATCC	0.415	2.088	2.068	1.982	1.931	0.538	0.210	99	94	91	7	-50	1.54E-6	6.73E-6	> 5.00E-5
EKVX	0.717	1.904	1.862	1.856	1.838	0.769	0.583	96	96	94	4	-19	1.56E-6	7.72E-6	> 5.00E-5
HOP-62	0.386	0.898	0.852	0.819	0.863	0.485	0.200	91	85	93	19	-48	1.92E-6	9.64E-6	> 5.00E-5
HOP-92	1.057	1.512	1.485	1.431	1.433	0.768	0.376	94	82	83	-27	-64	9.92E-7	2.82E-6	2.04E-5
NCI-H226	0.508	1.261	1.160	1.138	1.152	0.573	0.109	87	84	85	9	-79	1.45E-6	6.28E-6	2.35E-5
NCI-H23	0.483	1.631	1.590	1.557	1.521	0.498	0.117	96	94	90	1	-76	1.42E-6	5.20E-6	2.31E-5
NCI-H322M	0.767	1.697	1.679	1.641	1.718	0.870	0.463	98	94	102	11	-40	1.87E-6	8.26E-6	> 5.00E-5
NCI-H460	0.228	2.072	2.071	2.041	2.118	0.253	0.091	100	98	103	1	-60	1.65E-6	5.26E-6	3.43E-5
NCI-H522	0.614	1.548	1.385	1.373	1.434	0.620	0.247	82	81	88	1	-60	1.36E-6	5.12E-6	3.44E-5
Colon Cancer															
COLO 205	0.257	0.965	1.003	0.963	0.967	0.517	0.113	105	100	100	37	-56	3.09E-6	1.24E-5	4.29E-5
HCC-2998	0.948	2.808	2.845	2.722	2.763	1.152	0.049	102	95	98	11	-95	1.77E-6	6.34E-6	1.88E-5
HCT-116	0.267	1.841	1.879	1.796	1.810	0.222	0.054	102	97	98	-17	-80	1.31E-6	3.57E-6	1.68E-5
HCT-15	0.265	1.639	1.629	1.556	1.623	0.446	0.046	99	94	99	13	-83	1.86E-6	6.86E-6	2.28E-5
KM12	0.302	1.529	1.549	1.541	1.544	0.402	0.040	102	101	101	8	-87	1.77E-6	6.09E-6	2.05E-5
SW-620	0.186	1.193	1.197	1.128	1.194	0.327	0.036	100	93	100	14	-81	1.91E-6	7.02E-6	2.36E-5
CNS Cancer															
SF-268	0.379	1.389	1.366	1.327	1.352	0.609	0.210	98	94	96	23	-45	2.13E-6	1.09E-5	> 5.00E-5
SF-295	0.884	2.778	2.728	2.727	2.631	0.673	0.288	97	97	92	-24	-67	1.16E-6	3.11E-6	1.99E-5
SF-539	0.634	1.706	1.669	1.658	1.718	0.737	0.107	97	96	101	10	-83	1.81E-6	6.35E-6	2.20E-5
SNB-19	0.342	1.292	1.317	1.226	1.215	0.480	0.149	103	93	92	15	-56	1.74E-6	8.01E-6	4.06E-5
SNB-75	0.585	1.133	1.099	1.076	1.118	0.657	0.346	94	90	97	13	-41	1.83E-6	8.76E-6	> 5.00E-5
U251	0.264	1.416	1.341	1.373	1.325	0.366	0.160	93	96	92	9	-39	1.60E-6	7.62E-6	> 5.00E-5
Melanoma															
LOX IMVI	0.172	1.225	1.236	1.223	1.168	0.235	0.027	101	100	95	6	-84	1.59E-6	5.82E-6	2.08E-5
MALME-3M	0.593	1.019	1.005	0.992	1.021	0.478	0.244	97	94	100	-19	-59	1.32E-6	3.44E-6	2.98E-5
M14	0.304	1.075	1.105	1.050	1.113	0.426	0.085	104	97	105	16	-72	2.06E-6	7.55E-6	2.80E-5
MDA-MB-435	0.401	1.850	1.819	1.776	1.793	0.474	0.153	98	95	96	5	-62	1.60E-6	5.95E-6	3.33E-5
SK-MEL-2	0.896	1.739	1.694	1.700	1.790	0.573	0.193	95	95	106	-36	-78	1.24E-6	2.79E-6	1.06E-5
SK-MEL-28	0.383	0.891	0.914	0.896	0.948	0.519	0.102	104	101	111	27	-73	2.65E-6	9.25E-6	2.91E-5
SK-MEL-5	0.464	2.591	2.402	2.424	2.420	0.117	0.010	91	92	92	-75	-98	8.93E-7	1.78E-6	3.55E-5
UACC-257	0.669	1.647	1.619	1.585	1.523	0.621	0.456	97	94	87	-7	-32	1.24E-6	4.20E-6	> 5.00E-5
UACC-62	0.591	2.068	2.058	1.935	1.921	0.728	0.074	99	91	90	9	-88	1.57E-6	6.23E-6	2.05E-5
Ovarian Cancer															
IGROV1	0.575	1.727	1.778	1.746	1.782	0.935	0.412	104	102	105	31	-28	2.78E-6	1.67E-5	> 5.00E-5
OVCAR-3	0.341	1.026	1.038	1.021	1.016	0.357	0.056	102	99	99	2	-84	1.60E-6	5.31E-6	2.03E-5
OVCAR-4	0.589	1.033	1.029	1.009	1.102	0.600	0.288	99	95	116	2	-51	1.90E-6	5.56E-6	4.75E-5
OVCAR-5	0.463	1.188	1.171	1.149	1.178	0.593	0.271	98	95	99	18	-42	2.00E-6	1.00E-5	> 5.00E-5
OVCAR-8	0.122	0.590	0.583	0.554	0.528	0.190	0.126	99	92	87	15	1	1.62E-6	> 5.00E-5	> 5.00E-5
NCI/ADR-RES	0.406	1.497	1.504	1.455	1.486	0.446	0.298	101	96	99	4	-27	1.63E-6	6.59E-6	> 5.00E-5
SK-OV-3	0.639	1.561	1.548	1.515	1.503	0.643	0.348	99	95	94	.	-46	1.47E-6	5.11E-6	> 5.00E-5
Renal Cancer															
786-0	0.661	2.073	1.990	1.989	2.111	0.656	0.337	94	94	103	-1	-49	1.62E-6	4.92E-6	> 5.00E-5
A498	0.498	0.987	0.944	0.916	0.981	0.527	0.148	91	85	99	6	-70	1.67E-6	5.96E-6	2.71E-5
ACHN	0.497	1.852	1.907	1.817	1.881	0.560	0.337	104	97	102	5	-32	1.71E-6	6.67E-6	> 5.00E-5
CAKI-1	0.716	2.470	2.329	2.333	2.293	0.763	0.541	92	92	90	3	-25	1.43E-6	6.26E-6	> 5.00E-5
RXF 393	0.655	1.164	1.164	1.132	1.161	0.632	0.082	100	94	100	-4	-88	1.51E-6	4.62E-6	1.79E-5
SN12C	0.398	1.599	1.567	1.569	1.467	0.491	0.112	97	97	89	8	-72	1.51E-6	6.25E-6	2.65E-5
TK-10	0.548	1.229	1.166	1.154	1.314	0.654	0.266	91	89	112	15	-52	2.20E-6	8.51E-6	4.74E-5
UO-31	0.615	1.501	1.406	1.392	1.361	0.657	0.177	89	88	84	5	-71	1.35E-6	5.76E-6	2.62E-5
Prostate Cancer															
PC-3	0.451	1.716	1.698	1.722	1.637	0.515	0.305	99	100	94	5	-32	1.56E-6	6.80E-6	> 5.00E-5
DU-145	0.241	0.936	0.934	0.957	0.960	0.386	0.230	100	103	103	21	-5	2.22E-6	3.26E-5	> 5.00E-5
Breast Cancer															
MCF7	0.284	1.200	1.127	1.066	1.136	0.338	0.087	92	85	93	6	-69	1.56E-6	5.99E-6	2.76E-5
MDA-MB-231/ATCC	0.430	1.041	1.045	1.055	1.011	0.250	0.152	101	102	95	-42	-65	1.07E-6	2.47E-6	1.13E-5
HS 578T	0.812	1.671	1.639	1.602	1.642	0.834	0.574	96	92	97	3	-29	1.56E-6	5.99E-6	> 5.00E-5
BT-549	0.726	1.283	1.299	1.249	1.323	0.709	0.149	103	94	107	-2	-80	1.66E-6	4.76E-6	2.07E-5
T-47D	0.647	1.347	1.336	1.321	1.327	0.679	0.501	98	96	97	5	-23	1.62E-6	7.36E-6	> 5.00E-5
MDA-MB-468	0.575	1.135	1.088	1.069	1.100	0.491	0.069	92	88	94	-15	-88	1.27E-6	3.66E-6	1.51E-5

Mean Graphs

Report Date :February 01, 2011

Test Date :December 06, 2010

