

Supplementary Figure S1. Reverse cumulative distributions (logarithmic scale) of the number of restriction fragments that have at least one read mapped to them. The four dashed lines correspond to the four datasets generated using the *Myb* promoter as a 3C-seq viewpoint in fetal liver erythrocytes (FL) and fetal brain cells (FB). HindIII was used as a primary restriction enzyme. The seven solid curves correspond to seven datasets generated with various viewpoints in mouse pre-B cells, using BgIII as a primary restriction enzyme. A) Reverse cumulative distributions before normalization. B) Reverse cumulative distributions after normalization with the simple RPM calculation method. C) Reverse cumulative distributions after normalization with the reverse-cumulative fitted values from the power-law distributions.



Supplementary Figure S2. Reverse cumulative distributions (logarithmic scale) of the number of 5 kb binning windows that have at least one read mapped to them. The four dashed curves correspond to the four datasets generated using the *Myb* promoter as a 3C-seq viewpoint in fetal liver erythrocytes (FL) and fetal brain cells (FB). The seven solid curves correspond to seven datasets generated with various viewpoints in mouse pre-B cells. A) Reverse cumulative distributions before normalization. B) Reverse cumulative distributions after normalization with the simple RPM calculation method. C) Reverse cumulative distributions after normalization with the reverse-cumulative fitted values from the power-law distributions.





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Supplementary Figure S3. The MA plots of the log2 intensity ratio (M) versus the average log2 intensity values (A) for two sets of independently prepared samples using no normalization, a simple RPM normalization, or the reverse-cumulative fitted values normalization. A) MA plots of data points generated using the Myb promoter as a 3C-seq viewpoint in fetal liver erythrocytes (FL) against fetal brain cells (FB) in biological replicate 1. B) MA plots of data points generated using the Myb promoter as a 3C-seq viewpoint in fetal liver erythrocytes (FL) against fetal brain cells (FB) in biological replicate 2. The red line represents the loess smoothing of M-A values and the red dots show the data points located within ±200 kb of the viewpoint.



Distance relative to the viewpoint (Mbp)

Supplementary Figure S4. Scaling of background interaction signals using a nonparametric regression cubic smoothing spline algorithm. Interaction signals with the *Myb* promoter viewpoint in fetal liver erythrocytes (FL) were ranked based on the relative genomic distance to the viewpoint. The plot shows the observed interaction signals and the scaled interaction signal using different smoothing parameters within a ± 3 Mbp window around the viewpoint.



Supplementary Figure S5. Candidate interaction regions with the Myb promoter in both fetal liver erythrocytes (FL) and fetal brain cells (FB) explored in four different plots generated by r3Cseq. A) Plot generated by the plotOverviewInteractions function, which explores the distribution of candidate interaction regions on a genome-wide scale. B) Plot generated by the plotInteractionsPer-Chromosome function, showing the interaction regions along the selected *cis* chromosome. **C)** Plot generated by the plotInteractionsPerChromosome function, showing the interaction regions along the selected *trans* chromosome. D) Plot generated by the plotInteractionsNearViewpoint function, which is used to zoom in on the viewpoint region to visualize the distribution of interaction regions around it. It provides tracks with Refseq genes, restriction fragment locations, interaction signals of the experiment dataset, interaction signal of the control dataset and the ratio (log2) of normalized interaction signals between the experiment and control datasets.



Supplementary Figure S6. Comparison of detected interactions using a restriction fragment based or window based analysis method for the β-maj promoter 3C-seq data from fetal liver erythrocytes (FL). Gene locations are shown at the top followed by a map of restriction fragments. The line plots show detected *cis*-interaction regions (40 kb up- and 60 kb downstream of the viewpoint) for either the fragment based approach used in our previous study, or using the *r3Cseq* fragment based, 5 kb window and 10 kb window based methods respectively. Color gradients represent the range of significant interaction regions and binding sites of transcription factor complexes.



Supplementary Figure S7. *r3Cseq* analysis results using 4C-seq data generated by van de Werken HJ et al. using individual restriction fragments, 2 kb and 5 kb window-based analysis in the 100 kb β -globin domain. Gene locations are shown at the top followed by a map of restriction fragments. The line plots show detected cis-interaction regions (40 kb up- and 60 kb downstream of the viewpoint) for the fragment based, 5 kb window and 10 kb window based methods respectively. Color gradients represent the range of significant interaction signals (q-value). The domainogram was generated by the 4C-seq pipeline developed by van de Werken HJ et al. The light blue box highlights the LCR region with its hypersensitive sites (HS1-5) coinciding with several FL-specific significant interaction regions and binding sites of p300 and LDB1.



Supplementary Figure S8. Comparison of 3C-seq data analysis using both the *r3Cseq* package and the 4C-seq pipeline described in van de Weken HJ et al. using the *Myb* promoter data sets generated in both fetal liver erythrocytes (FL) and fetal brain (FB) cells. For the *r3Cseq* analysis, high signals on the viewpoint fragment or the immediately adjacent fragments were excluded. Line plots show detected *cis*-interaction regions 500 kb up- and 500 kb downstream of the *Myb* viewpoint. The domainograms show detected interactions using a window based analysis (running from 2 kb to 30 kb) for both datasets. Color gradients of domainograms represent the interaction signal strength detected for each run of the defined window (transformed *q*-value). For the 4C-seq pipeline analysis, domainograms were generated using default parameters provided by the software. Color gradients represent the normalized window coverage. The purple dashed box highlights the *Myb-Hbs1l* intergenic region, which shows strong and specific interaction signals in FL using both methods, coinciding with binding sites of LDB1 and p300.



Supplementary Figure S9. Application of 3C-seq/r3Cseq to analyze chromatin interaction dynamics during erythroid differentiation at the *Myb* locus. Gene locations are shown at the top followed below by a map of restriction fragments. The line plots show detected *cis*-interaction regions 500 kb up- and 500 kb downstream of the *Myb* viewpoint in MEL cells before and after induction of differentiation. High interaction signal on the viewpoint fragment and those immediately adjacent have been excluded. Domainograms show detected interactions using a window based analysis (running from 2 kb to 30 kb) for both datasets. Color gradients of domainograms represent the interaction signal strength detected for each run of the defined window (transformed *q*-value). The purple dashed box highlights the *Myb-Hbs11* intergenic region, which shows strong interaction signals coinciding with binding sites of LDB1 and p300.



Supplementary Figure S10. Exemplary analysis comparing single sample analysis and analysis with replicates using the Myb promoter viewpoint datasets from fetal liver erythrocytes (FL) and fetal brain cells (FB). A) The genome-wide distribution of significant interaction regions for a single r3Cseq dataset. B) The genome-wide distribution of significant interaction regions after analyzing *r3Cseq* biological replicates. C) Top 40 genes found in close proximity to significant interaction signals in the FL erythrocytes of single sample analysis. D) Genes detected in close proximity to significant interaction signals present in FL erythrocytes of biological replicates.

			Interactions			
			present only	present only	Interactions present	
Cis/Trans	Per region	Sample	in replicate1	in replicate2	in common	
					591 (28% for	
	per restriction	Myb viewpoint in			replicate1 and 24% for	
	fragment	fetal liver	1,499	1,888	replicate2)	
					672 (34% for	
	per 5 Kb	Myb viewpoint in			replicate1 and 28% for	
	window	fetal liver	1,277	1,626	replicate2)	
					727 (40% for	
	per 10 Kb	Myb viewpoint in			replicate1 and 36% for	
cis-interactions	window	fetal liver	1,065	1,294	replicate2)	
					208 (20% for	
	per restriction	Myb viewpoint in			replicate1 and 17% for	
	fragment	fetal brain	831	1,041	replicate2)	
					229 (24% for	
	per 5 Kb	Myb viewpoint in			replicate1 and 20% for	
	window	fetal brain	732	940	replicates2)	
					226 (25% for	
	per 10 Kb	Myb viewpoint in	<i></i>	0.50	replicate1 and 21% for	
	window	fetal brain	675	850	replicate2)	
					178 (0.02% for both	
	per restriction	Myb viewpoint in	5 501	0.005	replicated and	
	fragment	fetal liver	7,791	9,227	replicate2)	
	5 171				349 (0.04% for both	
<i>trans</i> -interactions	per 5 Kb	Myb viewpoint in	7.000	0.000	replicate I and	
	window	fetal liver	7,923	9,280	replicate2)	
		M. L			535 (0.06% for	
	per 10 Kb	Myb viewpoint in	7.021	0.041	replicate1 and	
	window	Ietal liver	/,831	9,041	replicate2)	
	per restriction	fotol broin	2 (12	4.570	4/ (0.01% for both	
	Iragment	Ietai orain	5,012	4,579	76 (0.020/ for both	
	per 5 Kb	fotol broin	3 632 4 570		70 (0.02% 10f b0th	
	willdow	ietai biain	5,022	4,370	111 (0.2% for	
	por 10 Kh	Muh viewneint in			111(0.5% 10f	
	window	fotal brain	3621	4502	replicate1 and 0.02%	
	window	ictal Utalli	5021	4303	Tephcate2)	

Supplementary Table1. The calculated reproducibility of 3C-seq data from replicate experiments using fragment-based and window-based analysis methods (interaction signals >=1 RPM).

			Interactions	Interactions	
			present only	present only	Interactions present
Cis/Trans	Per region	Sample	in replicate1	in replicate2	in common
					21 (57% for replicate1
<i>cis</i> -interactions within ± 500 kb relative to the viewpoint	per restriction	Myb viewpoint in			and 84% for
	fragment	fetal liver	16	4	replicate2)
					26 (67% for replicate1
	per 5 Kb	Myb viewpoint in			and 84% for
	window	fetal liver	13	5	replicate2)
					28 (70% for replicate1
	per 10 Kb	Myb viewpoint in			and 90% for
	window	fetal liver	12	3	replicate2)
					6 (50% for replicate1
	per restriction	Myb viewpoint in			and 67% for
	fragment	fetal brain	6	3	replicate2)
					8 (53% for replicate1
	per 5 Kb	Myb viewpoint in			and 80% for
	window	fetal brain	7	2	replicates2)
					10 (63% for replicate1
	per 10 Kb	Myb viewpoint in			and 83% for
	window	fetal brain	6	2	replicate2)

Supplementary Table 2. The calculated reproducibility of 3C-seq data from replicate experiments using fragment-based and window-based analysis methods (interaction signals >=500 RPM).

Viewpoint	Experiment	Control	Experim	Control	Number of	Number of	Processing
	file name	file name	ent file	file size	reads in	reads in the	time of
			size	(MB)	experiment	control	r3Cseq
			(MB)				(min)
Myb	MYB_Pro	MYB_Pr	134	145	2,950,203	3,253,519	3.06
promoter	m_12.5_F	om_12.5					
	L.bam	_FB.bam					
β-major	Beta_Pro_	Beta_Pro	248	296	6,281,237	7,241,107	5.77
promoter	FL.bam	_FB.bam					

Supplementary Table 3. Comparison of processing run times of the *r3Cseq* pipeline for different data sets. *r3Cseq* was performed using R version 2.15 running on a standard personal computer (MAC OS X with 2.53GHz Intel Core Duo and 4GB RAM).