Gel # 1	Gel # 2	Gel # 3	Gel # 4	Gel # 5	Gel # 6
Standard Cy2 BSA neg ctrl Cy5 15' Cort-BSA Cy3	Standard Cy2 BSA neg ctrl Cy5 15' Cort-BSA Cy3	Standard Cy2 BSA neg ctrl Cy5 15' Cort-BSA Cy3	Standard Cy2 5' Cort-BSA Cy5 90' Cort-BSA Cy3	Standard Cy2 5' Cort-BSA Cy5 90' Cort-BSA Cy3	Standard Cy2 5' Cort-BSA Cy5 90' Cort-BSA Cy3
Cytosolic fraction	Cytosolic fraction	Cytosolic fraction	Cytosolic fraction	Cytosolic fraction	Cytosolic fraction

Gel # 7	Gel # 8	Gel # 9	Gel # 10	Gel # 11	Gel # 12
Standard Cy2					
BSA neg ctrl Cy5	BSA neg ctrl Cy5	BSA neg ctrl Cy5	5' Cort-BSA Cy5	5' Cort-BSA Cy5	5' Cort-BSA Cy5
15' Cort-BSA Cy3	15' Cort-BSA Cy3	15' Cort-BSA Cy3	90' Cort-BSA Cy3	90' Cort-BSA Cy3	90' Cort-BSA Cy3
Nuclear fraction					

Cytosolic and nuclear fractions were fluorescence-labeled as follows: BSA negative control and 5min Cort-BSA by Cy5 dye, 15min and 90 min Cort-BSA by Cy3. A total of 3 analytical gels were performed for each of the following comparisons: CYTO neg vs. CYTO 15min; CYTO 5 min vs. CYTO 90min; NUCL neg vs. NUCL 15min; NUCL 5min vs. 90min. In each analytical gel a common Cy2-labelled control, obtained by pooling all the samples in equal proportion, was loaded and used as internal standard. To account for variations between the labeling, the dyes were swapped in one of the three replicatesCytosolic and nuclear fractions were fluorescence-labeled as follows: BSA negative control and 5min Cort-BSA by Cy5 dye, 15min and 90 min Cort-BSA by Cy3. A total of 3 analytical gels were performed for each of the following comparisons: CYTO neg vs. CYTO 15min; CYTO 5 min vs. CYTO 90min; NUCL neg vs. NUCL 15min; NUCL 5min vs. 90min. In each analytical gel a common Cy2-labelled control, obtained by pooling all the samples in equal proportion, was loaded and used as internal standard. To account for variations between the labeling, the dyes were swapped in one of the three replicatesCytosolic and nuclear fractions were fluorescence-labeled as follows: BSA negative control and 5min Cort-BSA by Cy5 dye, 15min and 90 min Cort-BSA by Cy3. A total of 3 analytical gels were performed for each of the following comparisons: CYTO neg vs. CYTO 15min; CYTO 5 min vs. CYTO 90min; NUCL neg vs. NUCL 15min; NUCL 5min vs. 90min. In each analytical gel a common Cy2-labelled control, obtained by pooling all the samples in equal proportion, was loaded and used as internal standard. To account for variations between the labeling, the dyes were swapped in one of the three replicates



Supplementary Fig. 2 Bidimensional gel showing the ovelay of cytosolic and nuclear fractions.



Supplementary Fig. 3

Schematic representation of the epitopes targeted by the four anti-GR antibodies used in the in situ PLA experiments.



Supplementary Fig. 4

Cells were washed three times in Dulbecco's phosphate-buffered saline and incubated for 24h in their growth medium containing 10% charcoalstripped FBS. Membrane GR expression was subsequently tested by in situ PLA using the P20 antibody. Glucocorticoid withdrawal significantly increased mGR expression in all cell lines, except Jurkat cells.



HADH in the cytosolic fraction

CoxVb in the nuclear fraction

Supplementary Fig. 5 GR signalling was pharmacologically blocked in CCRF-CEM cells. Prior to Cort-BSA stimulation cells were incubated with 10 μ M Ru486 for 15 minutes. The modulatory effects of Cort-BSA on both CoxVb and HADH were lost in treated cells, indicating that a functional GR is necessary for Cort-BSA to exert its