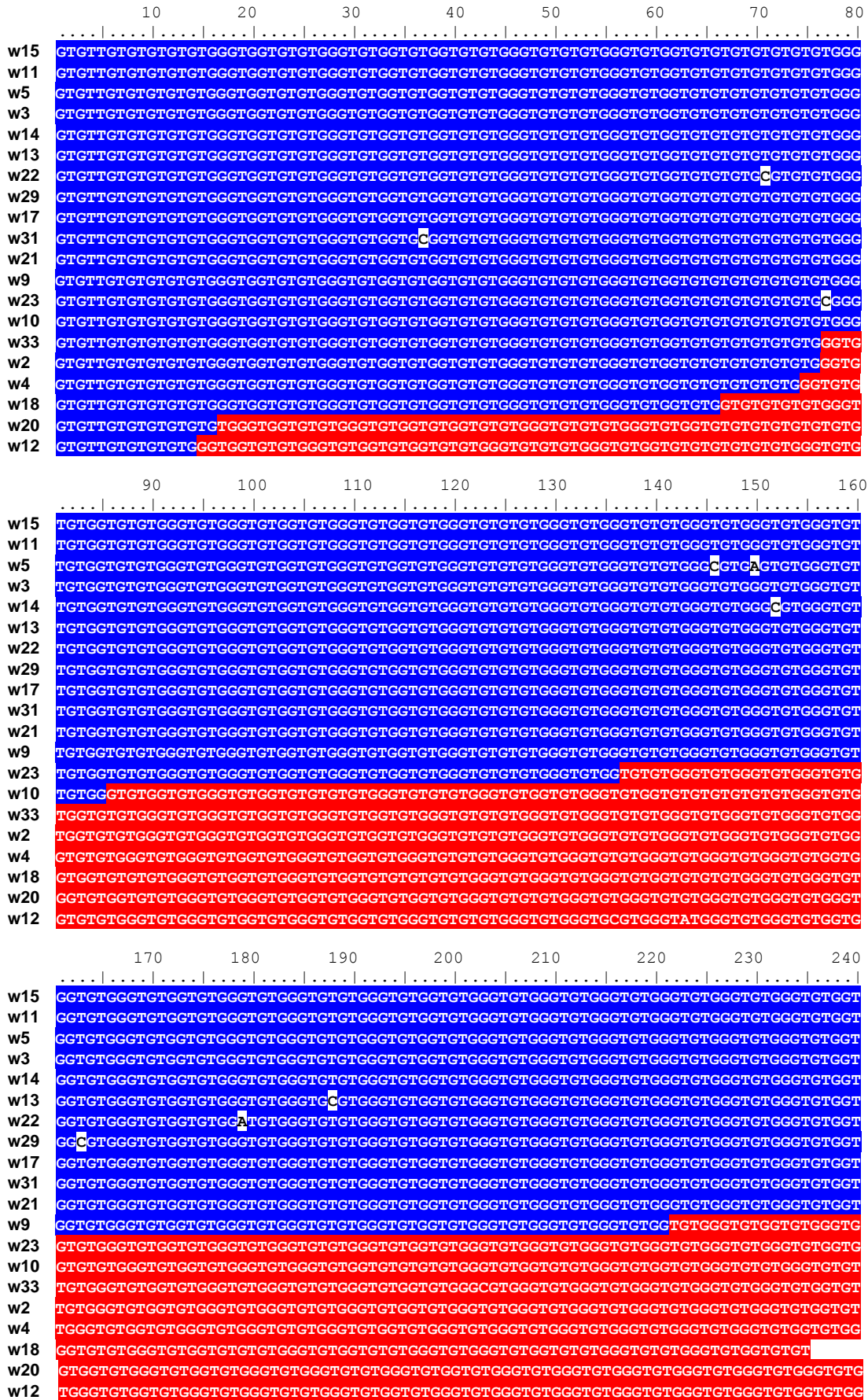


**Supplementary Figure 1. Deletion of the *TSA1* gene causes increase ROS in 20% oxygen condition.** The wild-type and *tsa1* spores were dissected from a *TSA1/tsa1* heterozygous strain. Immediately after dissection, one plate was allowed to germinate in a closed jar in which oxygen was depleted by GasPak EZ Gas Generating Sachets (low O<sub>2</sub>). Another plate was incubated at ambient oxygen conditions (20% O<sub>2</sub>). After 5 days, an aliquot of cells was stained with DHE and the fluorescence of 30,000 cells were counted using FACS.

A.



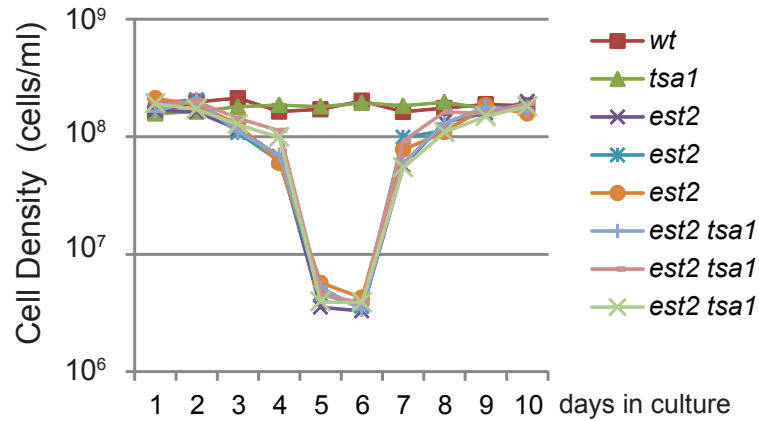




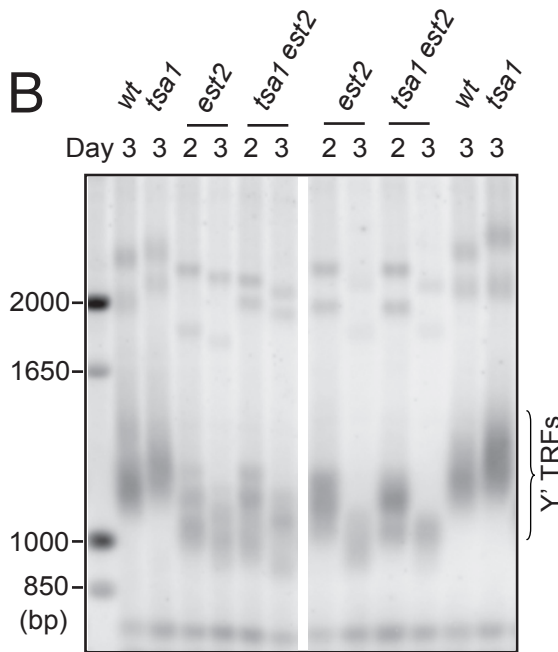


**Supplementary Figure 2. Sequence alignment of telomere XV-L from the wild type (A) and *tsa1* mutant (B).** Telomere XV-L (TEL15L) was PCR amplified, cloned, and sequenced from an early passage (~50 generations after spore germination) of the wild type and *tsa1* mutant. Chromosomal DNA was isolated from corresponding strains and multiple Cs were added to chromosome ends. Tel XV-L was amplified by telomere PCR using polyG oligonucleotide and a primer specific for Tel XV-L. PCR product was cloned into pCR2.1 vector and sequenced. Sequences of telomere repeats (TG<sub>1-3</sub>) were aligned. The internal identical sequences were shaded with blue. The terminal divergent sequences were shaded with red.

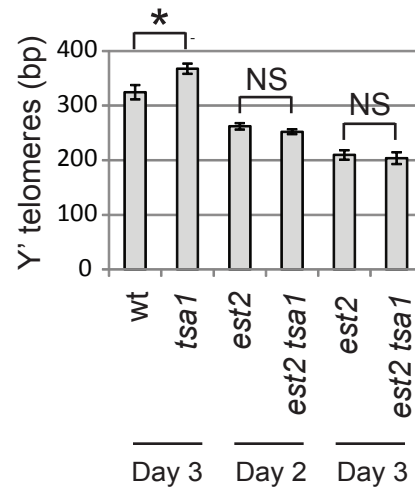
**A**



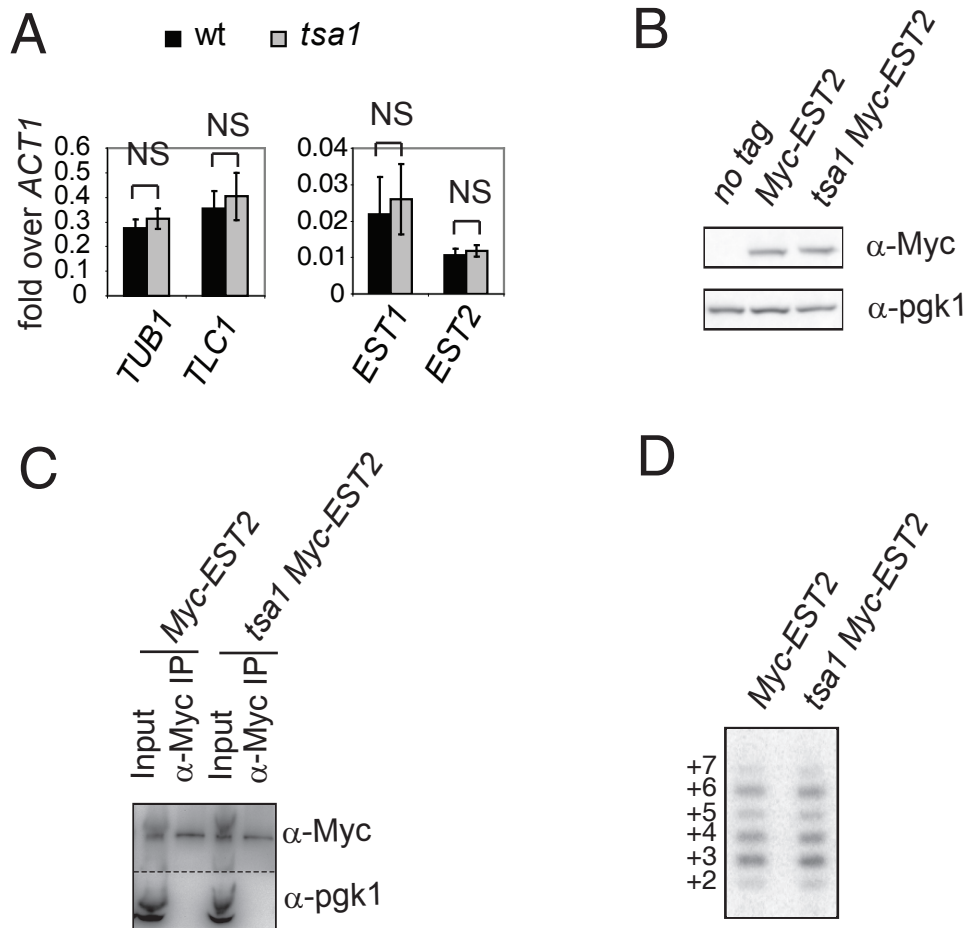
**B**



**C**

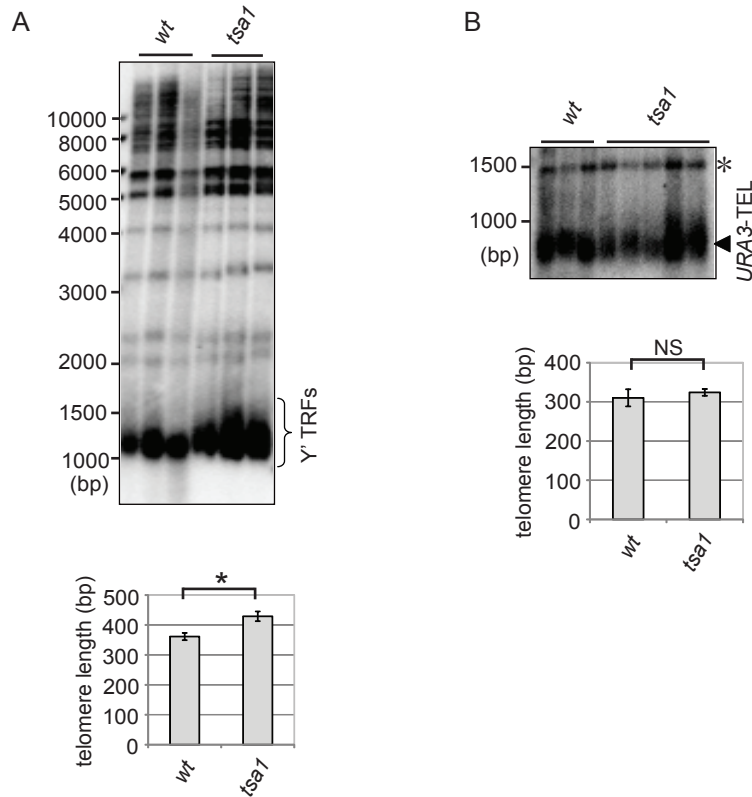


**Supplementary Figure 3. Senescence and telomere length are similar between the *tsa1* and *tsa1 est2* mutants.** (A) Cells with indicated genotype were picked from a fresh dissection plate and inoculated into YEPD media. Every 24 hr, cell density was measured using a hemacytometer. For wild type and *tsa1* cells, the culture was diluted to 10<sup>5</sup> cells/ml and grown to saturation (estimated ~11 generations daily for non-senescent cells). Strains carrying *est2* mutation were grown similarly as wild type before day 4 and after day 7. Between day 4 and day 7, the cultures were started with 10<sup>5</sup> cells/ml, but went through varied numbers of generations. (B) Telomere length of cultures at indicated time was analyzed by Southern blot using probe specific for telomere repeats (TG<sub>1-3</sub>). Typical clones were shown. (C) Telomere length was calculated by subtract subtelomere length from Y' TRFs and shown as mean±SD. Error bars indicate SD from the results of least three independent clones. \*, p<0.05; NS, not significant; Student's t-test.

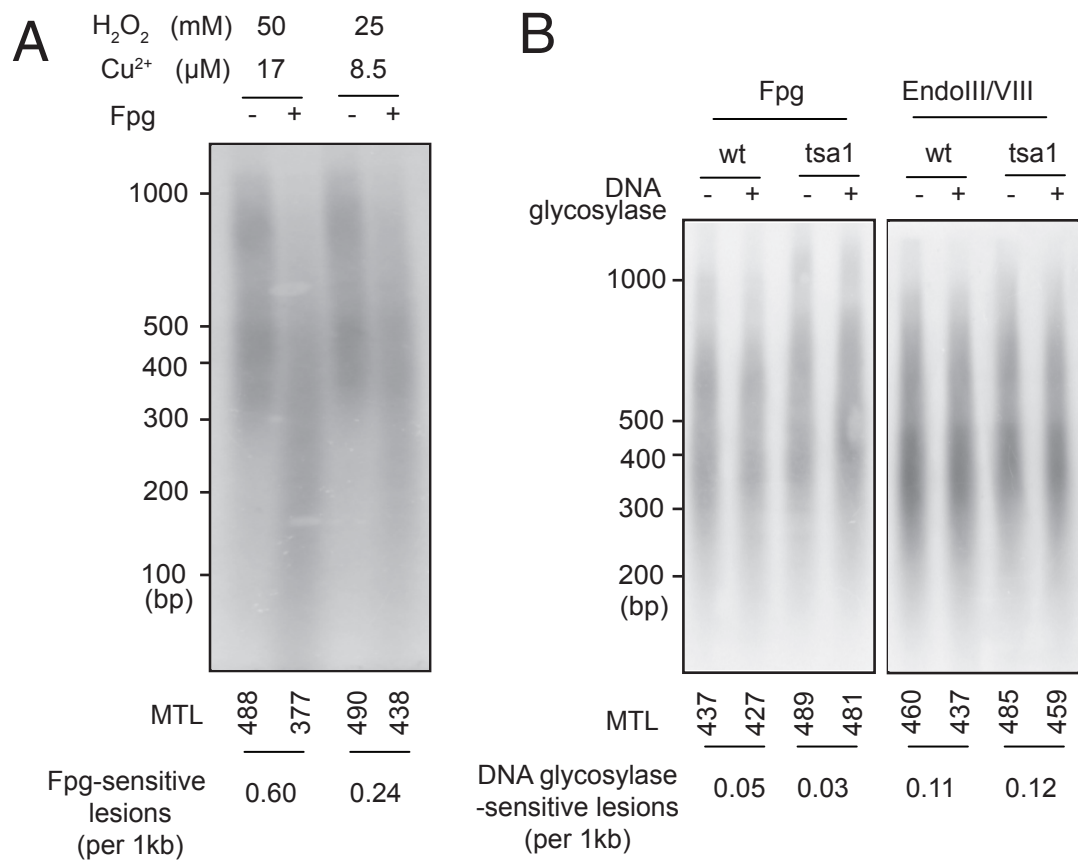


**Supplementary Figure 4. Expression of telomerase subunits and telomerase activity are not changed in the *tsa1* mutant.** (A) Total RNA was isolated from the wild-type (black bars) and *tsa1* (grey bars) strains. mRNA level of *EST1*, *EST2*, *TUB1*, *ACT1*, and *TLC1* RNA were monitored by real-time RT-PCR. The expression level of telomerase components and *TUB1* were normalized to the value for *ACT1*. (B) Whole cell extracts of *EST2* (no tag), *Myc-EST2*, and *tsa1 Myc-EST2* strains were analyzed by Western blot using  $\alpha$ -Myc and  $\alpha$ -pgk1 antibodies. Lack of any signal in the strain expressing *EST2* (no tag) in  $\alpha$ -Myc blot validates the specificity of Myc-tagged Est2. (C) Est2 protein was immunoprecipitated using  $\alpha$ -Myc antibody ( $\alpha$ -Myc IP) from whole cell lysate (Input) in a strain carrying Myc-tagged *EST2* allele or its *tsa1* derivative. The blot was probed with  $\alpha$ -Myc and  $\alpha$ -pgk1 antibodies. The wild-type and *tsa1* strains did not show any difference in Est2 level and IP efficiency. (D) Myc-EST2 and its associated telomerase components were mixed with a telomeric template oligonucleotide and radio-labeled dGTP. The wild-type and *tsa1* strains did not show any difference in primer-extension.

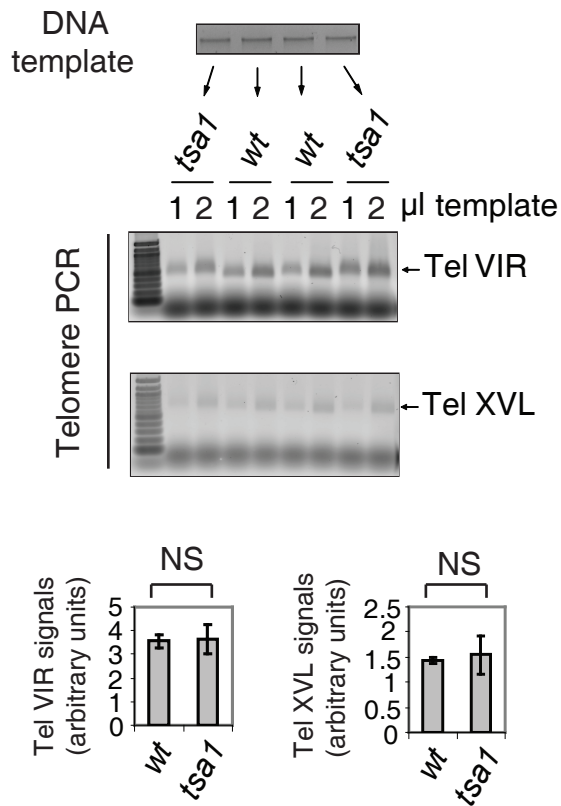




**Supplementary Figure 5. Telomere length analysis of a strain carrying URA3-Telomere.** Individual clones of UMY2585 (wt) and its *tsal* derivatives (*tsal*) were analyzed for Y' telomeres (A, telomere repeat probe) and URA3-TEL (B, *URA3* probe). \* indicated an unspecific band detected by URA3 probe as a control for DNA fragment migration. The length of TRFs was subtracted by that of *URA3* fragment (~424 bp) or Y' element fragment (~875 bp) to yield *URA3*-TEL or Y'-containing telomere length (shown under each lane). Representative clones are shown. The length of telomeres was compared between two groups using Student's t test. Error bars indicate SD from independent clones. NS, not significant; \*, p<0.05.



**Supplementary Figure 6. Measurement for oxidative DNA lesions in the *tsa1* mutant.** (A) For the positive control, genomic DNA from the wild-type cells was treated with H<sub>2</sub>O<sub>2</sub> plus Cu<sup>2+</sup> *in vitro*. Telomere DNA fragments were separated by alkaline gel electrophoresis and detected by Southern blot analysis using a telomere repeat-specific probe. A telomere DNA fragment with oxidative base lesions is converted into smaller fragments by a DNA glycosylase Fpg. The level of DNA glycosylase-sensitive sites were calculated by comparing medium telomere length (MTL) in the same sample treated with (+) or without (-) Fpg using the formula described in the Methods. Each experiment was repeated twice and representative data were shown. High oxidative exposure causes significant increase in Fpg-sensitive lesions that can be easily detected as shorter telomeric DNA fragments and MTL in Fpg (+) samples. (B) Genomic DNA from the wild type or *tsa1* mutant was restricted by 4-base cutters (AluI, HaeI, HinfI, and MspI), treated with (+) or without (-) DNA glycosylase (Fpg, left panel, and EndoIII/VIII, right panel) separated on alkaline gel. DNA lesions were analyzed as in (A).



**Supplementary Figure 7. Measurement for DNA blocking lesions in telomere DNA.** Genomic DNA was prepared from the wild-type and *tsa1* mutant cells and adjusted to the same concentration (top panel). Tel VIR and Tel XLV were amplified by PCR. Different amount of templates were used to ensure PCR cycles fall in linear detection range. PCR product was run on agarose gel and stained with Sybr Green. After subtraction of background, intensity of telomere PCR signals were calculated and average intensity (mean $\pm$ SD) of each telomere are shown at the bottom. NS, not significant; Student's t-test.

Table S1. Primers used in Quantitative PCR.

<b>Name</b>	<b>Sequence</b>
TLC1 forward	5'-CCATGGGAAGCCTACCAT-3'
TLC1 reverse	5'-GAACTCGTGCAAACCTTTGCTAA-3'
U2 forward	5'-TCGATGGGAAGAAATGGTGCTATAG-3'
U2 Reverse	5'-GATAAATAATGTGCCGGATCCCGAAAACTTCCTCTTGCAGC-3'
EST1 forward	5'-CTGCATTAAAGTATCATTTCAGGTAA-3'
EST1 reverse	5'-CGAGTTGTAAACATAAGATCGAAA-3'
EST2 Forward	5'-AACAGACCAACAGCAAGTGA-3'
EST2 Reverse	5'-GATAAATAATGTGCCGGATCCTCTCTATTGGCTTTCGCATTA-3'
ACT1 Forward	5'-TCCGGTGATGGTGTTACTCA-3'
ACT1 Reverse	5'-ATTCTCAAATGGCGTGAGG-3'