

DETERMINATION OF ABSOLUTE CONFIGURATION OF SECONDARY ALCOHOLS USING THIN-LAYER CHROMATOGRAPHY

Alexander J. Wagner and Scott D. Rychnovsky*

*Department of Chemistry, 1102 Natural Sciences II, University of California, Irvine,
California 92697-2025*

srychnov@uci.edu

SUPPORTING INFORMATION

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Reaction Conditions for Table 1, Entries 1-13.

The table below contains reaction conditions for all samples used in Table 1. The protocol listed in the experimental for using parallel reactions in the process of determining absolute configuration was followed for all of the entries.

| Entry | Original amount of alcohol (mg/ μ mol) | CDCl ₃ added to alcohol vial (μ L) | Alcohol solution added to each reaction (μ L) | <i>R</i> -HBTM solution added to <i>R</i> -HBTM reaction (μ L) | <i>S</i> -HBTM solution added to <i>S</i> -HBTM reaction (μ L) | <i>i</i> Pr ₂ NEt solution added to each reaction (μ L) | (EtCO) ₂ O solution added to each reaction (μ L) | Methanol-d ₄ added to quench each reaction (μ L) | CDCl ₃ added to give 500 μ L total volume for each reaction (μ L) |
|----------------|--------------------------------------------|----------------------------------------------------|----------------------------------------------------|---------------------------------------------------------------------|---------------------------------------------------------------------|-------------------------------------------------------------------------|------------------------------------------------------------------|------------------------------------------------------------------|---------------------------------------------------------------------------------------|
| 1 | 9.1/74 | 403.8 | 175.0 | 131.6 | 131.6 | 72.5 | 67.0 | 50.0 | 4.0 |
| 2 ^a | 10.4/60.9 | 403.8 | 175.0 | 108.3 | 108.3 | 59.7 | 55.1 | 50.0 | 51.9 |
| 3 ^a | 9.7/58 | 403.8 | 175.0 | 103.1 | 103.1 | 56.9 | 52.5 | 50.0 | 62.6 |
| 4 | 8.9/48 | 300.0 | 130.0 | 85.2 | 85.2 | 47.2 | 43.4 | 50.0 | 144.2 |
| 5 | 5.1/19 | 150.0 | 65.0 | 33.8 | 33.8 | 18.6 | 17.2 | 50.0 | 315.4 |
| 6 | 8.7/40 | 276.9 | 120.0 | 71.2 | 71.2 | 39.2 | 36.2 | 50.0 | 183.5 |
| 7 | 6.3/25 | 184.6 | 80.0 | 44.4 | 44.4 | 24.6 | 22.6 | 50.0 | 278.4 |
| 8 | 5.7/18 | 150.0 | 65.0 | 32.0 | 32.0 | 17.8 | 16.2 | 50.0 | 319.0 |
| 9 | 5.6/17 | 150.0 | 65.0 | 30.2 | 30.2 | 16.8 | 15.4 | 50.0 | 322.7 |
| 10 | 7.3/32 | 196.2 | 85.0 | 56.8 | 56.8 | 31.4 | 28.8 | 50.0 | 247.8 |
| 11 | 6.6/18 | 150.0 | 65.0 | 32.0 | 32.0 | 17.6 | 16.2 | 50.0 | 319.1 |
| 12 | 6.2/18 | 150.0 | 65.0 | 32.0 | 32.0 | 17.6 | 16.2 | 50.0 | 319.1 |
| 13 | 5.8/20 | 150.0 | 65.0 | 35.6 | 35.6 | 19.6 | 18.0 | 50.0 | 311.7 |

(a) The peak from methanol-d₄ overlaid a peak from the alcohol used in determining conversion in the crude ¹H NMR spectrum. The solution was concentrated under reduced pressure. Then, 500 μ L of CDCl₃ was added and a second ¹H NMR spectrum was taken. Both spectra are included.

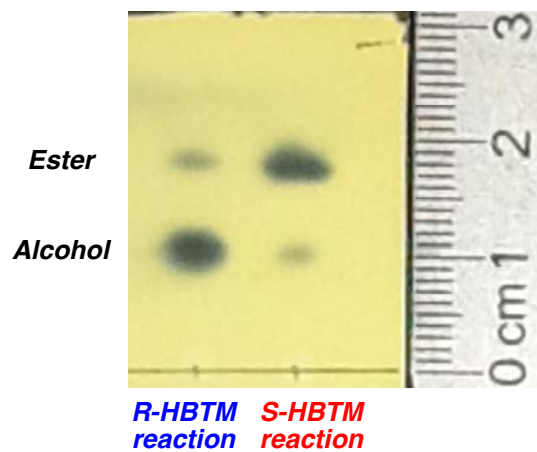
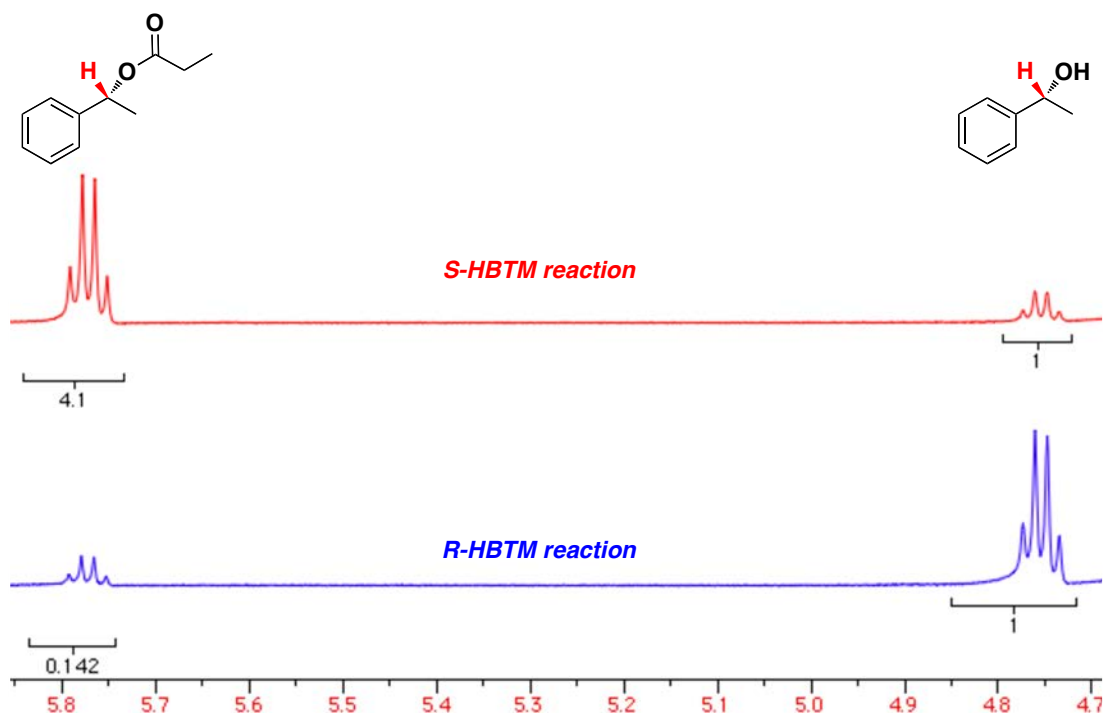
¹H NMR spectra were collected for both the *R*-HBTM and the *S*-HBTM crude reaction mixtures. Percent conversion for each reaction was measured according the integrals of each peak after phase correction, line broadening, and automatic baseline correction of all spectra (Formula 1).

Formula 1.

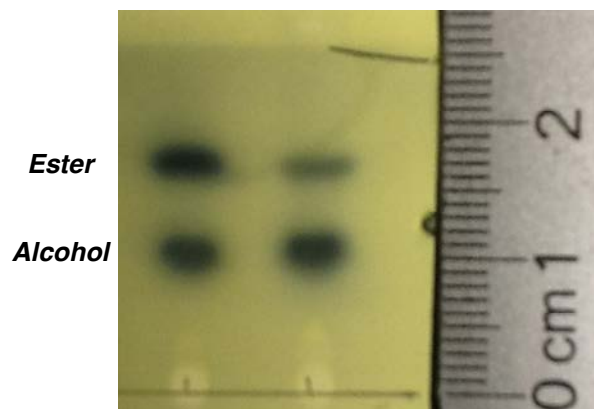
$$\text{Percent Conversion} = \frac{{}^1\text{H NMR integral}_{\text{ESTER}}}{{}^1\text{H NMR integral}_{\text{ESTER}} + {}^1\text{H NMR integral}_{\text{ALCOHOL}}} \times 100$$

Summary of TLC and ^1H NMR Spectral Data, Entries 1-13.

Entry 1. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.

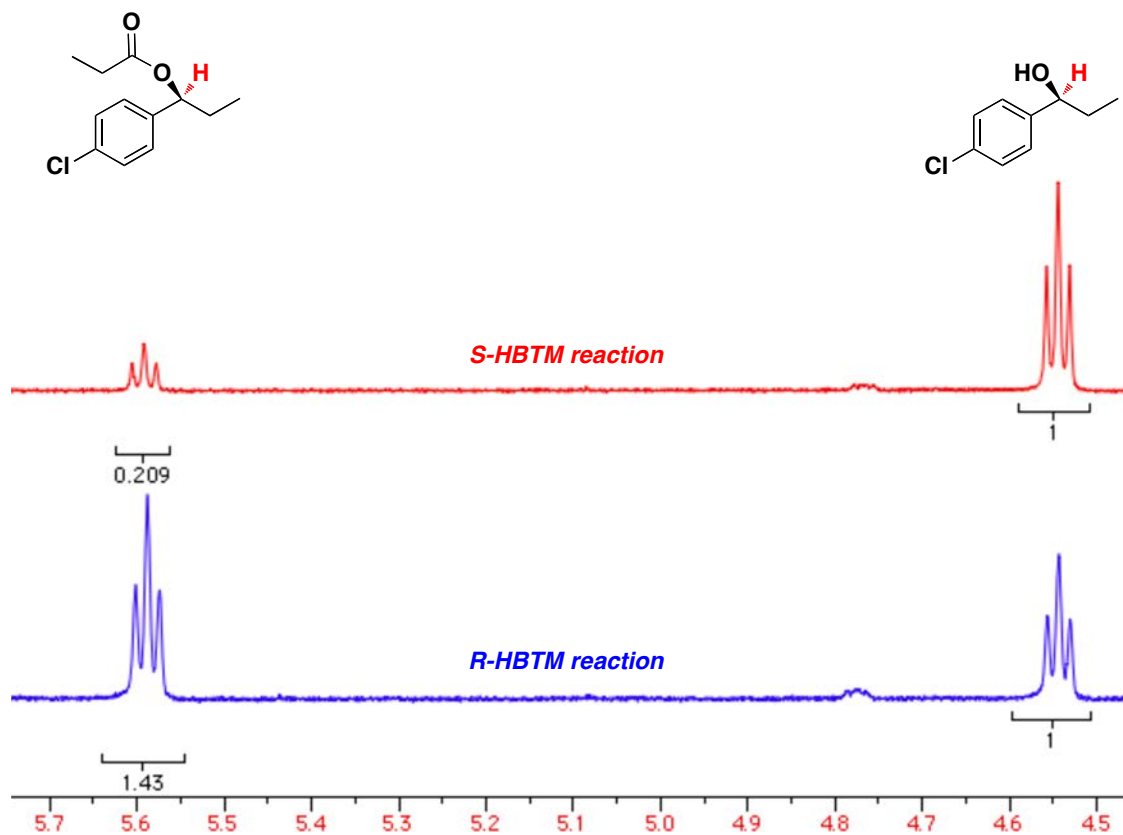
**(a)****(b)**

Entry 2. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



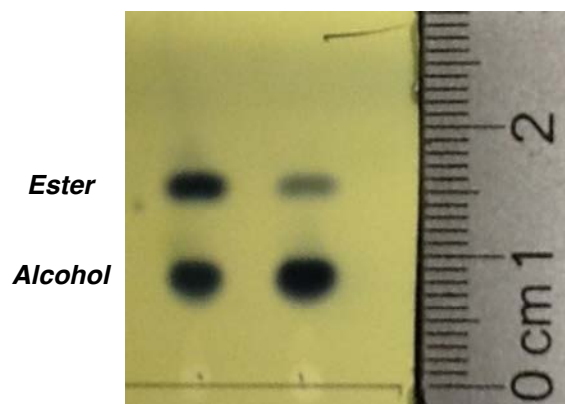
R-HBTM reaction **S-HBTM reaction**

(a)



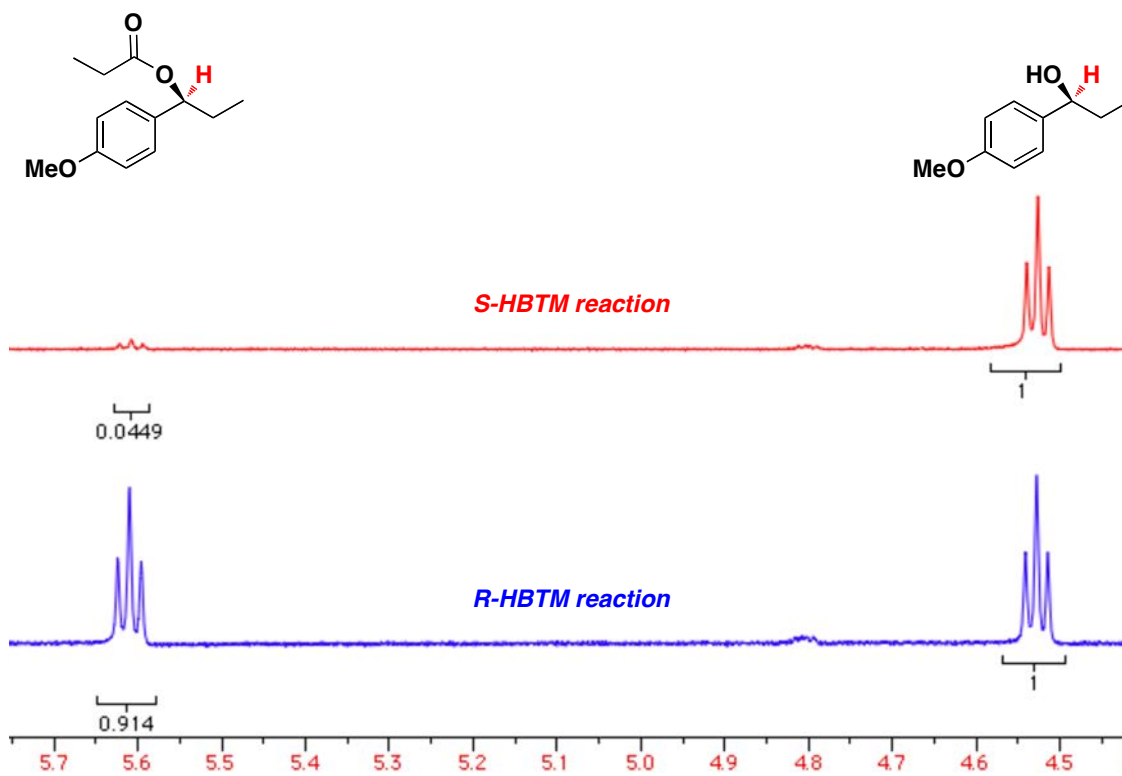
(b)

Entry 3. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



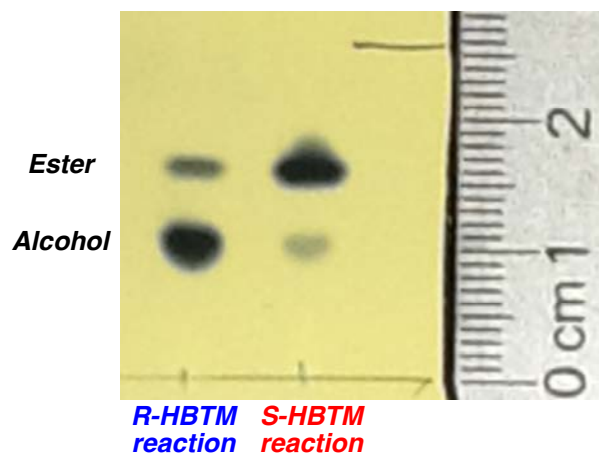
R-HBTM **S-HBTM**
reaction **reaction**

(a)

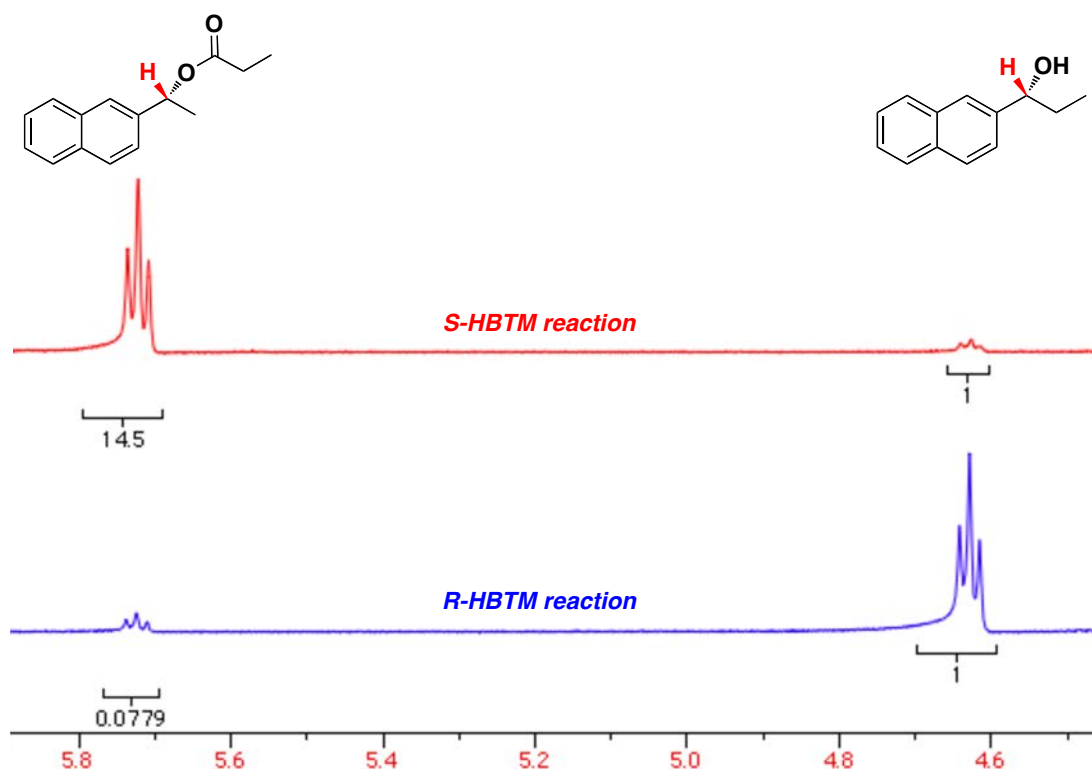


(b)

Entry 4. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.

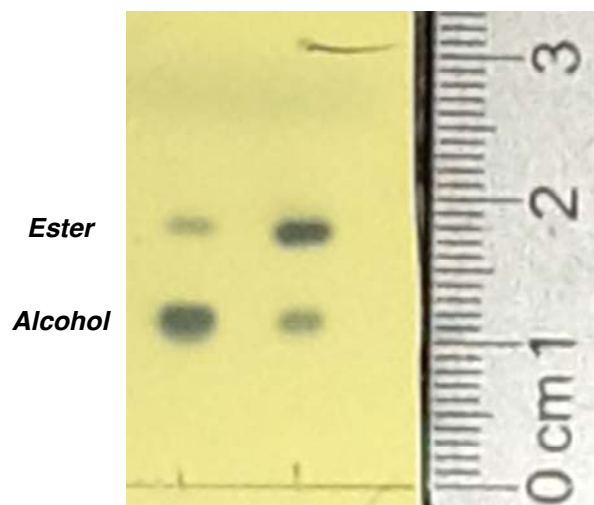


(a)



(b)

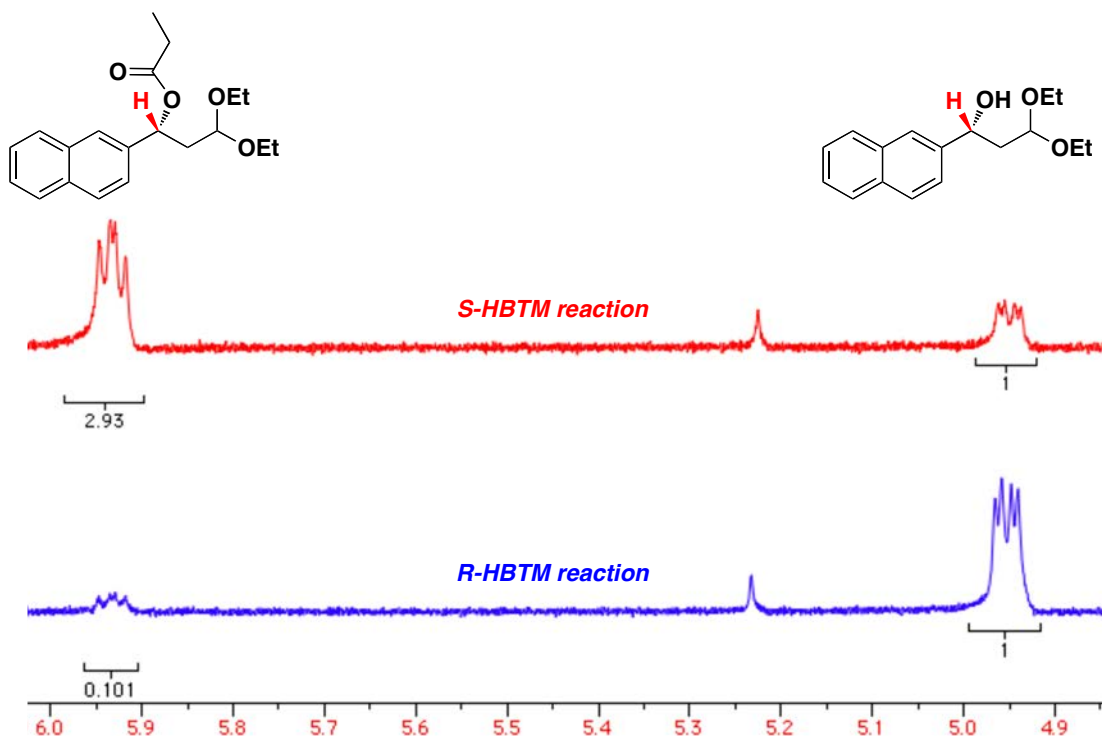
Entry 5. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



R-HBTM reaction

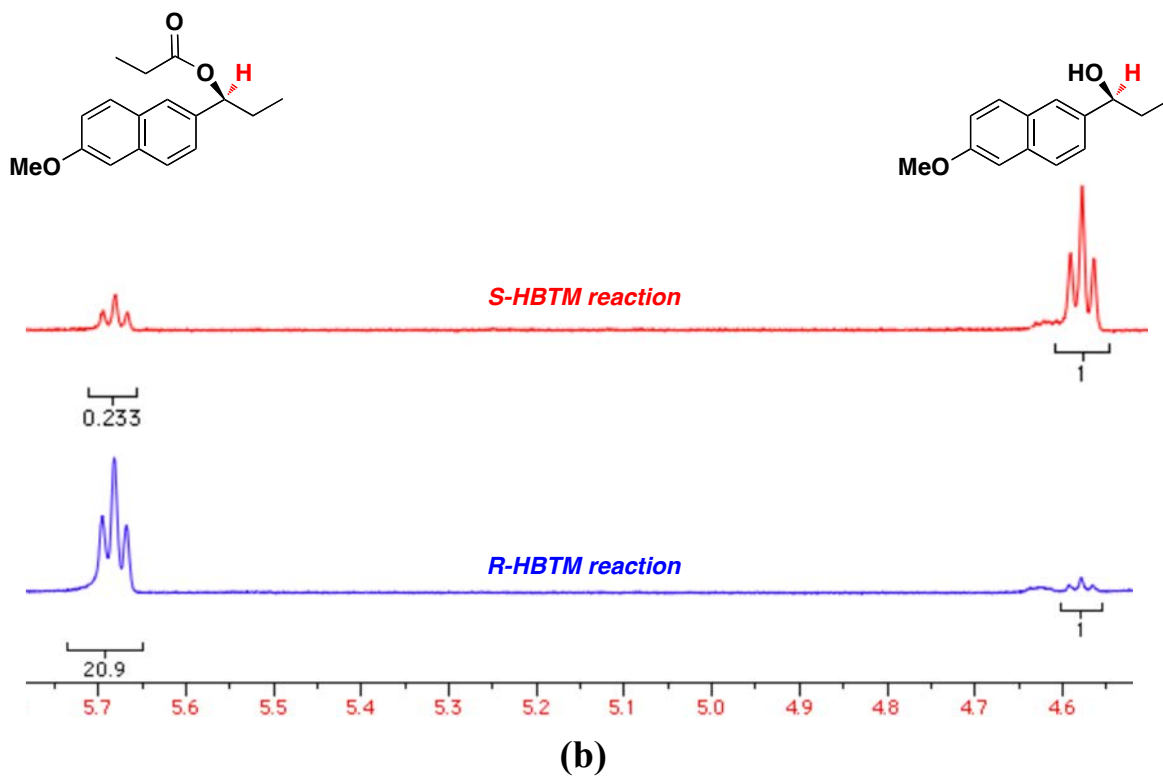
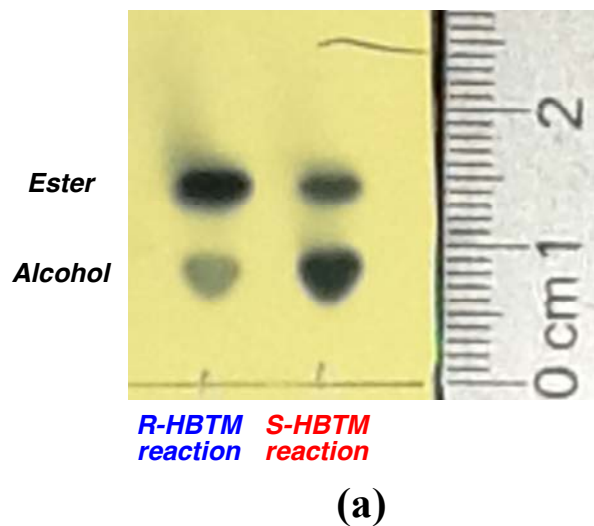
S-HBTM reaction

(a)

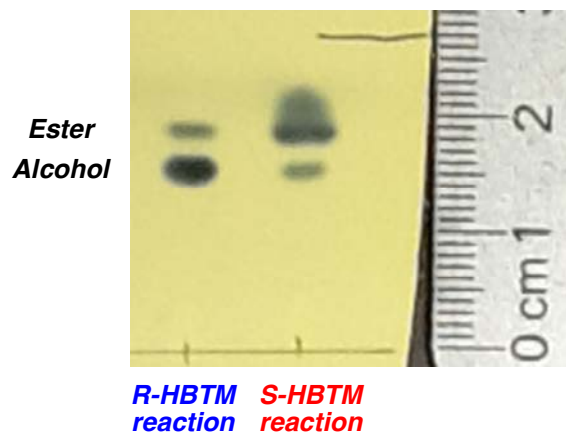


(b)

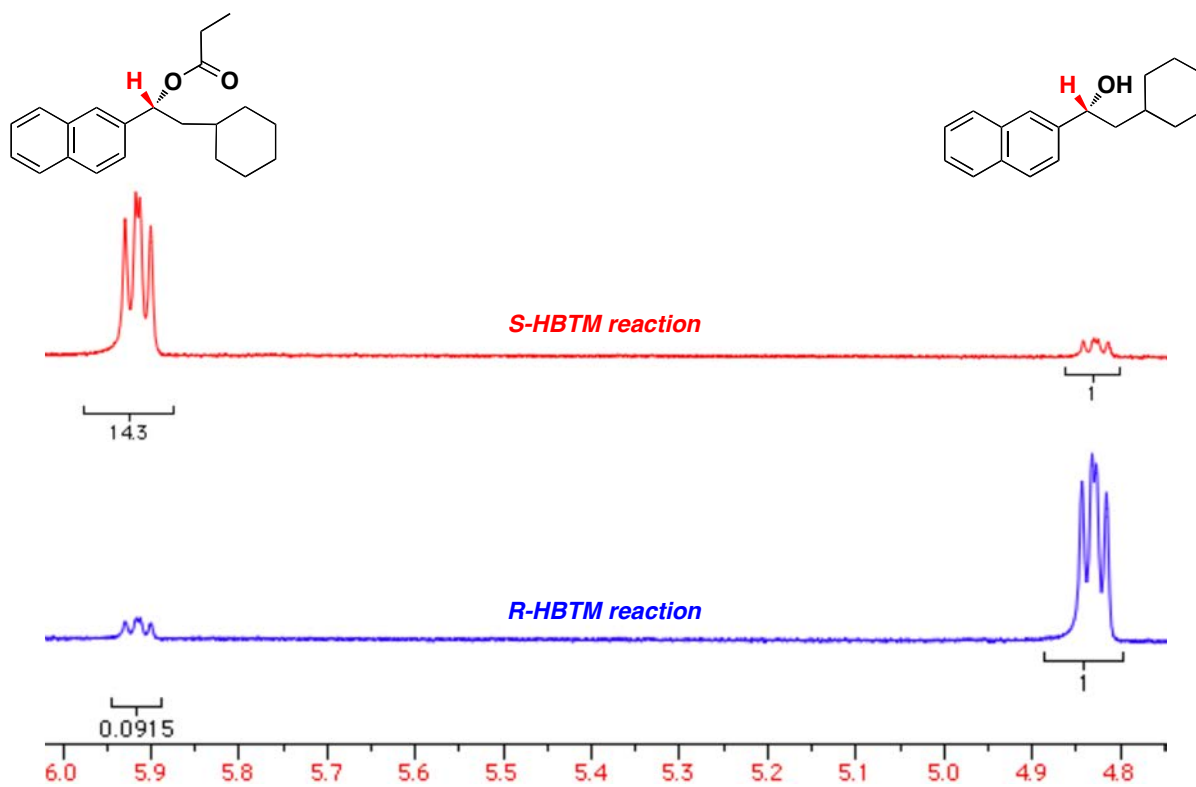
Entry 6. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



Entry 7. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.

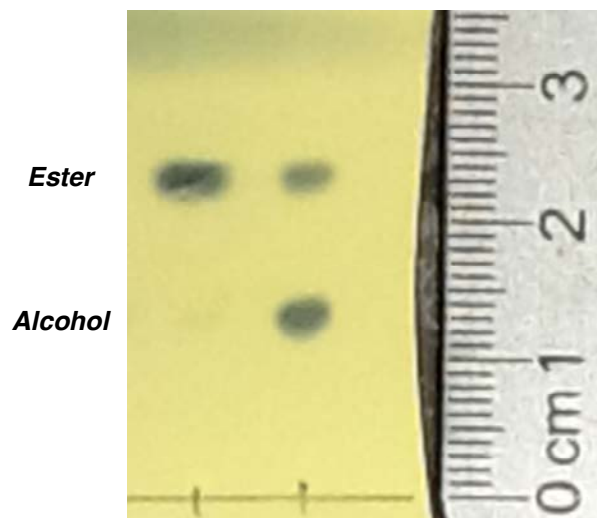


(a)



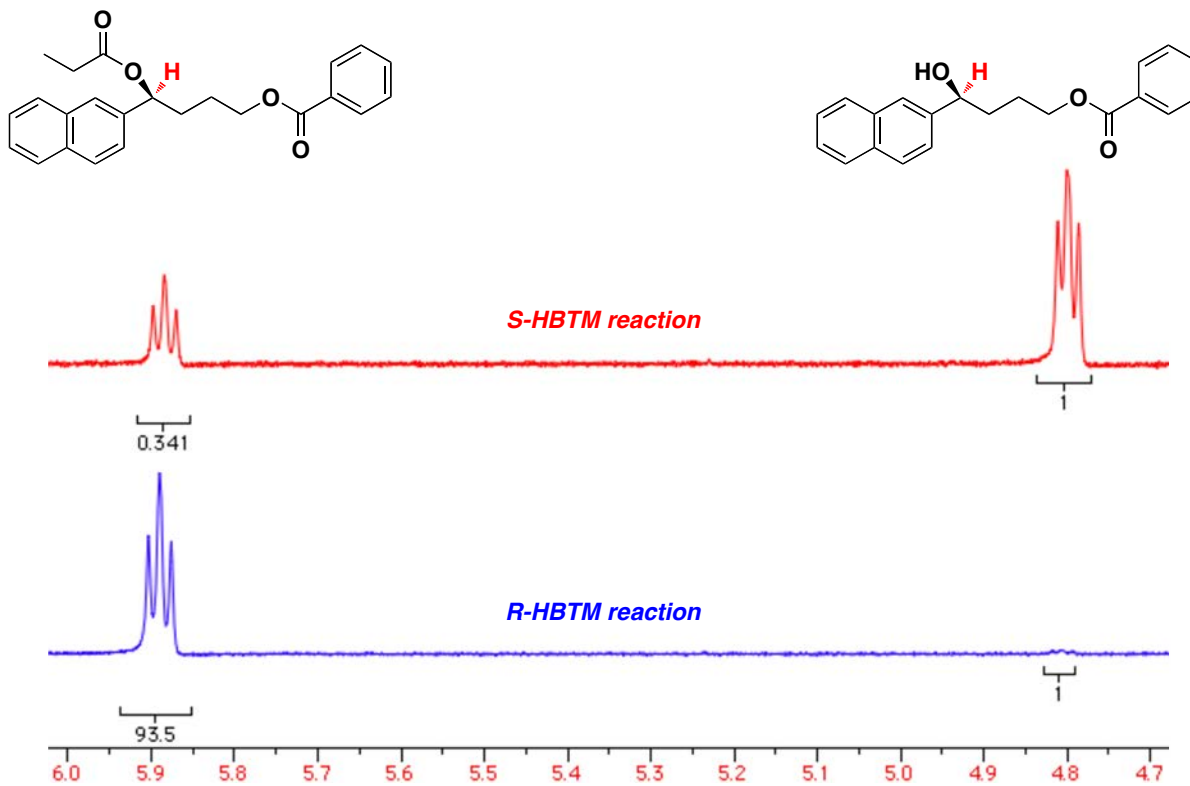
(b)

Entry 8. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



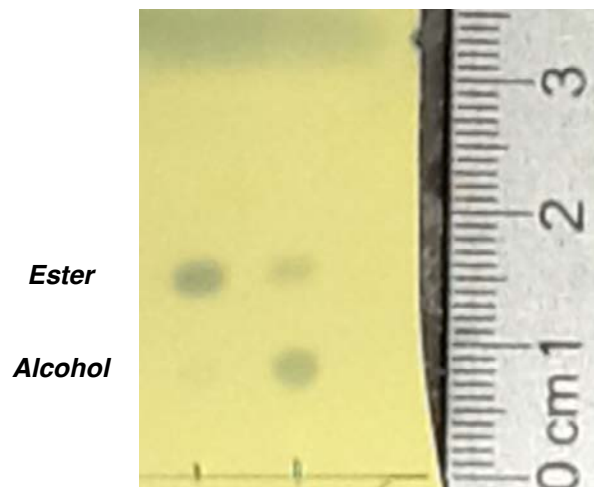
R-HBTM reaction *S*-HBTM reaction

(a)



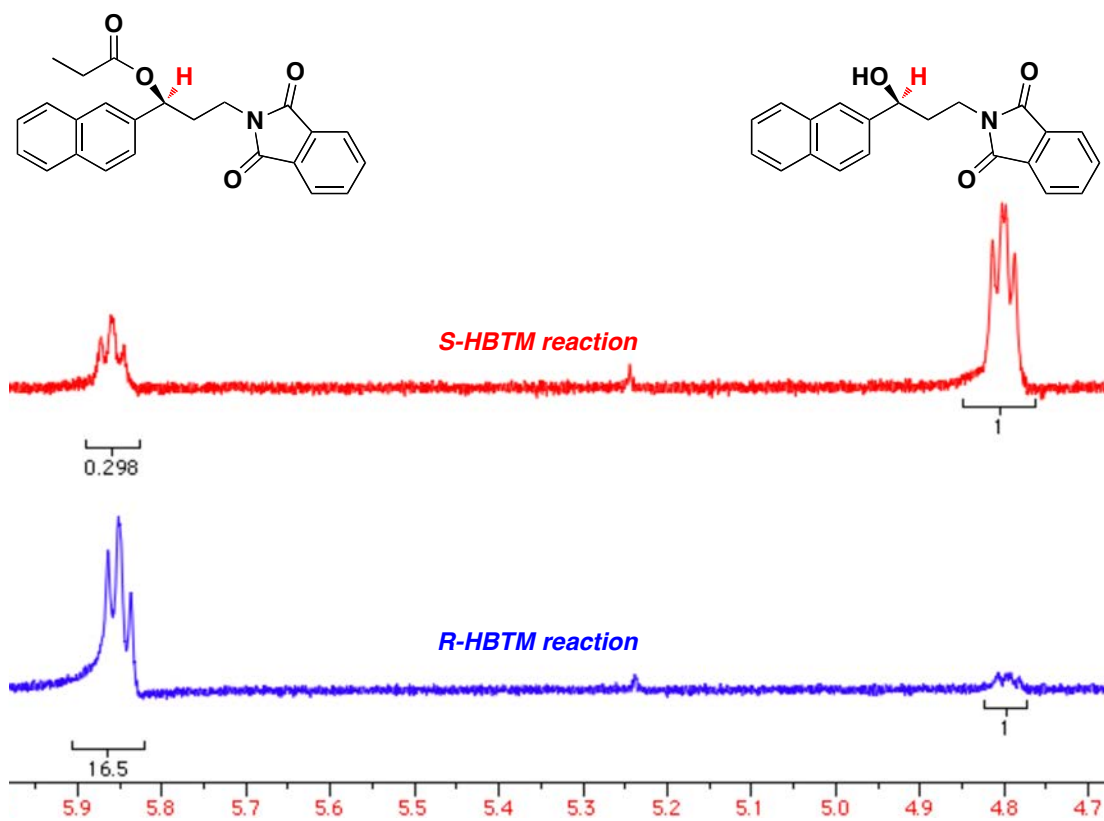
(b)

Entry 9. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



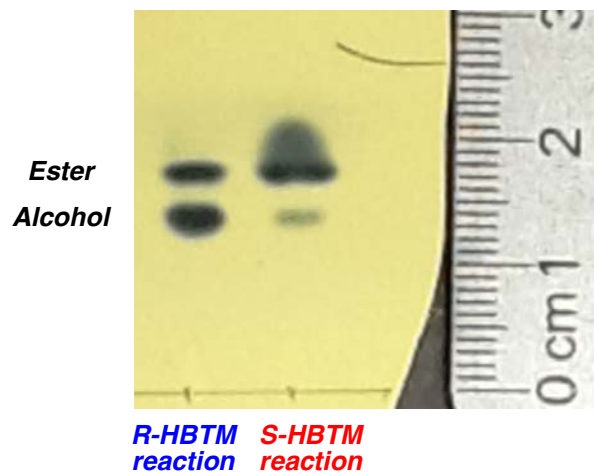
R-HBTM reaction
S-HBTM reaction

(a)

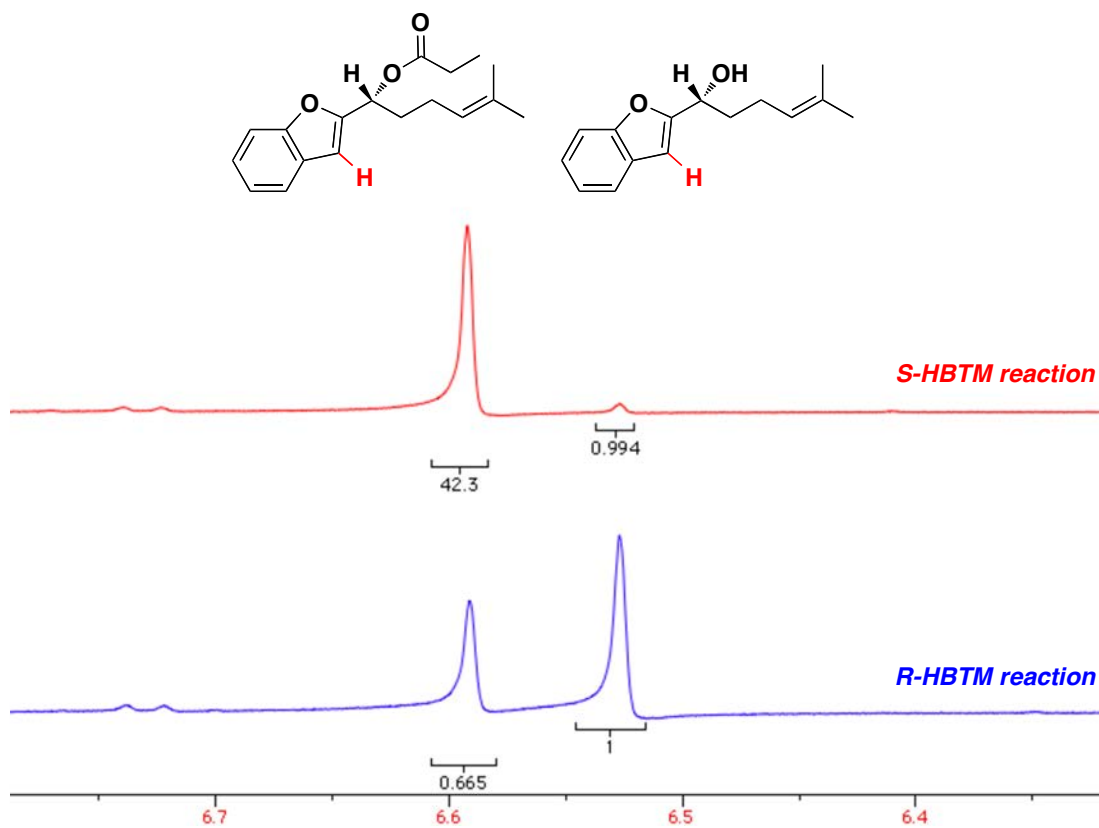


(b)

Entry 10. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.

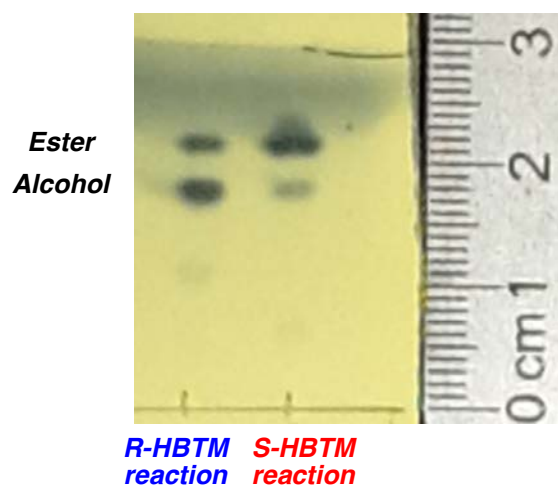


(a)

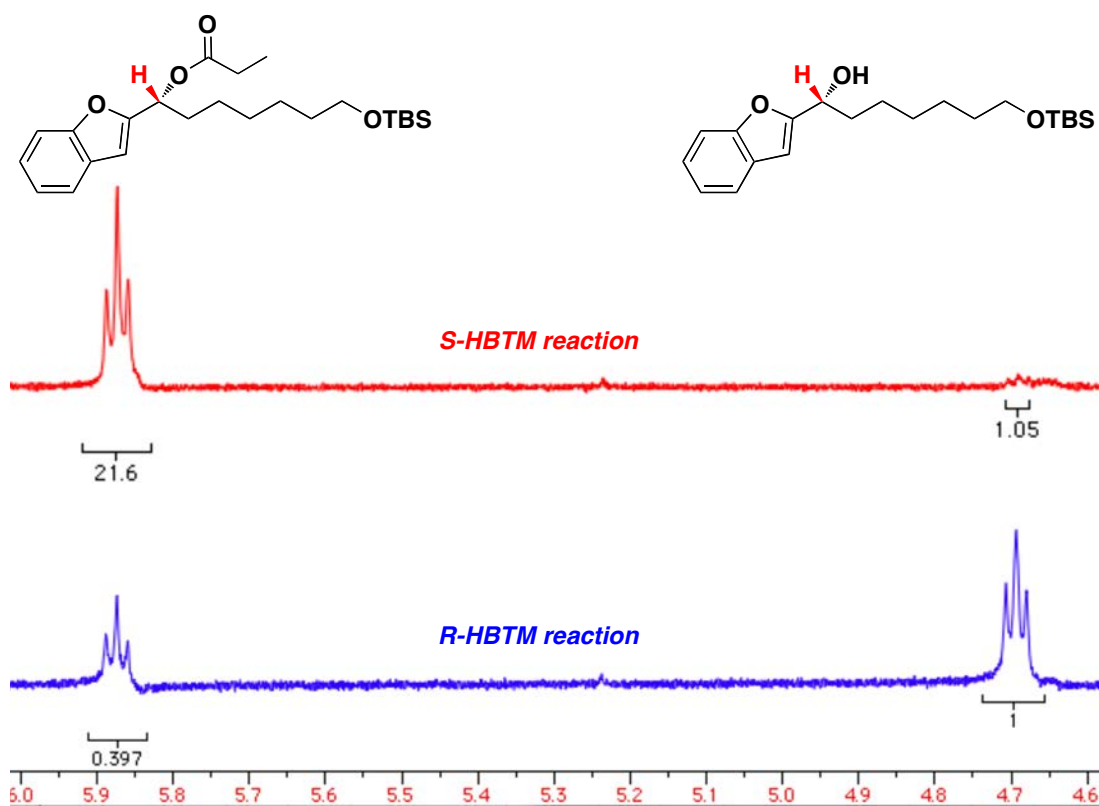


(b)

Entry 11. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.

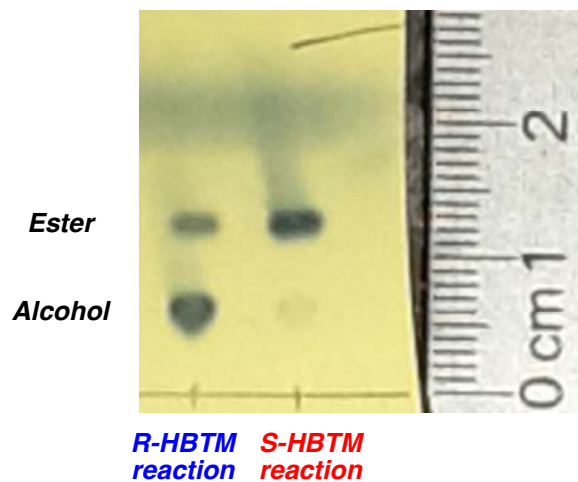


(a)

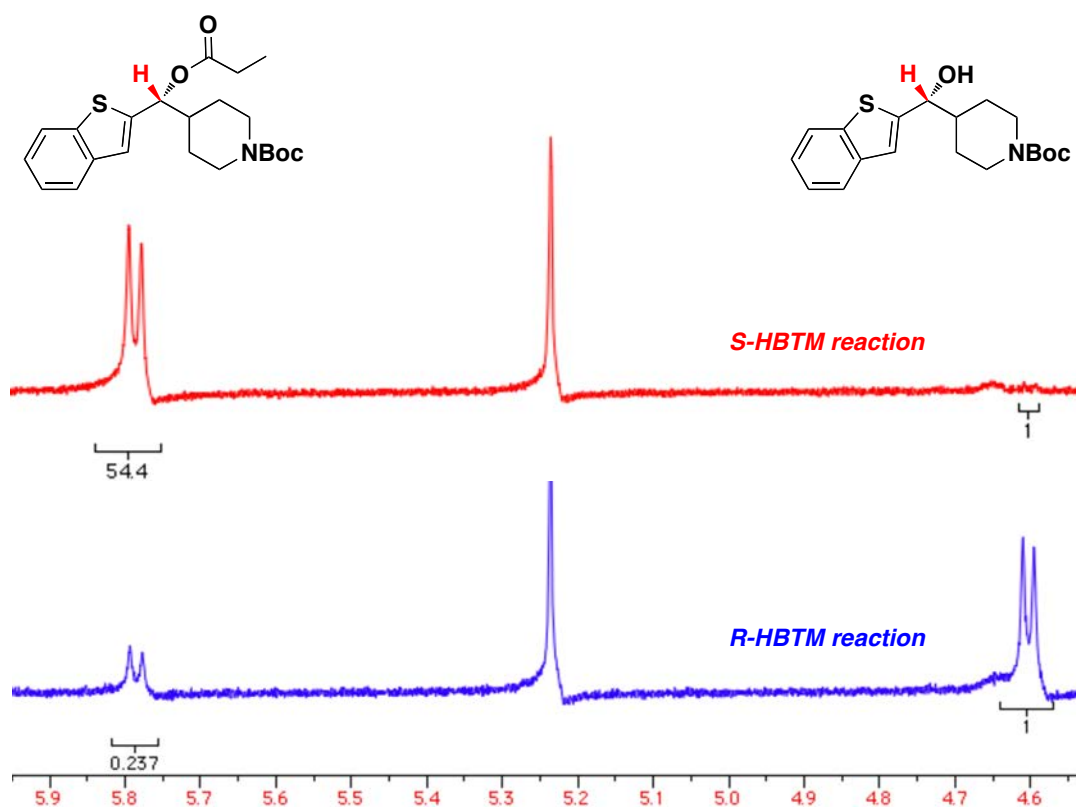


(b)

Entry 12. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.

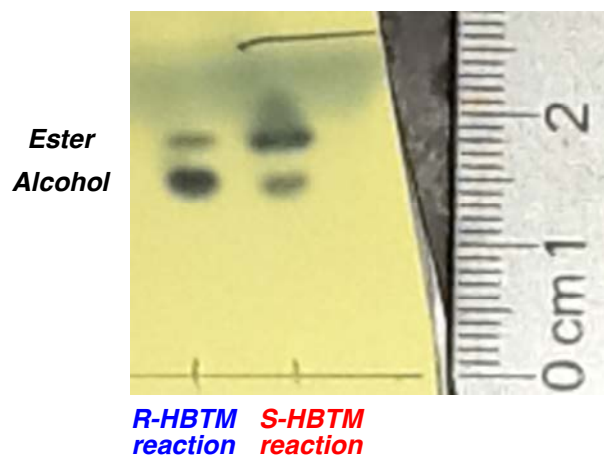


(a)

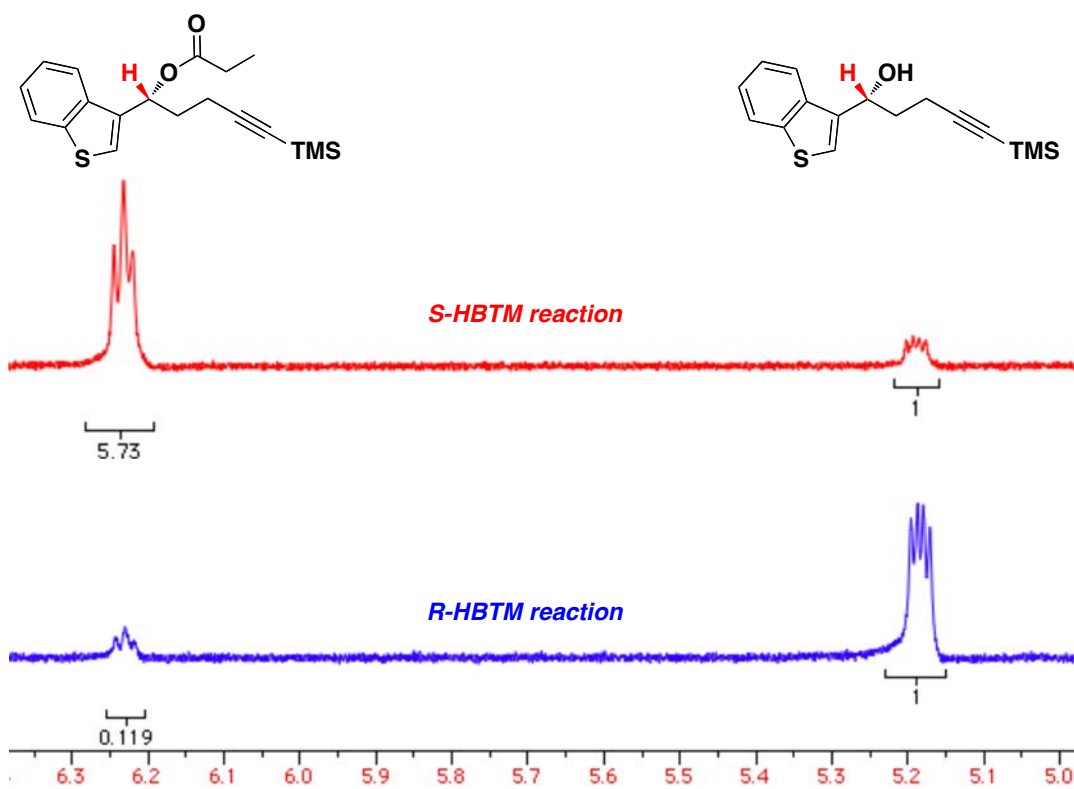


(b)

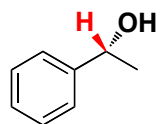
Entry 13. (a) TLC of parallel reactions (b) ^1H NMR Spectra of parallel reactions in CDCl_3 at 500 MHz.



(a)

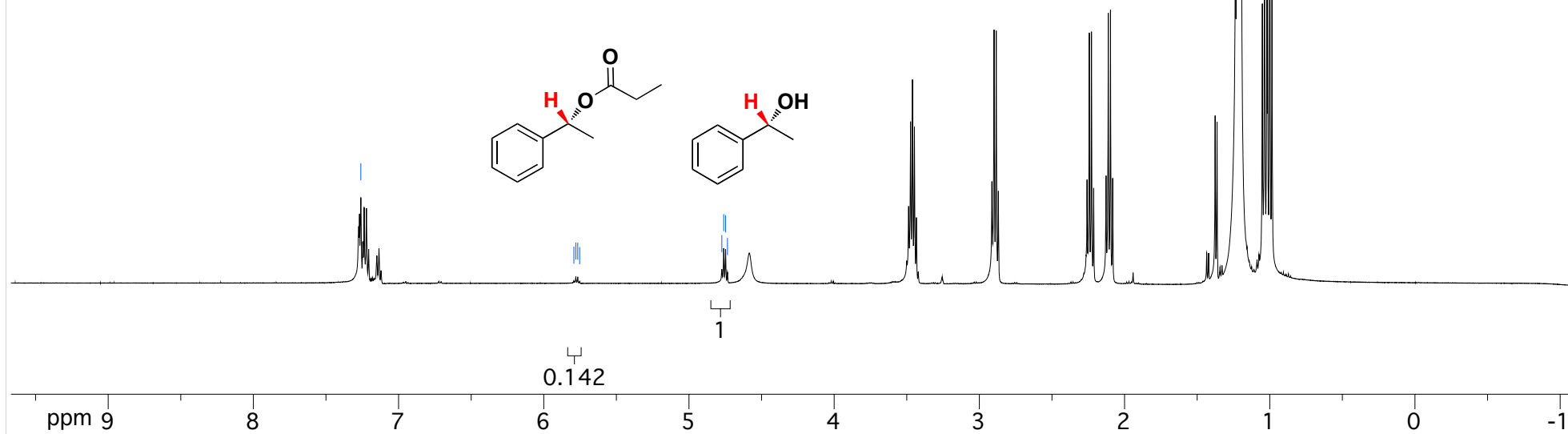


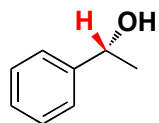
(b)

^1H NMR in CDCl_3 at 500 MHz

Entry 1 with R-HBTM

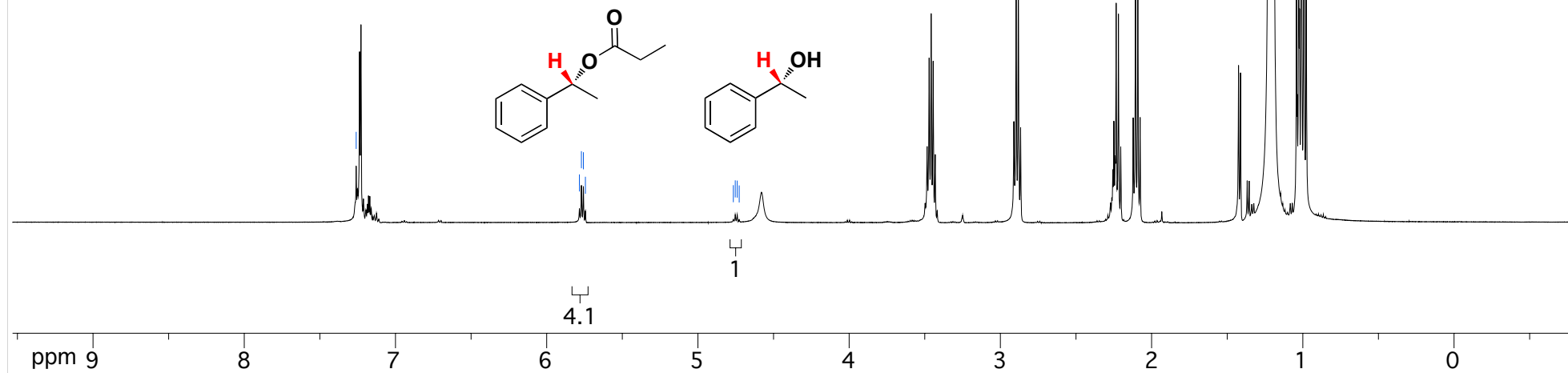
7.260

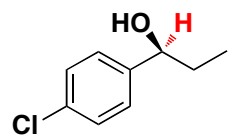
5.793
5.779
5.766
5.7534.774
4.761
4.748
4.735

^1H NMR in CDCl_3 at 500 MHz

Entry 1 with S-HBTM

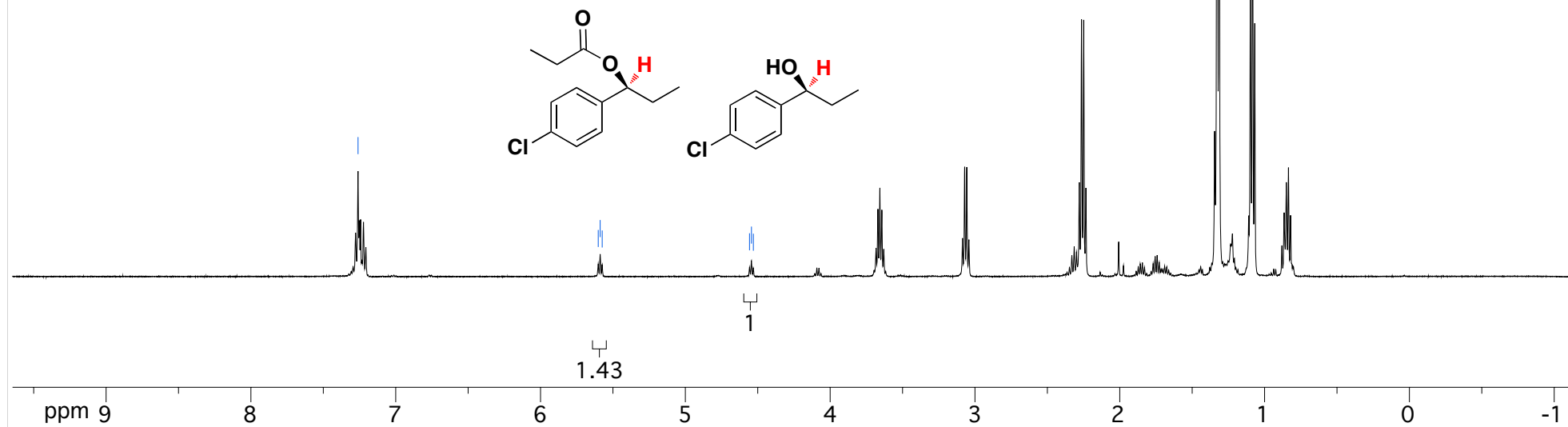
7.260

5.784
5.770
5.757
5.7444.766
4.753
4.740
4.727

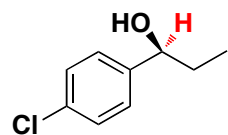
^1H NMR in CDCl_3 at 500 MHz

Entry 2 with R-HBTM

7.260

5.601
5.588
5.5744.557
4.543
4.530

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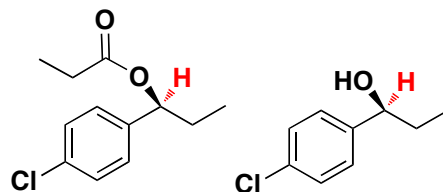
 ^1H NMR in CDCl_3 at 500 MHz

Entry 2 with R-HBTM

7.260

5.601
5.588
5.5744.557
4.543
4.530

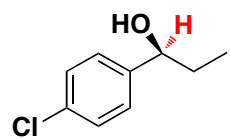
Crude NMR before concentration to remove methanol

Crude NMR after concentration and adding 0.500 mL CDCl_3

1.43

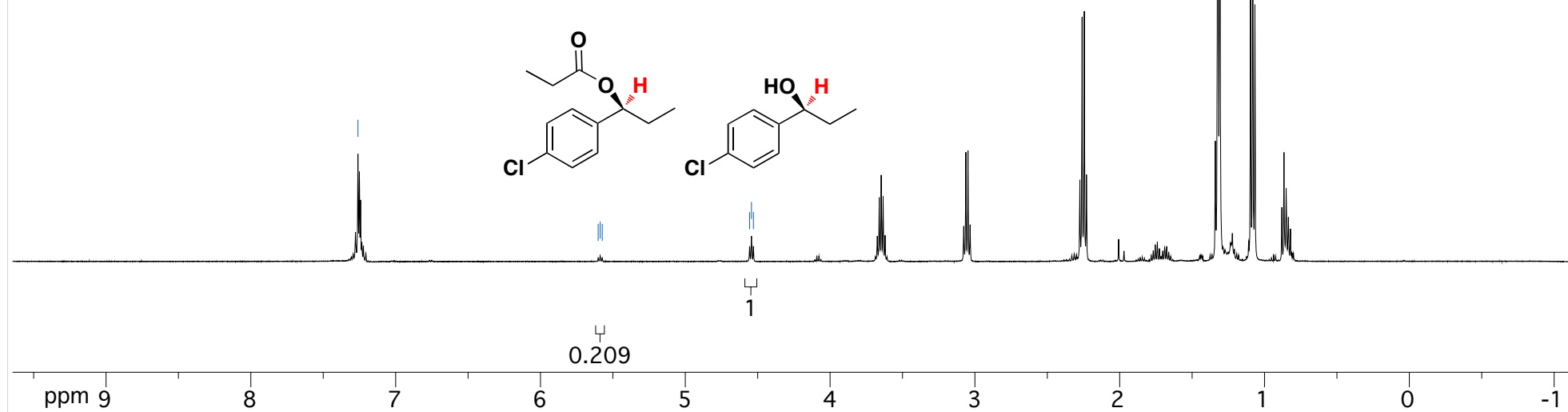
1

ppm 9 8 7 6 5 4 3 2 1 0 -1

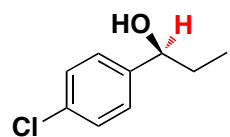
^1H NMR in CDCl_3 at 500 MHz

Entry 2 with S-HBTM

7.260

5.602
5.588
5.5734.556
4.543
4.529

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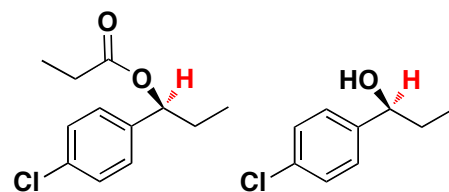
 ^1H NMR in CDCl_3 at 500 MHz

Entry 2 with S-HBTM

7.260

5.602
5.588
5.5734.556
4.543
4.529

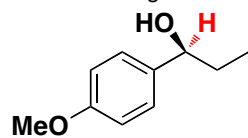
Crude NMR before concentration to remove methanol

Crude NMR after concentration and adding 0.500 mL CDCl_3

0.209

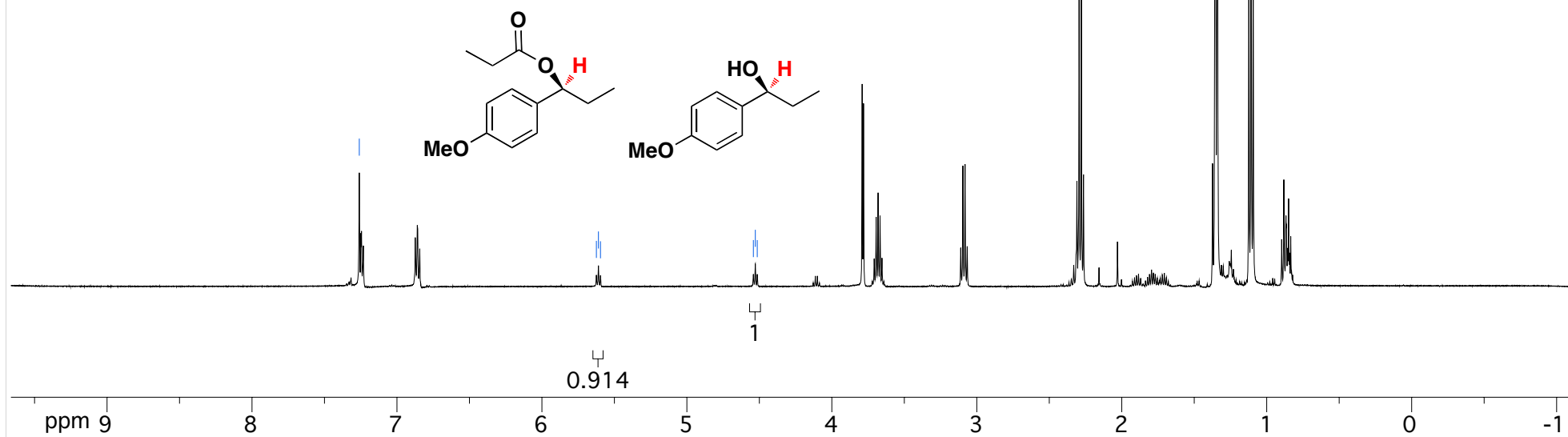
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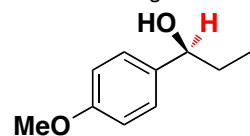
^1H NMR in CDCl_3 at 500 MHz

Entry 3 with R-HBTM

7.260

5.624
5.610
5.5964.541
4.528
4.514

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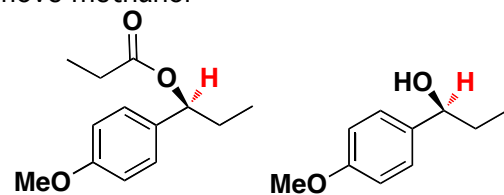
 ^1H NMR in CDCl_3 at 500 MHz

Entry 3 with R-HBTM

7.260

5.624
5.610
5.5964.541
4.528
4.514

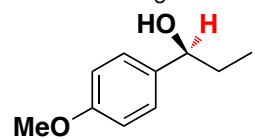
Crude NMR before concentration to remove methanol

Crude NMR after concentration and adding 0.500 mL CDCl_3

0.914

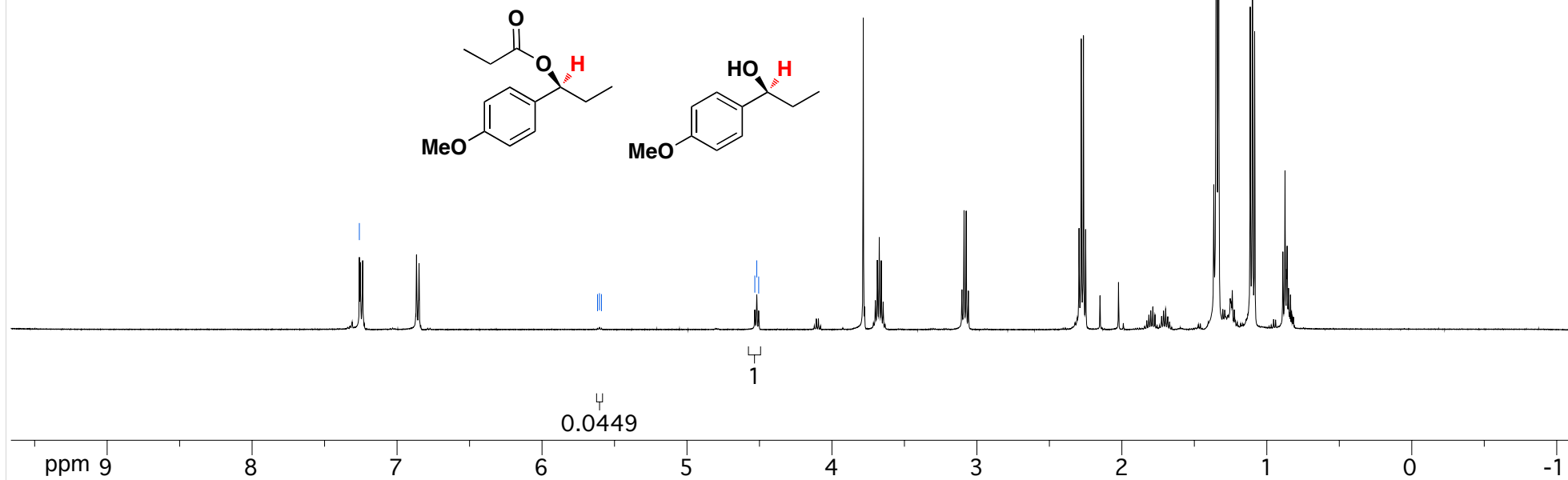
1

ppm 9 8 7 6 5 4 3 2 1 0 -1

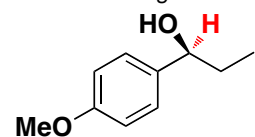
^1H NMR in CDCl_3 at 500 MHz

Entry 3 with S-HBTM

7.260

5.617
5.604
5.5904.532
4.519
4.505

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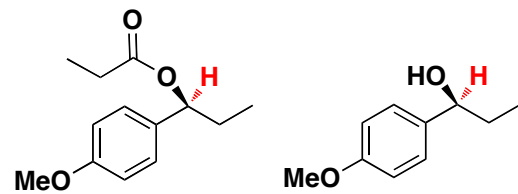
 ^1H NMR in CDCl_3 at 500 MHz

Entry 3 with S-HBTM

7.260

5.617
5.604
5.5904.532
4.519
4.505

Crude NMR before concentration to remove methanol

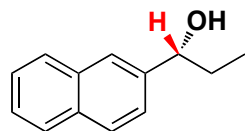
Crude NMR after concentration and adding 0.500 mL CDCl_3

0.0449

1

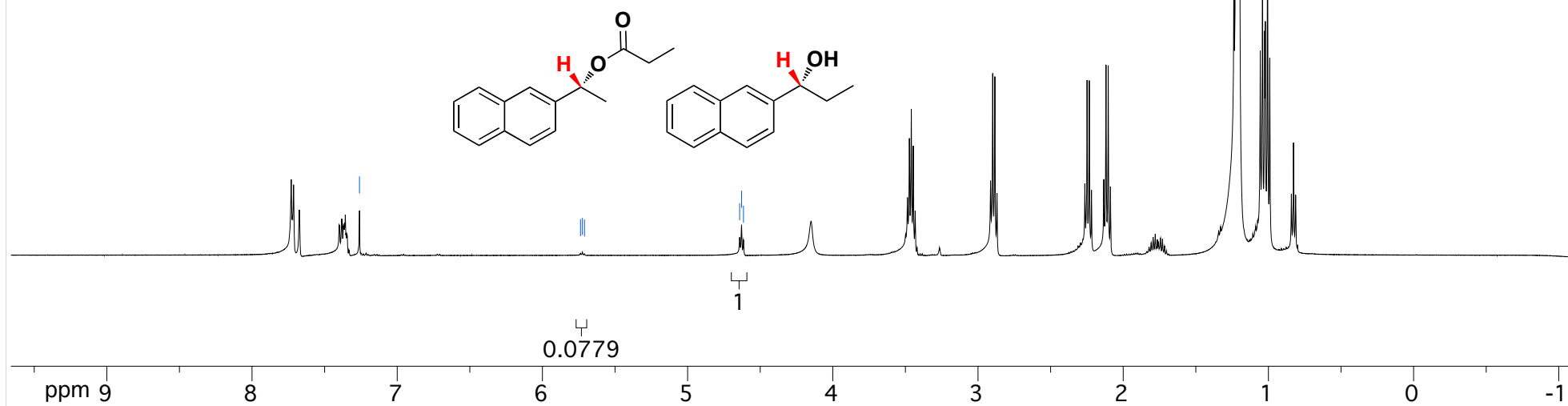
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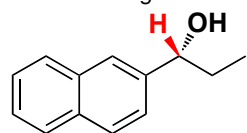
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 ^1H NMR in CDCl_3 at 500 MHz

Entry 4 with R-HBTM

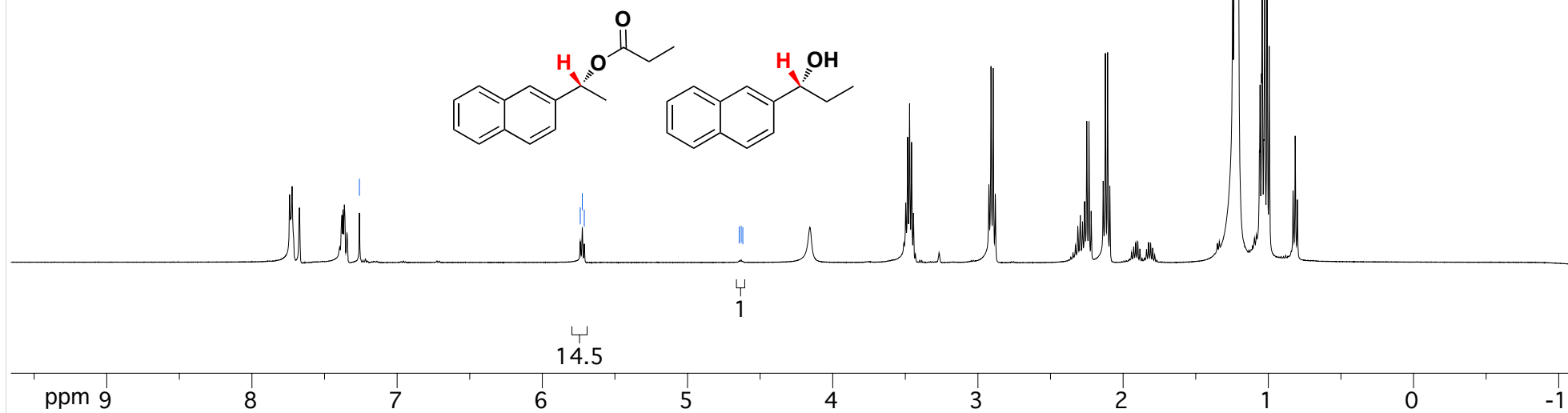
7.260

5.738
5.724
5.7104.642
4.628
4.615

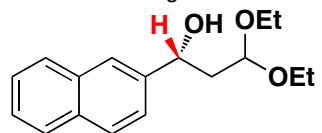
^1H NMR in CDCl_3 at 500 MHz

Entry 4 with S-HBTM

7.260

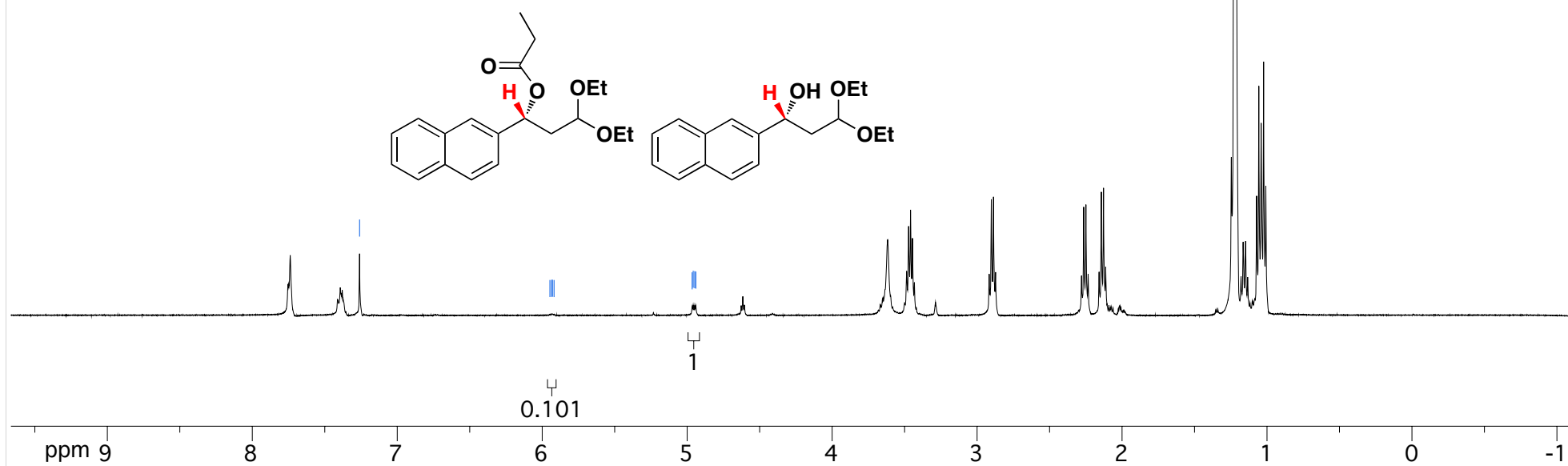
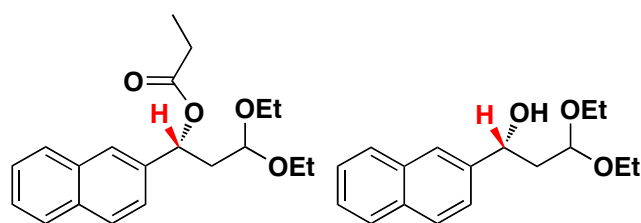
5.739
5.725
5.7114.644
4.631
4.619

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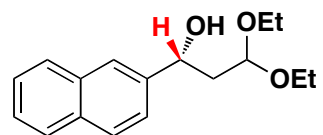
 ^1H NMR in CDCl_3 at 500 MHz

Entry 5 with R-HBTM

7.260

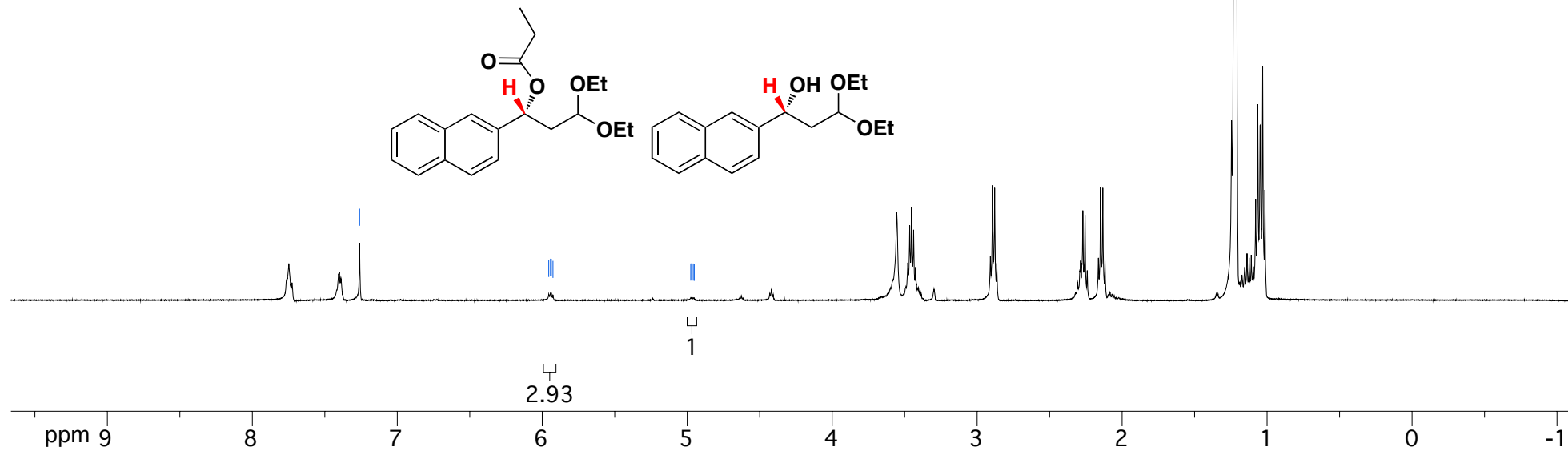
5.947
5.935
5.930
5.9184.966
4.958
4.948
4.940

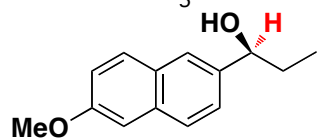
Wagner and Rychnovsky

 ^1H NMR in CDCl_3 at 500 MHz

Entry 5 with S-HBTM

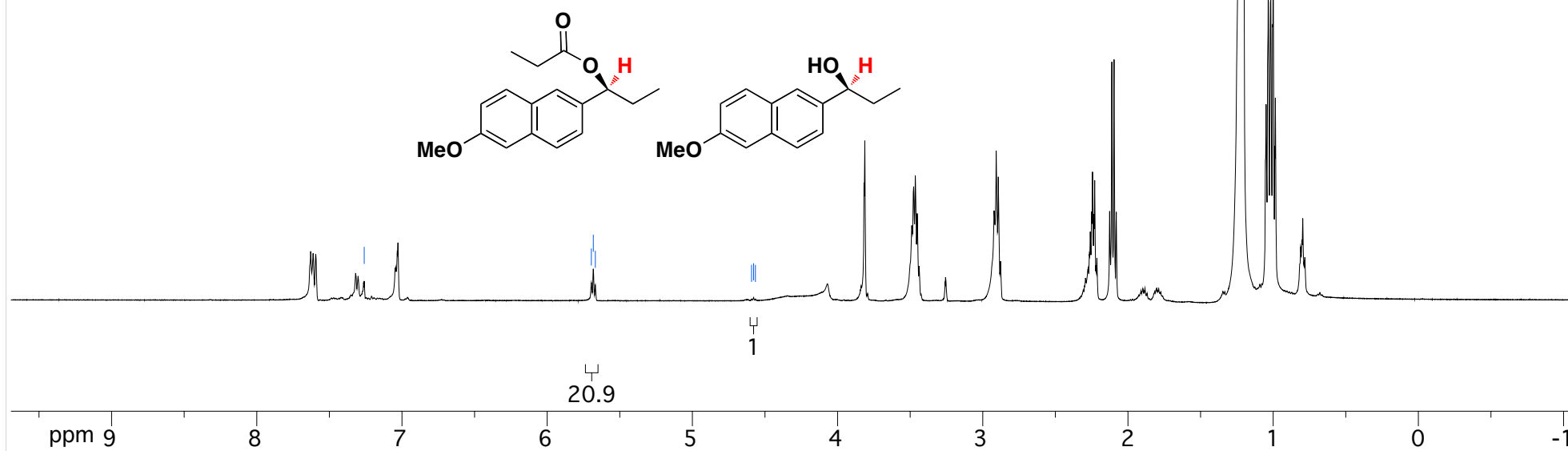
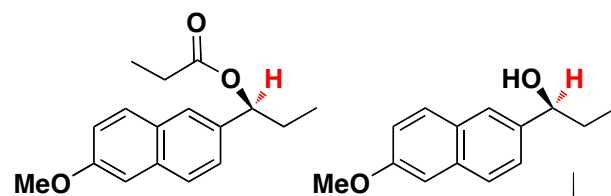
7.260

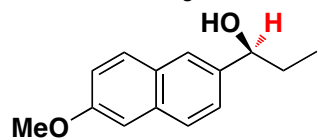
5.955
5.943
5.937
5.9264.976
4.969
4.959
4.952

^1H NMR in CDCl_3 at 500 MHz

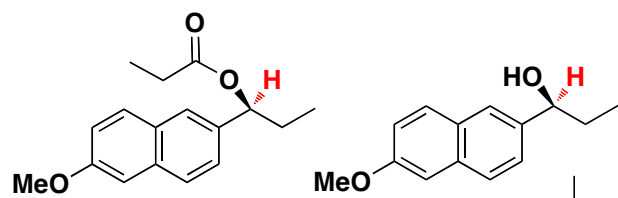
Entry 6 with R-HBTM

7.260

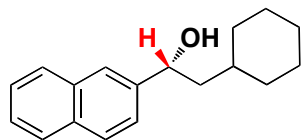
5.696
5.682
5.6684.592
4.578
4.566

^1H NMR in CDCl_3 at 500 MHz

Entry 6 with S-HBTM

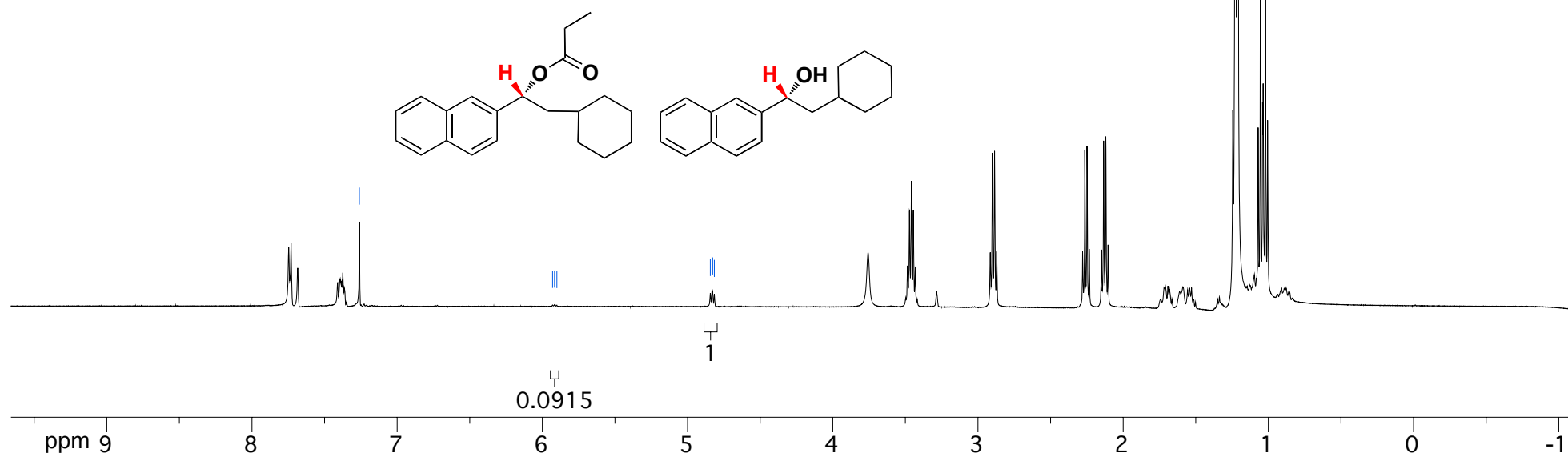
7.260
7.2595.710
5.696
5.6824.608
4.594
4.581

Wagner and Rychnovsky

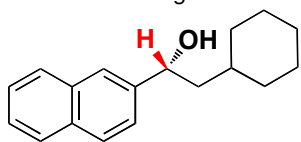
 ^1H NMR in CDCl_3 at 500 MHz

Entry 7 with R-HBTM

7.260

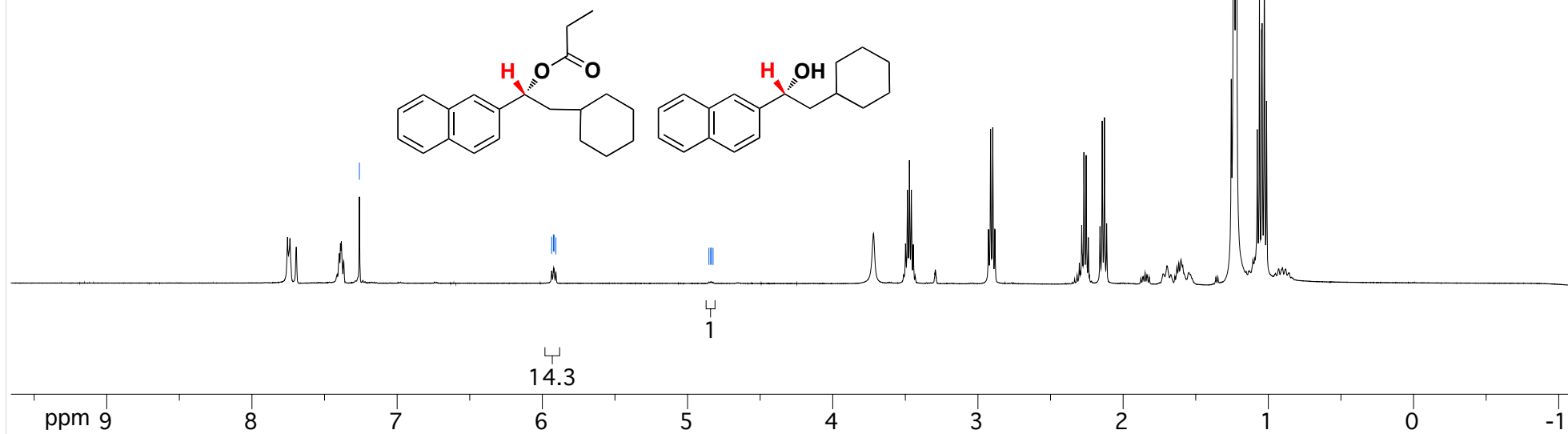
5.929
5.916
5.912
5.9004.843
4.832
4.827
4.816

Wagner and Rychnovsky

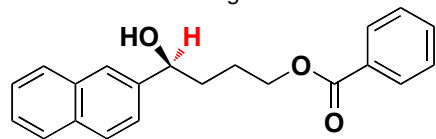
 ^1H NMR in CDCl_3 at 500 MHz

Entry 7 with S-HBTM

7.260

5.936
5.924
5.919
5.9074.853
4.841
4.836
4.824

Wagner and Rychnovsky

 ^1H NMR in CDCl_3 at 500 MHz

Entry 8 with R-HBTM

7.260

5.904

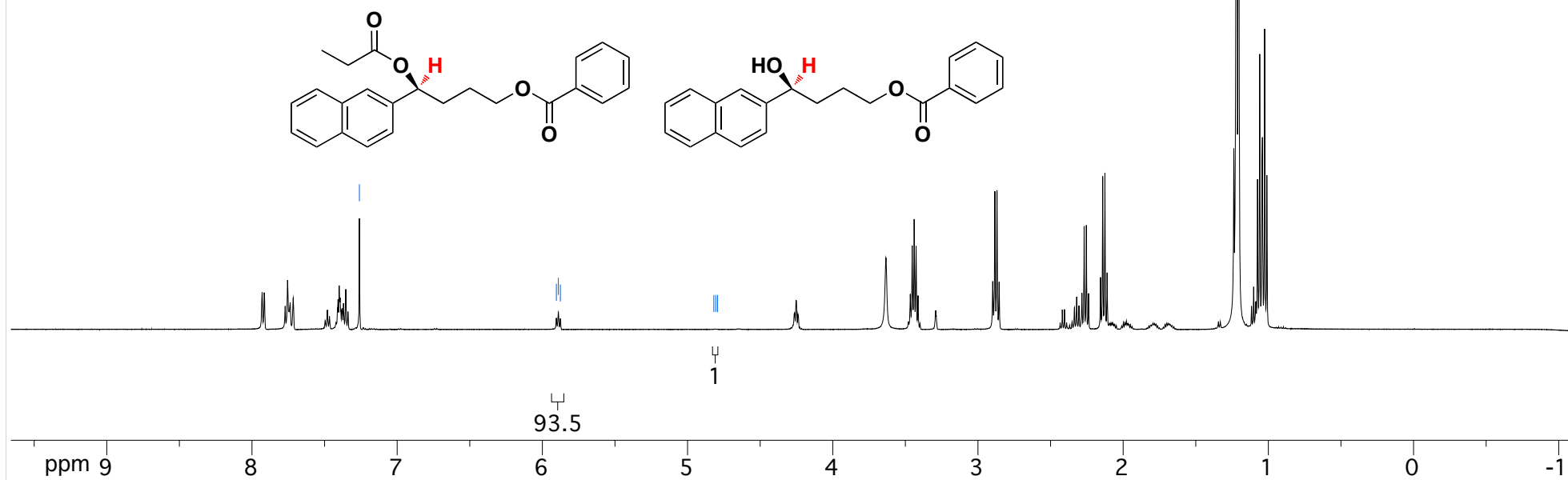
5.890

5.876

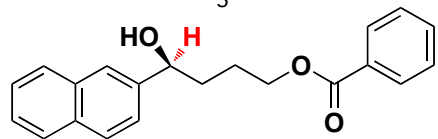
4.819

4.806

4.793

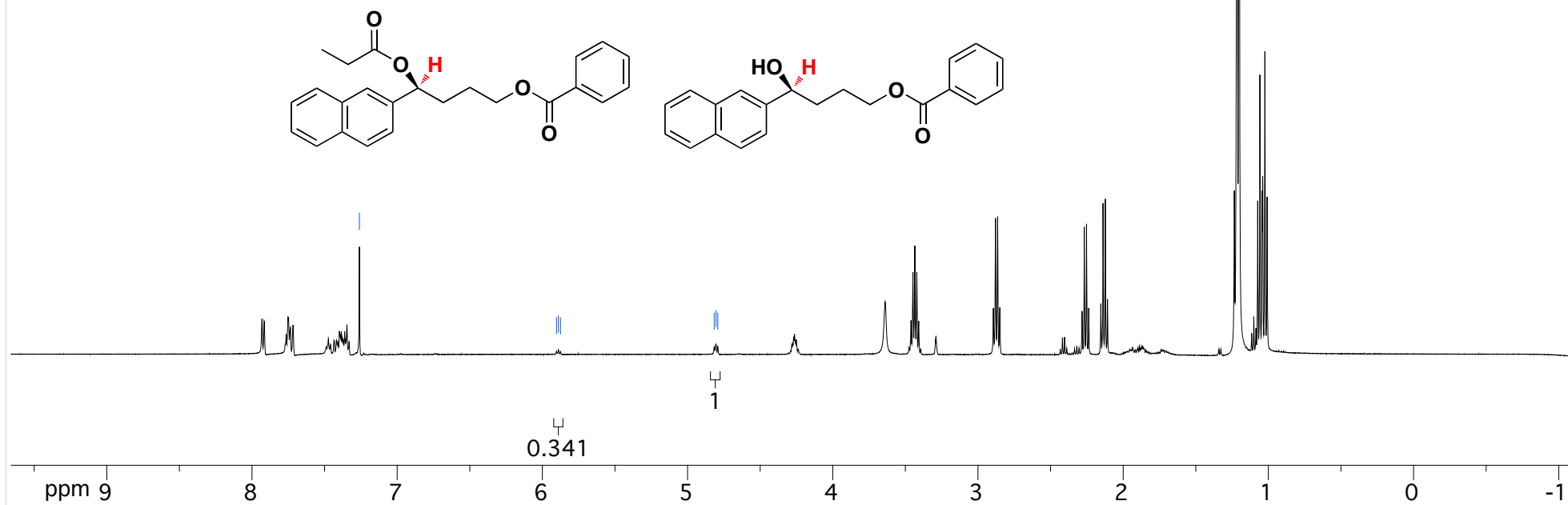


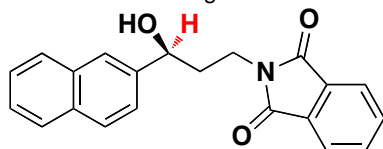
Wagner and Rychnovsky

 ^1H NMR in CDCl_3 at 500 MHz

Entry 8 with S-HBTM

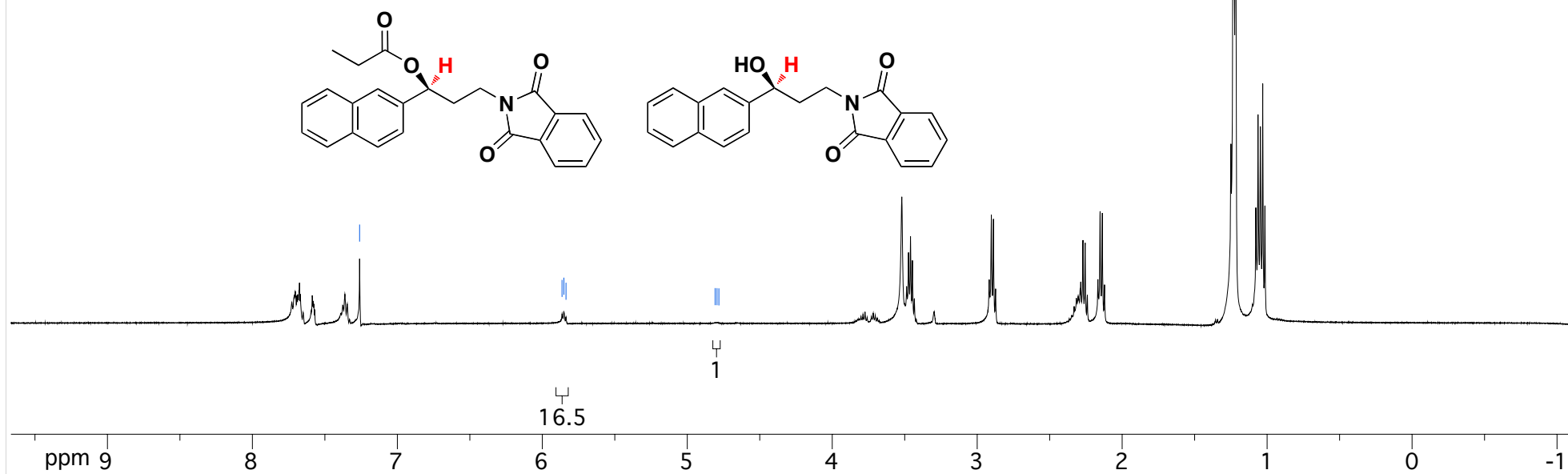
7.260

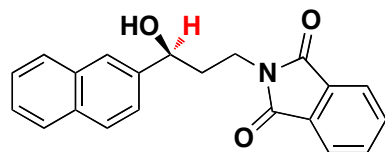
5.902
5.889
5.8754.817
4.804
4.792

^1H NMR in CDCl_3 at 500 MHz

Entry 9 with R-HBTM

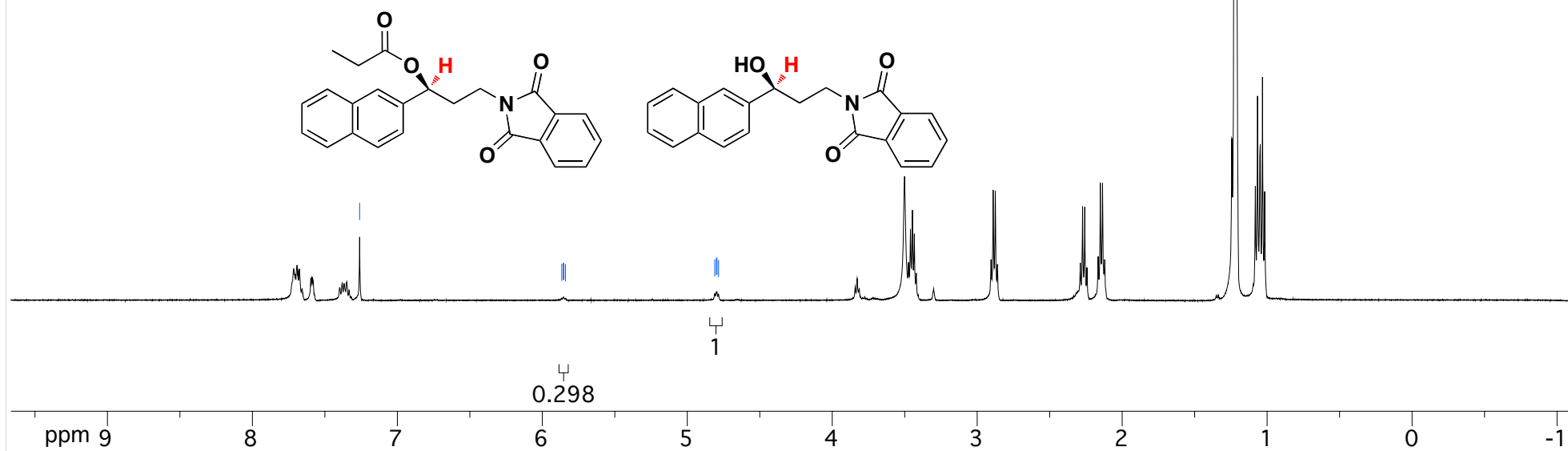
7.260

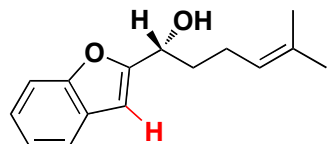
5.864
5.851
5.8374.808
4.795
4.782

^1H NMR in CDCl_3 at 500 MHz

Entry 9 with S-HBTM

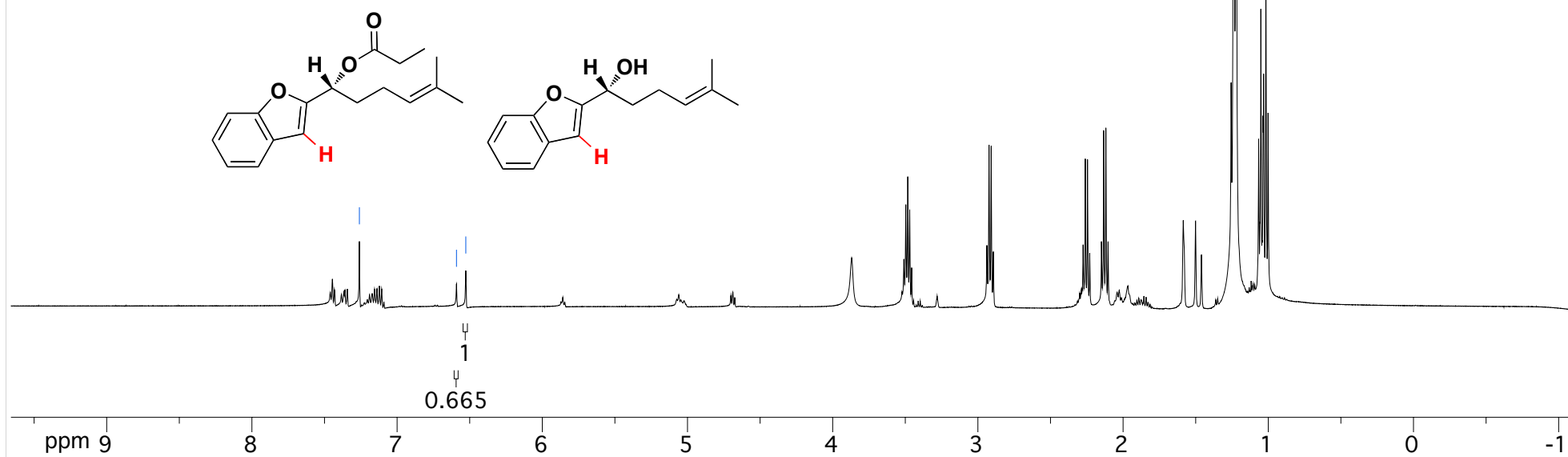
7.260

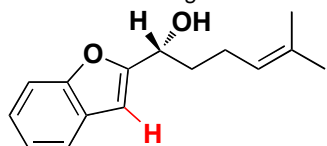
5.866
5.855
5.852
5.8404.810
4.799
4.795
4.784

^1H NMR in CDCl_3 at 500 MHz

Entry 10 with R-HBTM

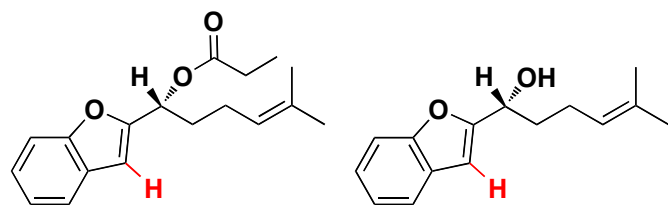
7.260

6.591
6.527

^1H NMR in CDCl_3 at 500 MHz

Entry 10 with S-HBTM

7.260

6.596
6.532

0.994

42.3

ppm 9

8

7

6

5

4

3

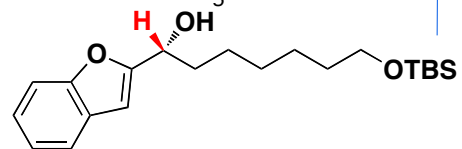
2

1

0

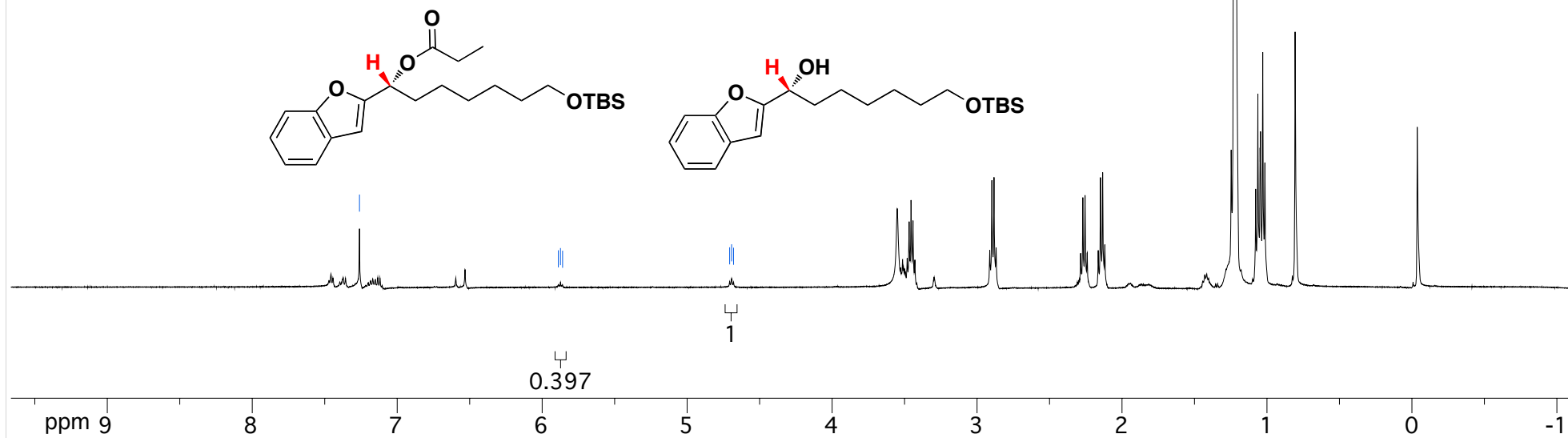
-1

Wagner and Rychnovsky

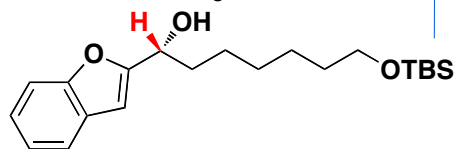
 ^1H NMR in CDCl_3 at 500 MHz

Entry 11 with R-HBTM

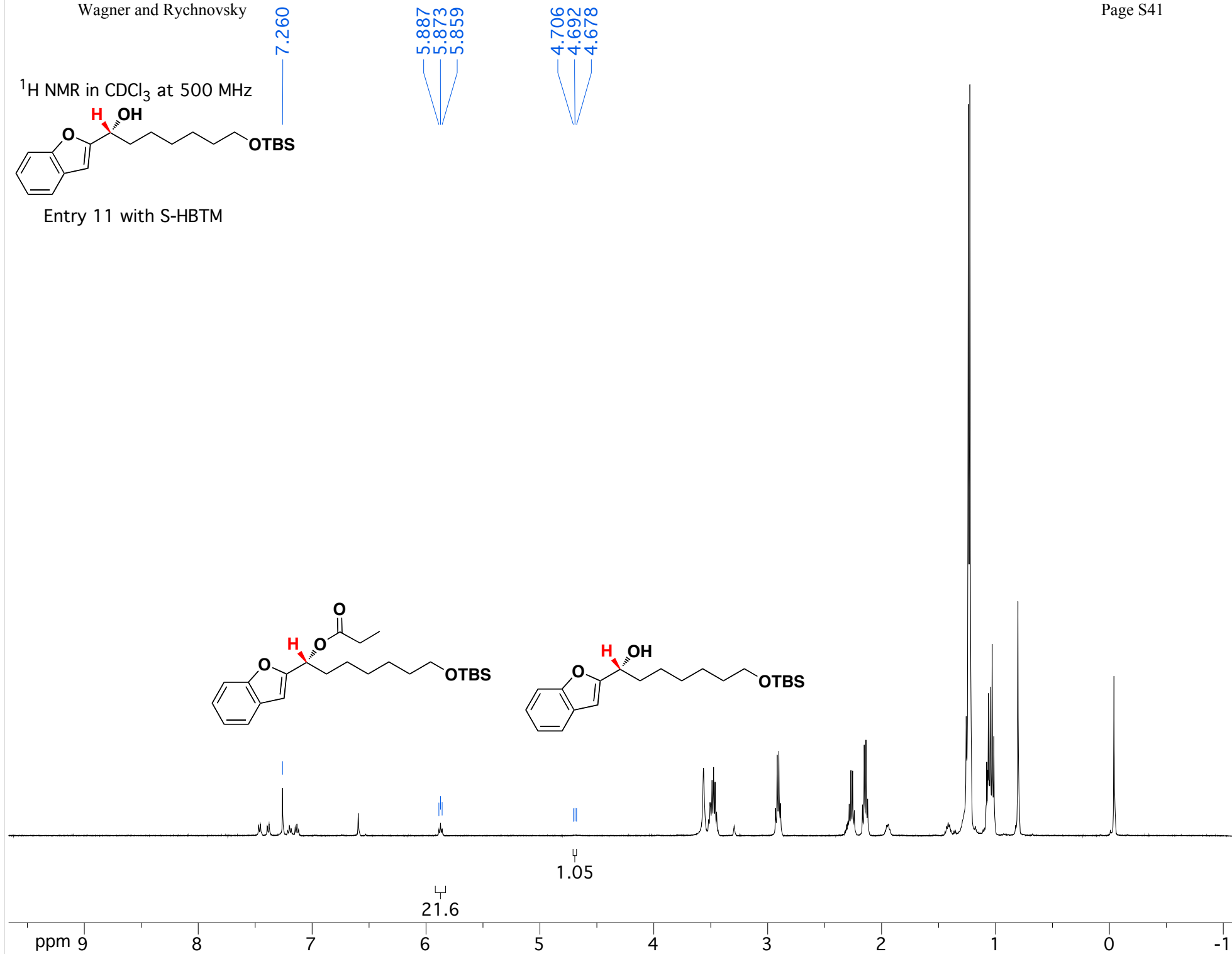
7.260

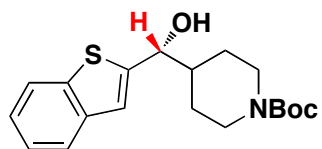
5.888
5.874
5.8594.707
4.693
4.680

Wagner and Rychnovsky

 ^1H NMR in CDCl_3 at 500 MHz

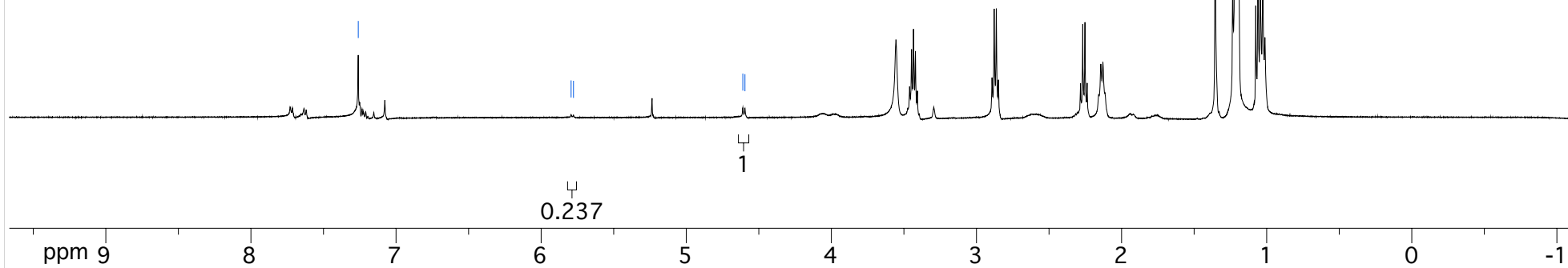
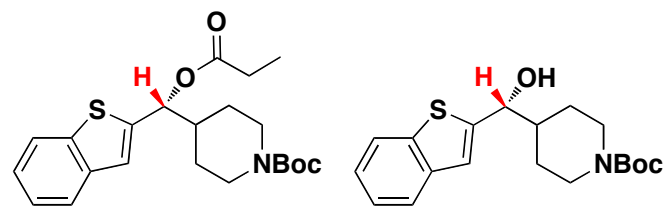
Entry 11 with S-HBTM

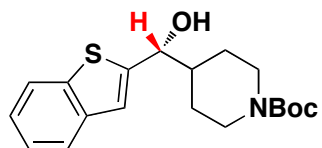


^1H NMR in CDCl_3 at 500 MHz

Entry 12 with R-HBTM

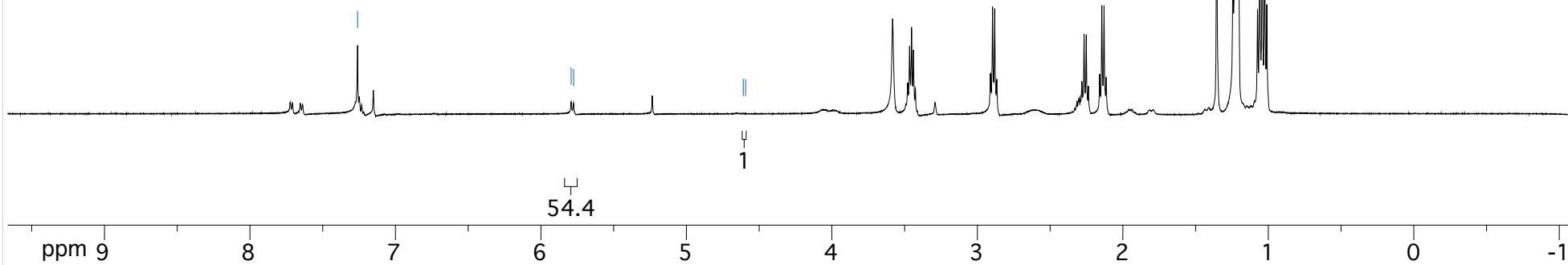
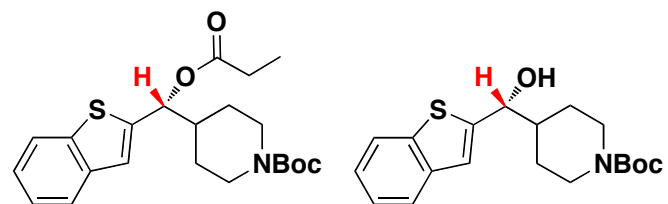
7.260

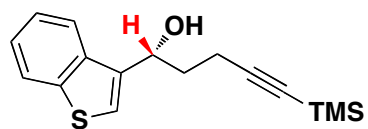
5.794
5.7774.610
4.596

¹H NMR in CDCl₃ at 500 MHz

Entry 12 with S-HBTM

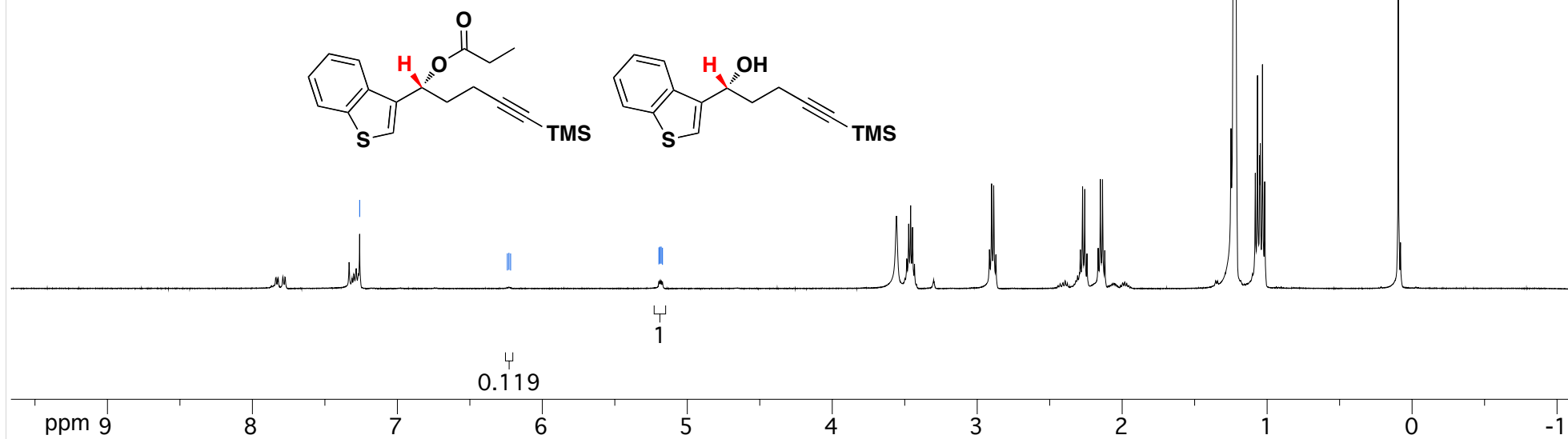
7.260

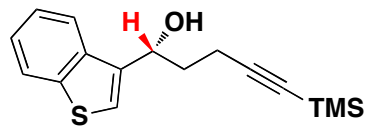
5.792
5.7764.609
4.593

^1H NMR in CDCl_3 at 500 MHz

Entry 13 with R-HBTM

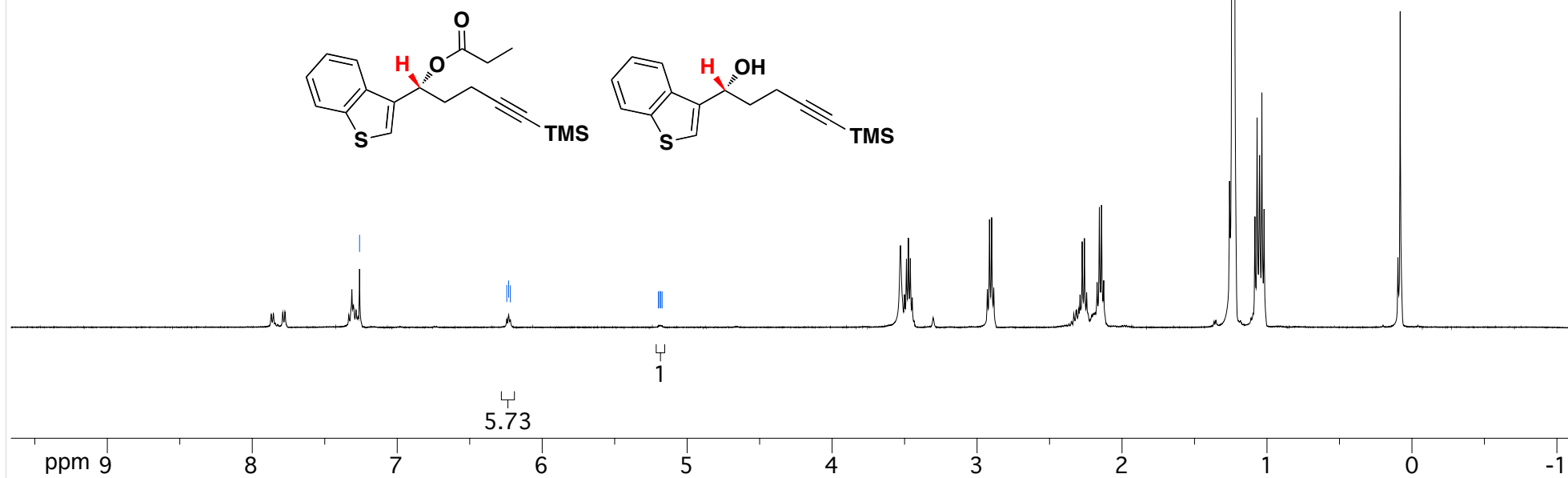
7.260

6.241
6.230
6.2195.196
5.188
5.180
5.171

¹H NMR in CDCl₃ at 500 MHz

Entry 13 with S-HBTM

7.260

6.244
6.231
6.2195.199
5.190
5.181
5.172

Chiral Analytical Traces for Entries 1-3, 6-7.

HPLC determinations of enantiopurity were performed on an Agilent 1100 Series instrument using isopropyl alcohol (OmniSolv®) and n-hexane (HPLC grade, 95% min) with Daicel™ columns.

Entry 1 (S47-S48):

HPLC analysis (Chiralcel® OD with OD guard, 5.0% *i*PrOH/n-Hexane, 1.0 mL/min) indicated >99:<1 er; t_R (major) = 9.6 minutes, t_R (minor) = 12.4 minutes.

Entry 2 (S49-S50):

HPLC analysis (Chiralpak® OB-H with OB guard, 5.0% *i*PrOH/n-Hexane, 0.80 mL/min) indicated 85:15 er; t_R (major) = 9.6 minutes, t_R (minor) = 11.1 minutes.

Entry 3 (S51-S52):

HPLC analysis (Chiralpak® AD with AD guard, 5.0% *i*PrOH/n-Hexane, 1.0 mL/min) indicated 98:2 er; t_R (major) = 12.0 minutes, t_R (minor) = 13.5 minutes.

SFC determinations of enantiopurity were performed on a Berger Analytical instrument using a Daicel™ column (100 bar, 50 °C, 215 nm).

Entry 6 (S53-S54):

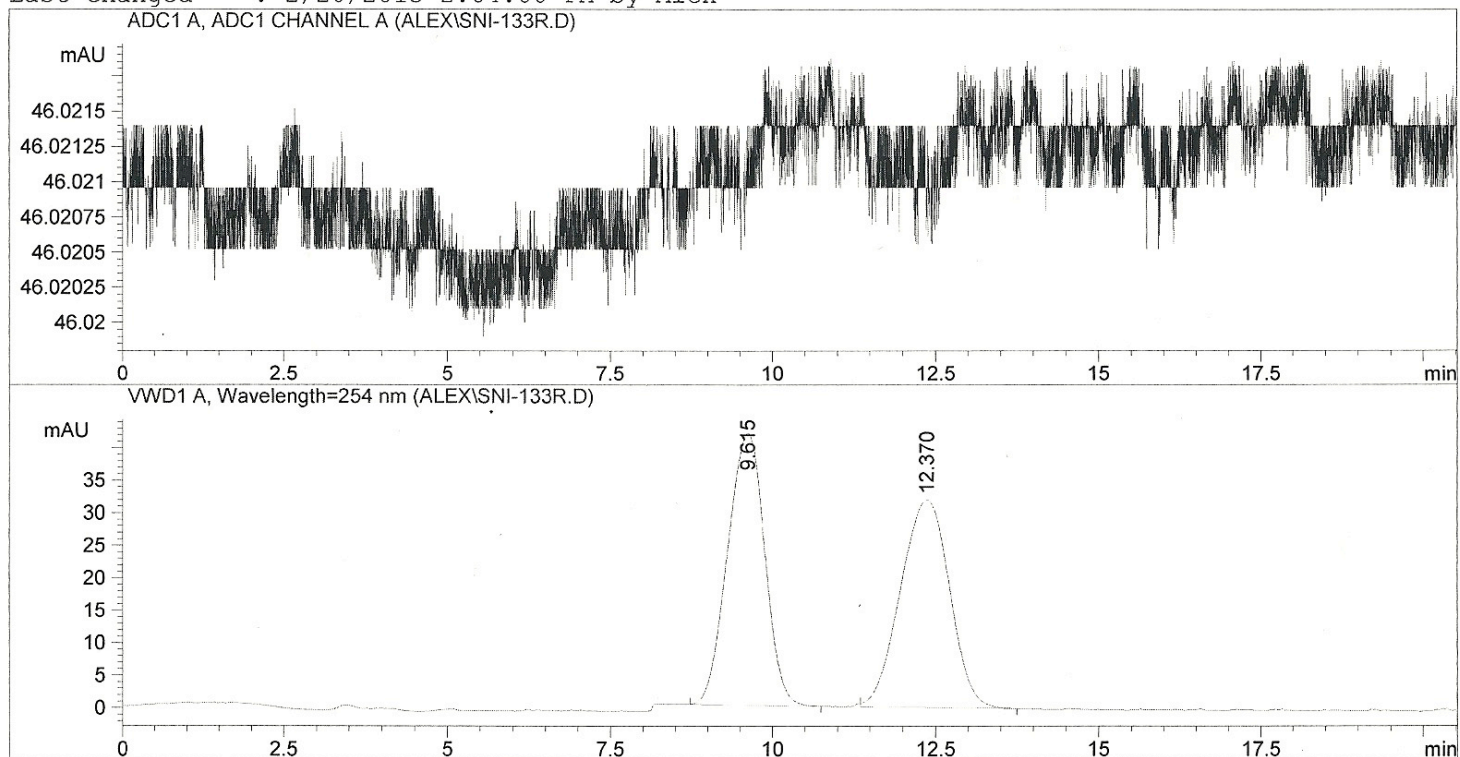
SFC analysis (Chiralpak® OD-H, 12.5% *i*PrOH, 3.0 mL/min) indicated 97:3 er; t_R (major) = 5.3 minutes, t_R (minor) = 6.5 minutes.

Entry 7 (S55-S56):

SFC analysis (Chiralpak® AS-H, 3.0% *i*PrOH, 3.0 mL/min) indicated 98:2 er; t_R (minor) = 12.2 minutes, t_R (major) = 12.8 minutes.

Column OD with guard 5% iPrOH/Hexanes 1.00 mL 38 bar

=====
 Injection Date : 9/28/2012 4:44:16 PM
 Sample Name : SN-1-133-RacOH Location : Vial 42
 Acq. Operator : Alex Inj Volume : 10 µl
 Acq. Method : C:\HPCHEM\1\METHODS\ALEX.M
 Last changed : 9/28/2012 4:44:17 PM by Alex
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\ALEX.M
 Last changed : 2/20/2013 2:04:00 PM by Alex



=====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Sample Amount : 10.00000 [ng/ul] (not used in calc.)
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: ADC1 A, ADC1 CHANNEL A

Signal 2: VWD1 A, Wavelength=254 nm

| Peak # | RetTime [min] | Type | Width [min] | Area mAU *s | Height [mAU] | Area % |
|--------|---------------|------|-------------|-------------|--------------|---------|
| 1 | 9.615 | BB | 0.6355 | 1675.50940 | 41.89484 | 50.0590 |
| 2 | 12.370 | VB | 0.7585 | 1671.56042 | 31.97147 | 49.9410 |

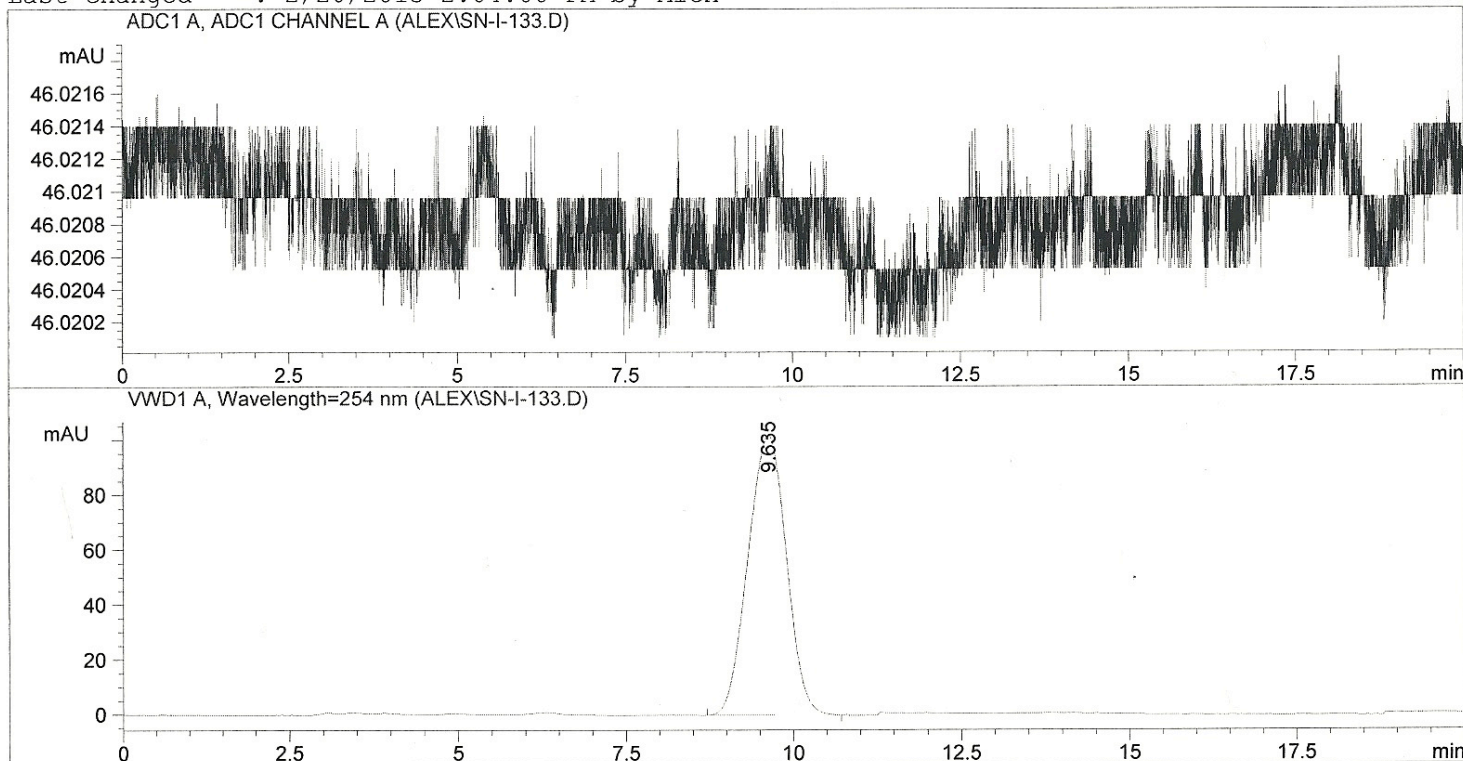
Totals : 3347.06982 73.86631

Results obtained with enhanced integrator!

=====
 *** End of Report ***

Column OD with guard 5% iPrOH/Hexanes 1.00 mL 38 bar

=====
 Injection Date : 9/28/2012 4:22:04 PM
 Sample Name : SN-1-133-OH Location : Vial 41
 Acq. Operator : Alex Inj Volume : 10 µl
 Acq. Method : C:\HPCHEM\1\METHODS\ALEX.M
 Last changed : 9/28/2012 3:34:42 PM by Alex
 (modified after loading)
 Analysis Method : C:\HPCHEM\1\METHODS\ALEX.M
 Last changed : 2/20/2013 2:04:00 PM by Alex



=====
 Area Percent Report
 =====

Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Sample Amount : 10.00000 [ng/ul] (not used in calc.)
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: ADC1 A, ADC1 CHANNEL A

Signal 2: VWD1 A, Wavelength=254 nm

| Peak # | RetTime [min] | Type | Width [min] | Area mAU | Height [mAU] | Area % |
|--------|---------------|------|-------------|------------|--------------|----------|
| 1 | 9.635 | BB | 0.6300 | 4094.94434 | 102.00067 | 100.0000 |

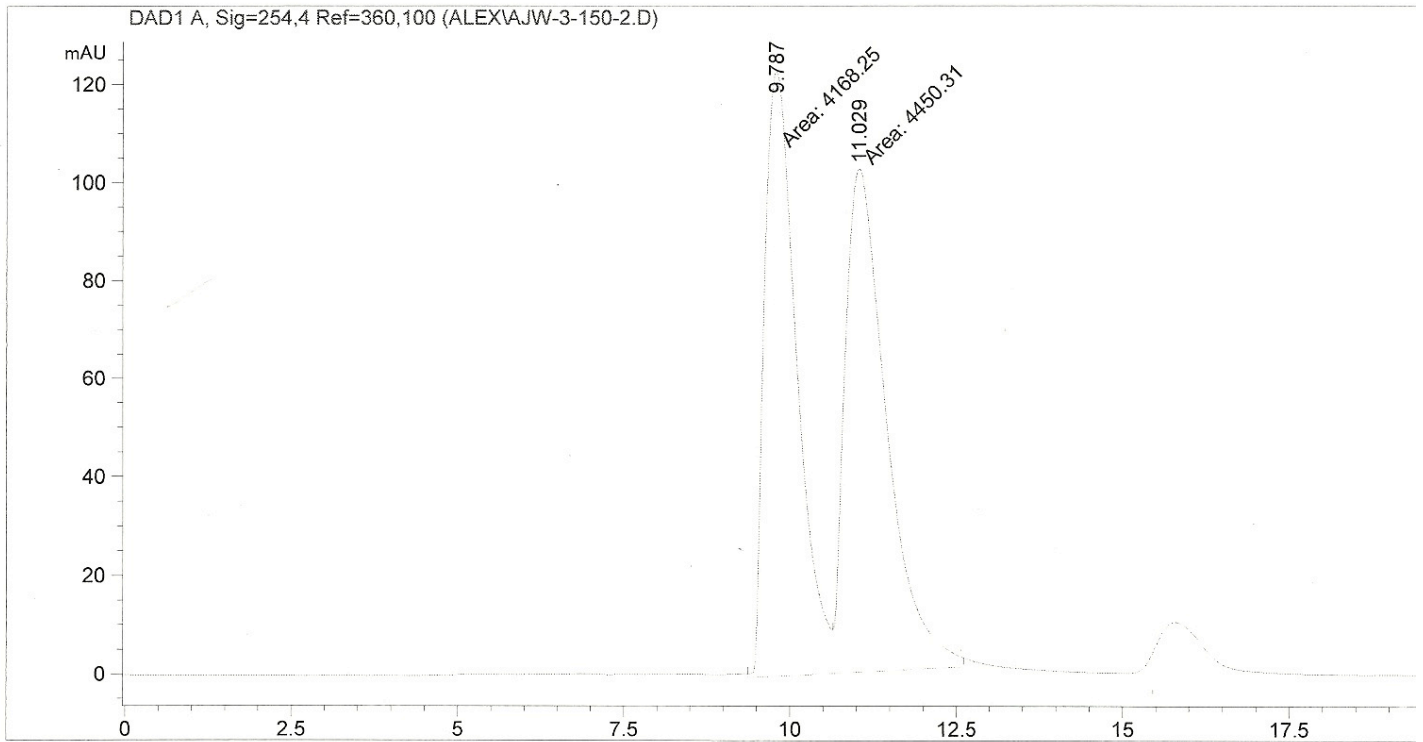
Totals : 4094.94434 102.00067

Results obtained with enhanced integrator!

=====
 *** End of Report ***

Wagner and Rychnovsky

=====
Acq. Operator : Alex
Acq. Instrument : Instrument 2 Location : Vial 61
Injection Date : 2/25/2013 2:32:16 PM Inj Volume : 30 µl
Acq. Method : C:\CHEM32\2\METHODS\JONLAM.M
Last changed : 2/25/2013 2:30:49 PM by Alex
(modified after loading)
Analysis Method : C:\CHEM32\2\DATA\JONLAM\JKL-04-154.D\DA.M (JONLAM.M)
Last changed : 2/25/2013 1:10:16 PM by Jon Lam
Sample Info : Chiralpak OB-H + OB Guard, 5% IPA/Hexanes, 0.8 ml/min,
42 bar



=====
Area Percent Report
=====

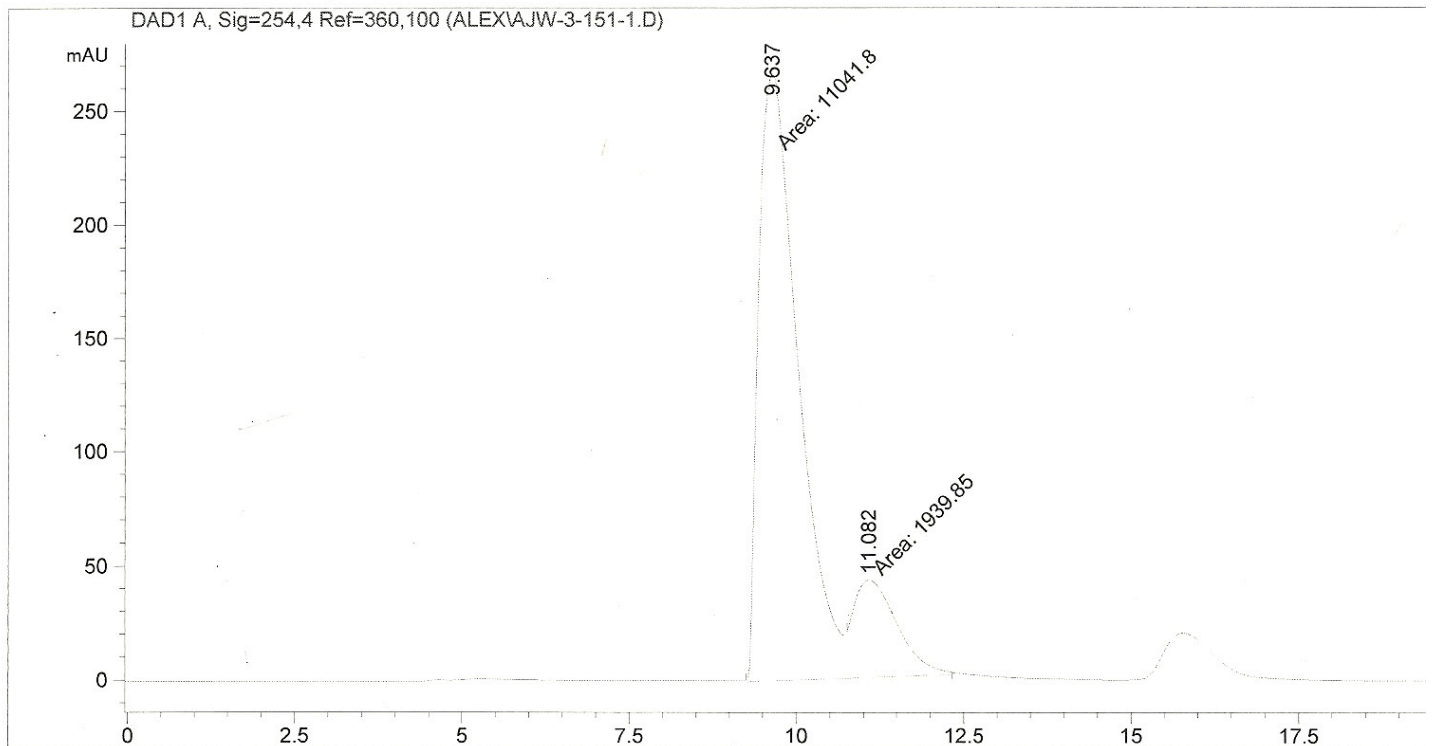
Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Sample Amount: : 1.00000 [ng/ul] (not used in calc.)
Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

| Peak # | RetTime [min] | Type | Width [min] | Area [mAU*s] | Height [mAU] | Area % |
|--------|---------------|------|-------------|--------------|--------------|---------|
| 1 | 9.787 | MF | 0.5648 | 4168.24658 | 123.00746 | 48.3636 |
| 2 | 11.029 | FM | 0.7247 | 4450.30811 | 102.34229 | 51.6364 |

Totals : 8618.55469 225.34975

=====
Acq. Operator : Alex
Acq. Instrument : Instrument 2
Injection Date : 2/25/2013 2:54:32 PM
Location : Vial 62
Inj Volume : 30 µl
Acq. Method : C:\CHEM32\2\METHODS\JONLAM.M
Last changed : 2/25/2013 2:52:37 PM by Alex
(modified after loading)
Analysis Method : C:\CHEM32\2\DATA\JONLAM\JKL-04-154.D\DA.M (JONLAM.M)
Last changed : 2/25/2013 1:10:16 PM by Jon Lam
Sample Info : Chiralpak OB-H + OB Guard, 5% IPA/Hexanes, 0.8 ml/min,
42 bar



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier: : 1.0000
Dilution: : 1.0000
Sample Amount: : 1.00000 [ng/ul] (not used in calc.)
Use Multiplier & Dilution Factor with ISTDs

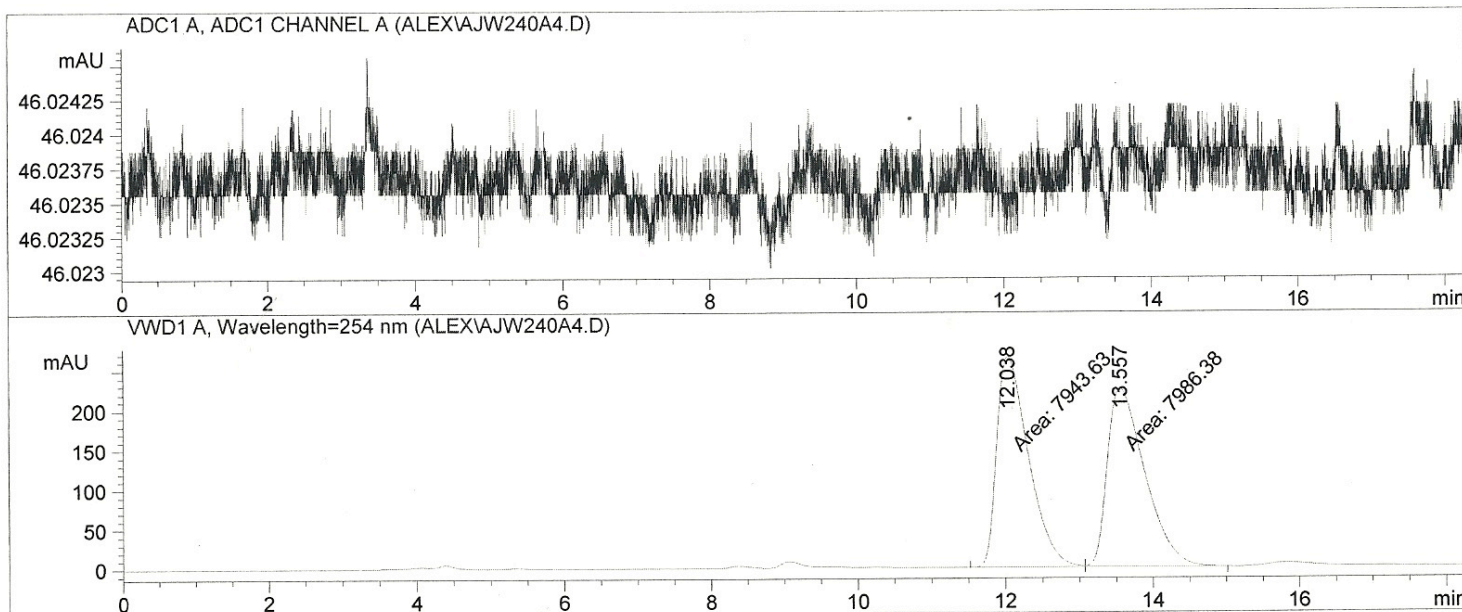
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

| Peak # | RetTime [min] | Type | Width [min] | Area [mAU*s] | Height [mAU] | Area % |
|--------|---------------|------|-------------|--------------|--------------|---------|
| 1 | 9.637 | MF | 0.6902 | 1.10418e4 | 266.61862 | 85.0570 |
| 2 | 11.082 | FM | 0.7567 | 1939.85461 | 42.72510 | 14.9430 |

Totals : 1.29817e4 309.34372

=====
Injection Date : 3/9/2012 2:18:48 PM
Sample Name : AJW-2-40A4 Location : Vial 3
Acq. Operator : Alex Inj Volume : 10 µl

Acq. Method : C:\HPCHEM\1\METHODS\ALEX.M
Last changed : 3/9/2012 12:49:39 PM by Alex
(modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\ALEX.M
Last changed : 3/9/2012 2:22:25 PM by Alex
(modified after loading)
=====



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Sample Amount : 10.00000 [ng/ul] (not used in calc.)
Use Multiplier & Dilution Factor with ISTDs

Signal 1: ADC1 A, ADC1 CHANNEL A

Signal 2: VWD1 A, Wavelength=254 nm

| Peak # | RetTime [min] | Type | Width [min] | Area mAU *s | Height [mAU] | Area % |
|--------|---------------|------|-------------|-------------|--------------|---------|
| 1 | 12.038 | MF | 0.4986 | 7943.62842 | 265.53699 | 49.8658 |
| 2 | 13.557 | FM | 0.5909 | 7986.37891 | 225.27487 | 50.1342 |

Totals : 1.59300e4 490.81186

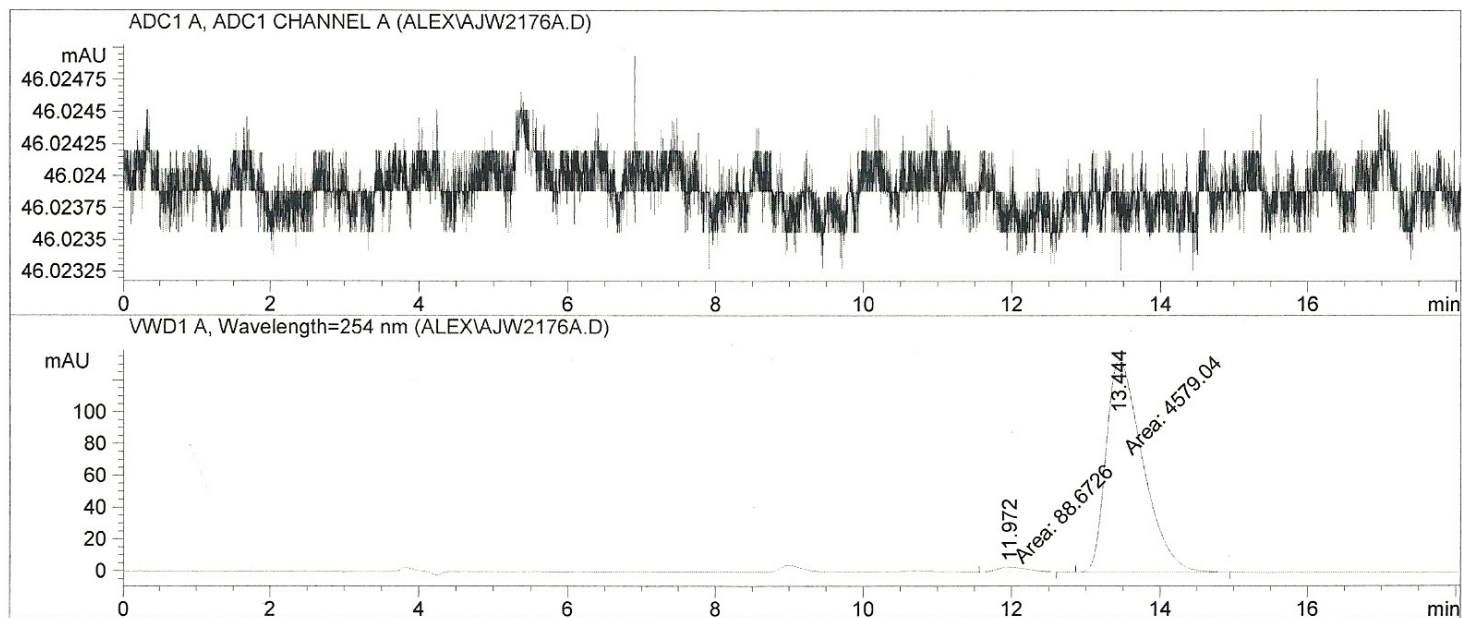
Results obtained with enhanced integrator!

=====
Summed Peaks Report
=====

Signal 1: ADC1 A, ADC1 CHANNEL A

=====
Injection Date : 3/9/2012 1:53:06 PM
Sample Name : AJW-2-176A4 Location : Vial 4
Acq. Operator : Alex Inj Volume : 10 µl

Acq. Method : C:\HPCHEM\1\METHODS\ALEX.M
Last changed : 3/9/2012 12:49:39 PM by Alex
(modified after loading)
Analysis Method : C:\HPCHEM\1\METHODS\ALEX.M
Last changed : 12/13/2011 8:09:06 PM by Alex
=====



=====
Area Percent Report
=====

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Sample Amount : 10.00000 [ng/ul] (not used in calc.)
Use Multiplier & Dilution Factor with ISTDs

Signal 1: ADC1 A, ADC1 CHANNEL A

Signal 2: VWD1 A, Wavelength=254 nm

| Peak # | RetTime [min] | Type | Width [min] | Area mAU *s | Height [mAU] | Area % |
|--------|---------------|------|-------------|-------------|--------------|---------|
| 1 | 11.972 | MM | 0.4641 | 88.67263 | 3.18407 | 1.8997 |
| 2 | 13.444 | MM | 0.5731 | 4579.04443 | 133.17099 | 98.1003 |

Totals : 4667.71706 136.35506

Results obtained with enhanced integrator!

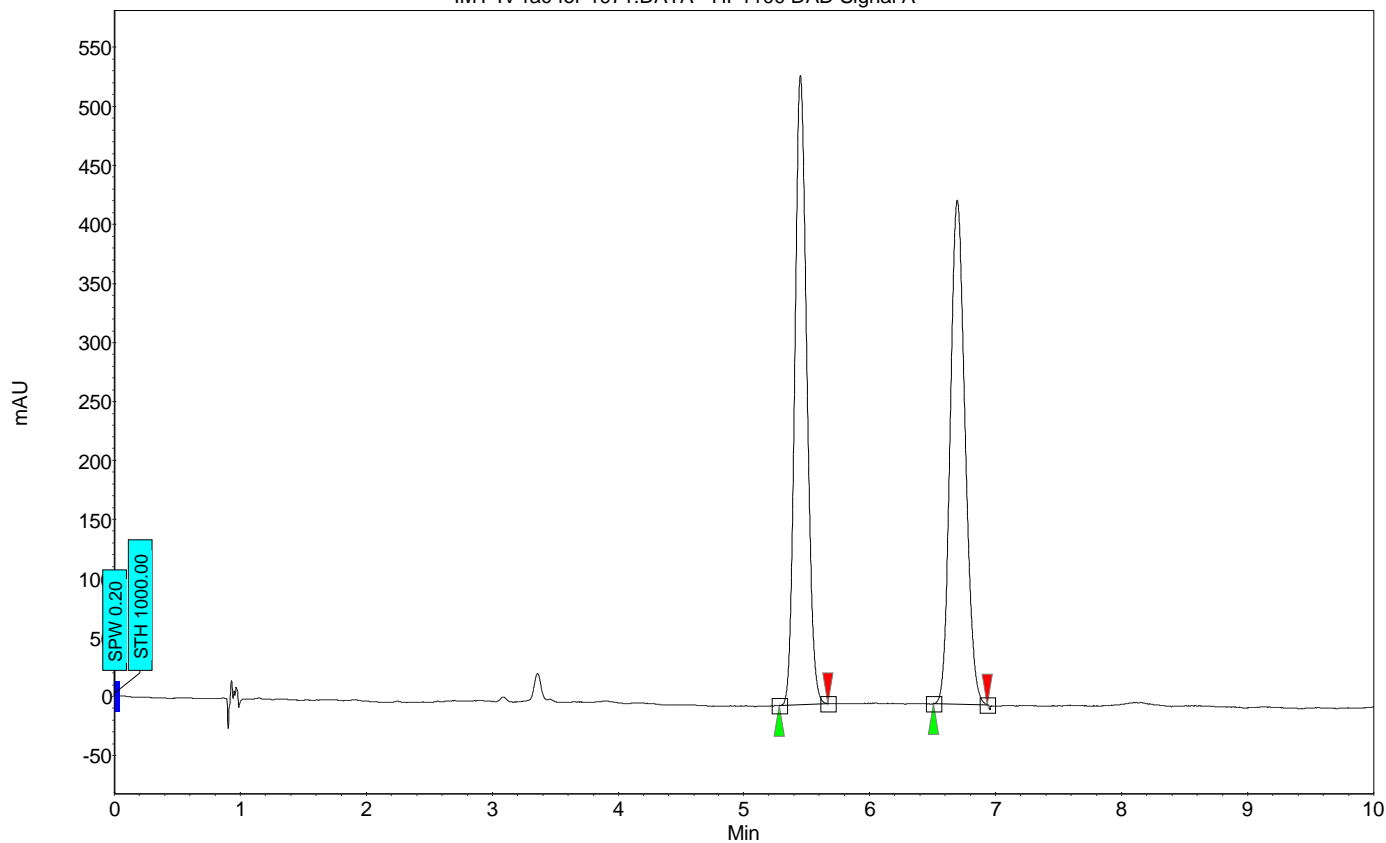
=====
Summed Peaks Report
=====

Signal 1: ADC1 A, ADC1 CHANNEL A

Method Name:IMY-6-OMeNapEt-alcohol
 Run Name:IMY-IV-rac-for-1971

Date:2/25/2013
 Time:5:07:26 PM

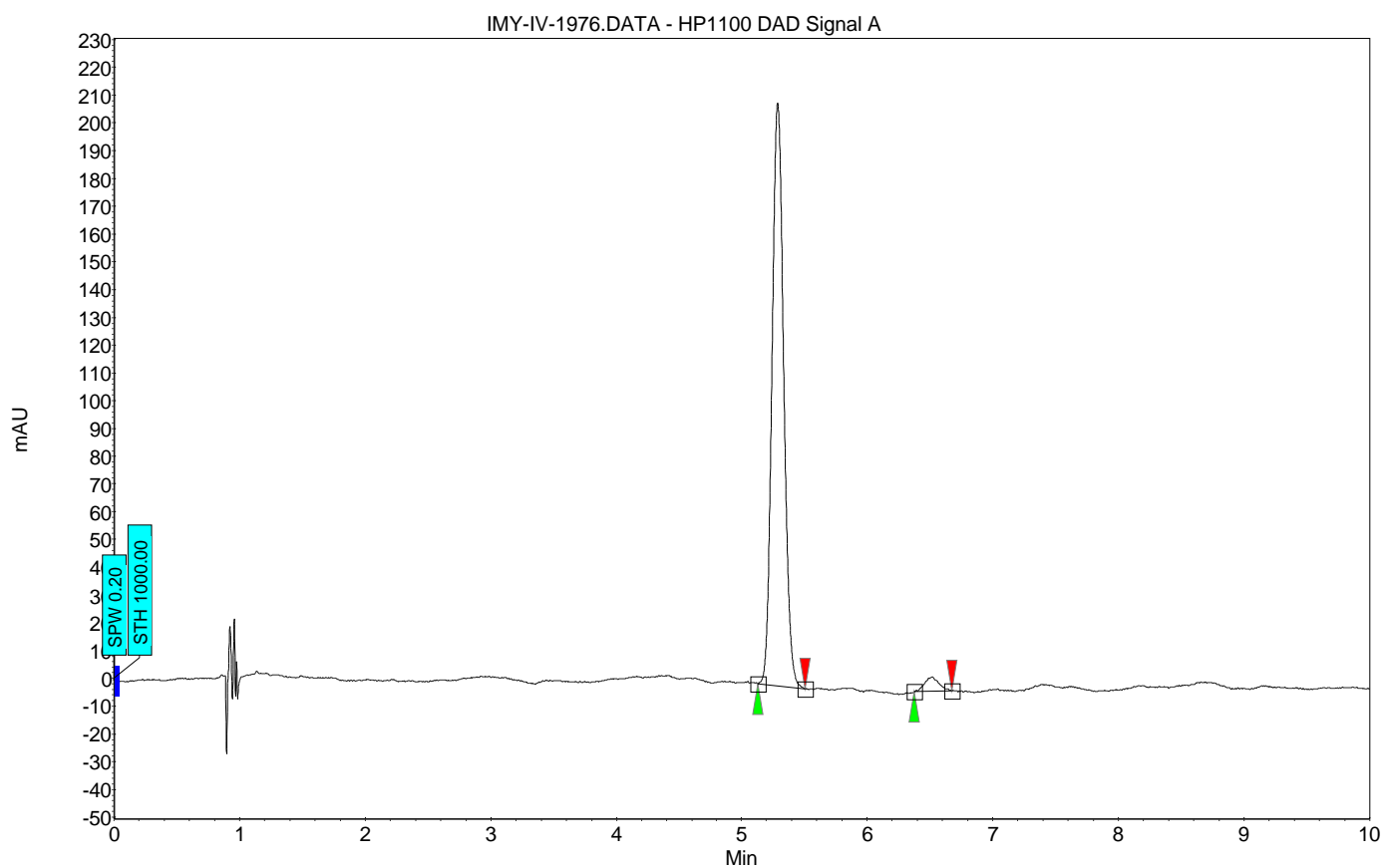
IMY-IV-rac-for-1971.DATA - HP1100 DAD Signal A



| Index | Name | Start | Time | End | RT Offset | Quantity | Height | Area | Area |
|-------|---------|-------|-------|-------|-----------|----------|------------|----------------|---------|
| | | [Min] | [Min] | [Min] | [Min] | [% Area] | [μ V] | [μ V.Min] | [%] |
| 1 | UNKNOWN | 5.28 | 5.45 | 5.67 | 0.00 | 50.08 | 533.2 | 58.7 | 50.078 |
| 2 | UNKNOWN | 6.51 | 6.69 | 6.93 | 0.00 | 49.92 | 427.1 | 58.5 | 49.922 |
| Total | | | | | | 100.00 | 960.2 | 117.1 | 100.000 |

Method Name:IMY-6-OMeNapEt-alcohol
 Run Name:IMY-IV-1976

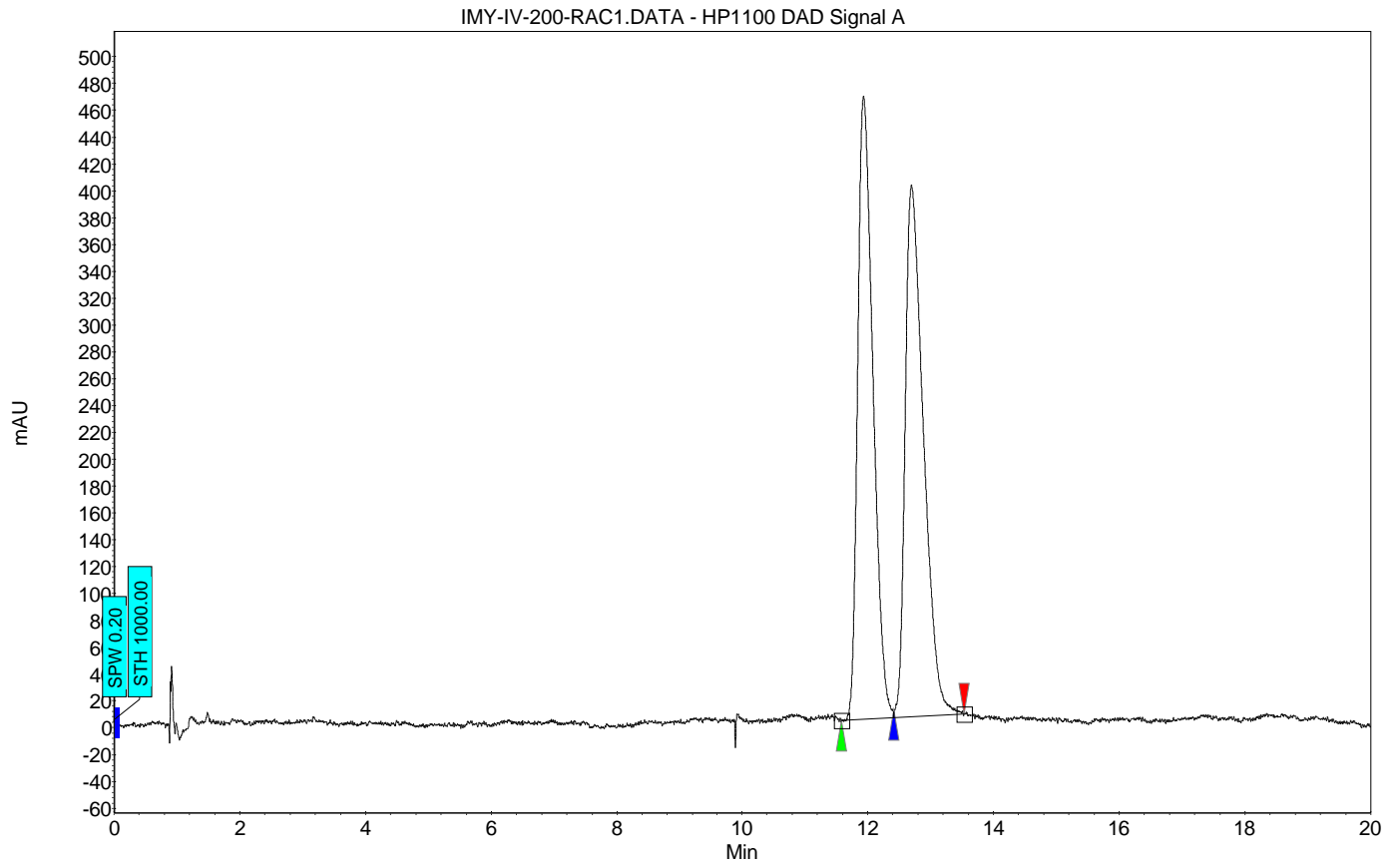
Date:2/25/2013
 Time:5:05:59 PM



| Index | Name | Start | Time | End | RT Offset | Quantity | Height | Area | Area |
|-------|---------|-------|-------|-------|-----------|----------|------------|----------------|---------|
| | | [Min] | [Min] | [Min] | [Min] | [% Area] | [μ V] | [μ V.Min] | [%] |
| 1 | UNKNOWN | 5.13 | 5.29 | 5.51 | 0.00 | 97.12 | 209.9 | 22.7 | 97.119 |
| 2 | UNKNOWN | 6.38 | 6.52 | 6.67 | 0.00 | 2.88 | 5.2 | 0.7 | 2.881 |
| Total | | | | | | 100.00 | 215.0 | 23.4 | 100.000 |

Method Name:AGJ-nap-cyclohex_OH
 Run Name:IMY-IV-200-RAC1

Date:2/25/2013
 Time:5:08:40 PM

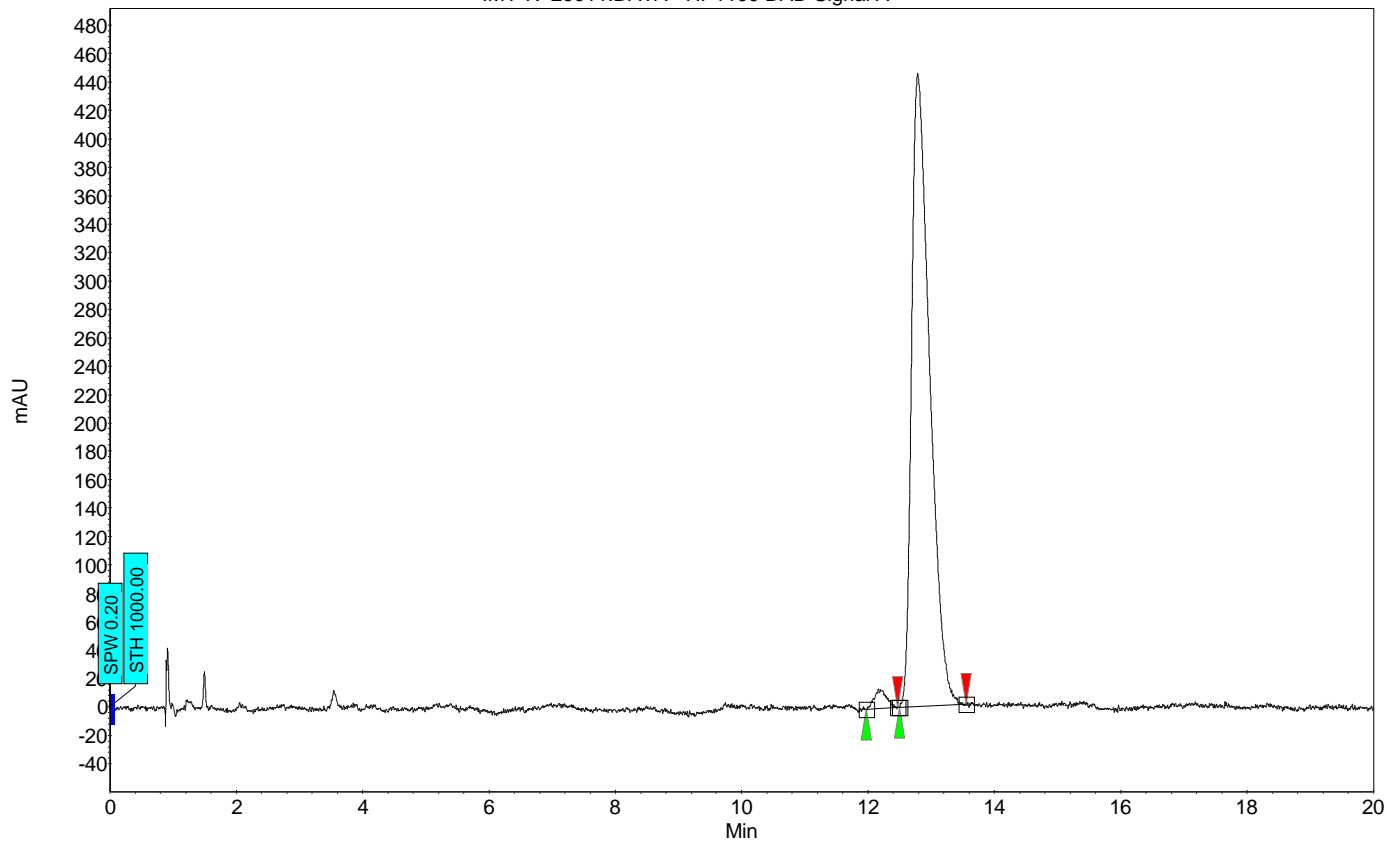


| Index | Name | Start | Time | End | RT Offset | Quantity | Height | Area | Area |
|-------|---------|-------|-------|-------|-----------|----------|------------|----------------|---------|
| | | [Min] | [Min] | [Min] | [Min] | [% Area] | [μ V] | [μ V.Min] | [%] |
| 1 | UNKNOWN | 11.58 | 11.93 | 12.41 | 0.00 | 49.52 | 464.2 | 131.0 | 49.517 |
| 2 | UNKNOWN | 12.41 | 12.69 | 13.53 | 0.00 | 50.48 | 396.3 | 133.6 | 50.483 |
| Total | | | | | | 100.00 | 860.5 | 264.6 | 100.000 |

Method Name:AGJ-nap-cyclohex_OH
 Run Name:IMY-IV-20011

Date:2/25/2013
 Time:5:08:07 PM

IMY-IV-20011.DATA - HP1100 DAD Signal A



| Index | Name | Start | Time | End | RT Offset | Quantity | Height | Area | Area |
|-------|---------|-------|-------|-------|-----------|----------|------------|----------------|---------|
| | | [Min] | [Min] | [Min] | [Min] | [% Area] | [μ V] | [μ V.Min] | [%] |
| 1 | UNKNOWN | 11.98 | 12.17 | 12.47 | 0.00 | 2.01 | 13.9 | 3.0 | 2.010 |
| 2 | UNKNOWN | 12.50 | 12.79 | 13.55 | 0.00 | 97.99 | 445.8 | 147.3 | 97.990 |
| Total | | | | | | 100.00 | 459.7 | 150.3 | 100.000 |