

SUPPORTING INFORMATION

Structure, Kinetic, and Chemical Mechanism of Isocitrate Dehydrogenase-1 from

Mycobacterium tuberculosis

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TABLE S1: “The Effect of Different Divalent Cations on the Rate of ICDH-1.”

Metal [5 mM]	V/E sec ⁻¹	V/E sec ⁻¹	V/E sec ⁻¹
	Replicate 1	Replicate 2	Average
None	0.08	0.09	0.085
MnCl ₂	16.18	15.86	16.02
MgCl ₂	14.94	13.76	14.35
CaCl ₂	0.18	0	0.09
ZnCl ₂	0.18	0.25	0.215

Assays were performed with 100 mM HEPES pH 7.0, 600 μ M isocitrate, 300 μ M NADP⁺, and 5 mM of the metal tested. Note that the maximum rate is decreased from other experiments since subsequent experiments were performed with ICDH-1 in which 5 mM MnCl₂ had been added to the gel filtration and storage buffers.

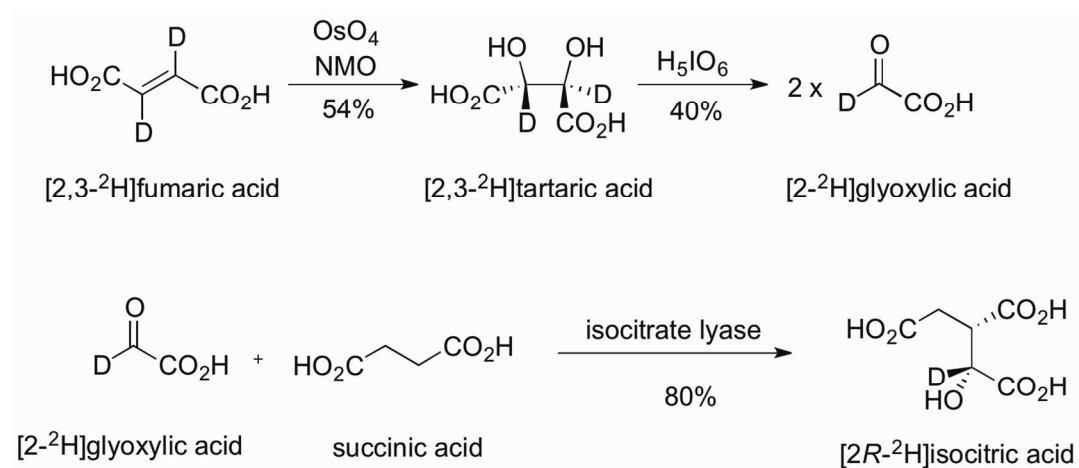
SCHEME S1: "Synthesis of [2R-²H]isocitrate."

FIGURE S1: “Transfer of ^2H from $[2R\text{-}^2\text{H}]$ isocitrate to $[4R\text{-}4\text{-}^2\text{H}]$ NADPH.” (A) ^1H NMR of $[4R\text{-}4\text{-}^2\text{H}]$ NADPH. (B) ^1H NMR of $[4R\text{-}4\text{-}^1\text{H}]$ NADPH.

FIGURE S2: “Confirmation of αHG by Mass Spectrometry.” Mass spectrum of secondary reaction product, αHG , $m/z = 147.0301$ in negative mode. The peak to the left $m/z = 147.0187$ is αKG in negative mode $A + 2$. Data were collected on a 12 Tesla Fourier transform ion cyclotron resonance mass spectrometer (Agilent).

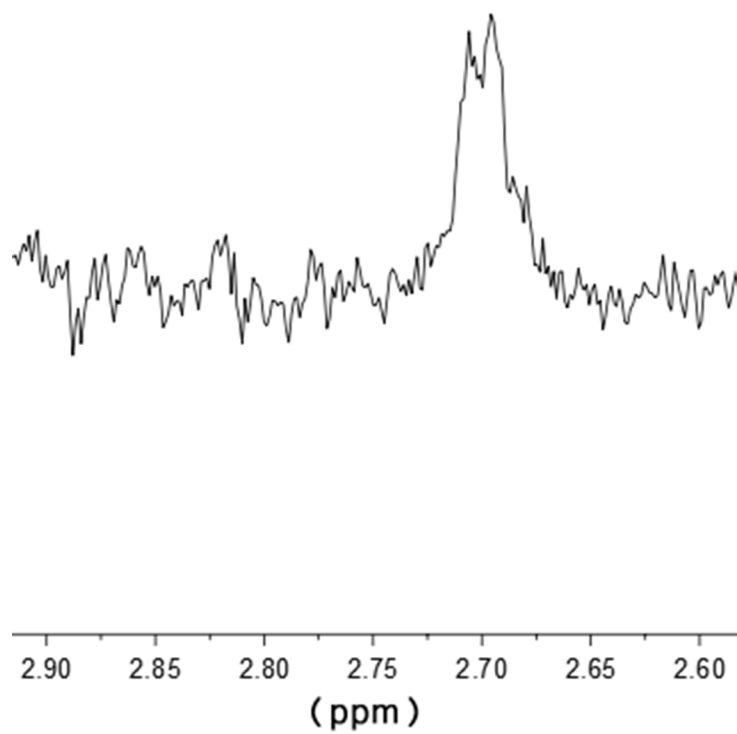
FIGURE S3: “Preliminary Kinetics of Secondary Reaction.” (A) The rate of the secondary ICDH-1 reaction was measured with saturating NADPH (100 μM) and varying αKG (1-50 mM). Fitting to eq 1 $k_{\text{cat}} = 0.12 \pm 0.02 \text{ sec}^{-1}$, $K_{\alpha\text{KG}} = 8.3 \pm 1.2 \text{ mM}$. (B) The rate of the secondary ICDH-1 reaction was measured with constant αKG (40 mM) and varying NADPH (1-50 μM). Fitting to eq 1 $k_{\text{cat}} = 0.10 \pm 0.02 \text{ sec}^{-1}$, $K_{\text{NADPH}} = 0.9 \pm 0.8 \mu\text{M}$. (C) A replicate of panel (B). Fitting to eq 1 $k_{\text{cat}} = 0.17 \pm 0.02 \text{ sec}^{-1}$, $K_{\text{NADPH}} = 2.3 \pm 1.0 \mu\text{M}$.

FIGURE S4: “Sequence Alignment of Relevant ICDHs.” ClustalO sequence alignment using EMBL-EBI server of the top three BLAST hits in the PDB against Mtb ICDH-1, plus *E. coli*. UniProt identifiers are on the right. Orange highlights are Lys 215 and Tyr 142 in the proposed chemical mechanism. Yellow highlighted residues are NADPH contacts, purple are Mn^{2+} contacts, blue are residues referenced in the discussion of regulation, and green is the analogous Hc R132 mutation location.

FIGURE S5: “Contacts between NADPH and Mtb ICDH-1.” A LigPlot⁽¹⁾ generated figure where red spheres represent oxygen, blue spheres represent nitrogen, black spheres represent carbon. The gold lines are the backbone of Mtb ICDH-1, and the purple lines are the backbone of NADPH. The red spokes are hydrophobic interactions, and the green numbers are distances in angstroms.

Figure S1

A



B

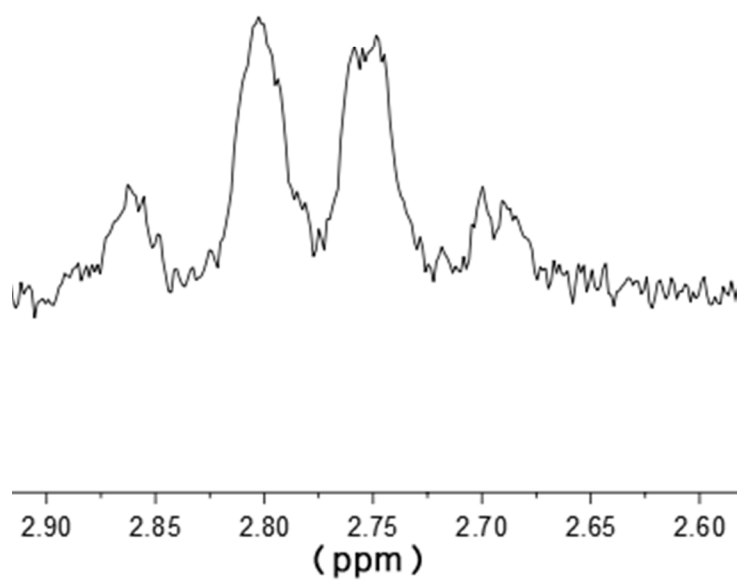


Figure S2

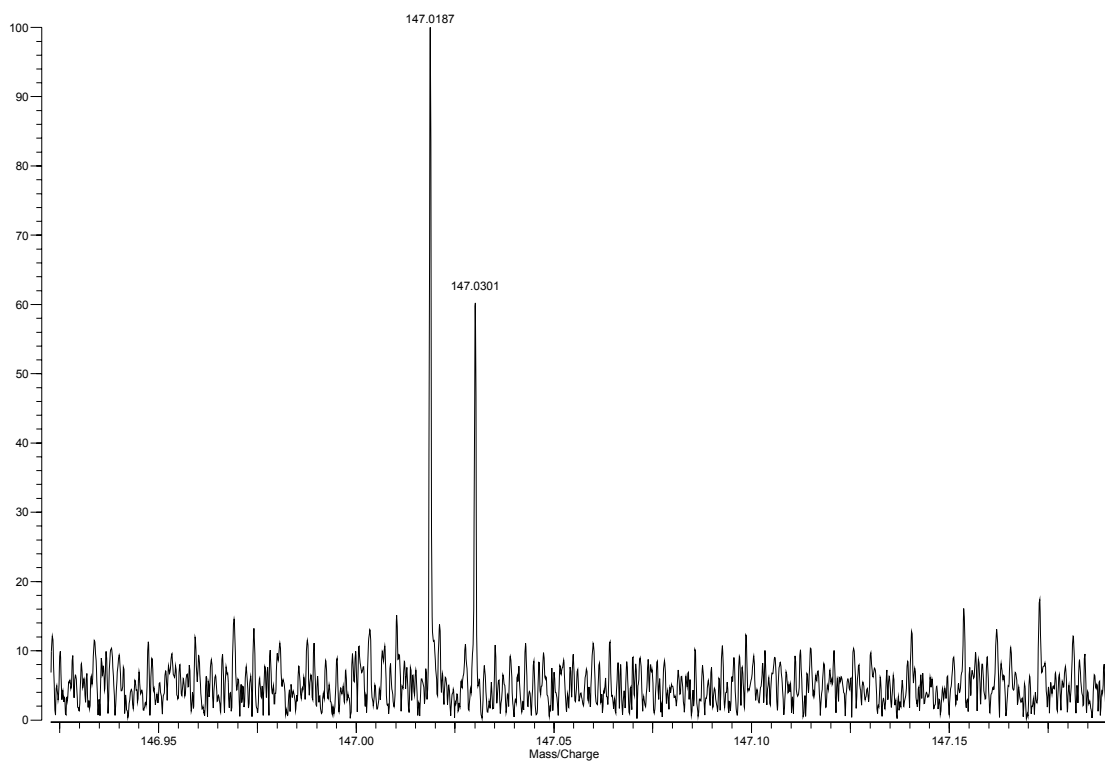


Figure S3

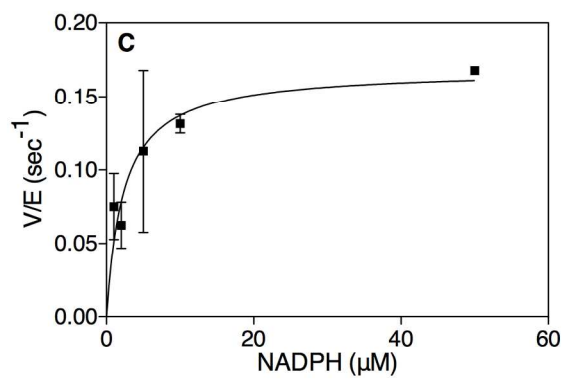
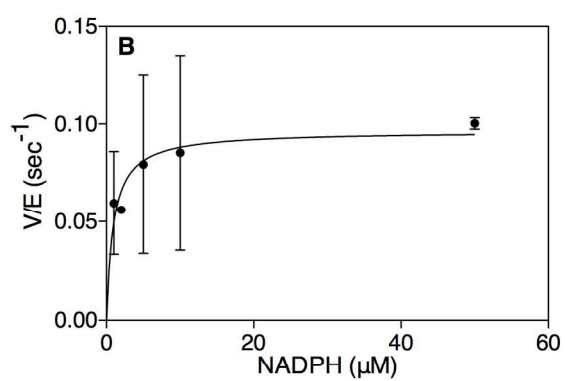
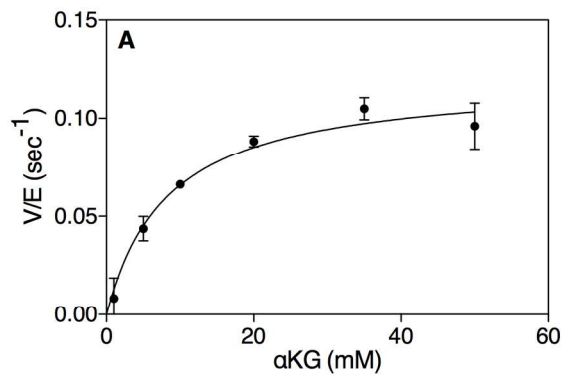
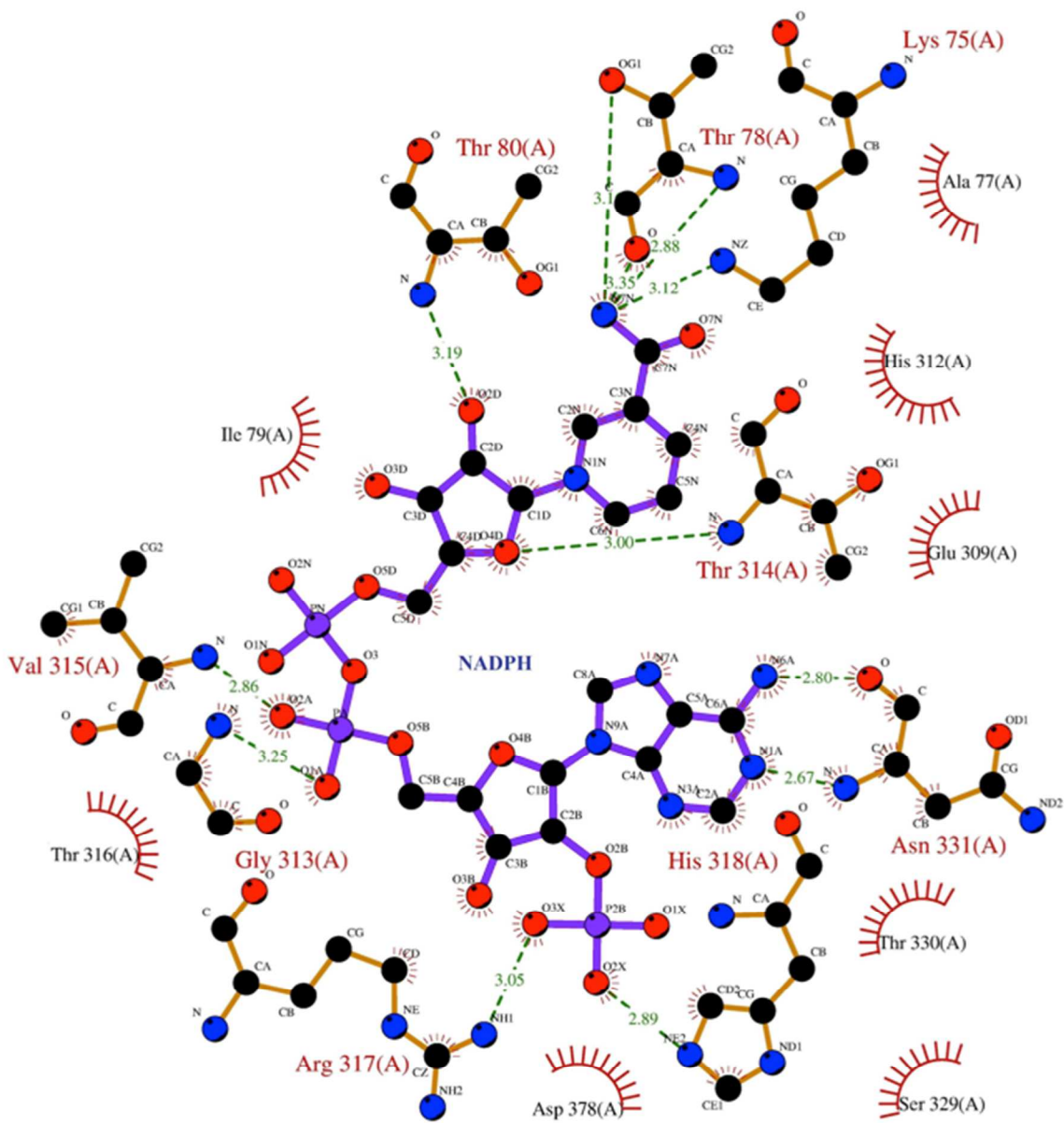


Figure S4

P65097	<i>M. tuberculosis</i>	-----MSNAPKIKVSGPVVELDGDDEMTRVIWK-LIKDMLILPYLDIRLDYYDL-----	47
Q92PG6	<i>R. meliloti</i>	-----MAKIKVANPVVELDGDDEMTRIIWQ-FIKDKLIHPYLDLDLEYDYL-----	44
075874	<i>H. sapiens</i>	-----MSKKISGGSVVEMOGDDEMTRIIWE-LIKEKLIFFYVELDLHSYDL-----	44
P33198	<i>S. scrofa</i>	ARAAARHYADQRIVKAPVEMDGDDEMTRIIWQ-FIKEKLIIPHVDVQLKYFDL-----	53
P08200	<i>E. coli</i>	-----MESKVVVPAQGKKITLQNGKLNVPENPIIPYIEGDGIGVDVTPAMLK	47
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P65097	<i>M. tuberculosis</i>	----GIEHRDATD-----DQVTIDAAYAIAKKHGIVGVKCATITPD	82
Q92PG6	<i>R. meliloti</i>	----GVENRDATD-----DQVTIDAANAIAKKHGIVGVKCATITPD	79
075874	<i>H. sapiens</i>	----GIENRDATN-----DQVTKDAAEAIAKKHNVGVKCATITPD	79
P33198	<i>S. scrofa</i>	----GLPNRDQTN-----DQVTIDSALATQKYSVAVKCATITPD	88
P08200	<i>E. coli</i>	VVDAAVEKAYKGERKISWMEIYTGEKSTQVYGDVWLPALDLDLIREYRVAIKGPLTTPV	107
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P65097	<i>M. tuberculosis</i>	EARVEEFNLKKMWLSPNGTIRNILGGTIFREPIVISNVPRVLPVGTWKPI-----VIG	134
Q92PG6	<i>R. meliloti</i>	EGRVEEFKLKMWKSPNGTIRNILGGVIFREPIICKNVPRVLPVGTWKPI-----IVG	131
075874	<i>H. sapiens</i>	EKRVEEFKQKMWKSPNGTIRNILGGTVFREAIICKNIPRLVSGWVKPI-----IIG	131
P33198	<i>S. scrofa</i>	EARVEEFKLKMWKSPNGTIRNILGGTIFREPIICKNIPRLVLPVGTWKPI-----TIG	140
P08200	<i>E. coli</i>	GGGIRSLNV-----ALRQE-----LDLYICLRPVRYQGTSPVVKHPELTDMMVIF	152
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P65097	<i>M. tuberculosis</i>	RHAFGDQYRATNFKVDQPGTVT-LTFTPADGSAPIVHEMVSIPEDGGVVLGMYNFKESIR	193
Q92PG6	<i>R. meliloti</i>	RHAFGDQYRATDFKFPKGGKLS-IKFVGEDGQT-IEHDVYD-APGAGVALAMYNLDESIT	188
075874	<i>H. sapiens</i>	RHAYGDQYRATDFVVPVPGPKVE-ITYTPSDGTQKVTYLVHNFEEGGVAMGMYNQDKSIE	190
P33198	<i>S. scrofa</i>	RHAHGDQYKATDFVVDVDRAGTFK-IVFTPKDGSSAKQWEVYNFPA-GVGMGMINTDESIS	198
P08200	<i>E. coli</i>	RENSEDIYAGIEWKADSADAEKVIKFLREE----MGVKKIRFPHEHCIGIKPCS-EEGTK	207
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P65097	<i>M. tuberculosis</i>	DFARASFSYGLNA-KWPVYLSTKNTILKAYDGMFKDEFERVYEEEFKAQFEAAGLTYEHR	252
Q92PG6	<i>R. meliloti</i>	EFARASFNGLQR-KVPVYLSTKNTILKAYDGRFKDIFQKVFDEEFAAQFKAEKLWYEHR	247
075874	<i>H. sapiens</i>	DFAHSSSQMALS-KWPLYLSTKNTILKAYDGRFKDIFQEIYDKQYKYSQFEAQKIWEHR	249
P33198	<i>S. scrofa</i>	GFAHSCFYAIQK-KWPLYLSTKNTILKAYDGRFKDIFQEIPEKHVKDFDKYKIWEHR	257
P08200	<i>E. coli</i>	RLVRAAIEYAIANDRDSVTLVHKNGNIMKFTTEGAFKDWGYQLAREEFGGLIDGGPWLVKVK	267
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P65097	<i>M. tuberculosis</i>	L-----IDDMV-----AAC-LKWSGGYVWACKNYDGDVQSDTVAQGYGSLGLMSTSVL	298
Q92PG6	<i>R. meliloti</i>	L-----IDDMV-----ASA-LKWSGGYVWACKNYDGDVQSDIVAQGYGSLGLMSTSVL	293
075874	<i>H. sapiens</i>	L-----IDDMV-----AQA-MKSEGGFIWACKNYDGDVQSDSVAQGYGSLGMSTSVL	295
P33198	<i>S. scrofa</i>	L-----IDDMV-----AQV-LKSSGGFVWACKNYDGDVQSDILAQGYGSLGLMSTSVL	303
P08200	<i>E. coli</i>	NPNTGKEIVIKDVIADAFLLQQLLRPAEYDVIACMNLNGDYISDALAAQVGGIGIAPGAN	327
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P65097	<i>M. tuberculosis</i>	MTADGKTVEAEAAGTIVTRHYRQYQAGKPTST----NPIASIFAWTRGLQHRGKLDGTPE	354
Q92PG6	<i>R. meliloti</i>	MTPDGKTVEAEAAGTIVTRHYRQHQKGEETST----NSIASIFAWTRGLAHRKLDGNAE	349
075874	<i>H. sapiens</i>	VCPDGKTVEAEAAGTIVTRHYRMYQKQGETST----NPIASIFAWTRGLAHRKLDNNKE	351
P33198	<i>S. scrofa</i>	VCPDGKTIEAEAAGTIVTRHYREHQKGRPTST----NPIASIFAWTRGLEHRKLDGNQD	359
P08200	<i>E. coli</i>	IGDECAL--FEATHGTAPKY-AGQDKVNPGSIIILSAEMMLRHMGWTEAADLIVK-----	378
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P65097	<i>M. tuberculosis</i>	VIDFAHKLESVVIATVESGKMTKDLAILIG-----PEQDWLNSEEFLDIAIDNLEKELA	408
Q92PG6	<i>R. meliloti</i>	LAKFSETLERVCVDTVESGFMTKDLALLIG-----PDQPWLSTTGFLDKIDENLRKAMA	403
075874	<i>H. sapiens</i>	LAFFANALEEVSIIETIEAGFMTKDLAICKGLPNVQRS-DYLNTFEMDKLGENLKIKLA	410
P33198	<i>S. scrofa</i>	LIRFAQTLEKVCVETVESGAMTKDLAGCIGHLSNVKLNEHFLNTSDFLDTIKSNLDRALG	419
P08200	<i>E. coli</i>	-----GMEGAINAKTVTYDFERLMD-----GAKLLKCESEFGDAIENM-----	416
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P65097	<i>M. tuberculosis</i>	N---	409
Q92PG6	<i>R. meliloti</i>	A---	404
075874	<i>H. sapiens</i>	QAKL	414
P33198	<i>S. scrofa</i>	RQ--	421
P08200	<i>E. coli</i>	----	416

Figure S5



1. Wallace, A. C., Laskowski, R. A., and Thornton, J. M. (1995) LIGPLOT: a program to generate schematic diagrams of protein-ligand interactions, *Protein engineering* 8, 127-134.