

Supplementary Information

Tetrasaccharide iteration synthesis of a heparin-like dodecasaccharide and radiolabelling for *in vivo* tissue distribution studies

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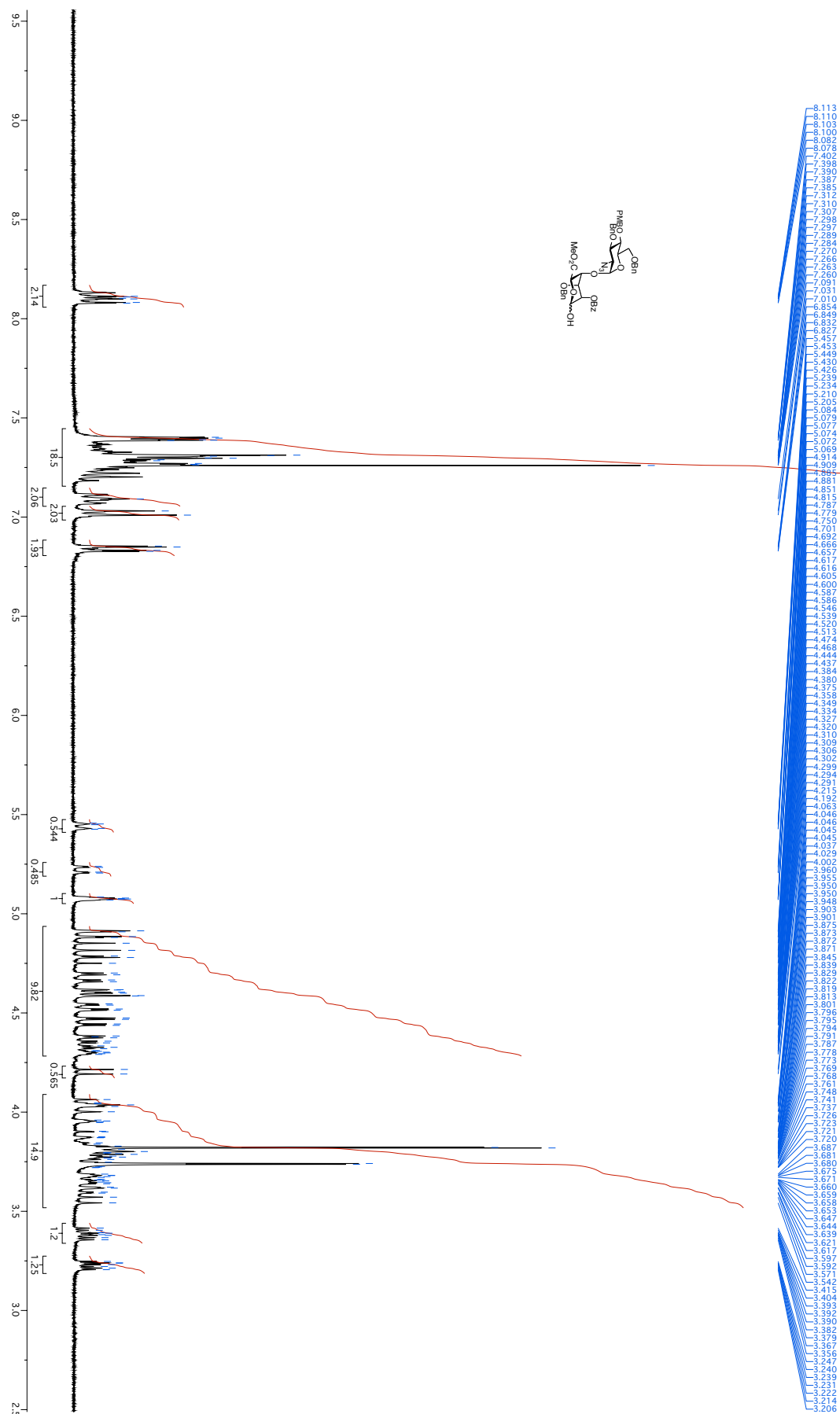
Supplementary figures

NMR and Mass spectra

Supplementary methods

Synthesis of oligosaccharides

Supplementary figures



Supplementary Figure S1. ¹H NMR (400 MHz; CDCl₃) spectrum for 3.



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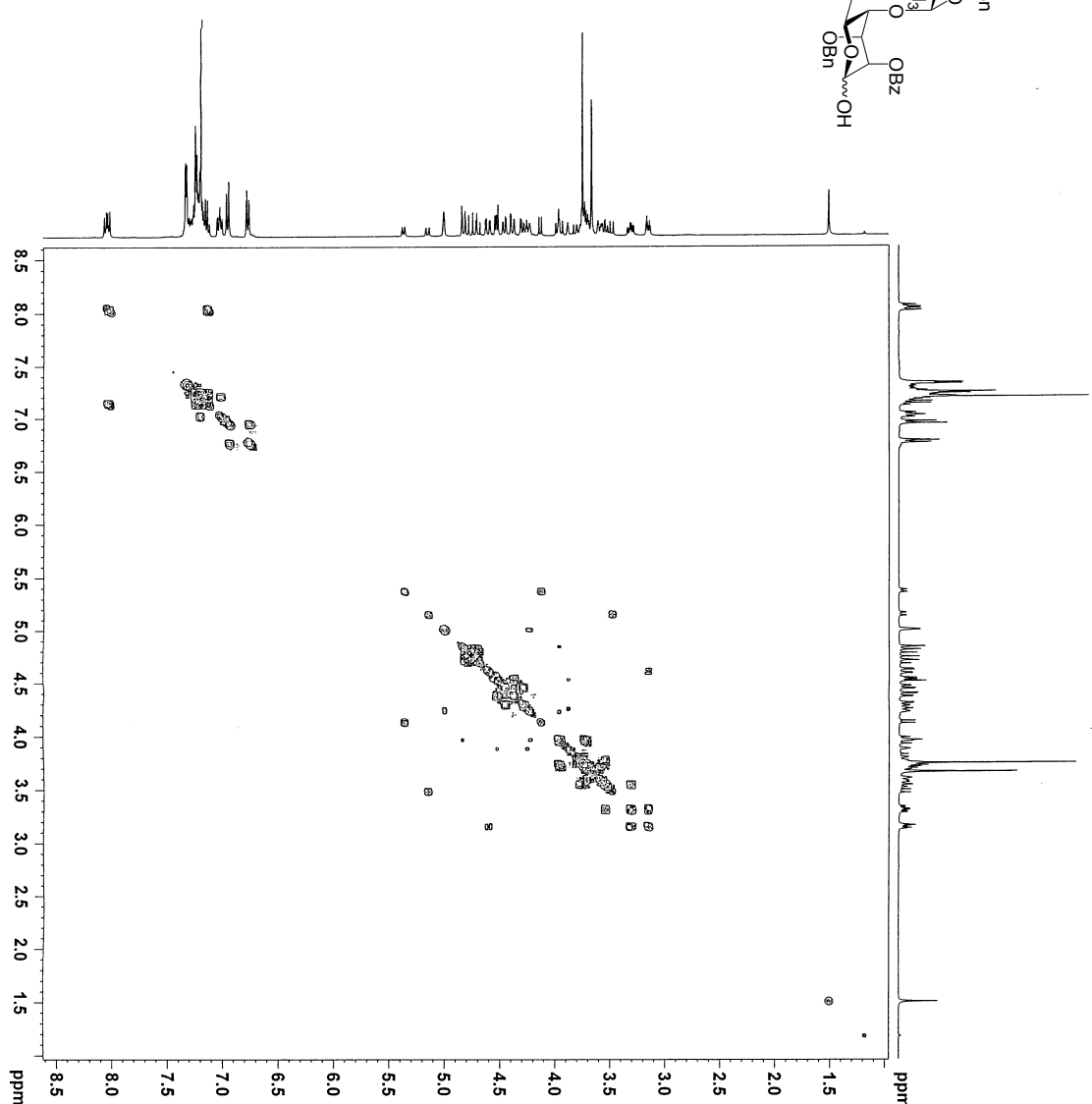
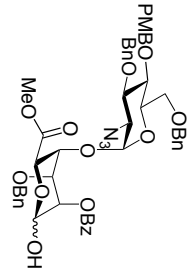
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 GEZ2 10.00 %
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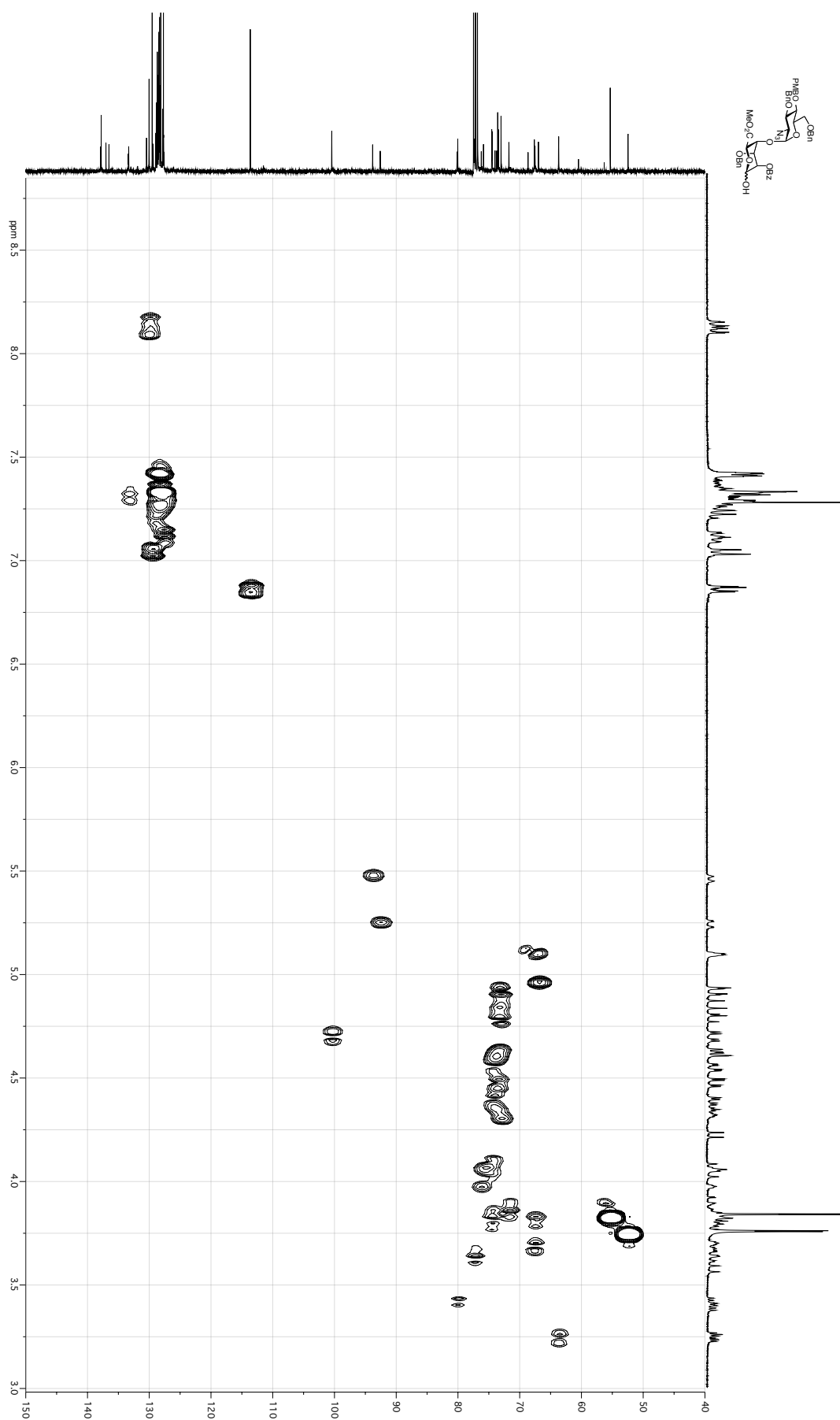
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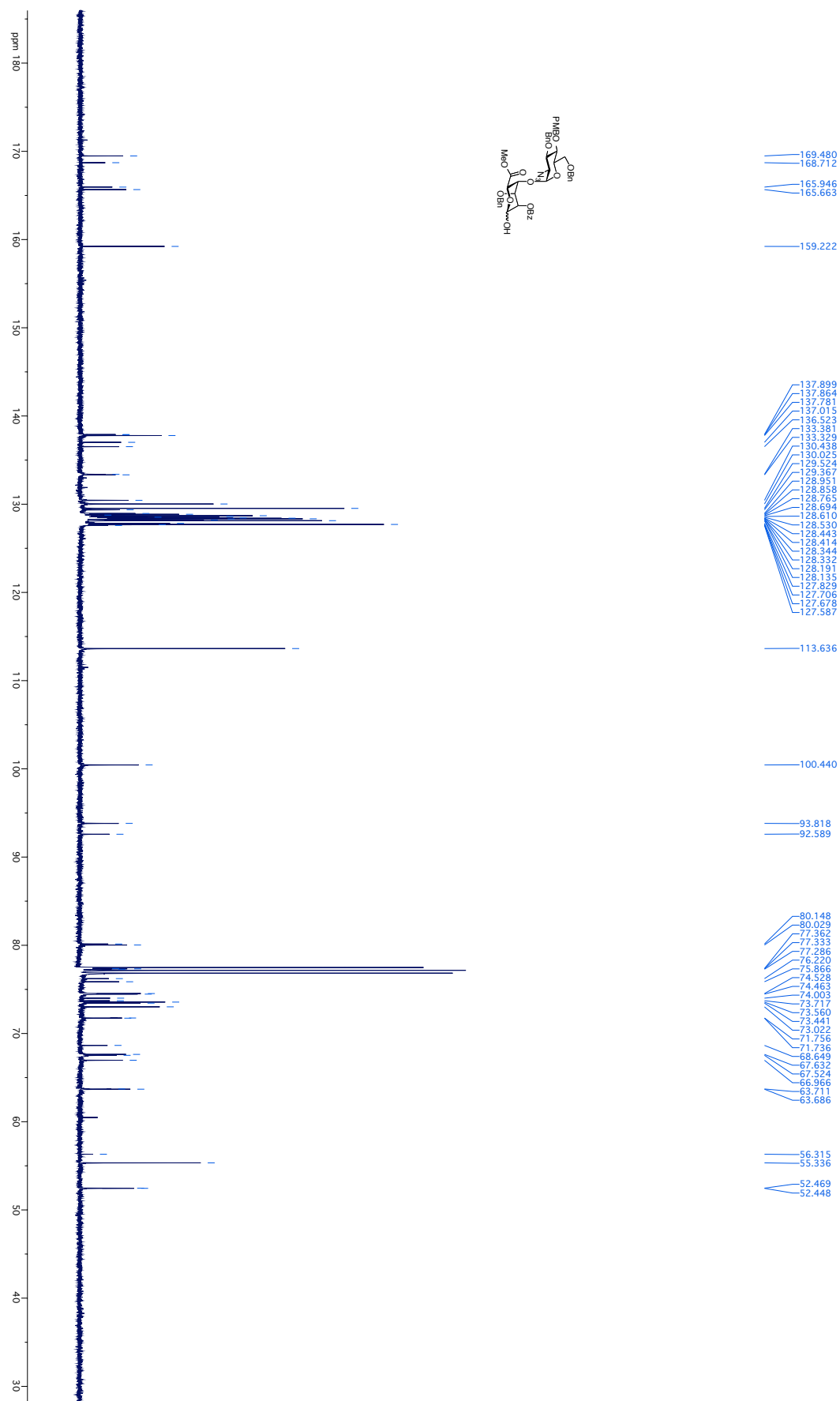
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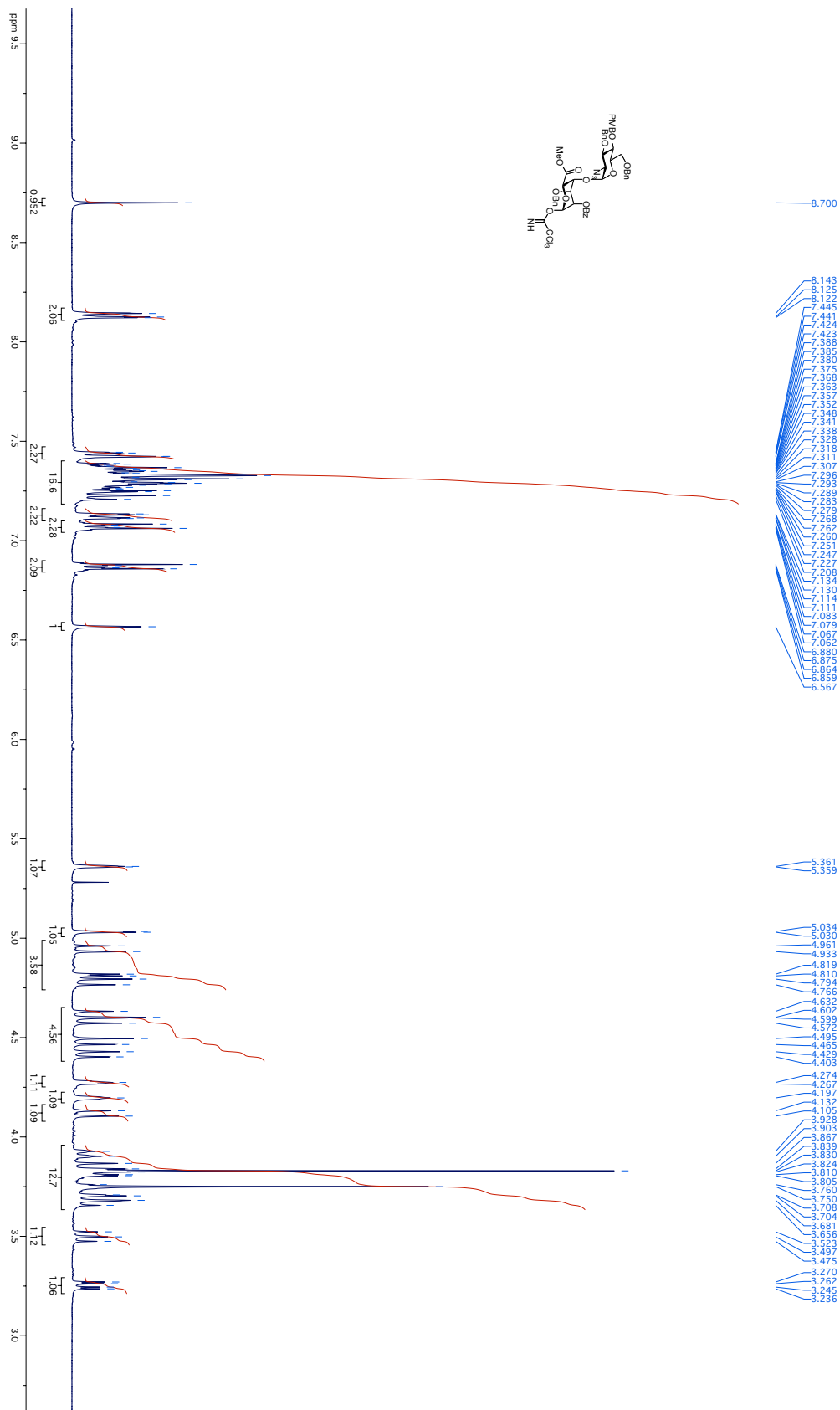
Supplementary Figure S2. COSY NMR spectrum for 3.



Supplementary Figure S3. HMQC NMR spectrum for 3.

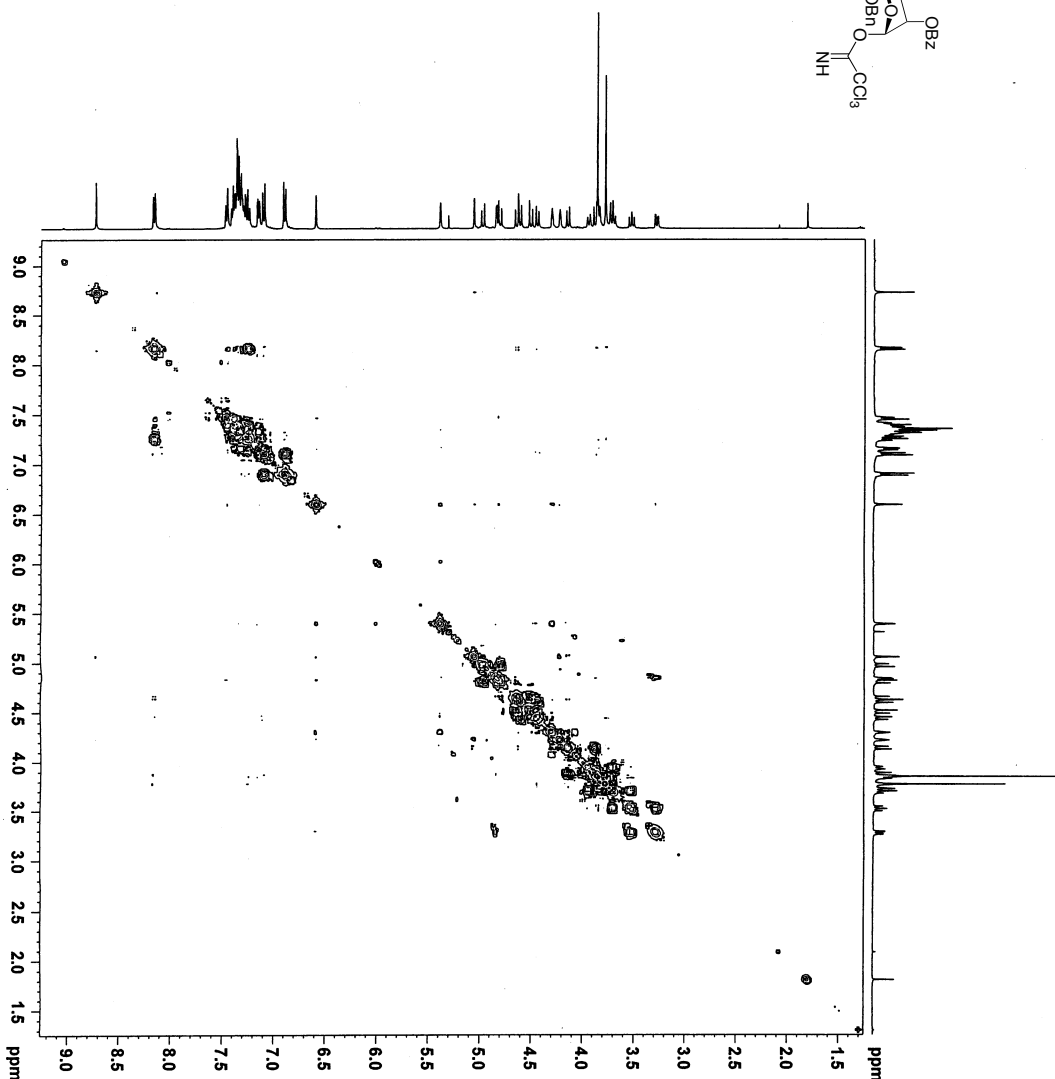
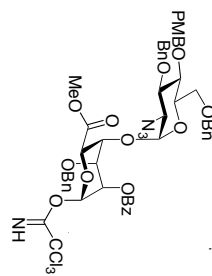


Supplementary Figure S4. ^{13}C NMR (100 MHz; CDCl_3) spectrum for 3.



Supplementary Figure S5. $^1\text{H NMR}$ (400 MHz; CDCl_3) spectrum for α -4.

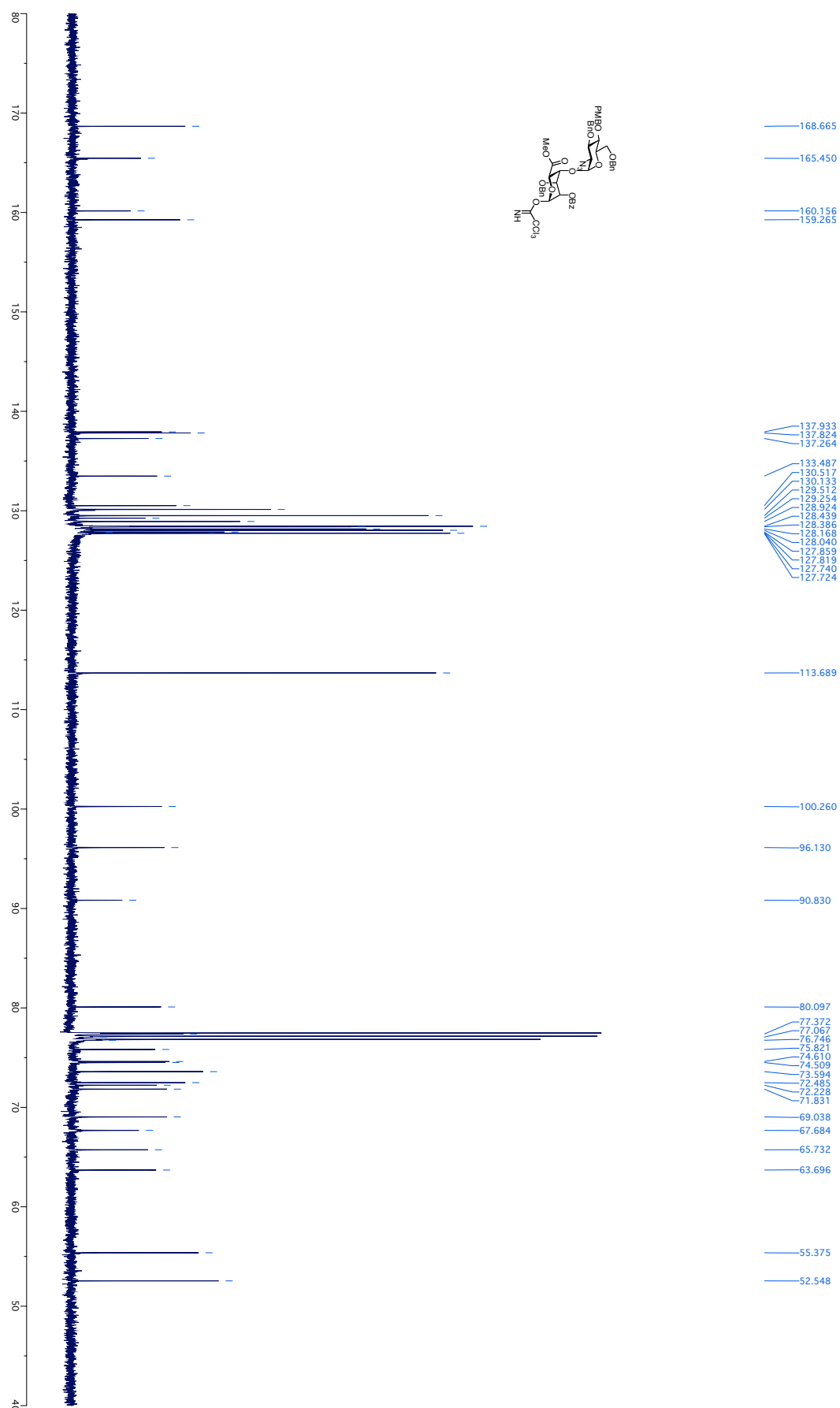
SU97A



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 RG 1024
 INEPT 25.4
 DE 156.000
 TE 300.0 K
 D1 0.00000300 sec
 D13 0.00000400 sec
 D15 0.00020000 sec
 INO 0.00031200 sec
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 PL1 -0.10 dB
 SFO1 400.1321048 MHz
 GRAB1 GRAB1 CHANNEL
 GRAB2 SINE 100
 GP42 10.00 usec
 P16 1000.00 usec
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 SFO1 400.1321048 MHz
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 SW 8.010 ppm
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 F2 - Processing Parameters
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 SF 400.1300000 MHz
 SE 3196
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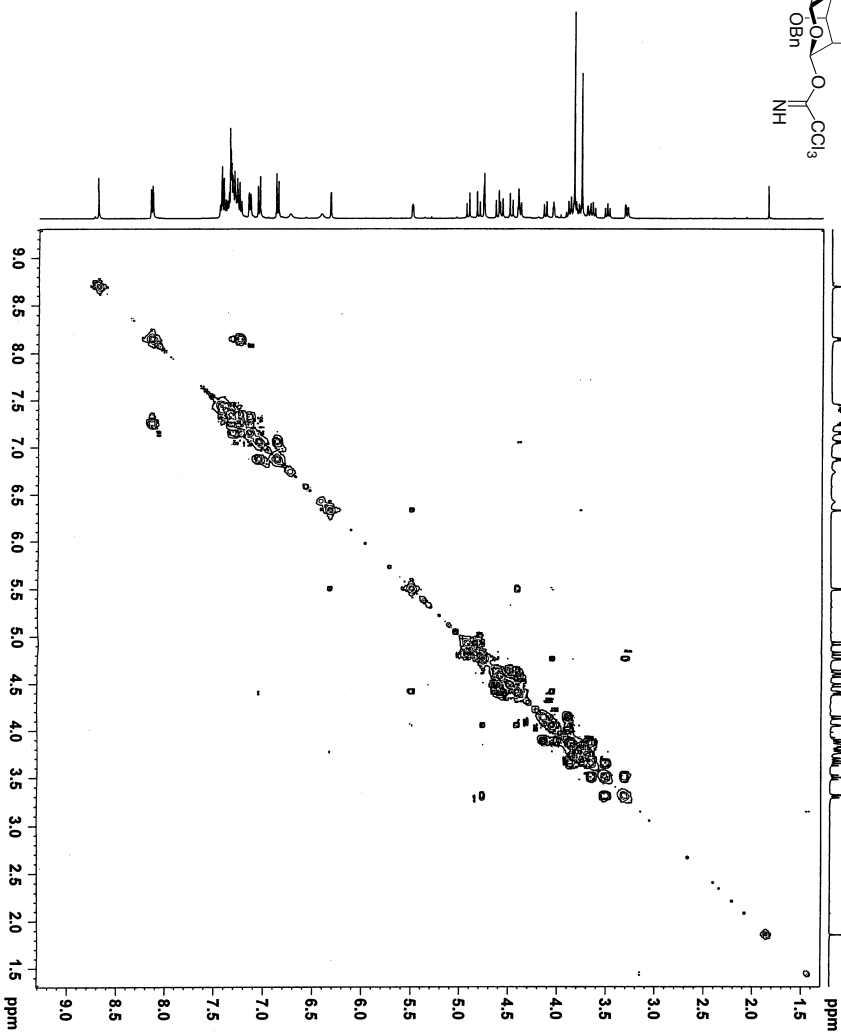
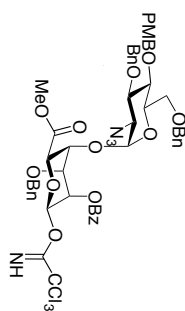


Supplementary Figure S6. COSY NMR spectrum for α -4.



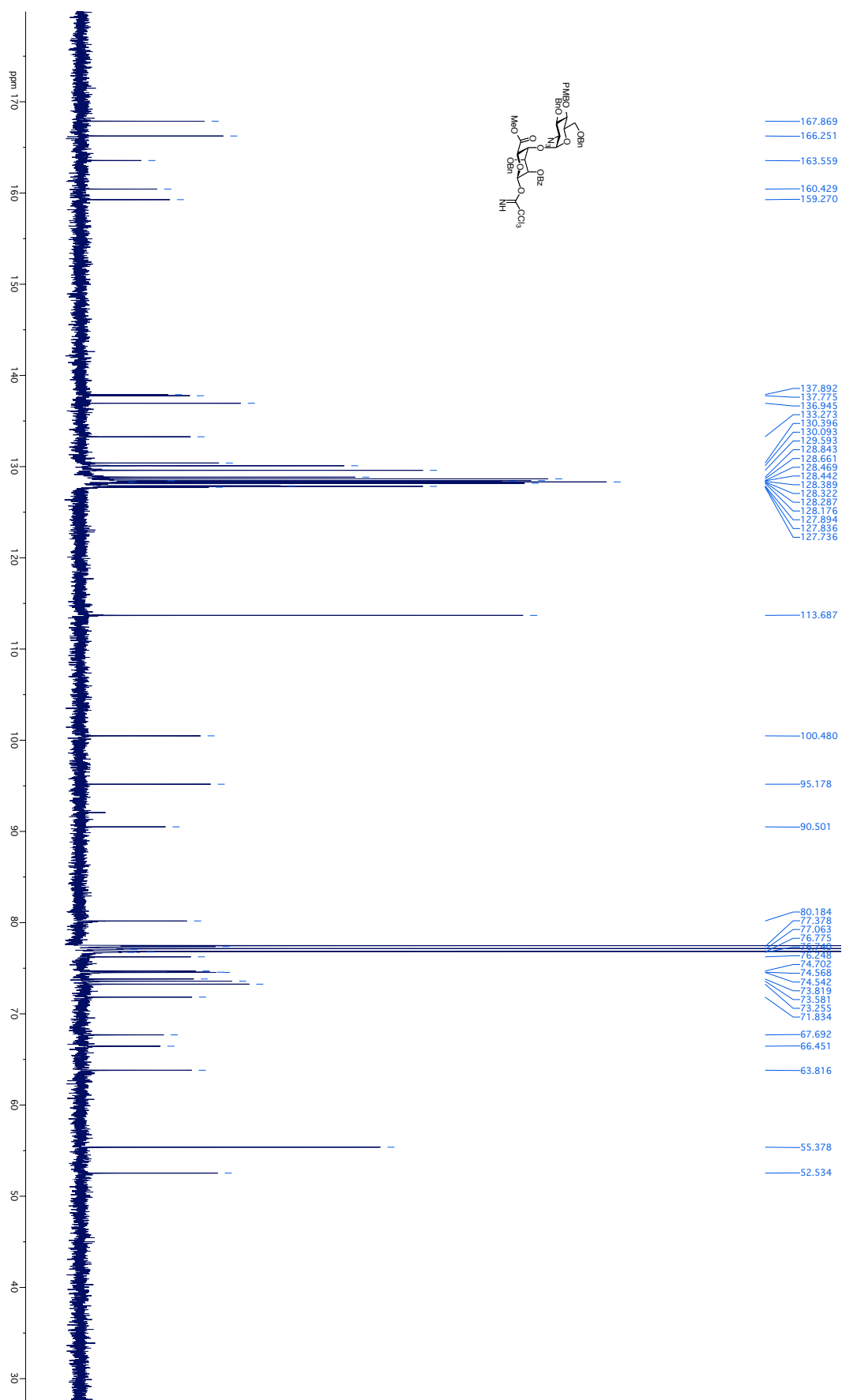
Supplementary Figure S8. ^{13}C NMR (100 MHz; CDCl_3) spectrum for α -4.

SU997B

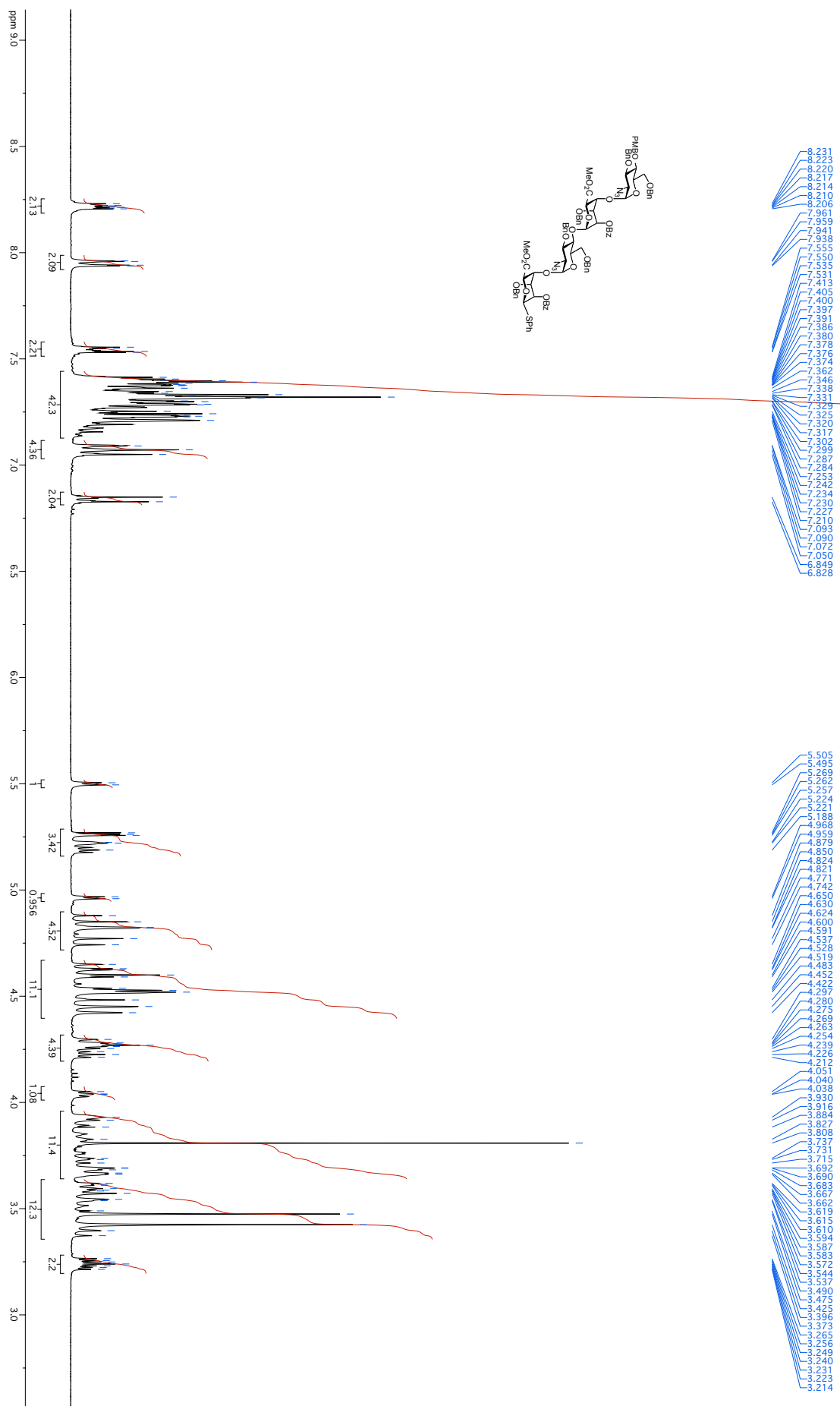


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 DS 4
 SFR 1205.128 Hz
 AQ 0.01666667 sec
 HZ 0.319380 sec
 RG 26.5
 DE 15.00 usec
 TE 300.0 K
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 DI 0.00000000 sec
 D16 0.00020000 sec
 D10 0.19025000 sec
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 P1 10.00 usec
 SFO1 400.132133 MHz
 GRADIENT CHANNELS
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 GRNAM2 SINE100
 STW1 10.00 usec
 STW2 10.00 usec
 P16 1000.00 usec
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 SFOBS 204.0641 MHz
 SF 8.010 Ppm
 FWHM 0.9
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 SF 400.1321 MHz
 MDW 0.00 Hz
 SSB 0.00 Hz
 LB 1.40
 PC 1.40
 F1 - Processing parameters
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 LB 0.00 Hz
 GB 0.00 Hz

Supplementary Figure S10. COSY NMR spectrum for β -4

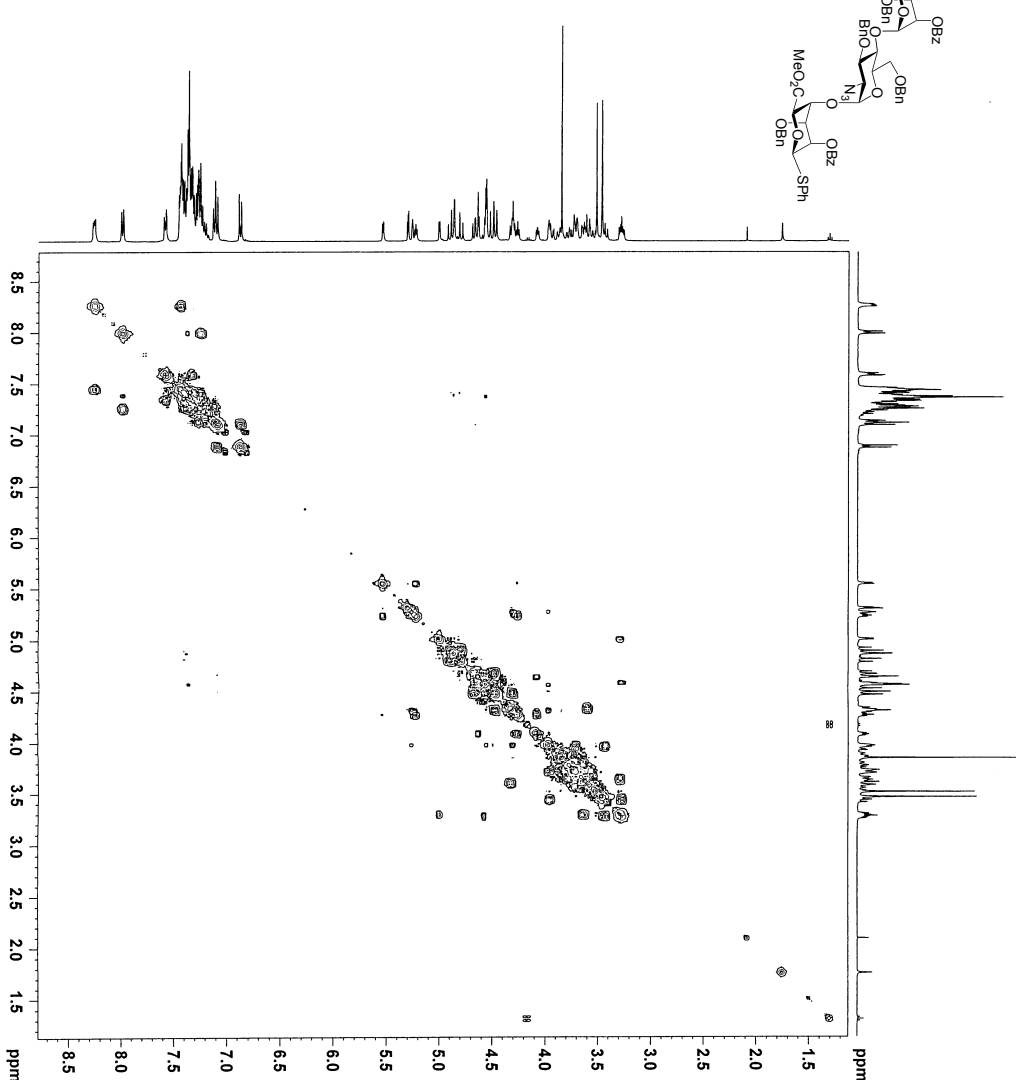
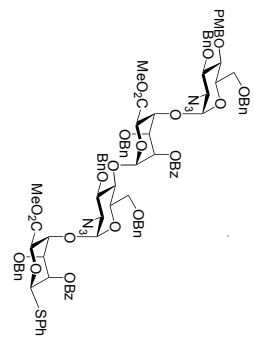


Supplementary Figure S12. ^{13}C NMR (100 MHz; CDCl_3) spectrum for β -4



Supplementary Figure S13. ^1H NMR (400 MHz; CDCl_3) spectrum for β -6

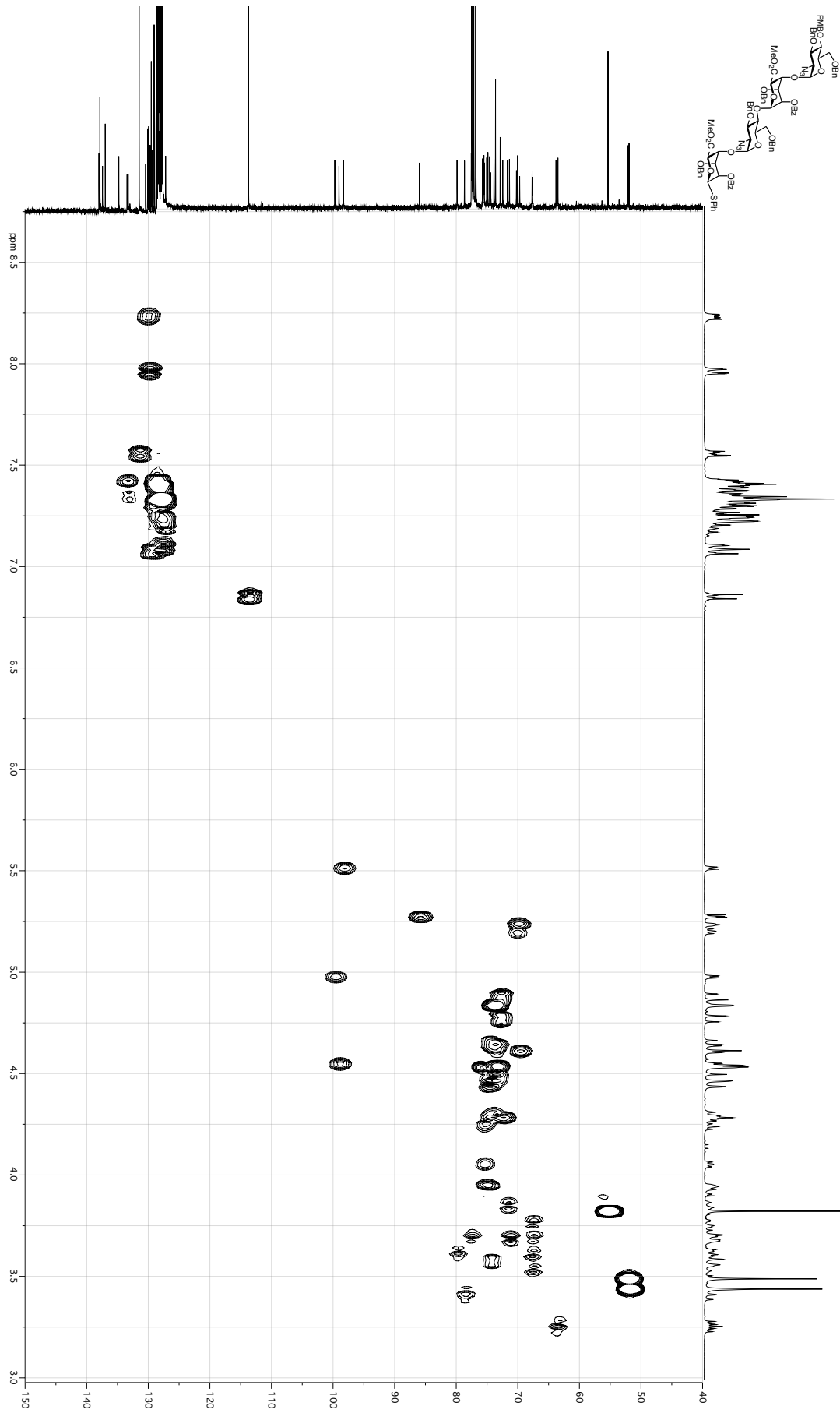
SU998



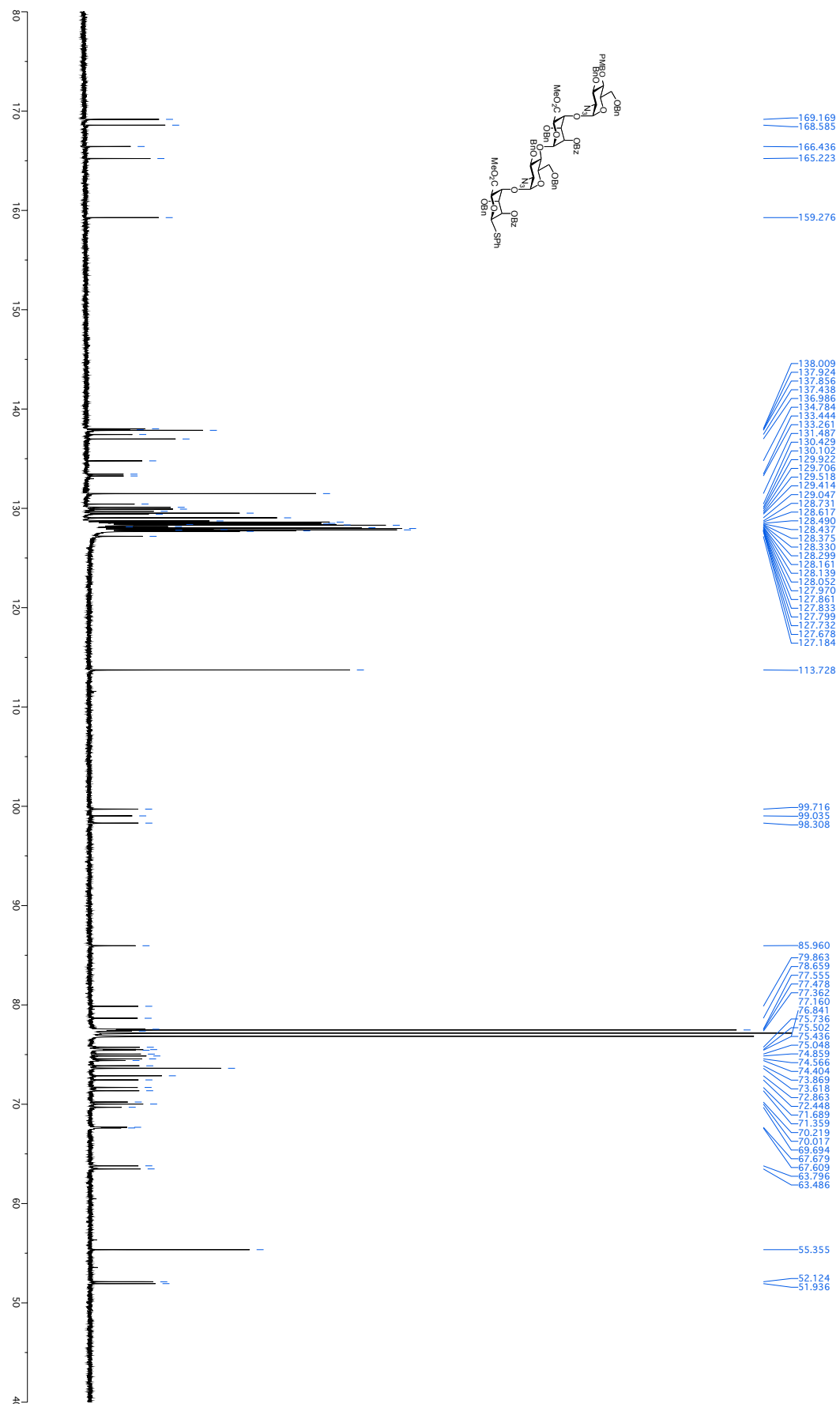
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DS        8
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AQ        0.3347836 sec
RG         22.6
WDW        163.200 usec
SSB         0.00000000 sec
GB          0.00000000 sec
PC          233.2 K
TE          0.00000000 sec
D16        0.00020000 sec
D18        0.00026400 sec
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PL         -0.10 dB
SFO1       400.1319839 MHz
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SINE      100
GAMMA2    0.00 %
GR22      10.00 %
P16       1000.00 usec
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PL         -0.10 dB
SFO1       400.1319839 MHz
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GR22      10.00 %
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SF         400.1300000 MHz
WDW        0.00 Hz
SSB         0.00 Hz
GB          0.00 Hz
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F1 - Processing parameters
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Supplementary Figure S14. COSY NMR spectrum for β -6

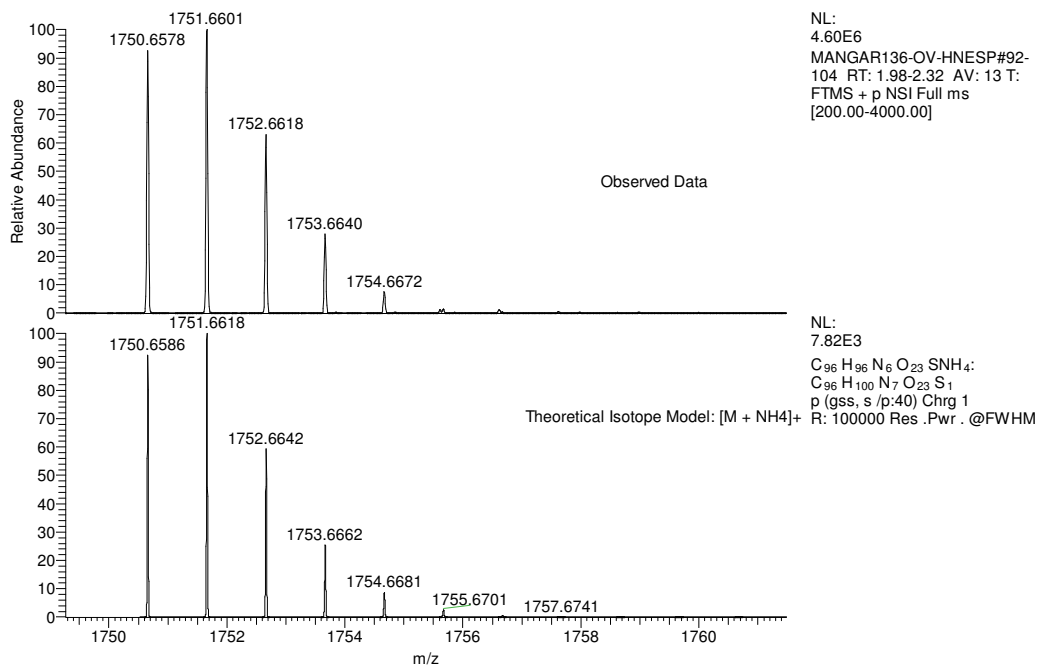
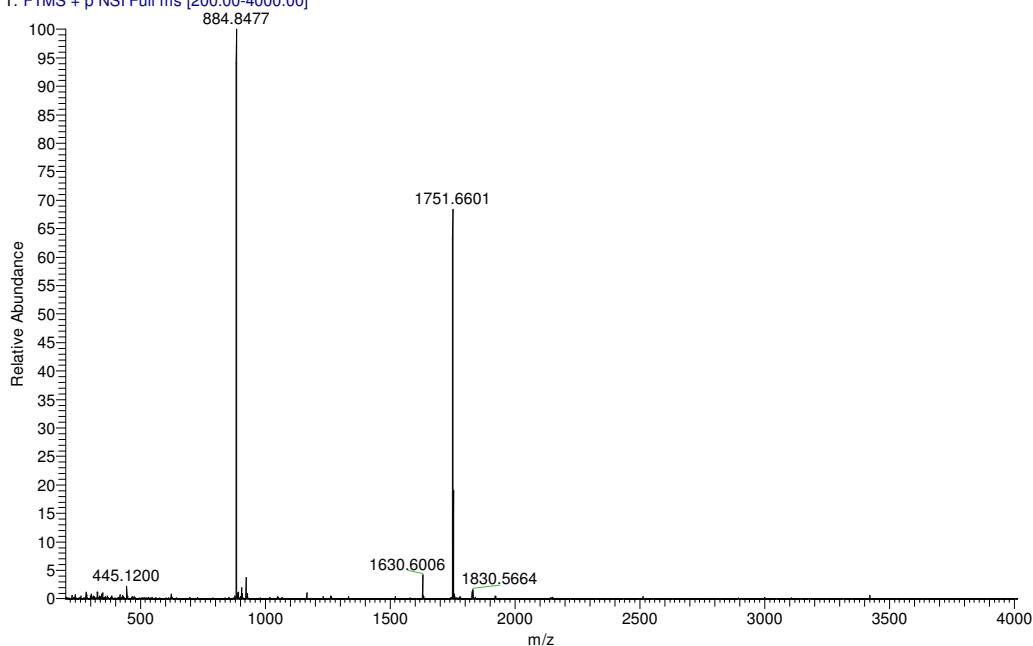


Supplementary Figure S15. HMQC NMR spectrum for β -6

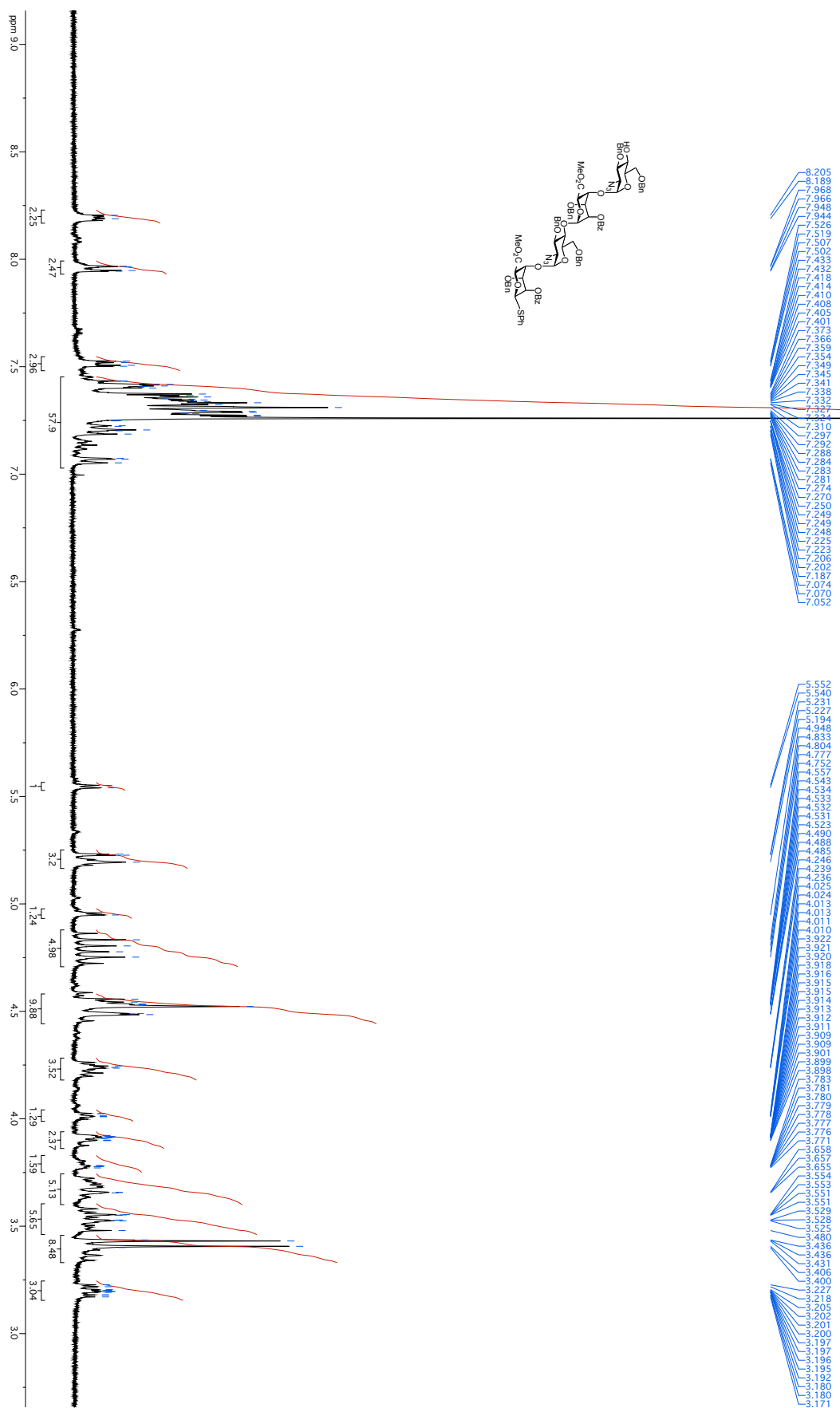


¹ Supplementary Figure S16. ¹³C NMR (100 MHz; CDCl₃) spectrum for β -6

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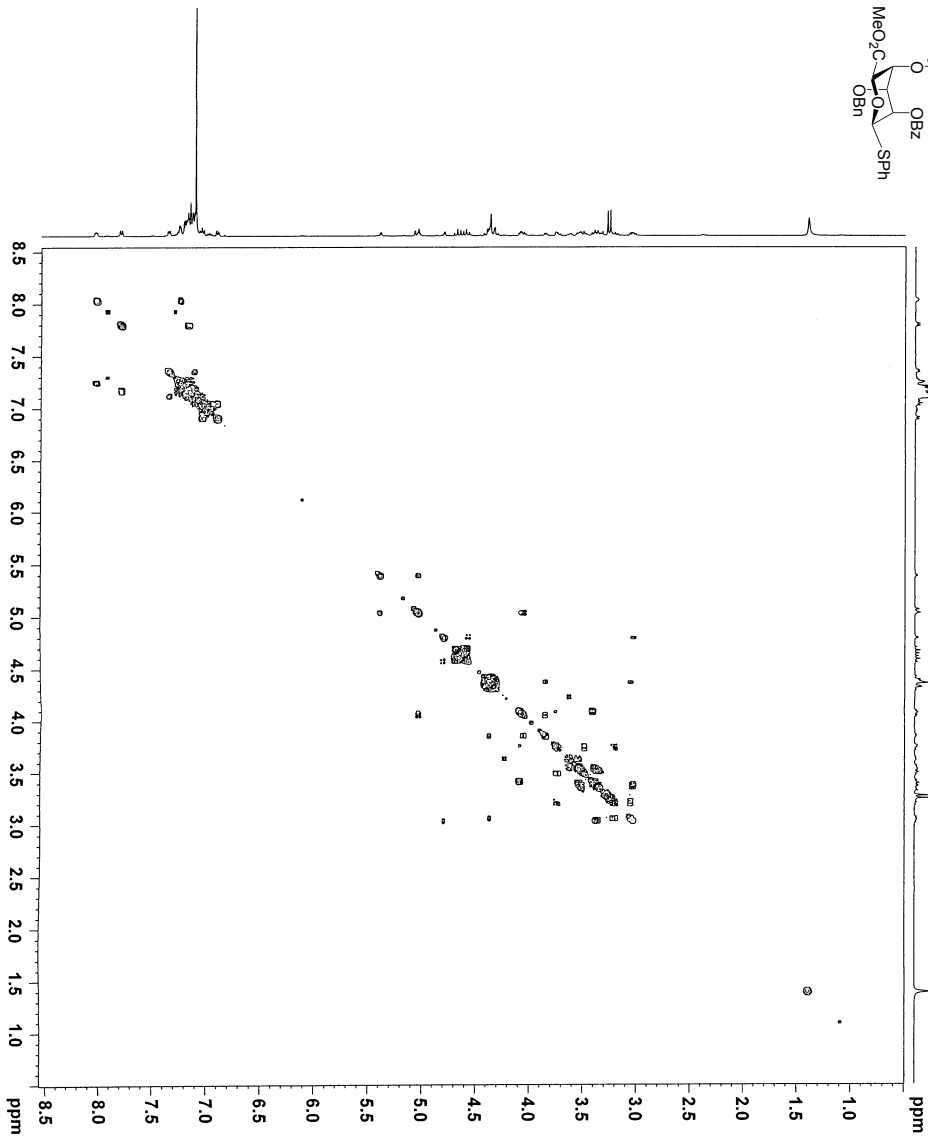
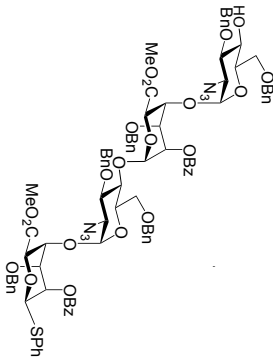


Supplementary Figure S17. HFT MS spectrum for β -6



Supplementary Figure S18. ^1H (400 MHz, CDCl_3) NMR spectrum for β -7

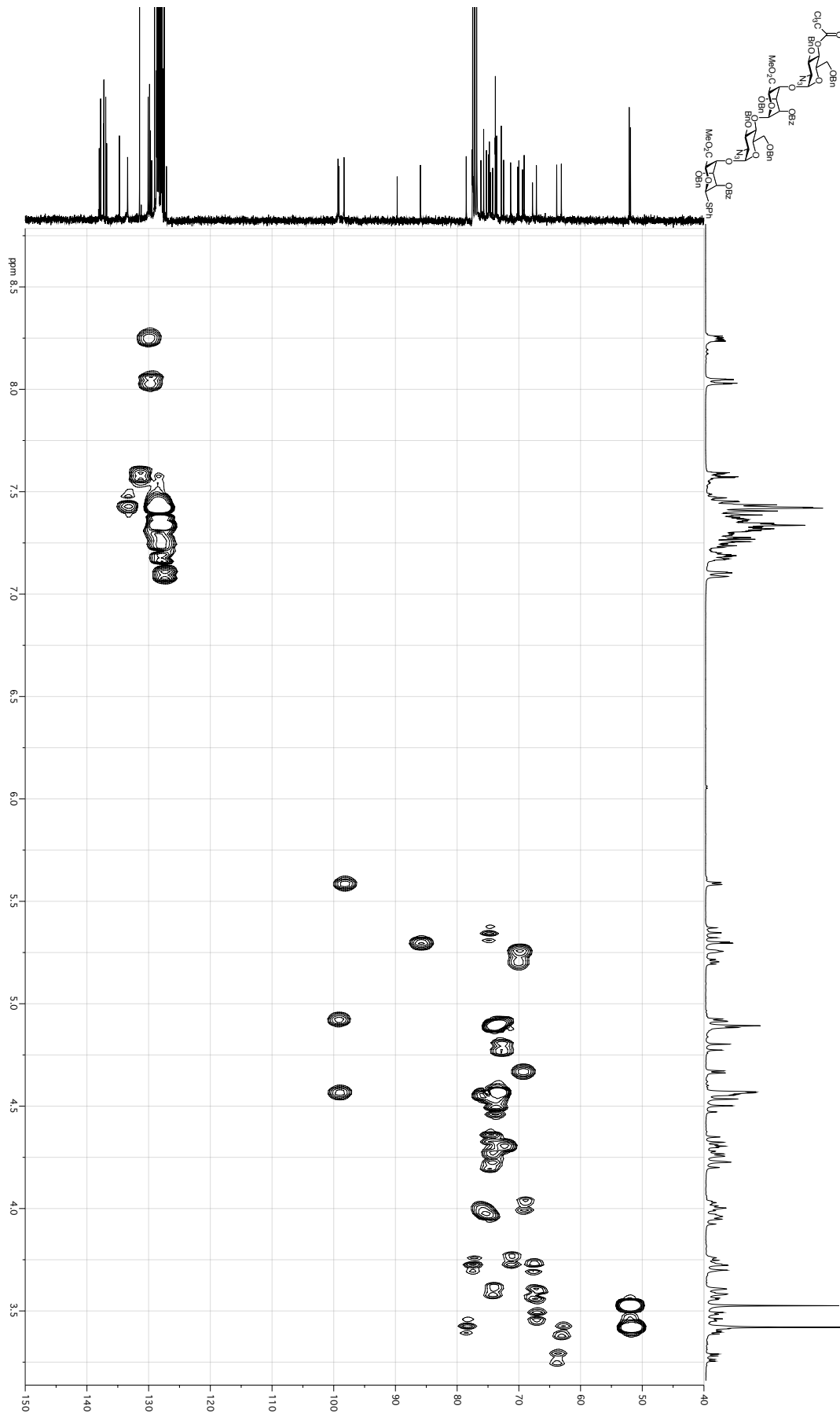
SU1246



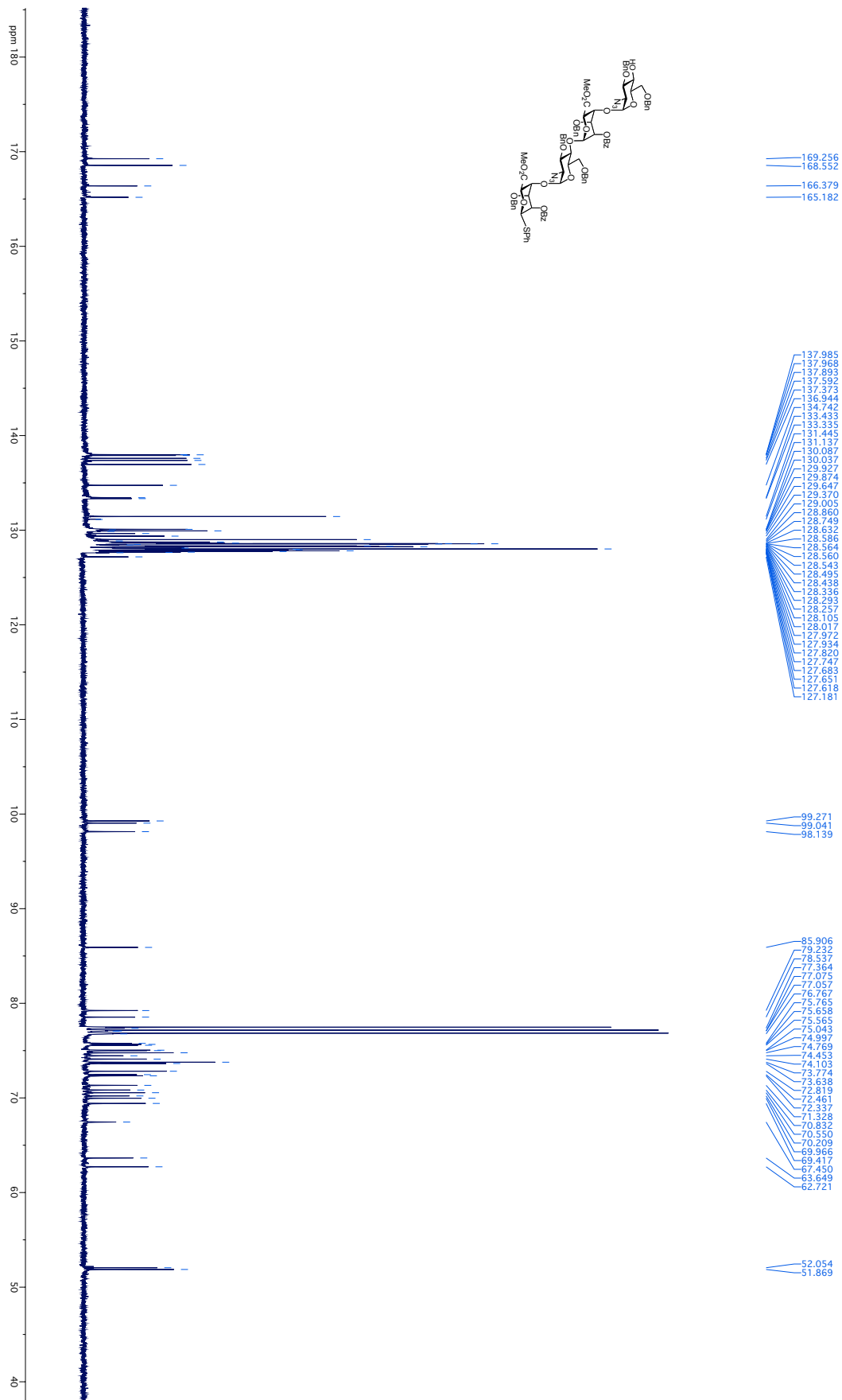
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 DS 4
 SWH 3221.649 Hz
 FIDRES 1.573071 Hz
 AQ 0.3178926 sec
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 TE 300.2 K
 TR 0.0002000 sec
 D1 1.36134502 sec
 d13 0.00000400 sec
 D16 0.00020000 sec
 INO 0.00031040 sec
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 GPCP2 SINE 10.00 usec
 GPCP3 SINE 10.00 usec
 P15 1000.00 usec
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 SF01 400.1319 MHz
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 SM 8.051 ppm
 RMK006 Q1
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 SSB 0
 LB 0.00 Hz
 GB 0
 TC 1.40
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 MDW SINE
 SSB 0
 LB 0.00 Hz
 GB 0
 TC 1.40



Supplementary Figure S19. COSY NMR spectrum for β -7

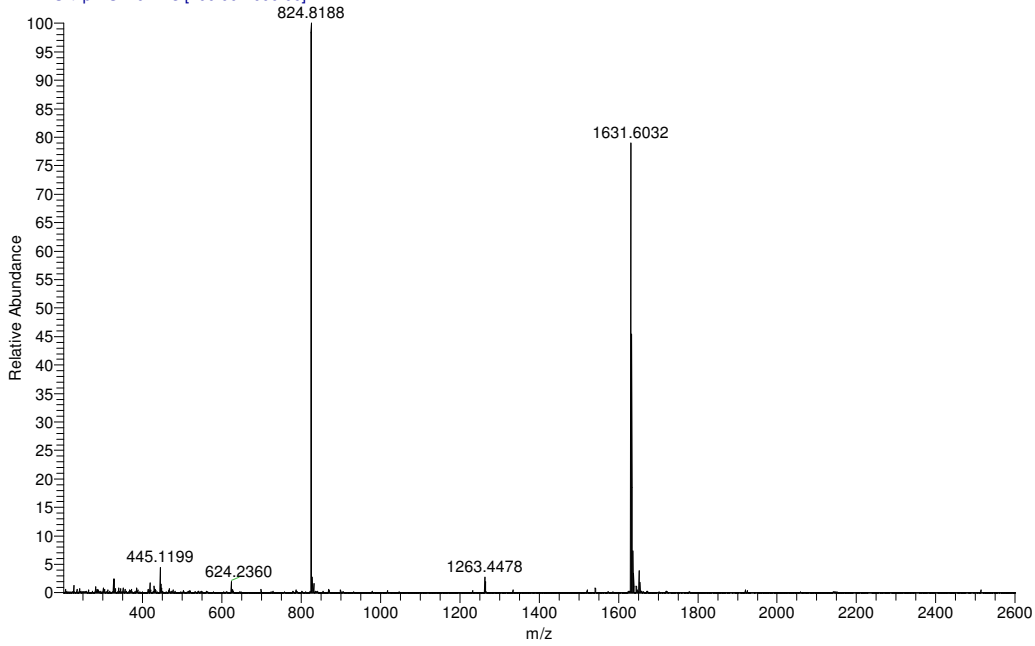


Supplementary Figure S20. HMBC NMR spectrum for β -7

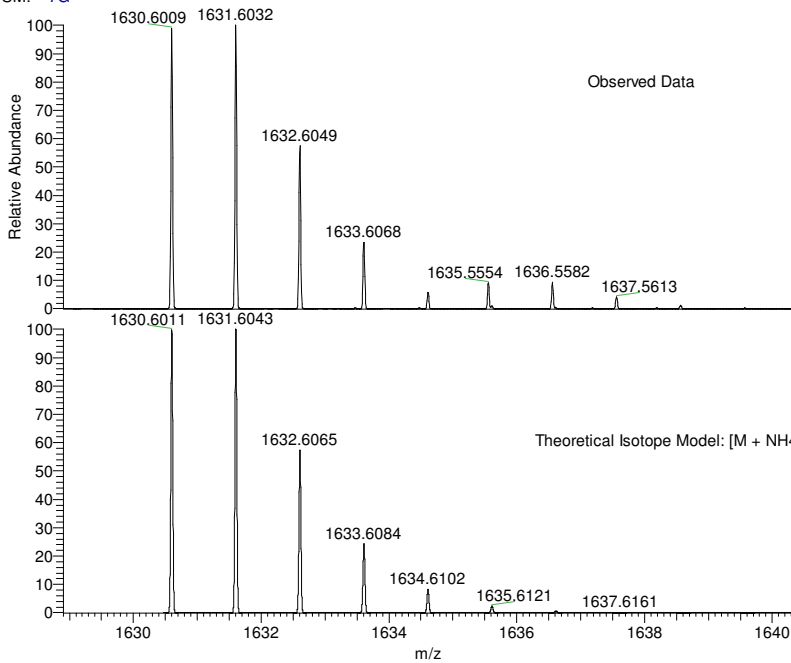


Supplementary Figure S21. $H^{13}C$ NMR (100 MHz; $CDCl_3$) spectrum for β -7

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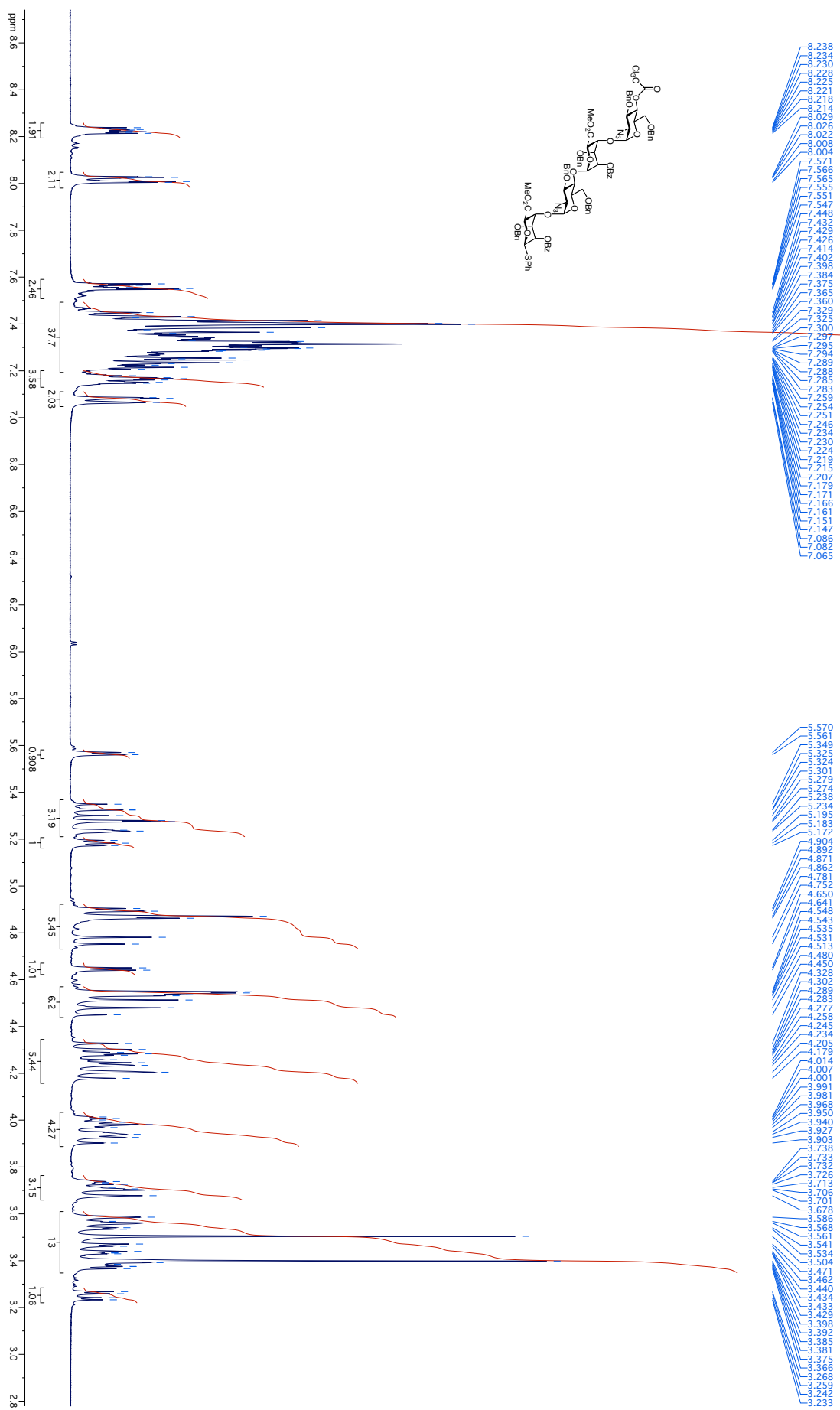
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NL:
2.49E6
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FTMS + p NSI Full ms
[200.00-4000.00]

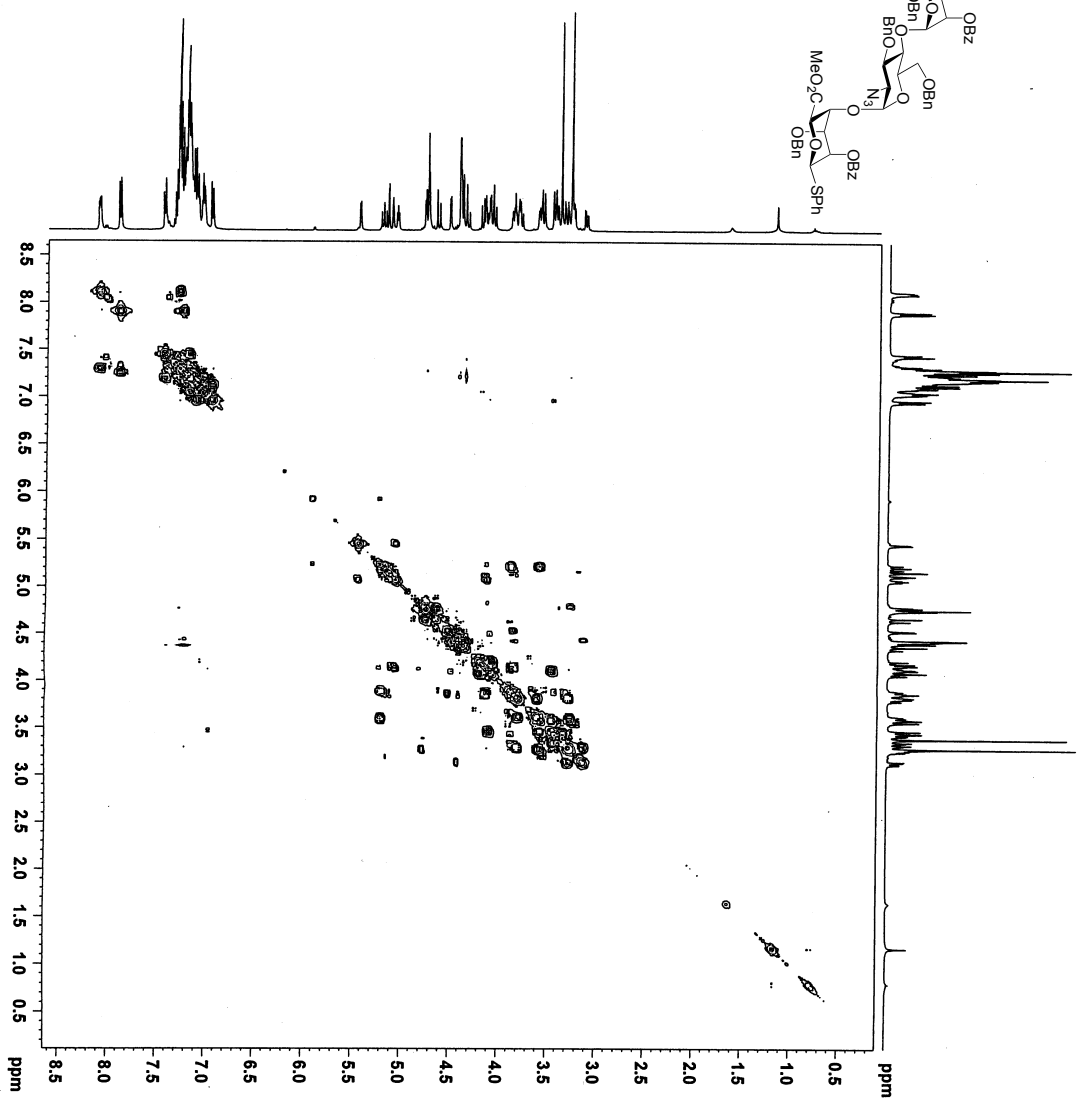
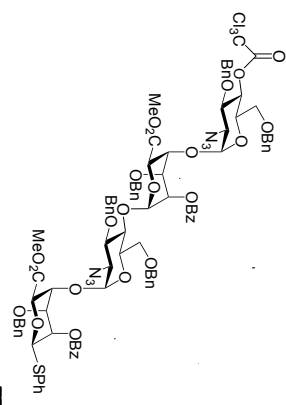
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7.91E3
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C₈₈H₉₂N₇O₂₂S₁
p (gss, s /p:40) Chrg 1
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Supplementary Figure S22. HFT MS spectrum for β -7.



Supplementary Figure S23. ^1H NMR (400 MHz; CDCl_3) spectrum for β -8

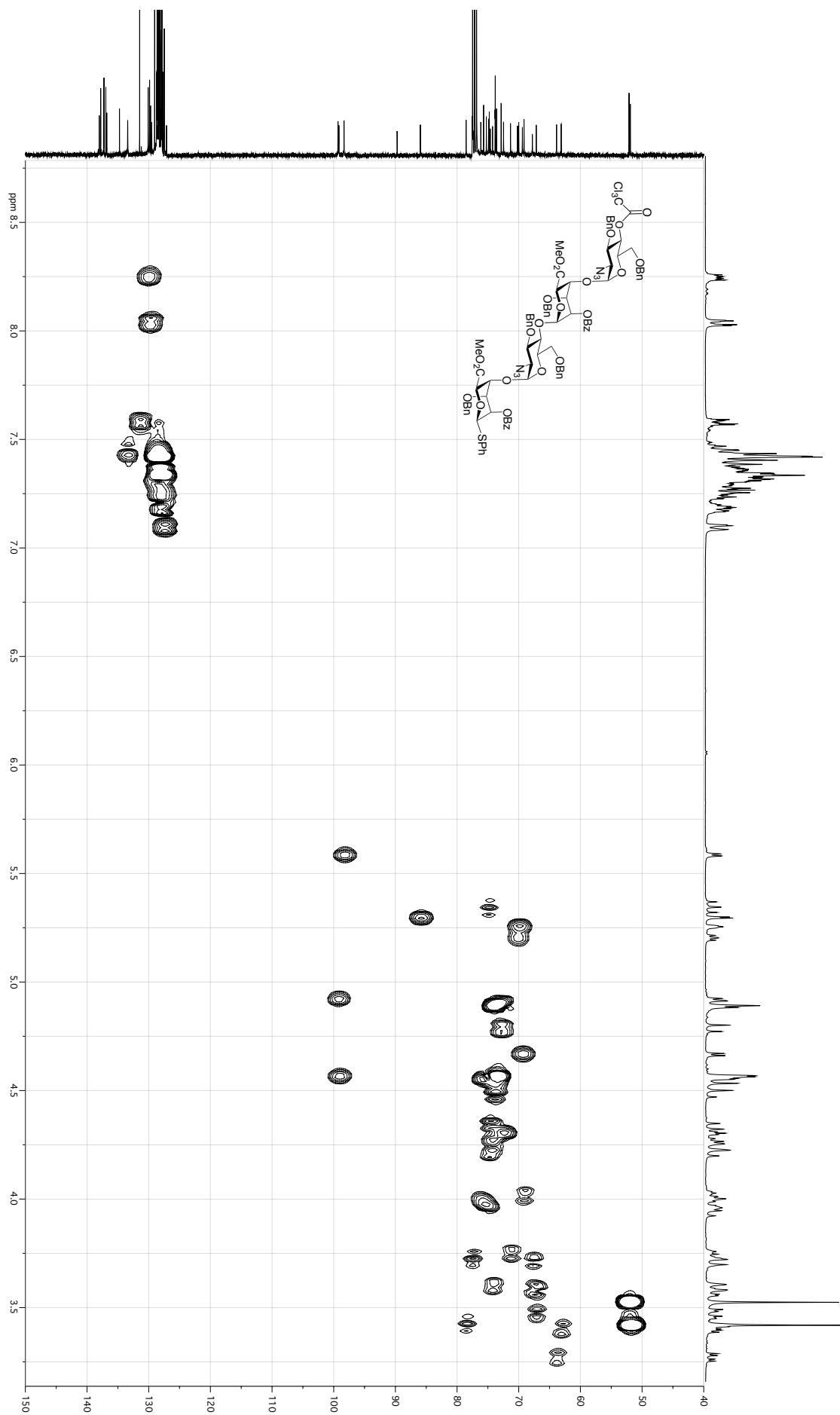
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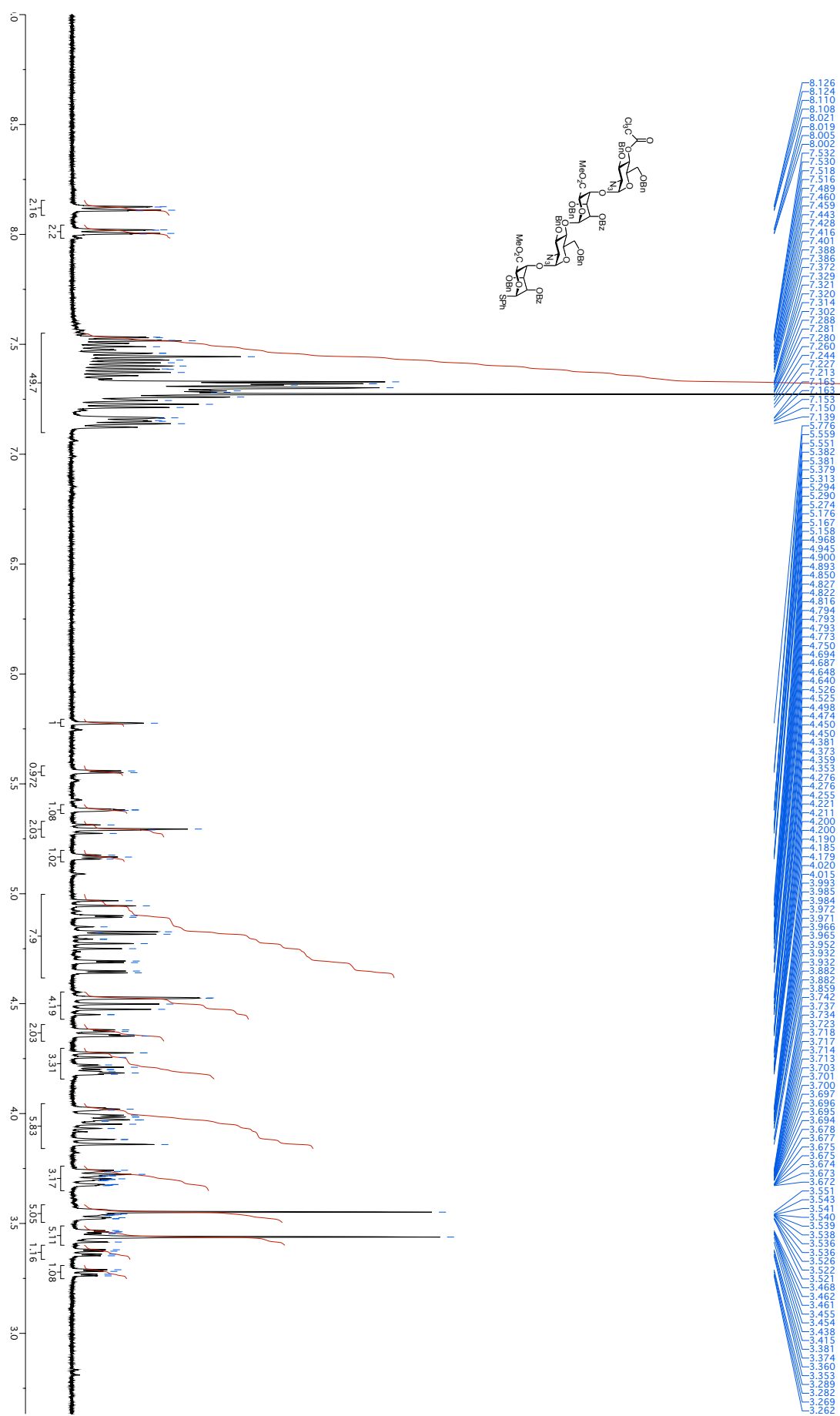
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 DE 6.00 usec
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 GPM1T 10.00 usec
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 GPM1E 10000.00 usec
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 FIDRES 26.682035 Hz
 F2 - Processing parameters
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 WDW EM
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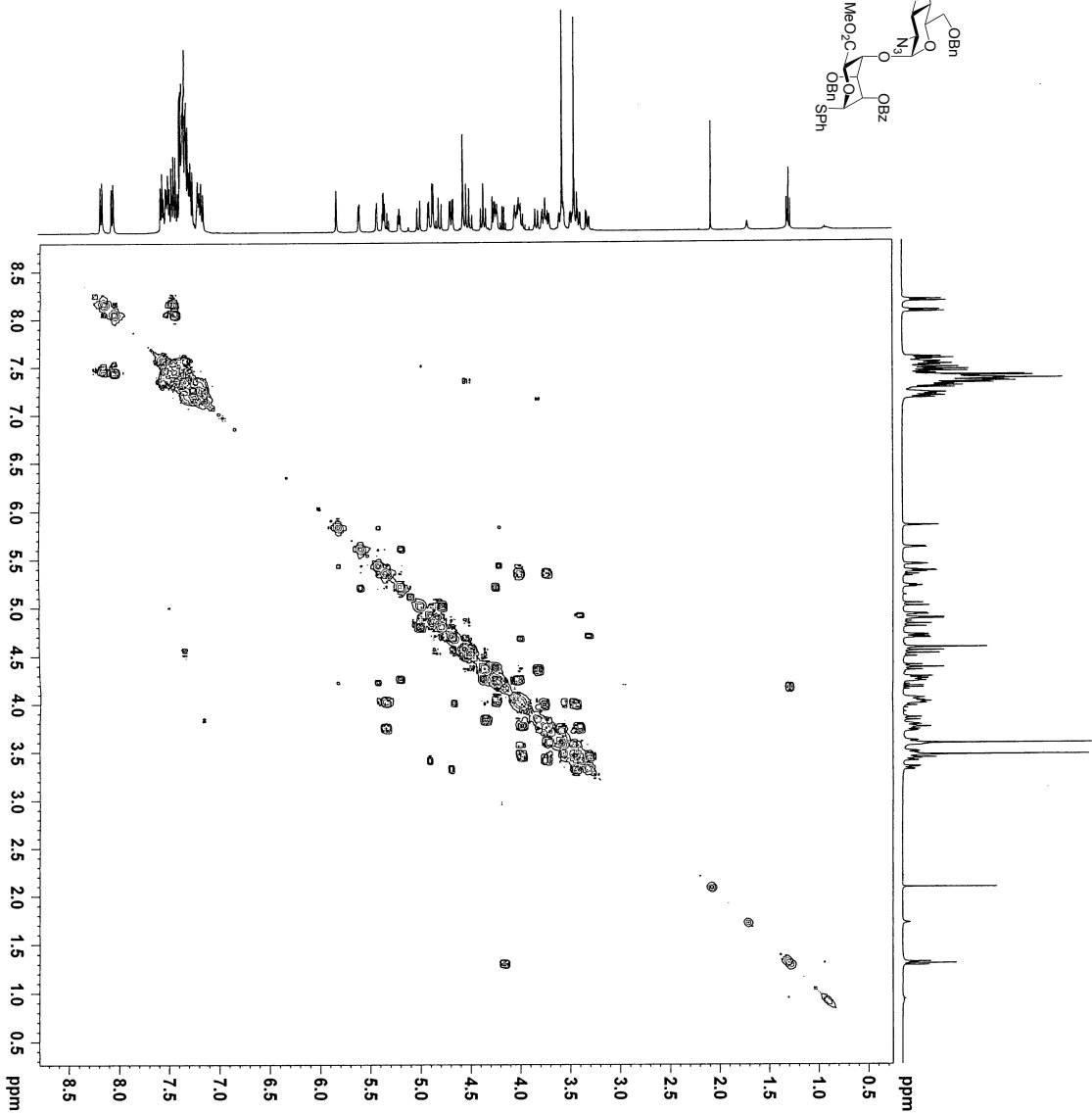
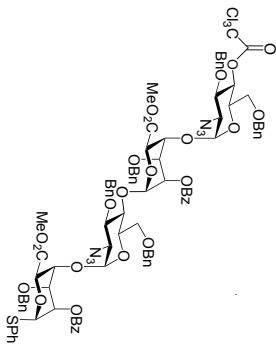
Supplementary Figure S24. COSY NMR spectrum for β -8



Supplementary Figure S25. HMPC NMR spectrum for β -8



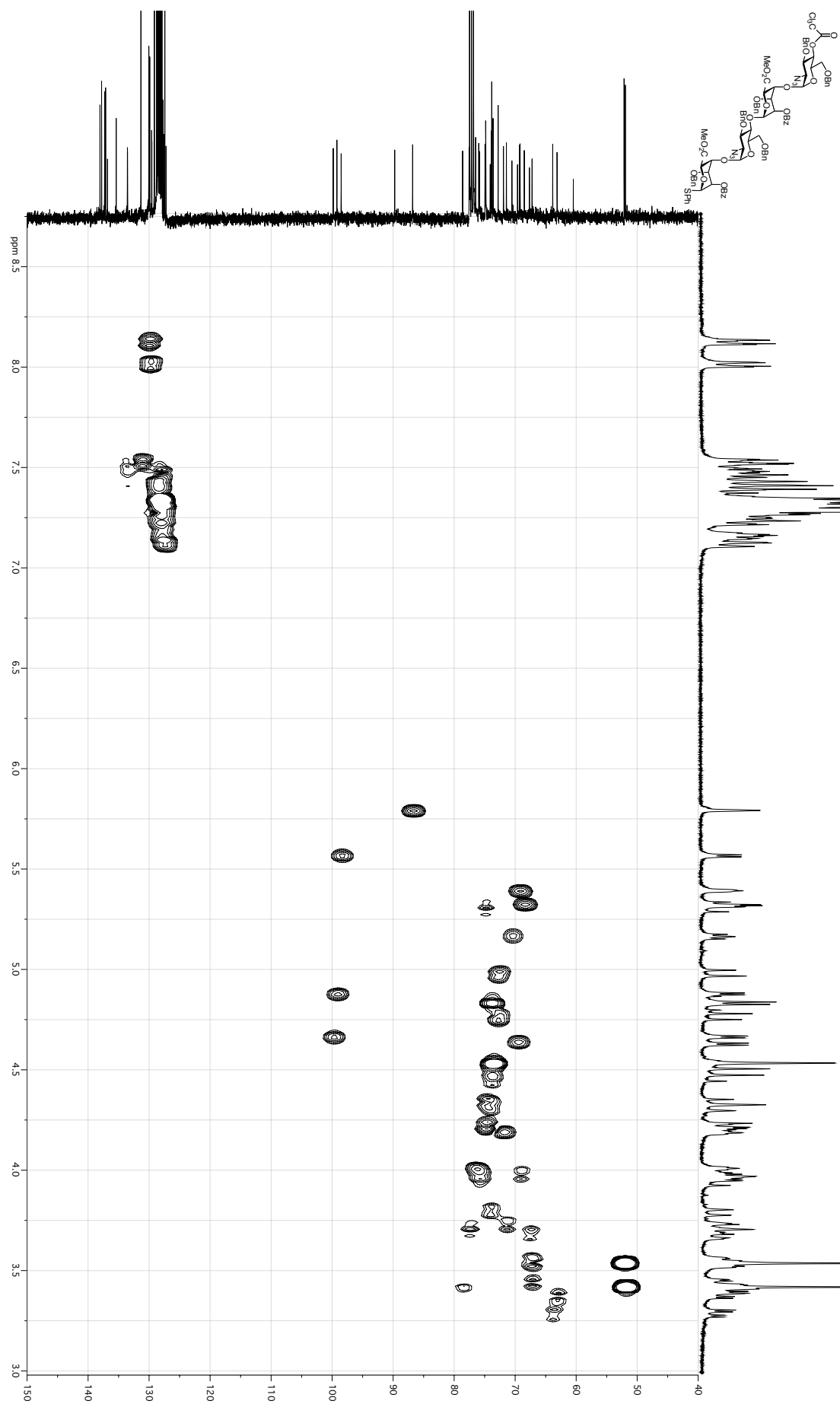
SU1297



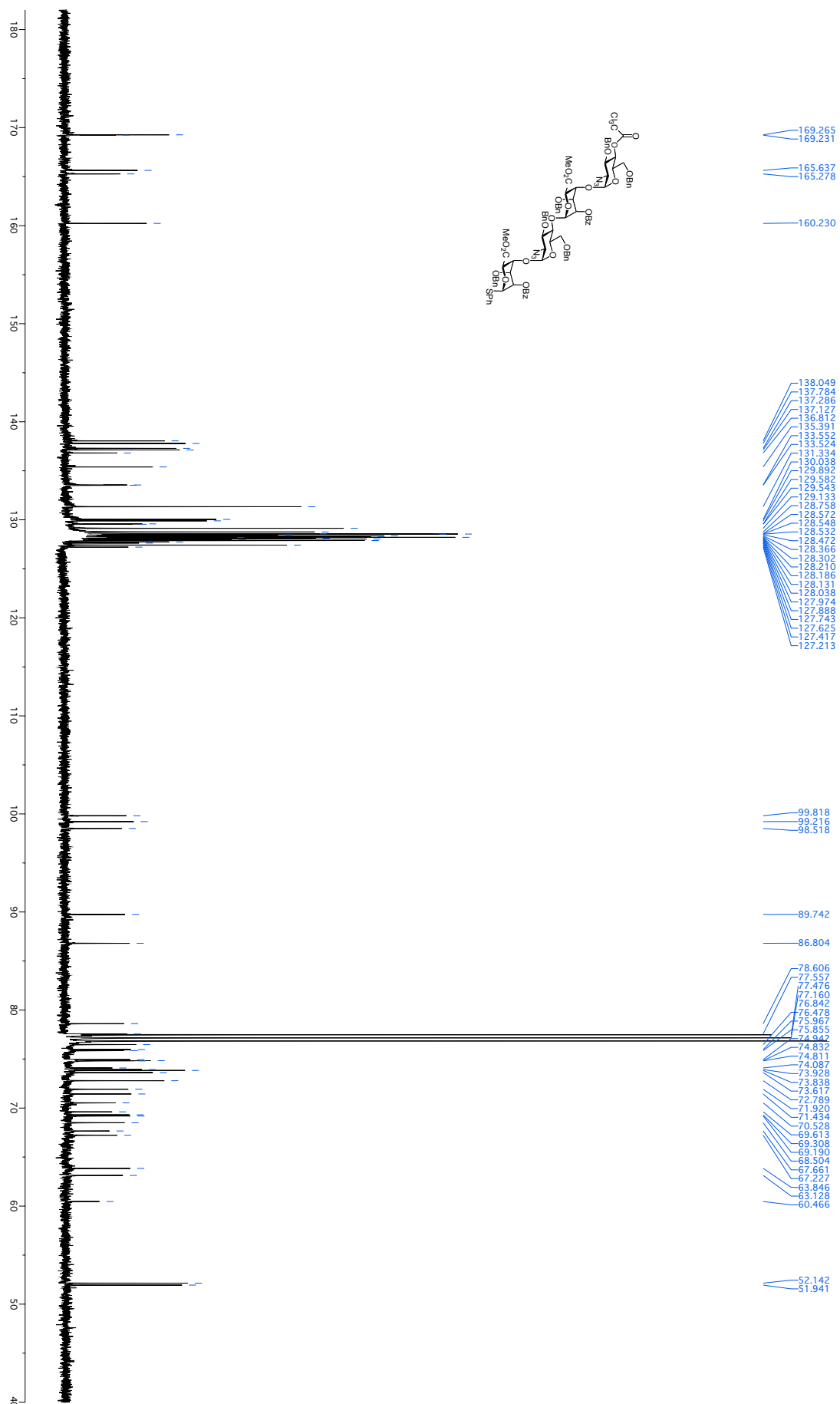
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 F2: 400.1318116 MHz
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 D16: 0.00020000 sec
 INO: 0.00029280 sec
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 GRNAM1: GRANDIRP CHANNEL
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 GRNAM3: SINE:100
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 NRG: 3100
 SSB: 0
 LB: 0.00 Hz
 GB: 0
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Supplementary Figure S28. COSY NMR spectrum for α -8



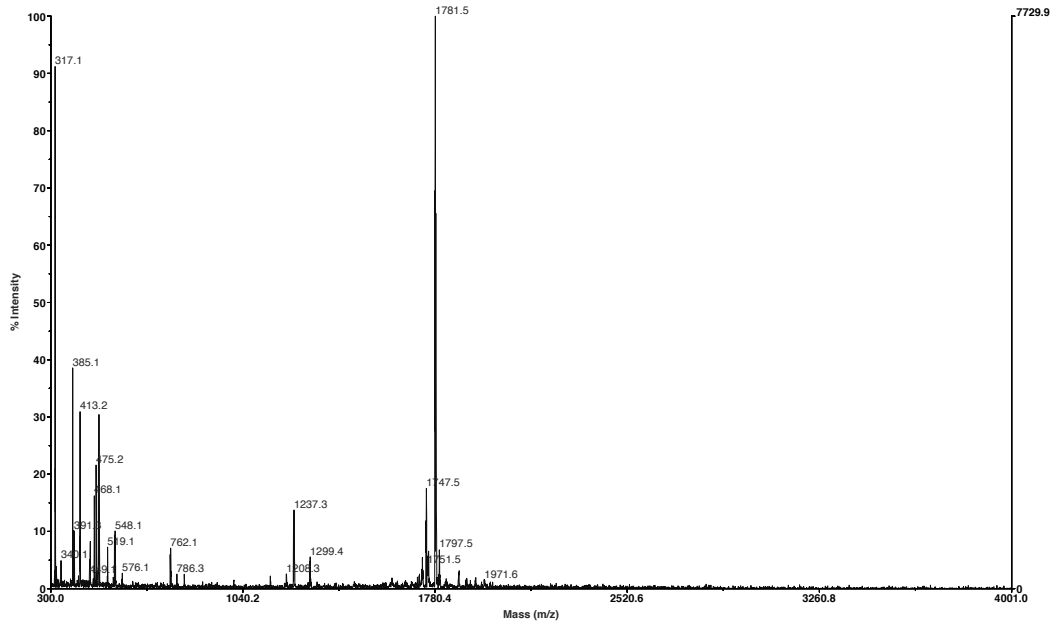
Supplementary Figure S29. HMQC NMR spectrum for α -8



Supplementary Figure S30. ^{13}C NMR (100 MHz; CDCl_3) spectrum for α -8

EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea

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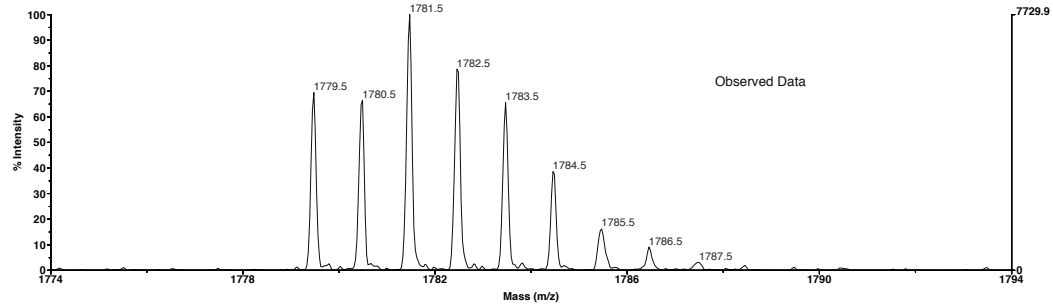


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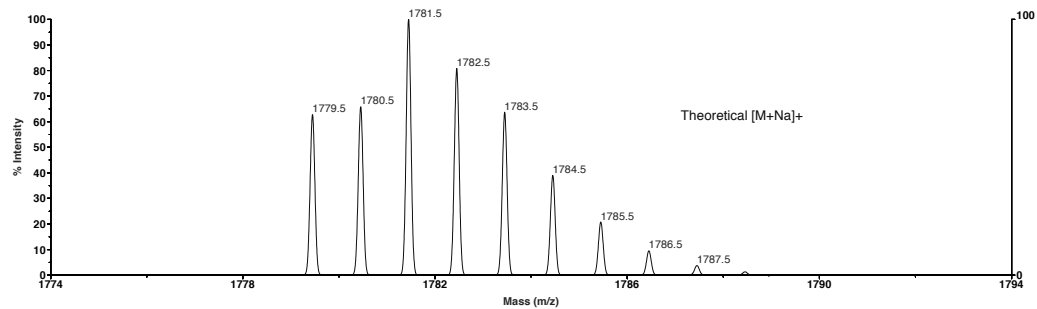
Printed: 09:35, May 10, 2012

EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea

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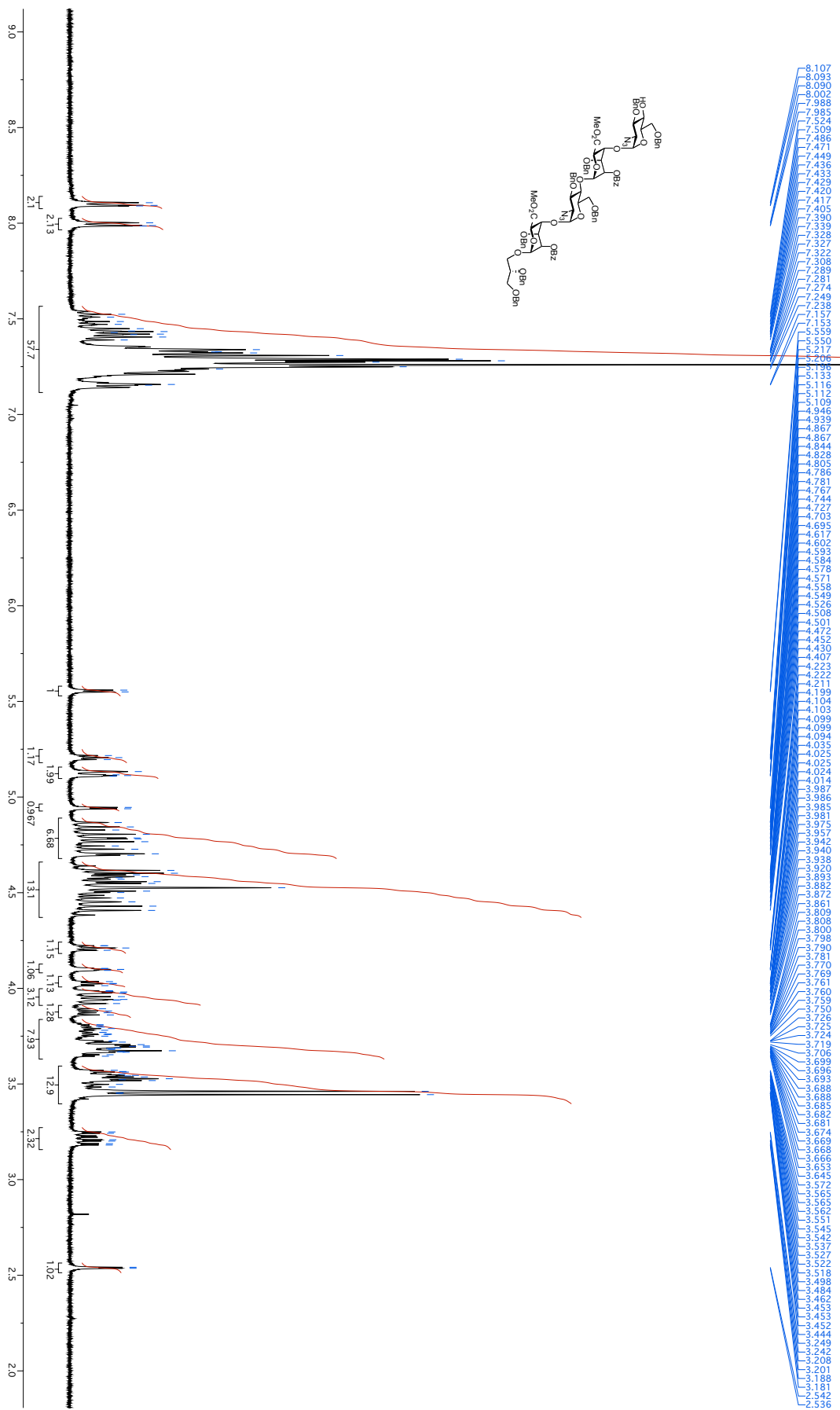
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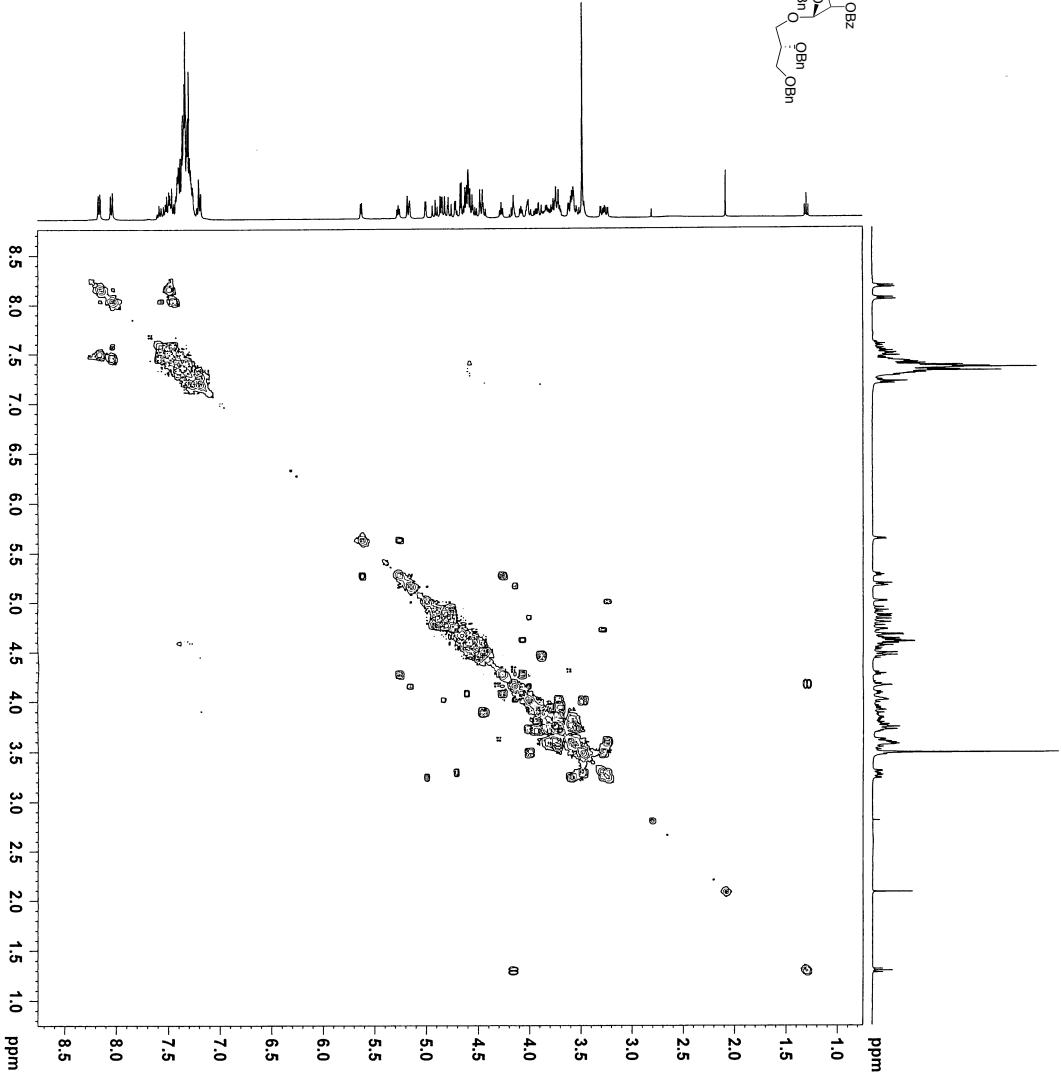
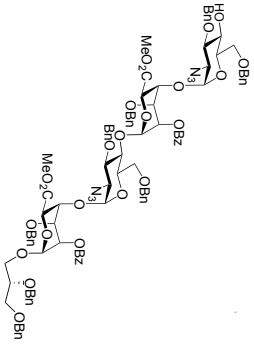
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Supplementary Figure S31. MALDI-TOF spectrum for α -8



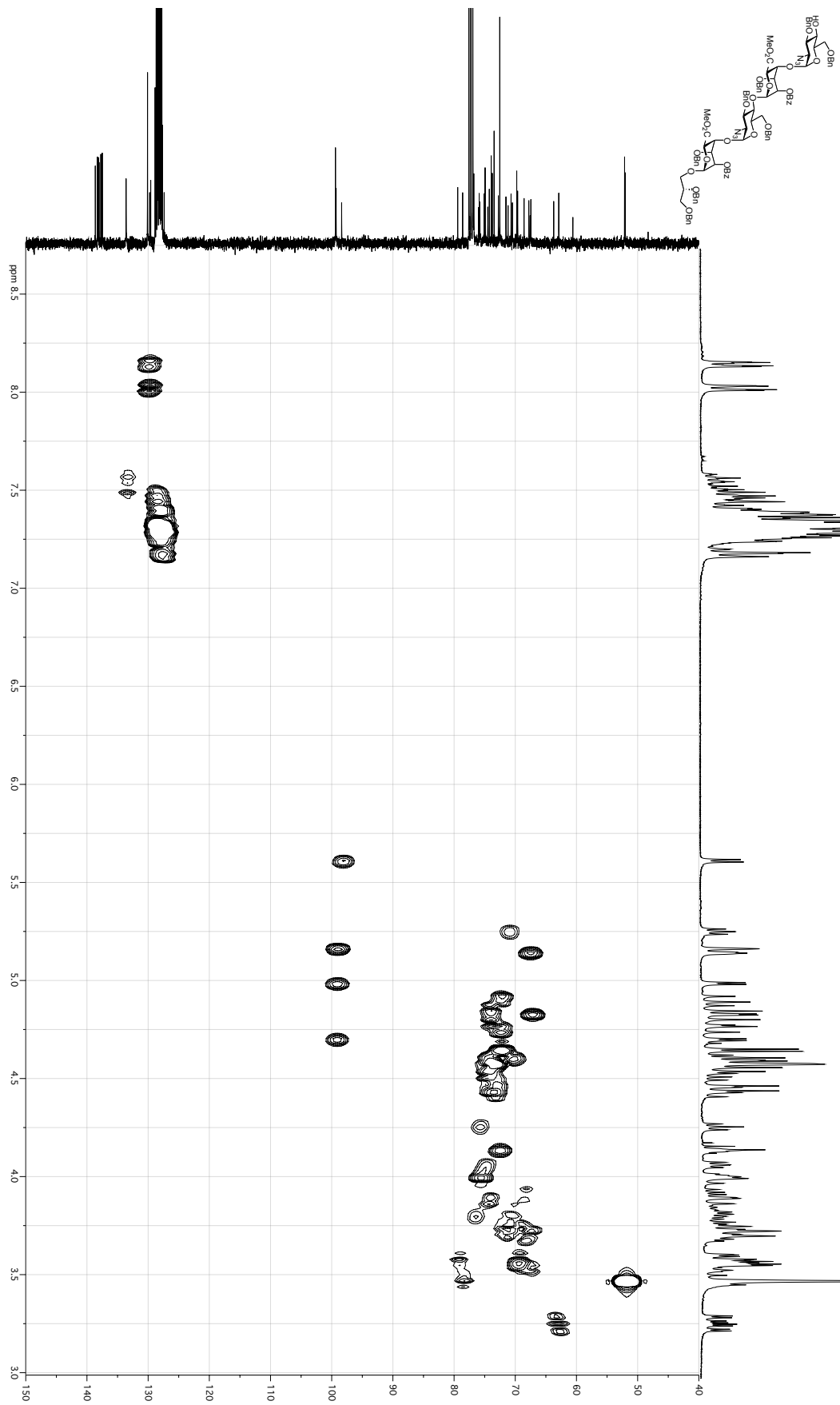
Supplementary Figure S32. ¹H NMR (500 MHz; CDCl₃) spectrum for 9

SU1290B

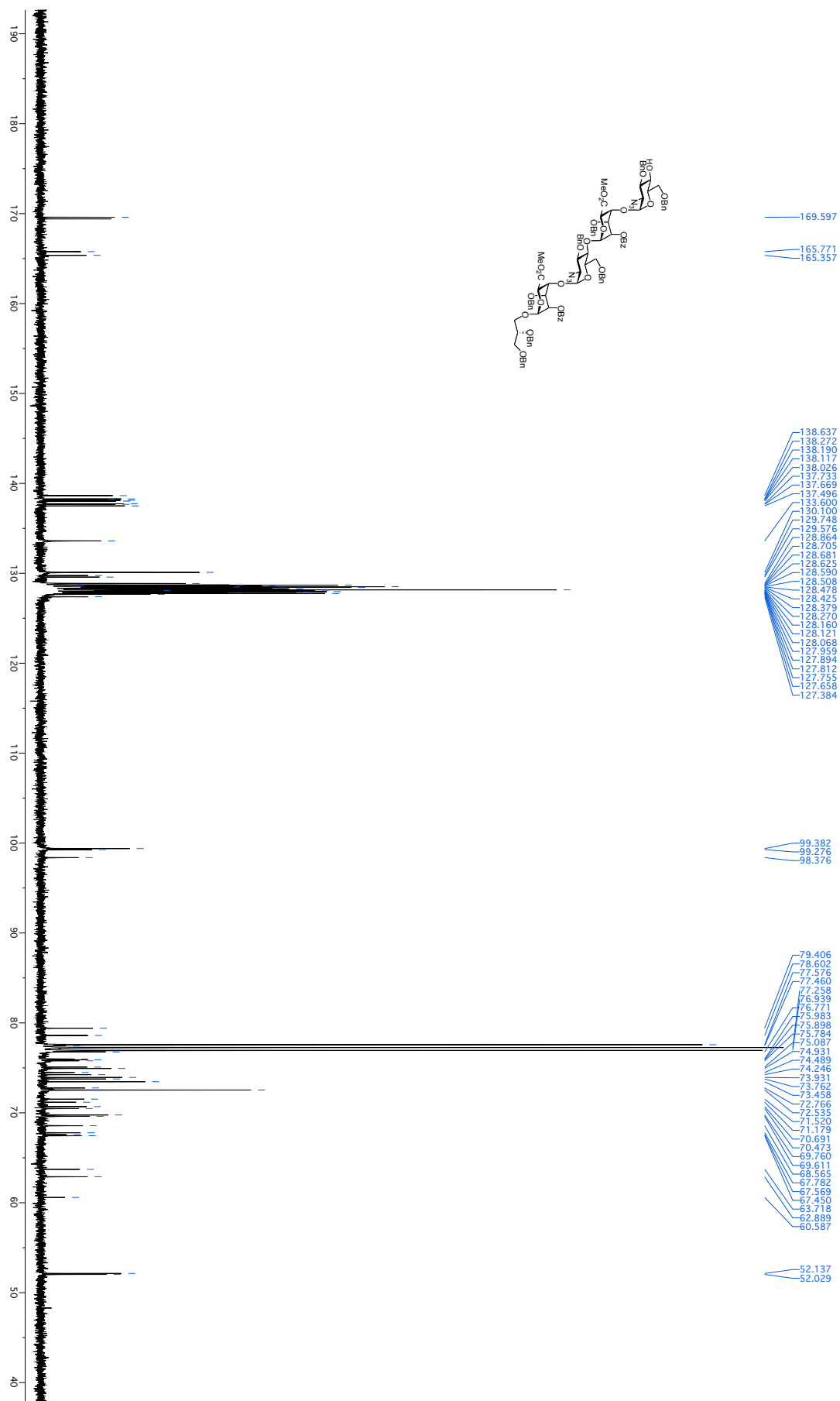


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 NAME May98-2010-5-G
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20100508
 Time 19.41
 INSTRUM spect
 PULPROG zgpg30
 TD 2048
 SOLVENT CDCl₃
 NS 6
 DS 8
 SWH 3205.128 Hz
 FIDRES 0.1355380 Hz
 RG 25.4
 OX 156.000 usec
 PZ 294.2 K usec
 TE 294.2 K usec
 D0 0.00000300 sec
 D1 0.25000000 sec
 D16 0.00000000 sec
 INO 0.00031200 sec
 CHANNEL f1
 NUC1 ¹H
 P0 10.00 usec
 PL1 -0.10 dB
 SFO1 400.1319029 MHz
 ----- GRADIENT CHANNEL -----
 GRNNA1 SINE:100
 GRNZ1 10.00 %
 GRZ2 10.00 %
 P16 1000.00 usec
 F1 - Acquisition parameters
 NQ0
 TD 128
 SFO 400.1319 MHz
 SM FIDRES 23.0010 ppm
 F2 - Processing parameters
 SI 1024
 SF 400.1300000 MHz
 WDW SINC
 LB 0.00 Hz
 GB 0
 PC 1.40
 F1 - Processing parameters
 SI 1024
 SF 400.1300000 MHz
 WDW SINC
 SSB 0
 CB 0

Supplementary Figure S33. COSY NMR spectrum for 9

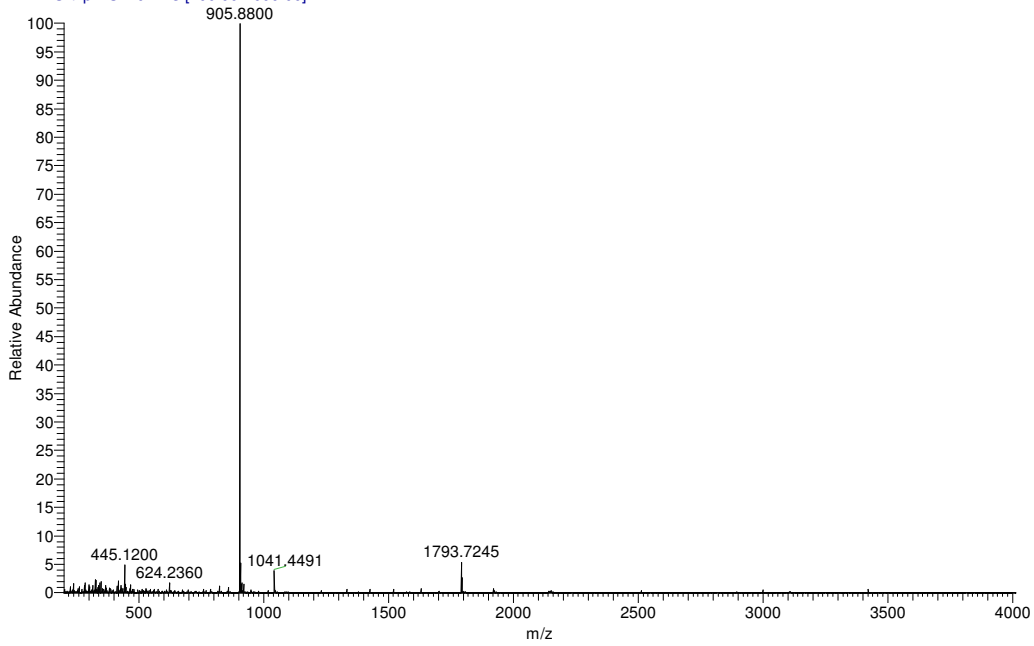


Supplementary Figure S34. HMBC NMR spectrum for 9

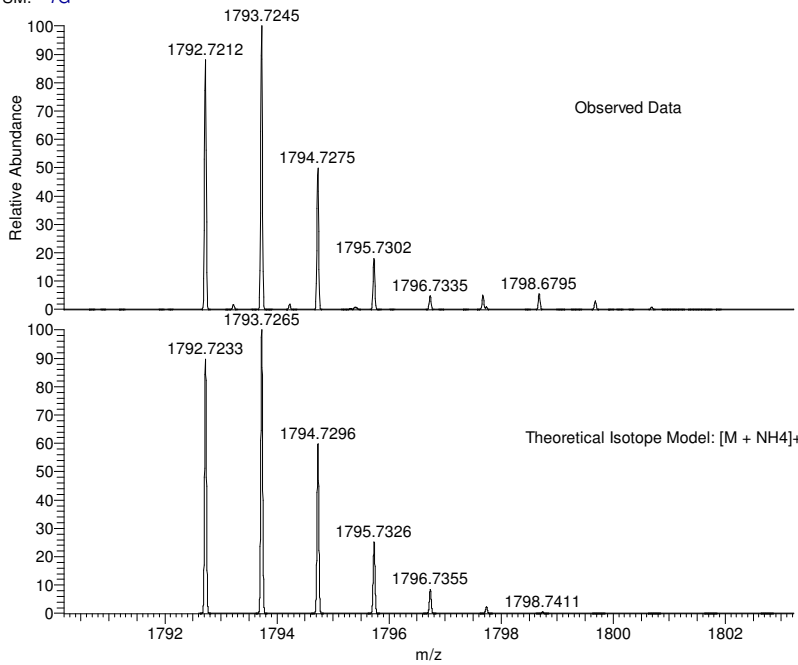


Supplementary Figure S35. ¹³C NMR (100 MHz; CDCl₃) spectrum for 9

MANGAR139-OV-HNESP #92-106 RT: 2.03-2.43 AV: 15 SM: 7G NL: 2.95E6
T: FTMS + p NSI Full ms [200.00-4000.00]



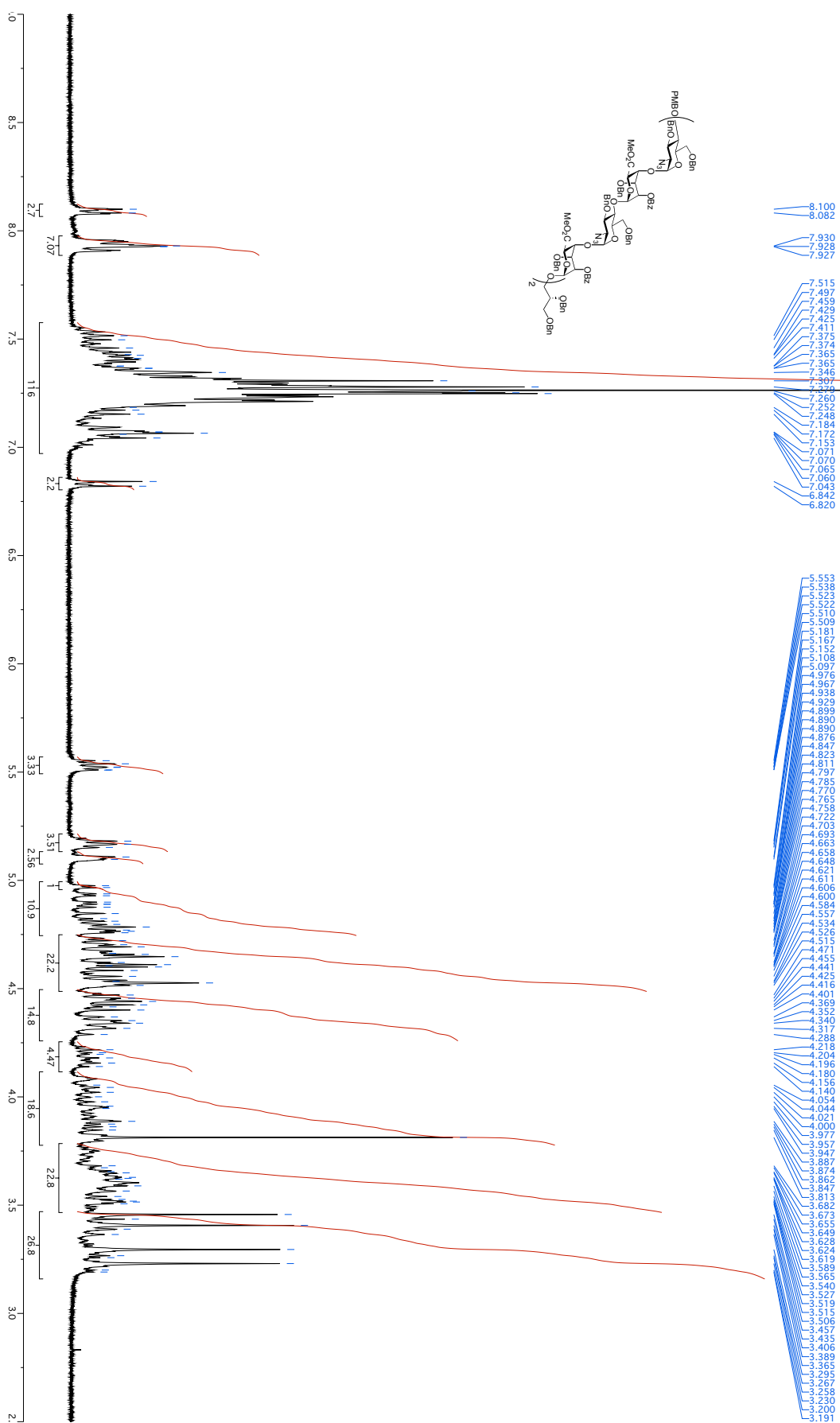
SM: 7G



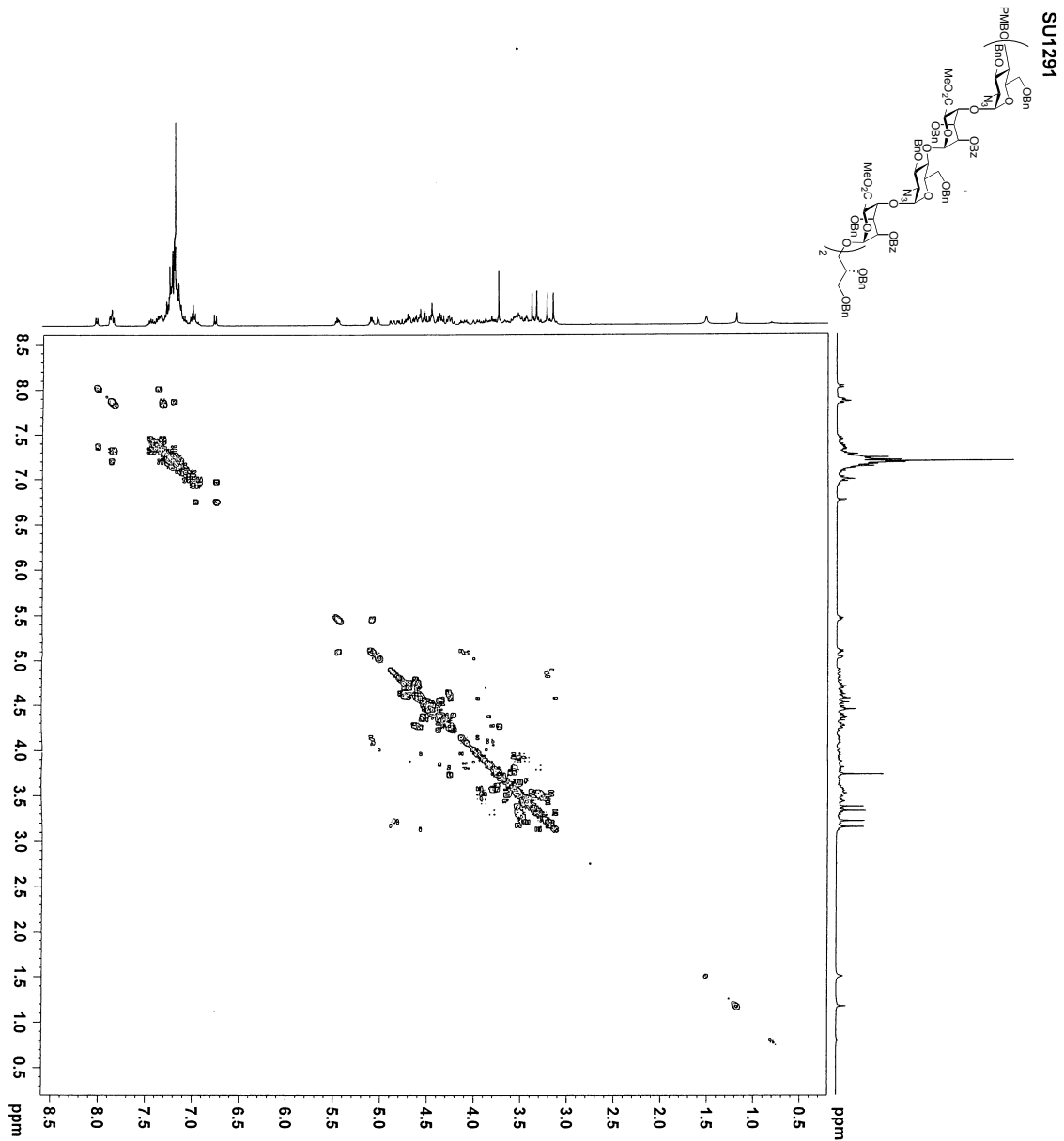
NL:
1.57E5
MANGAR139-OV-HNESP#92-
106 RT: 2.03-2.43 AV: 15 T:
FTMS + p NSI Full ms
[200.00-4000.00]

NL:
8.18E3
C₉₉H₁₀₂N₆O₂₅NH₄:
C₉₉H₁₀₆N₇O₂₅
p (gss, s /p:40) Chrg 1
R: 50000 Res .Pwr .@FWHM

Supplementary Figure S36. FT MS spectrum for 9



Supplementary Figure S37. ¹H NMR (400 MHz; CDCl₃) spectrum for 10



BRUKER

Current Data Parameters
 Name: SU1291
 EXNO: 31
 PROCNO: 1

F2 - Acquisition Parameters
 Date_: 20120228
 Time: 11.55
 INSTRUM: spect
 PULPROG: zgpg30
 PU1PROG: ccsygpqf
 TD: 2048
 NS: 2048
 DS: 8
 SWH: 3460.215 Hz
 FWHM: 12.000 Hz
 AQ: 0.3047924 sec
 RG: 114
 DM: 148.800 usec
 DE: 5.000 usec
 TE: 300.0 K
 D0: 0.00000000 sec
 d1: 1.37000000 sec
 d11: 0.00000000 sec
 D16: 0.00020000 sec
 INO: 0.00029760 sec

===== CHANNEL f1 =====
 NUC1: 1H
 P1: 10.00 usec
 PL1: 0.00 dB
 SFO1: 400.1317998 MHz

===== GRADIENT CHANNEL =====
 GGRM1: SINE:100
 GPR1: 10.00 usec
 GPR2: 10.00 usec
 PL2: 0.00 dB
 PL6: 1000.00 usec

F1 - Acquisition parameters
 SI: 128
 TD: 128
 SFO1: 400.1318 MHz
 FIDRES: 26.251680 Hz
 SF: 400.1318 MHz
 OF: 0.00000000 ppm
 FMODE: OF

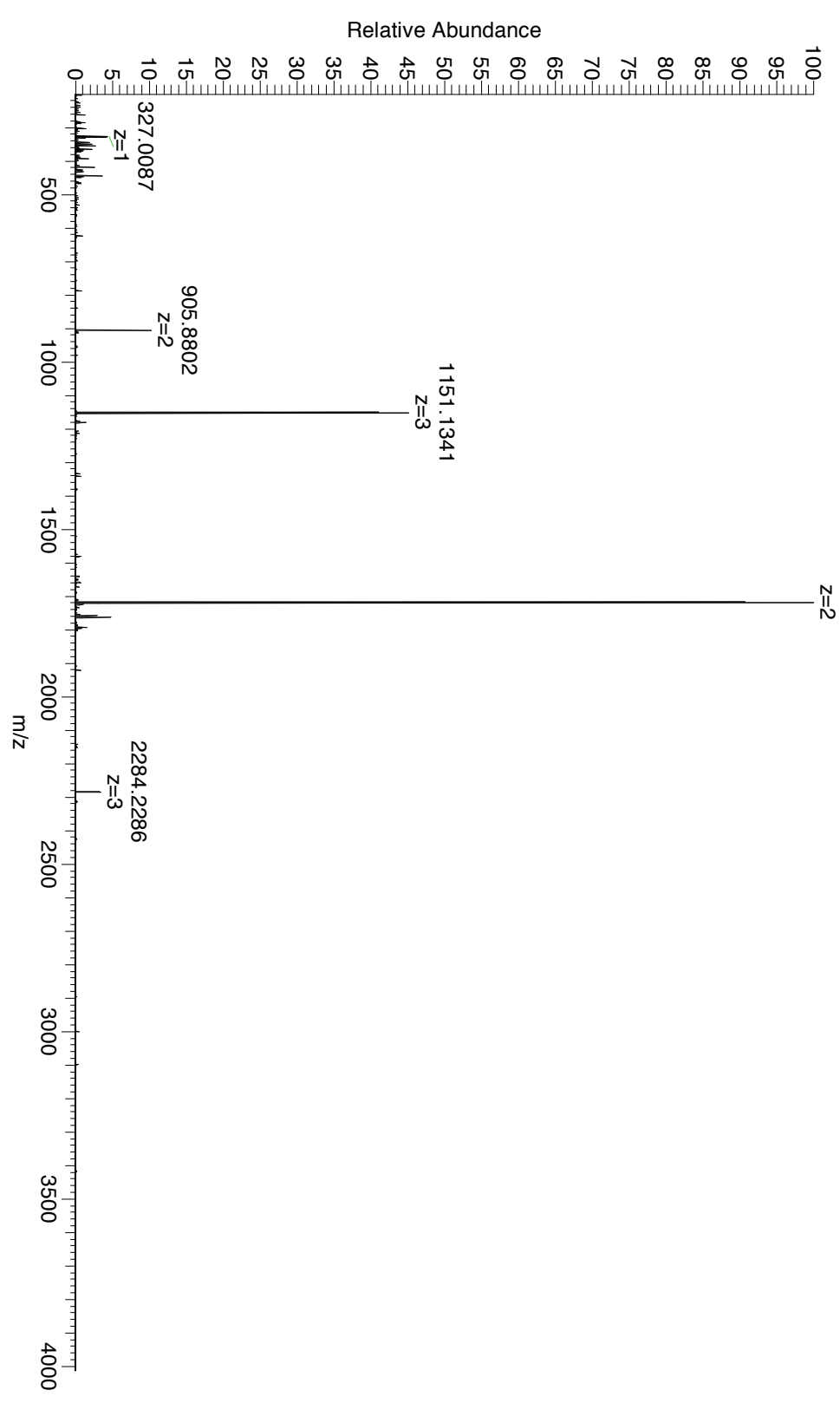
F2 - Processing parameters
 SI: 128
 SF: 400.1300389 MHz
 WTM: SINE
 SSB: 0.00 Hz
 LB: 1.40 Hz
 GB: 0.00 Hz
 PC: 1.40

F1 - Processing parameters
 SI: 1024
 MC2: OF
 SF: 400.1300389 MHz
 WTM: SINE
 SSB: 0.00 Hz
 LB: 0.00 Hz
 GB: 0

Supplementary Figure S38. COSY NMR spectrum for 10

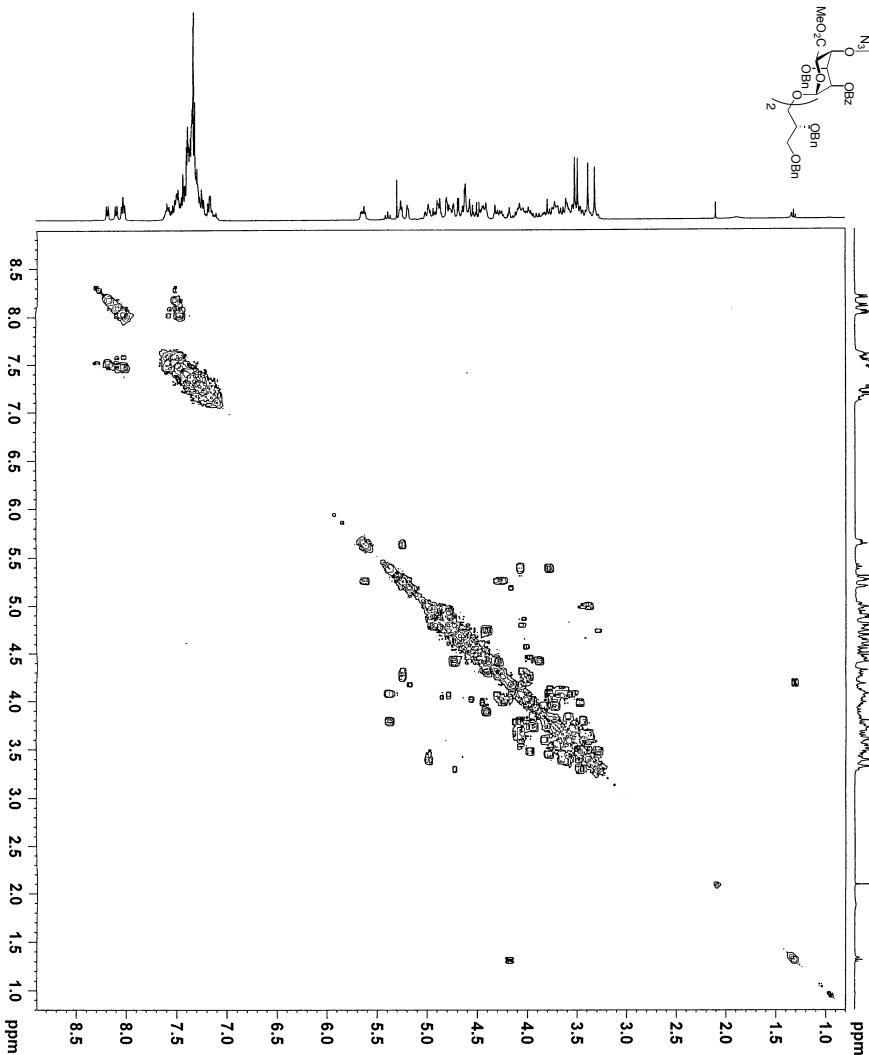
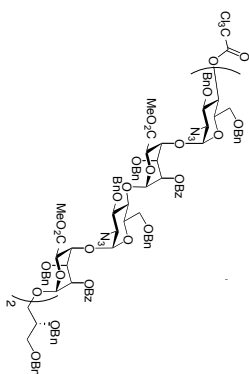
SU1291 MW=3397?
(DCM)/MeOH + NH4OAc
EP SRC National Centre Swansea
LTQ Orbitrap XL
Steen Hansen
01/05/2012 13:24:09

MANGAR156-OJ-HNESP #89-109 RT: 1.92-2.49 AV: 21 SM: 7G NL: 2.26E6
T: FTMS + p NSI Full ms [200.00-4000.00]



Supplementary Figure S39. FT MS spectrum for 10

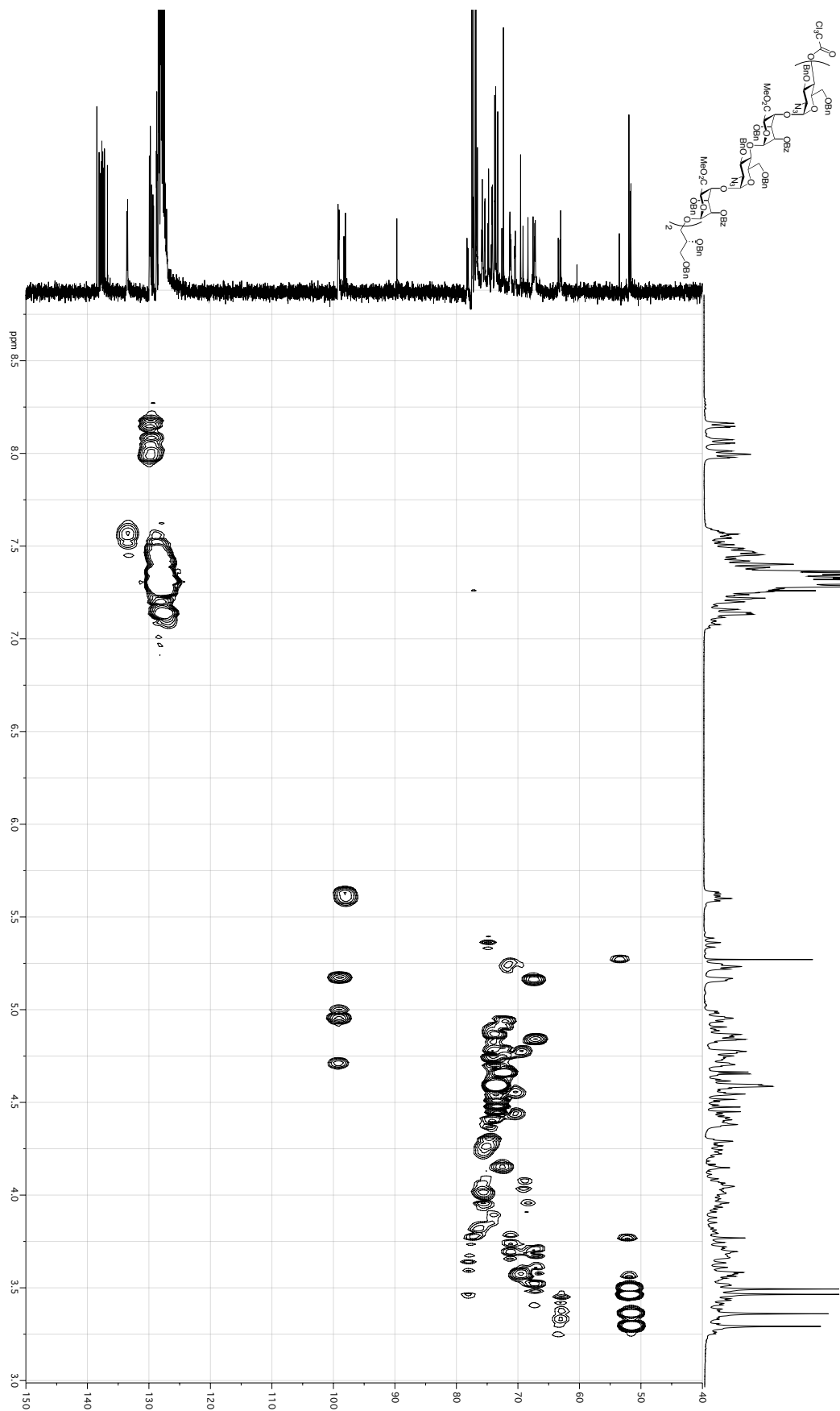
SU1303



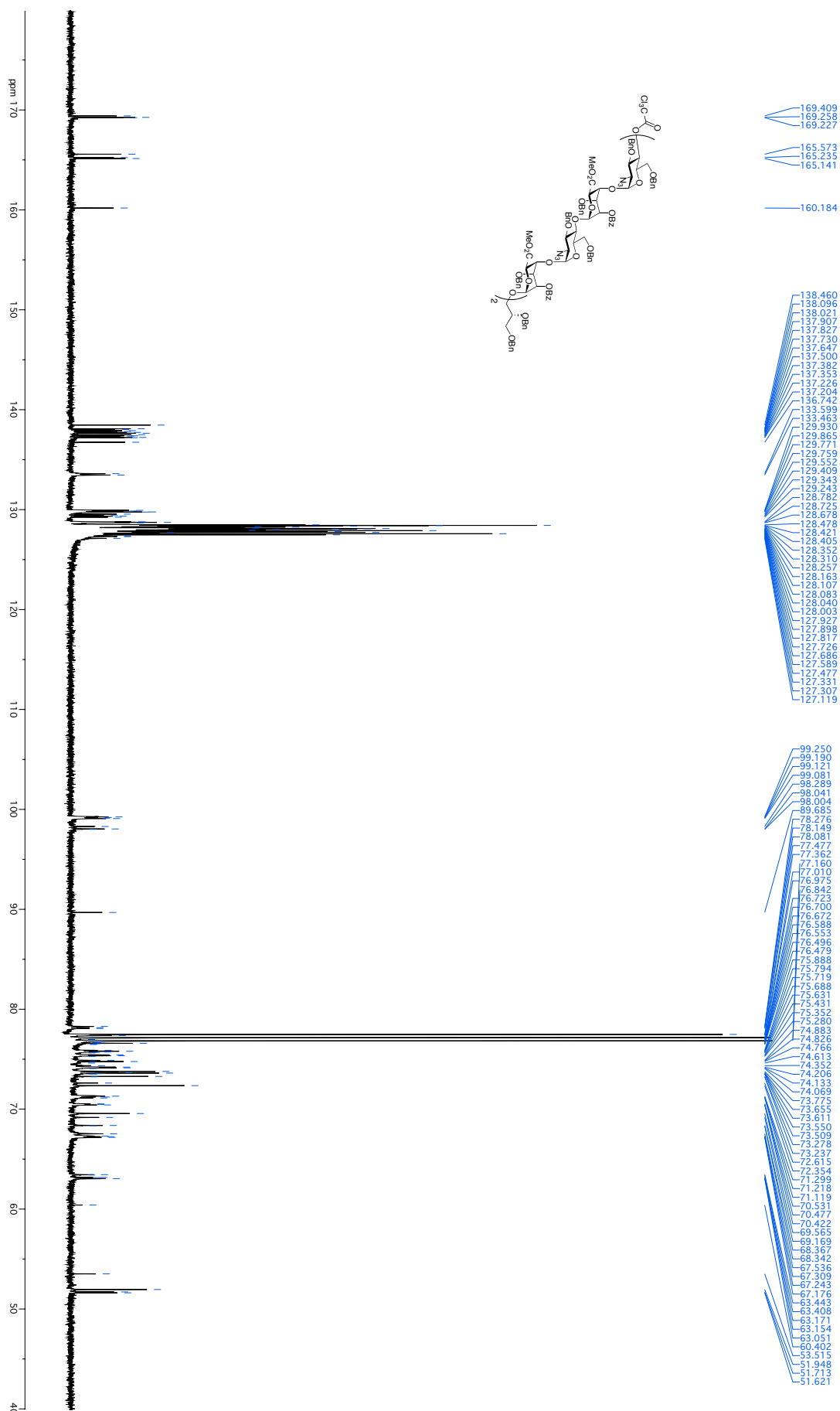
Current Data Parameters
 EXNO 10
 PROCNO 11
 F2 - Acquisition Parameters
 Date_ 20100605
 Time 16.58
 Run 1
 Prgnm 5 mm hmltnc1
 Prbnd 5 mm hmltnc1
 PULPROG carygpgf
 TD 6548
 SFO 400
 SOLVENT cdcl3
 NS 1
 DS 1
 FIDRES 328.348 Hz
 FTRES 1.581222 Hz
 AQ 0.3162612 sec
 AC 14.43
 DC 14.43
 DE 5.00 usec
 TE 294.2 K
 T1 0.60000000 sec
 T1RHO 0.00000000 sec
 d13 1.3619304 sec
 D16 0.00020000 sec
 ANS 0.00000000 sec
 ===== CHANNEL f1 =====
 NU1 1
 P1 10.00 usec
 PL 0.00 dB
 PR1 9.10 dB
 SFO1 400.13992 MHz
 ===== CHANNEL =====
 GRADIENT CHANNEL =====
 GPCP1 10.00 usec
 GPCP2 10.00 usec
 GPCP3 10.00 usec
 GPCP4 10.00 usec
 GPCP5 10.00 usec
 F1 - Acquisition parameters
 TD 128
 SFO1 400.1319 MHz
 SF 25.229547 Hz
 FIDRES 0.147 ppm
 OF 0
 F2 - Processing parameters
 SI 1024
 SF 400.130000 MHz
 WDW SINE
 SSB 0
 GB 0
 PC 1.40
 F1 - Processing parameters
 SI 1024
 SF 400.130000 MHz
 WDW SINE
 SSB 0
 GB 0



Supplementary Figure S41. COSY NMR spectrum for 11

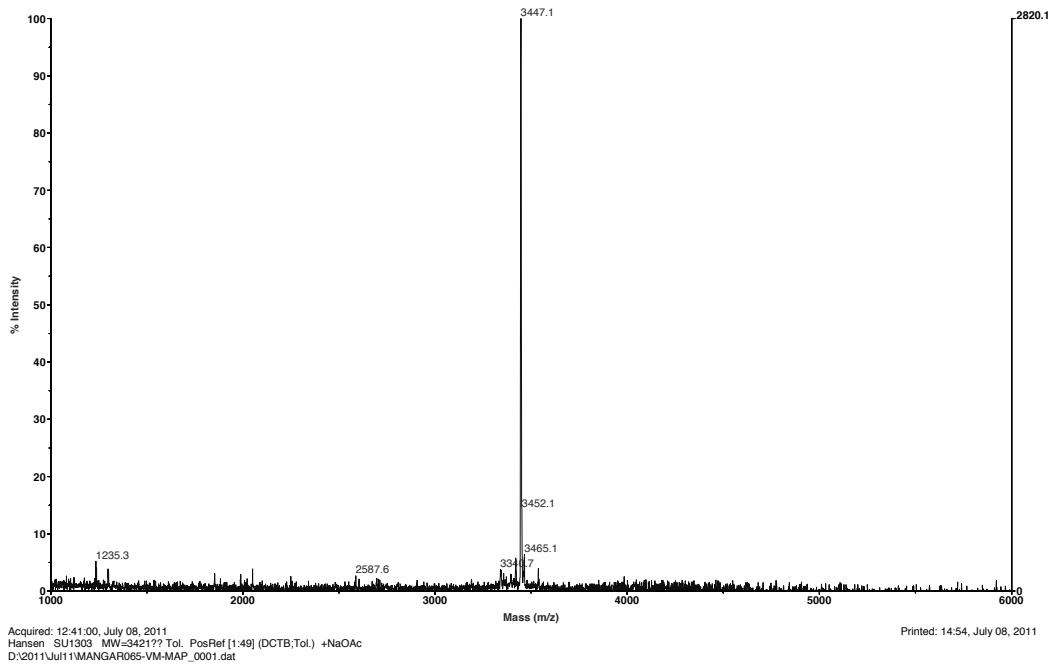


Supplementary Figure S42. HMQC NMR spectrum for 11

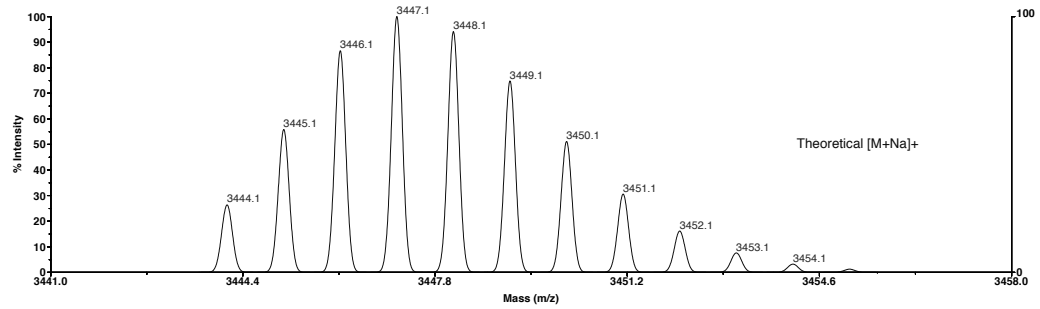


Supplementary Figure S43. ¹³C NMR (100 MHz; CDCl₃) spectrum for **11**

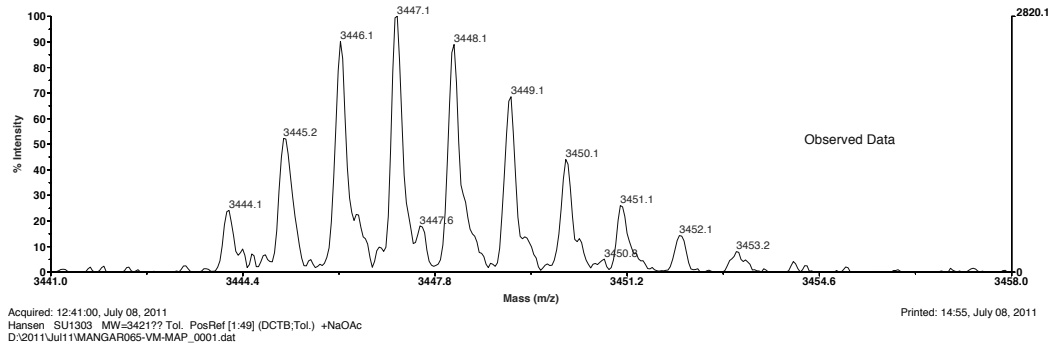
EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea
<<MANGAR065-VM-MAP_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)=>NF0.7[BP = 3447.1, 2820]



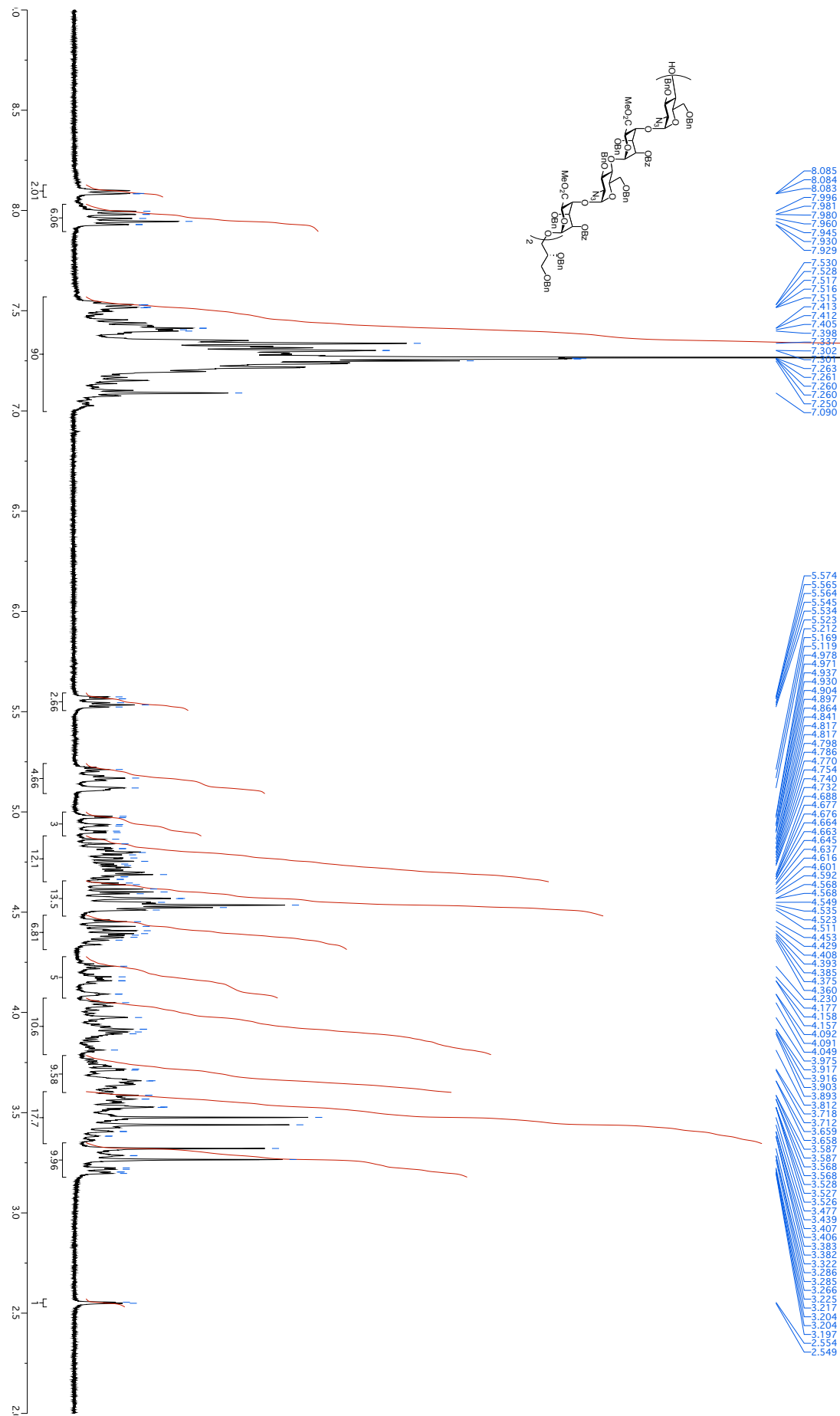
EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea
ISO:C183H183N12O48Cl3 + (Na)



<<MANGAR065-VM-MAP_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)=>NF0.7[BP = 3447.1, 2820]

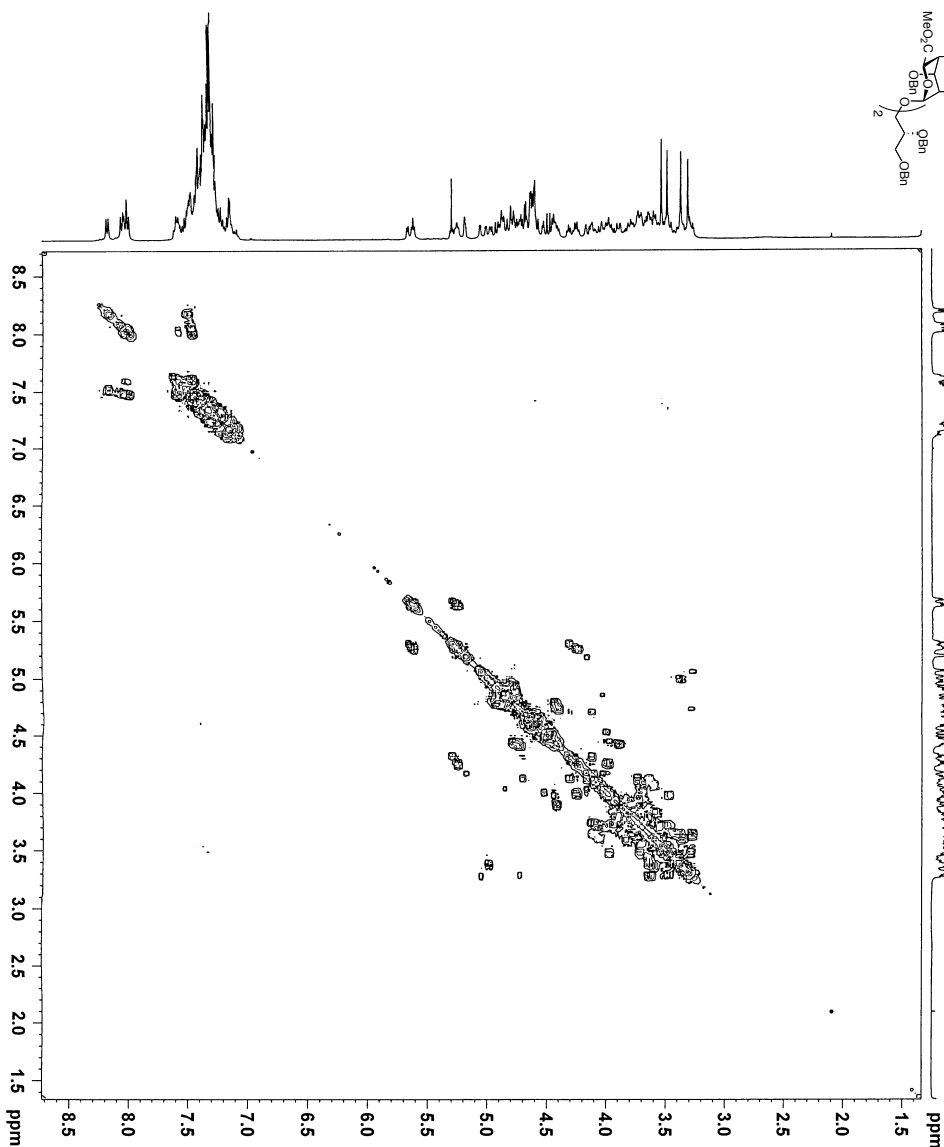
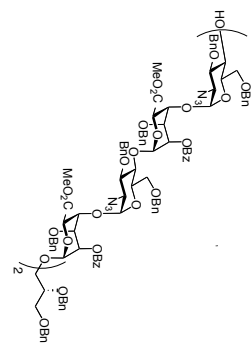


Supplementary Figure S44. MALDI-TOF MS spectrum for 11



Supplementary Figure S45. ¹H NMR (500 MHz; CDCl₃) spectrum for **12**

SU1294D1



```

Current Data Parameters
NAME      SU1294D1
EXPNO     1
PROCNO    1
Date_     20100531
Time      20:27
INSTRUM   spect
PROBHD    5 mm Multinuc1
PULPROG   cosypprf
SOLVENT   CDCl3
NS         1
DS         8
SWH        295.083 Hz
F2         147.542 MHz
AQ         0.3465716 sec
RG         16
DM         169.200 usec
TE         295.2 K
dD         0.00000300 sec
d13        1.31247304 sec
D13        0.00000400 sec
DELTA     0.00000000 sec
IN0        0.00038840 sec

===== CHANNEL f1 =====
NUC1      13C
P0         10.00 usec
P1         10.00 usec
PL1        -0.10 dB
SFO1      400.1320130 MHz

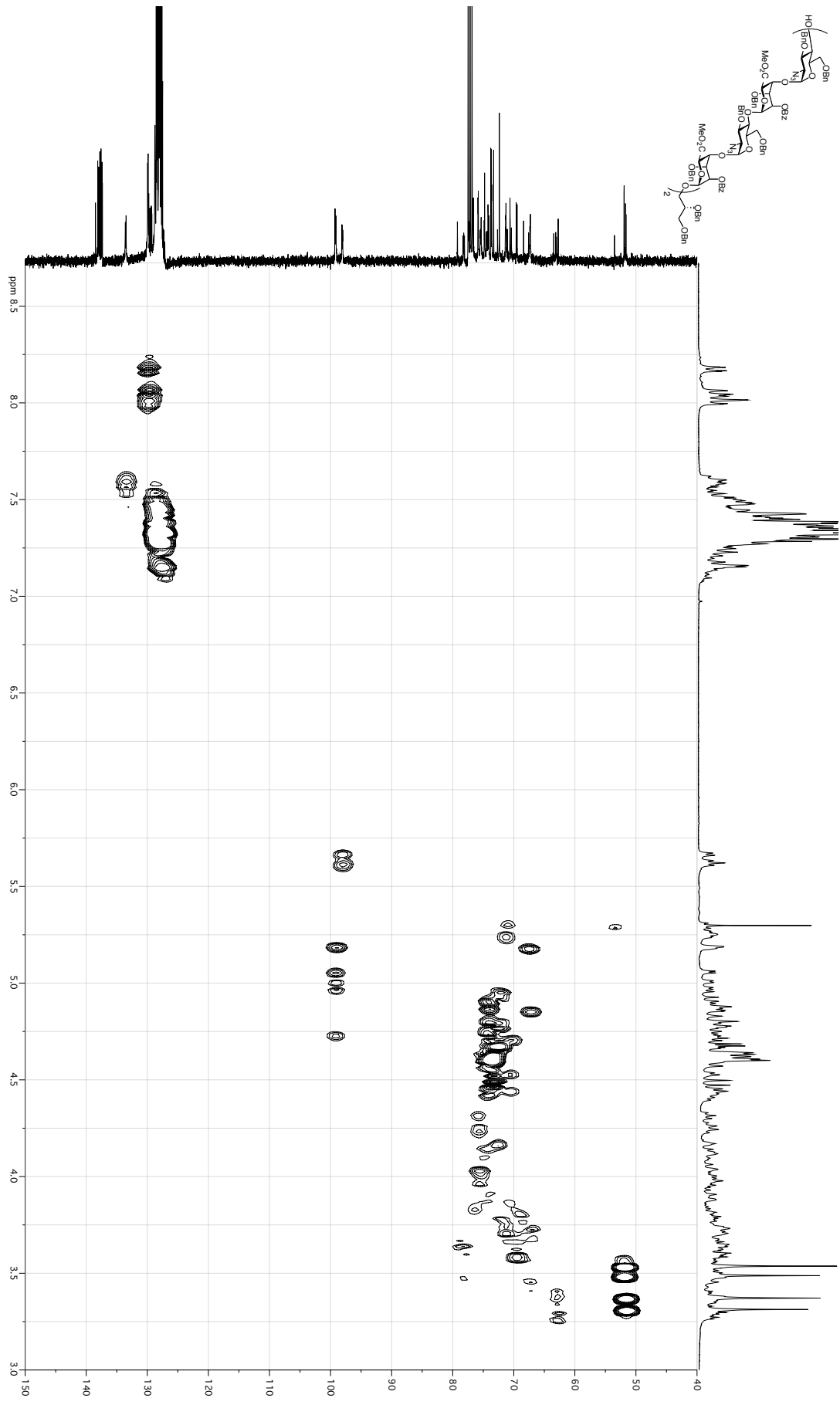
===== GRADIENT CHANNEL =====
GRNAM1    SINE_100
GRNAM2    SINE_100
GPRG1     10.00 %
GPRG2     10.00 %
PI6        1000.00 usec

F1 - Acquisition parameters
NUC1      13C
TD         128
SFO1      400.132 MHz
SF02      23.086584 Hz
FIDRES    7.282 ppm
RMODE     0

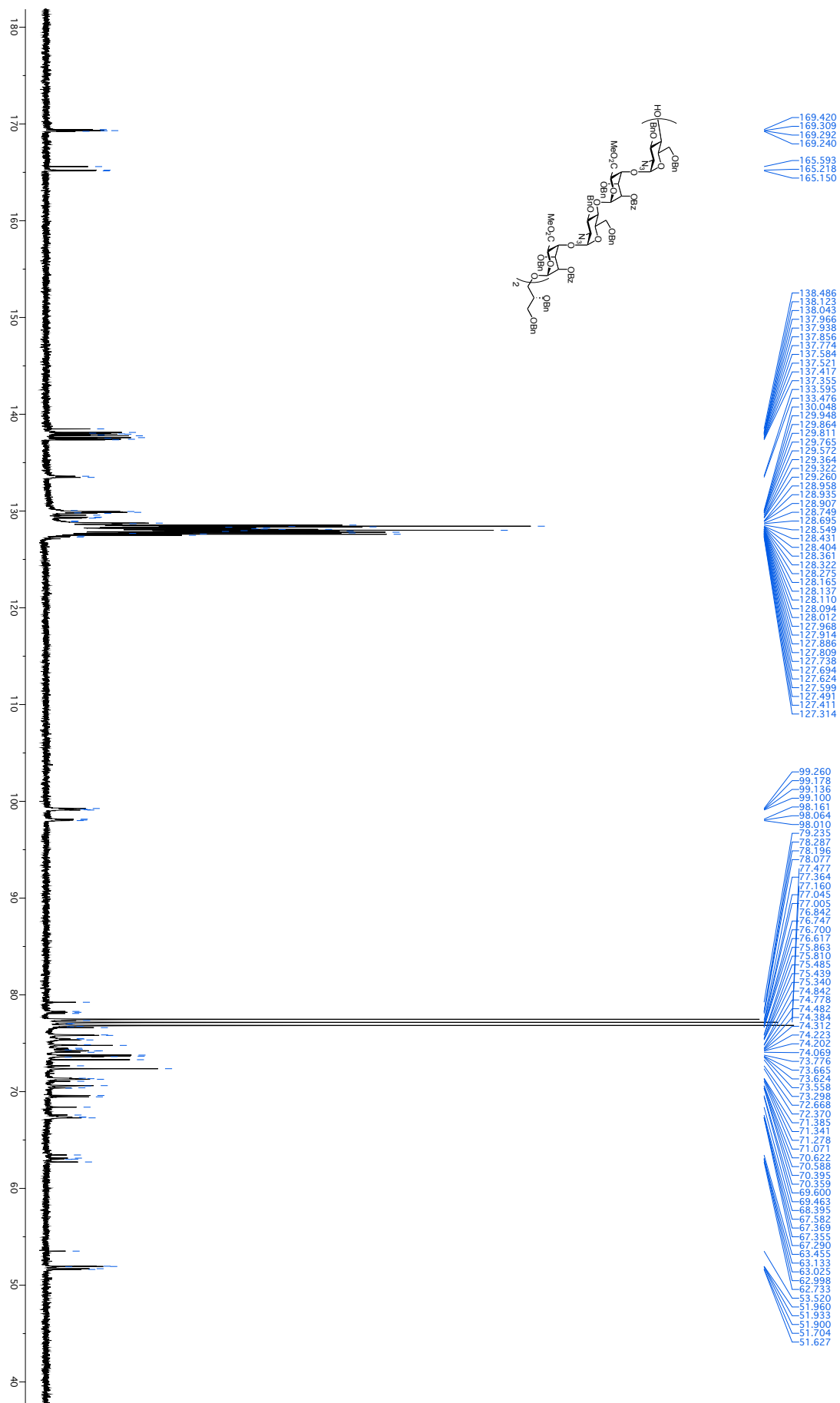
F2 - Processing parameters
SI         32768
SF         400.130000 MHz
WDW        SSB
SSB        0
LB         0.00 Hz
GB         0
PC         1.40

F1 - Processing parameters
SI         1024
SF         400.130000 MHz
WDW        SSB
SSB        0
LB         0.00 Hz
GB         0
  
```

Supplementary Figure S46. COSY NMR spectrum for 12

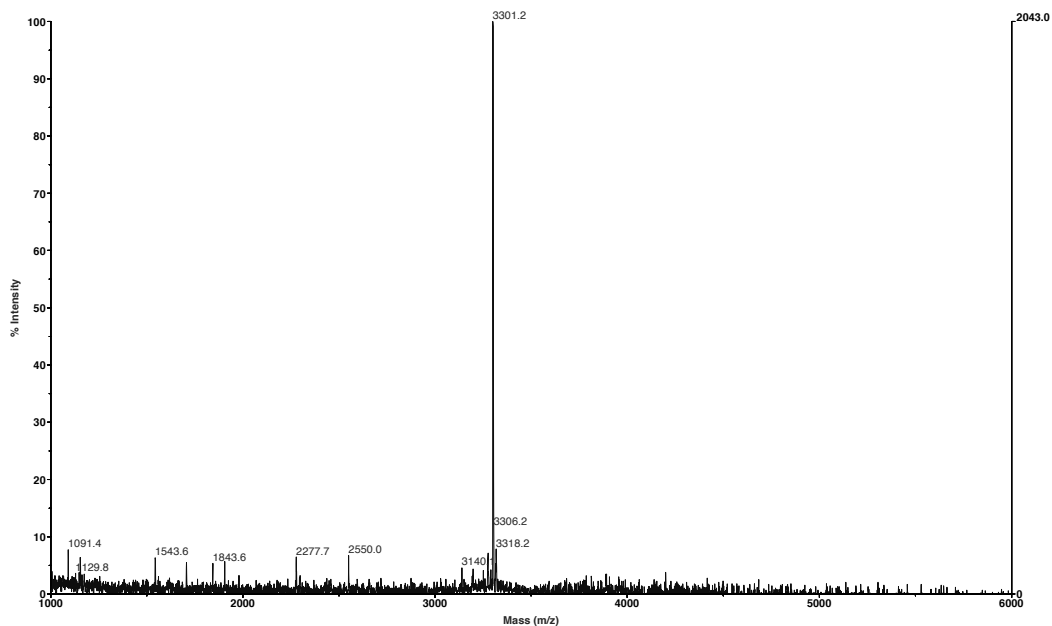


Supplementary Figure S47. HMQC NMR spectrum for 12



Supplementary Figure S48. HMQC NMR spectrum for **12** ¹³C NMR (100 MHz; CDCl₃) spectrum for **12**

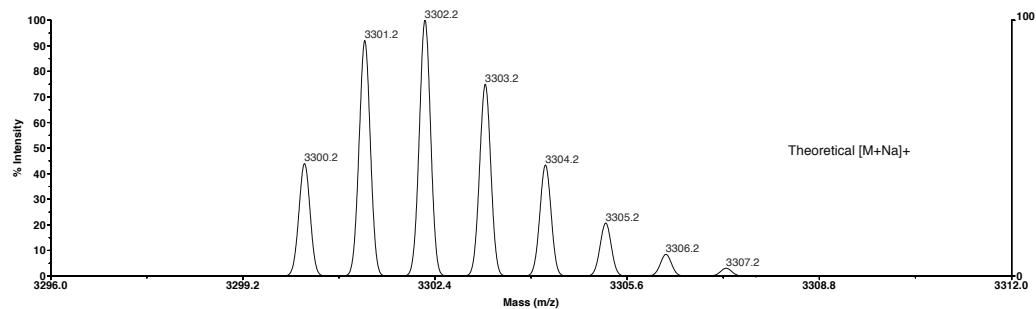
EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea
<<MANGAR066-VM-MAP_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)=>NF0.7[BP = 3301.2, 2043]



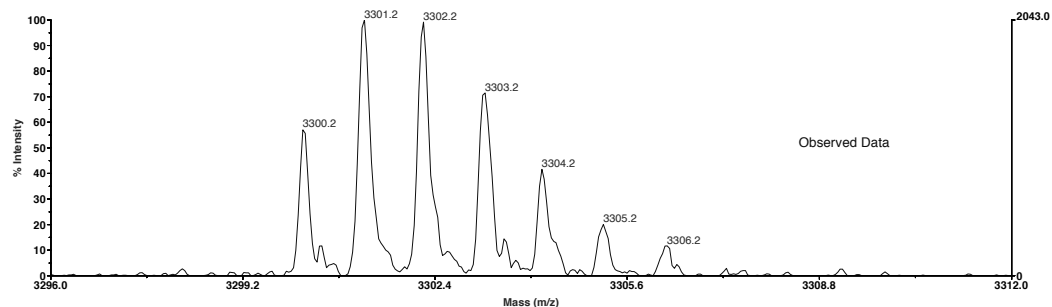
Acquired: 12:49:00, July 08, 2011
Hansen SU1304 MW=3277?? Tol. PosRef [1:49] (DCTB;Tol.) +NaOAc
D:\2011\Jul11\MANGAR066-VM-MAP_0001.dat

Printed: 15:03, July 08, 2011

EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea
ISO:C181H184N12O47 + (Na)¹



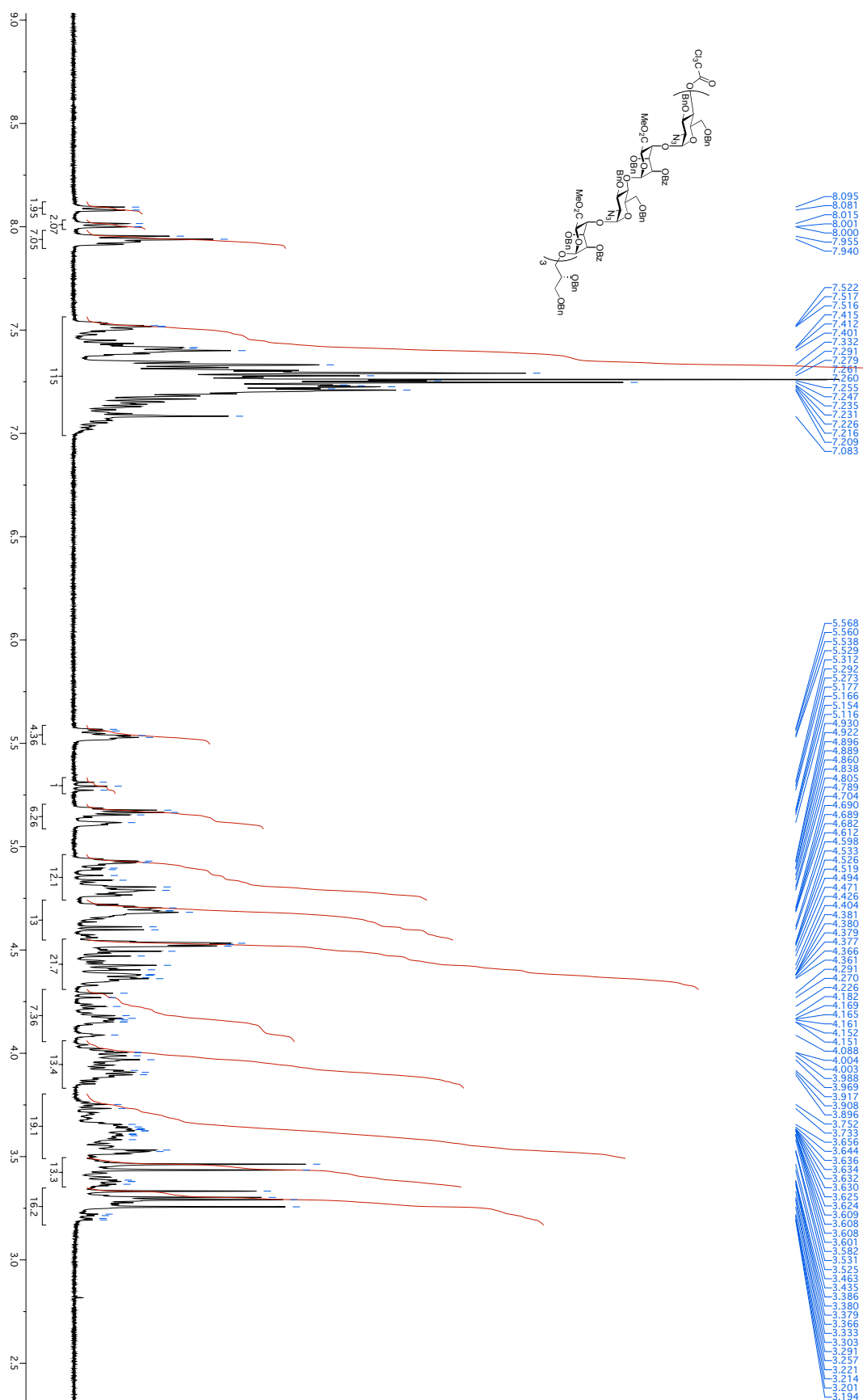
<<MANGAR066-VM-MAP_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)=>NF0.7[BP = 3301.2, 2043]



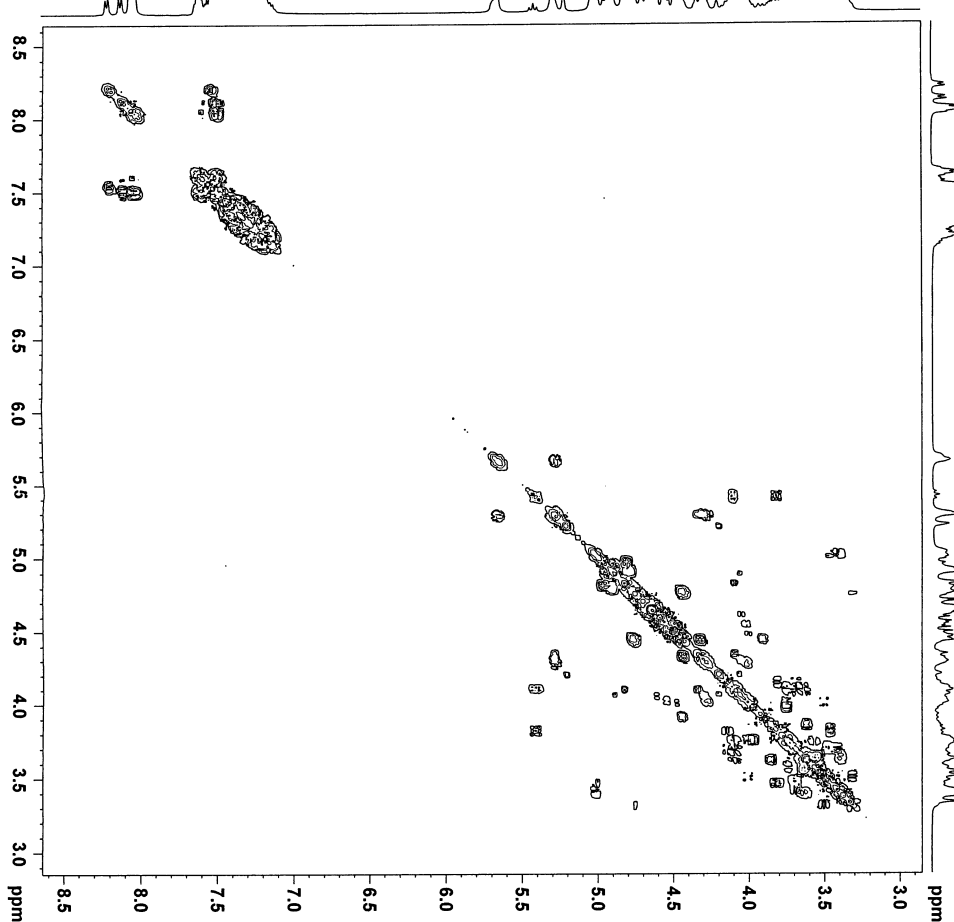
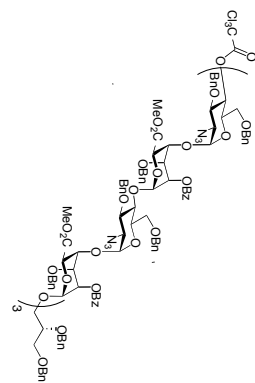
Acquired: 12:49:00, July 08, 2011
Hansen SU1304 MW=3277?? Tol. PosRef [1:49] (DCTB;Tol.) +NaOAc
D:\2011\Jul11\MANGAR066-VM-MAP_0001.dat

Printed: 15:04, July 08, 2011

Supplementary Figure S49. MALDI-TOF MS spectrum for 12



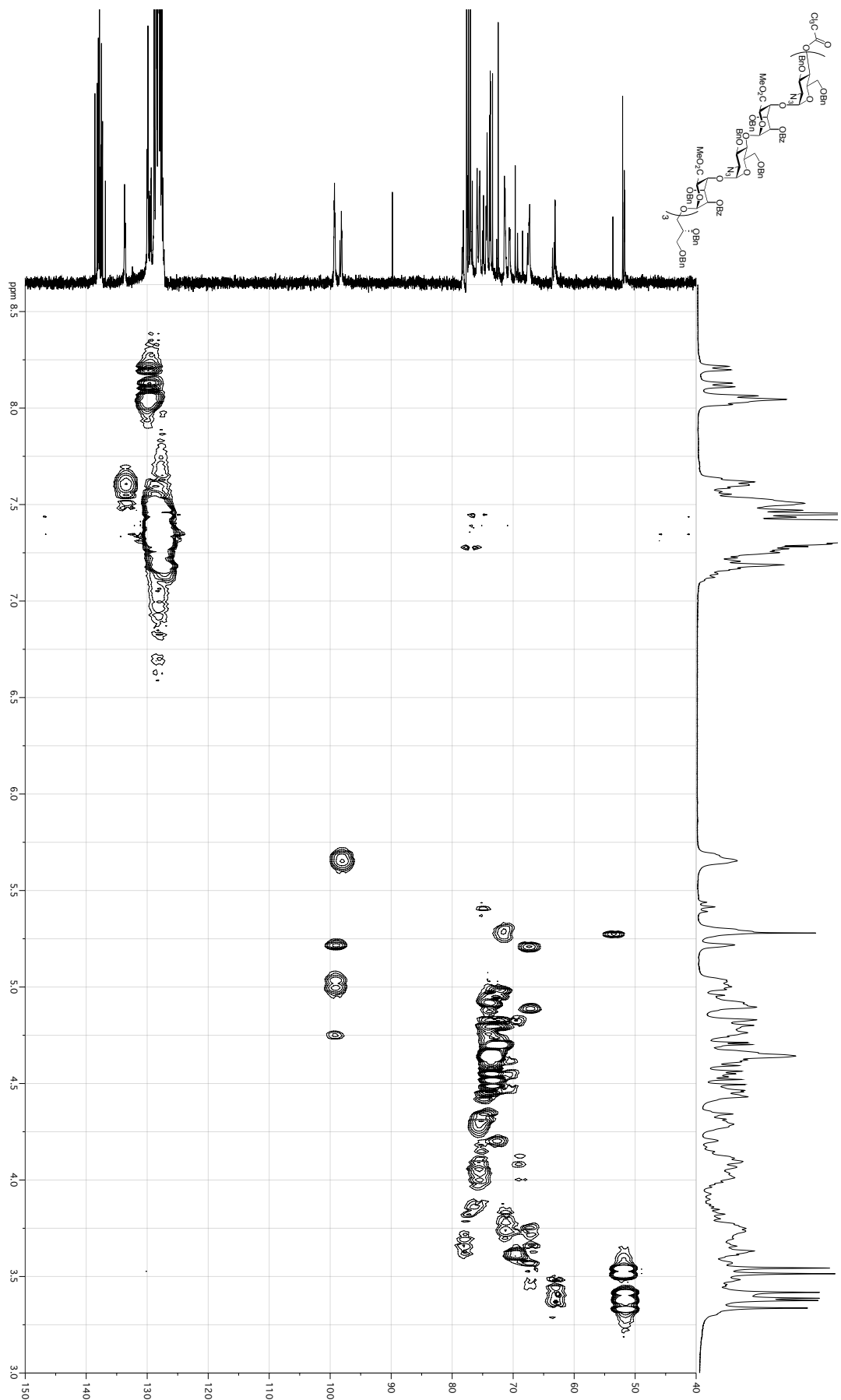
Supplementary Figure S50. ^1H NMR (500 MHz; CDCl_3) spectrum for **13**



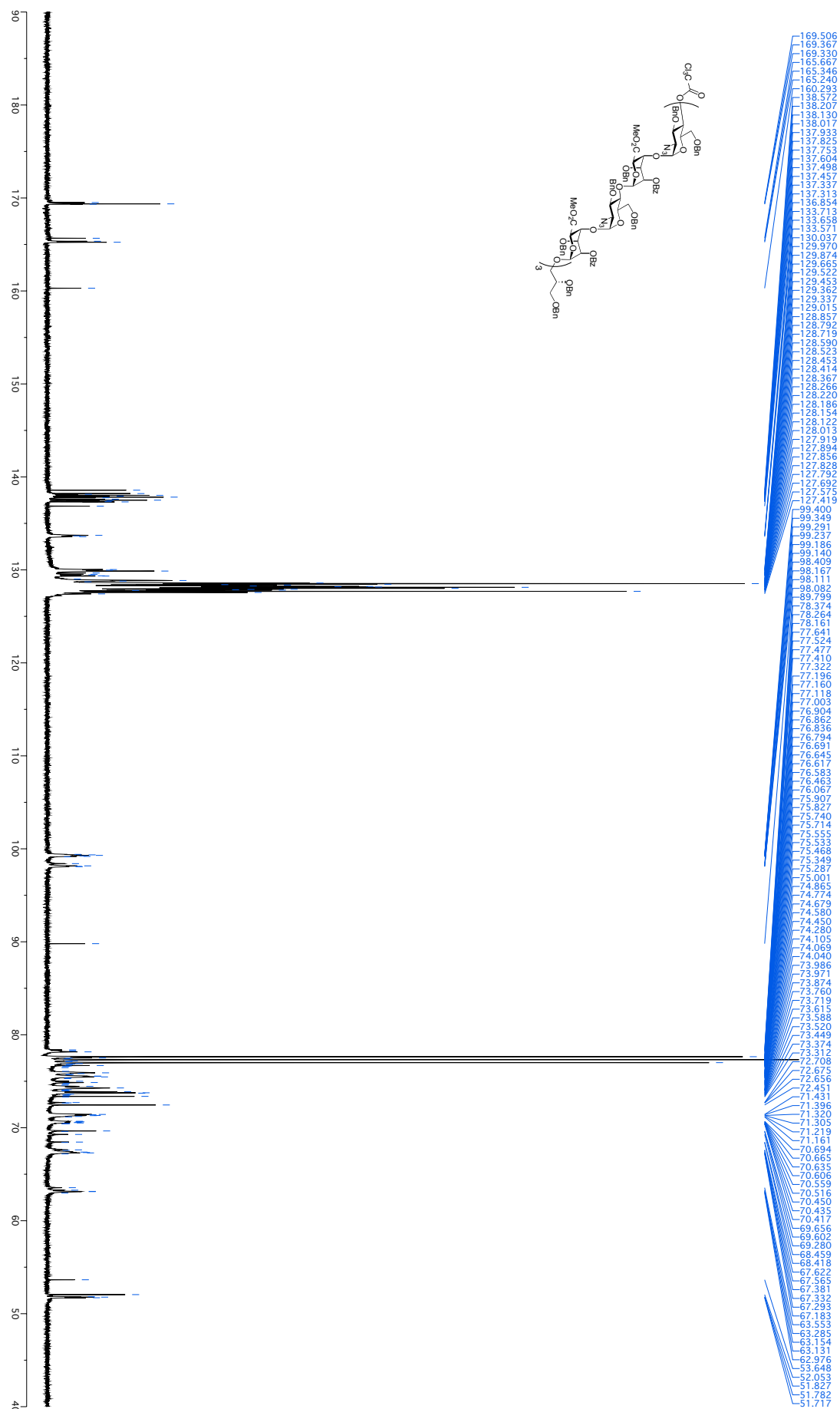
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 PROCNO: 1
 P2 - Acquisition Parameters
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 INSTRUM: spect
 PROBRD: 5 mm MULTISQ
 PULPROG: zgpg30
 TD: 655376
 SOLVENT: CDCl3
 NS: 1
 DS: 1
 SWH: 2314.815 Hz
 FIDRES: 1.130281 Hz
 AQ: 0.4424180 sec
 RG: 327.5
 ACQ: 216.013 usec
 DE: 6.00 usec
 TE: 294.2 K
 D0: 0.0000000 sec
 d13: 0.0000000 sec
 d16: 0.0002000 sec
 INO: 0.00043200 sec
 ===== CHANNEL f1 =====
 NUC1: 1H
 P0: 10.00 usec
 P1: 10.00 usec
 P2: 10.00 usec
 SFO1: 400.1322955 MHz
 ===== GRADIENT CHANNEL =====
 GBRM1: SINE 100
 GRNM1: SINE 100
 GP21: 10.00 %
 GP22: 10.00 %
 P16: 1000.00 usec
 F1 - Acquisition parameters
 ND0: 1
 TD: 128
 FIDRES: 400.1322955 Hz
 SW: 5.785 ppm
 FWHM: 5.785 ppm
 QF: 0
 P2 - Processing parameters
 SI: 1024
 SF: 400.1300000 MHz
 WDM: SINE
 SSB: 0 Hz
 GB: 0 Hz
 PC: 1.40
 F1 - Processing parameters
 S: 655376
 MC2: 1024
 SF: 400.1300000 MHz
 WDM: SINE
 L8: 0.00 Hz
 GB: 0



Supplementary Figure S51. COSY NMR spectrum for 13



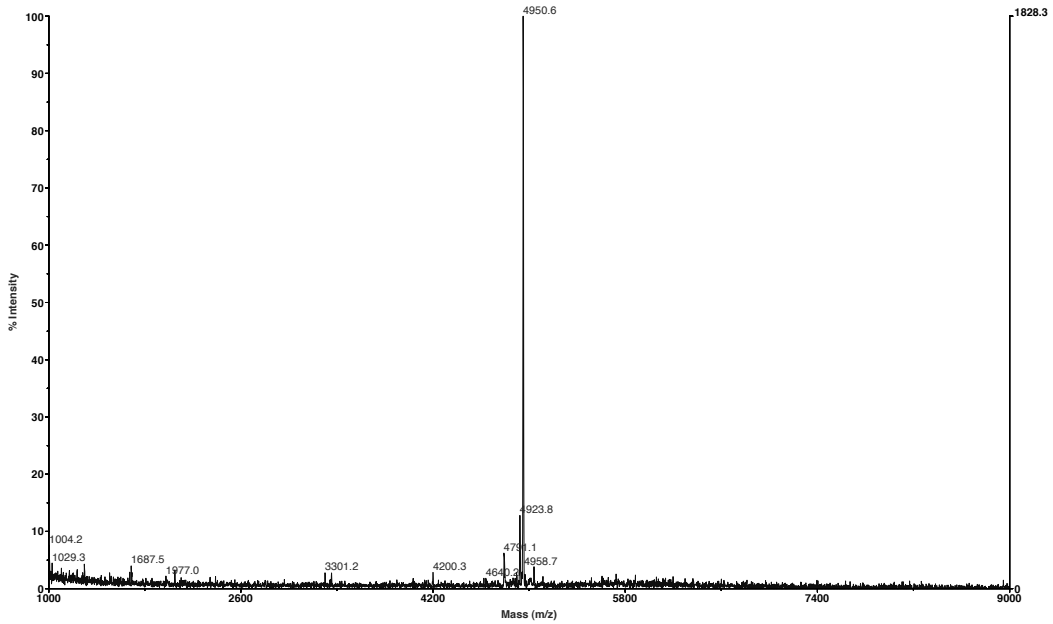
Supplementary Figure S52. HMQC NMR spectrum for 13



Supplementary Figure S53. ^{13}C NMR (100 MHz; CDCl_3) spectrum for **13**

EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea

<<MANGAR040_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)=>SM5[BP = 569.9, 4307]

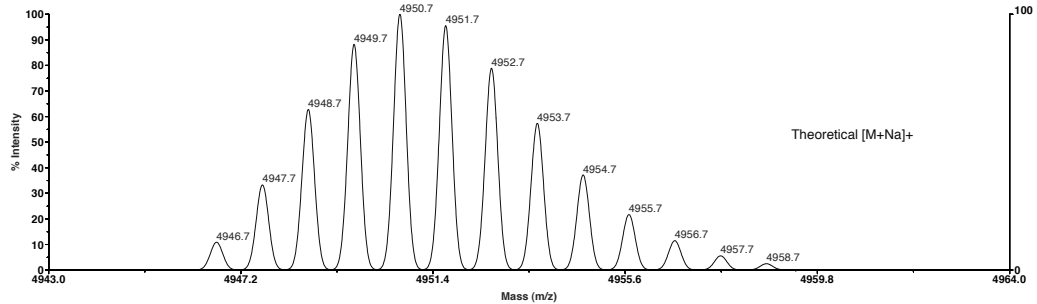


Acquired: 18:03:00, July 20, 2010
Hansen SU1305 MW=4923? DCM PosRef [1:49] (DCTB:DCM) +NaOAc
D:\2010\Jul10\MANGAR040_0001.dat

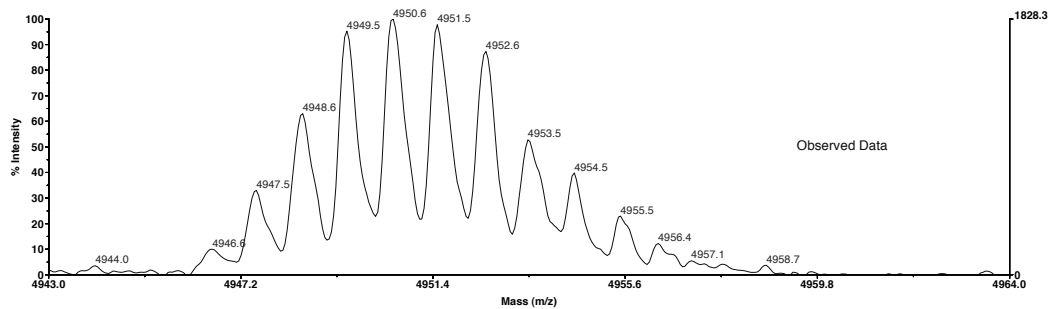
Printed: 10:00, July 21, 2010

EPSRC National Mass Spectrometry Service Centre (NMSSC), Swansea

ISO:C265H265Cl3N18O70 + (Na)1



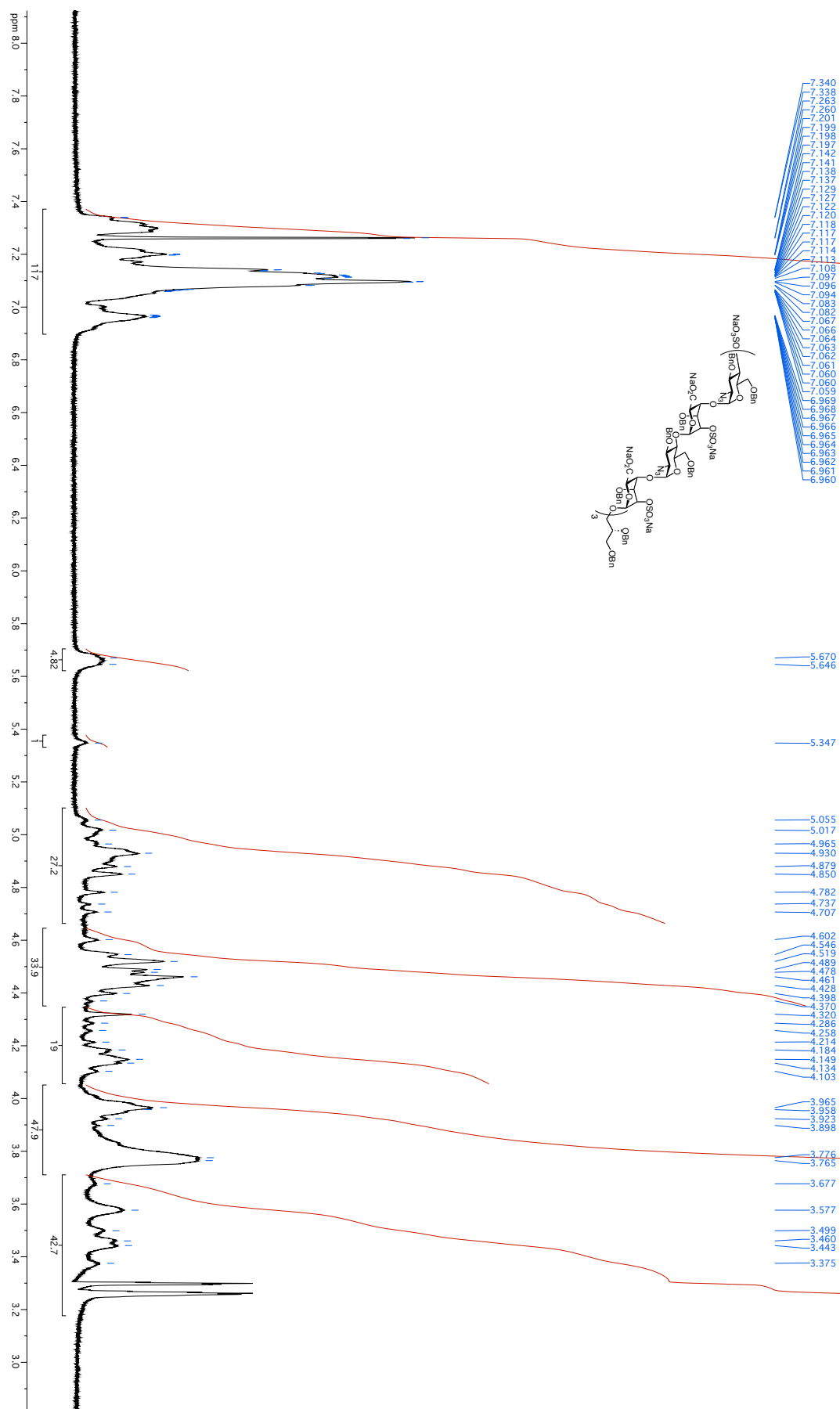
<<MANGAR040_0001>> Voyager Spec #1=>AdvBC(64,0.5,0.1)=>SM5[BP = 569.9, 4307]



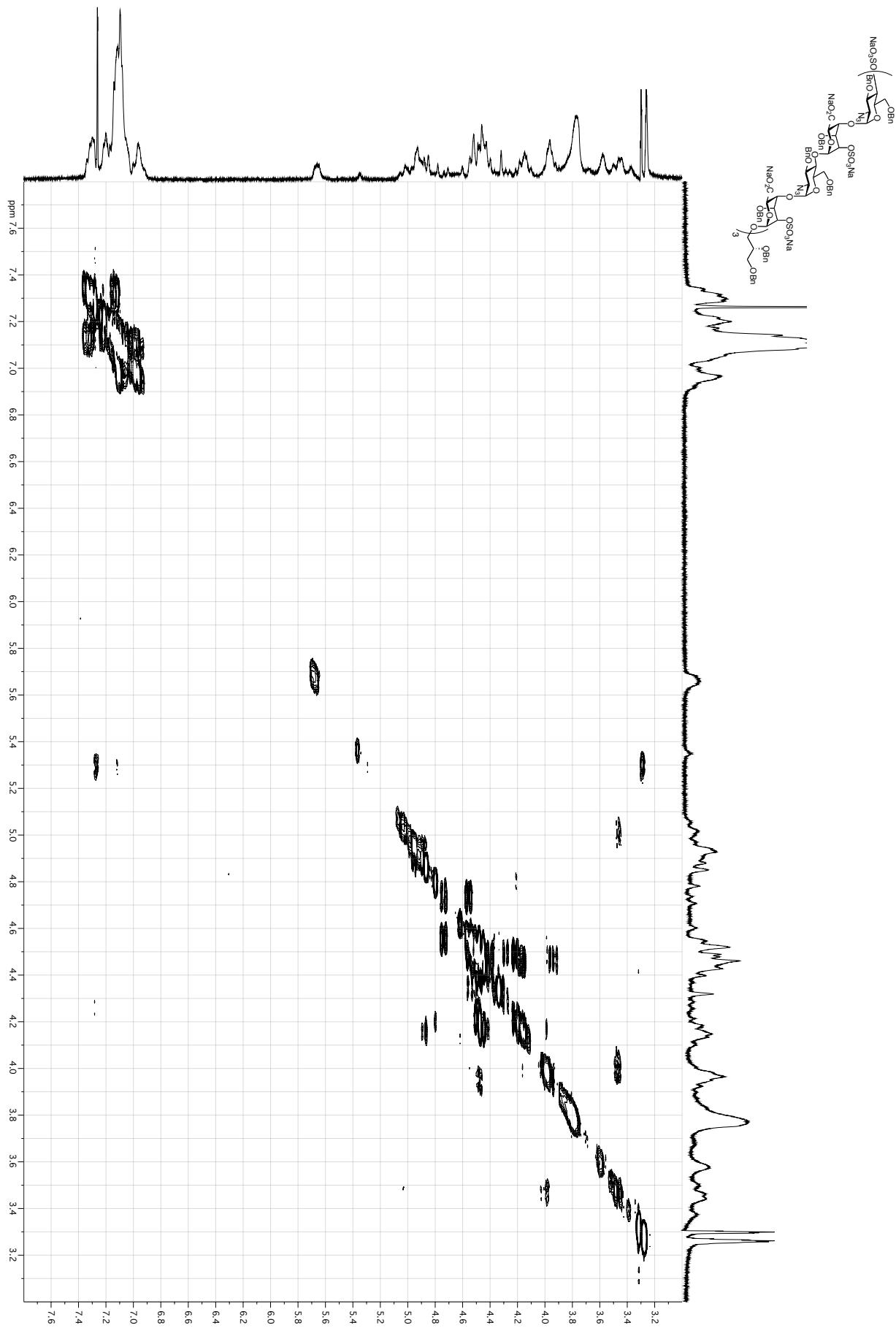
Acquired: 18:03:00, July 20, 2010
Hansen SU1305 MW=4923? DCM PosRef [1:49] (DCTB:DCM) +NaOAc
D:\2010\Jul10\MANGAR040_0001.dat

Printed: 10:01, July 21, 2010

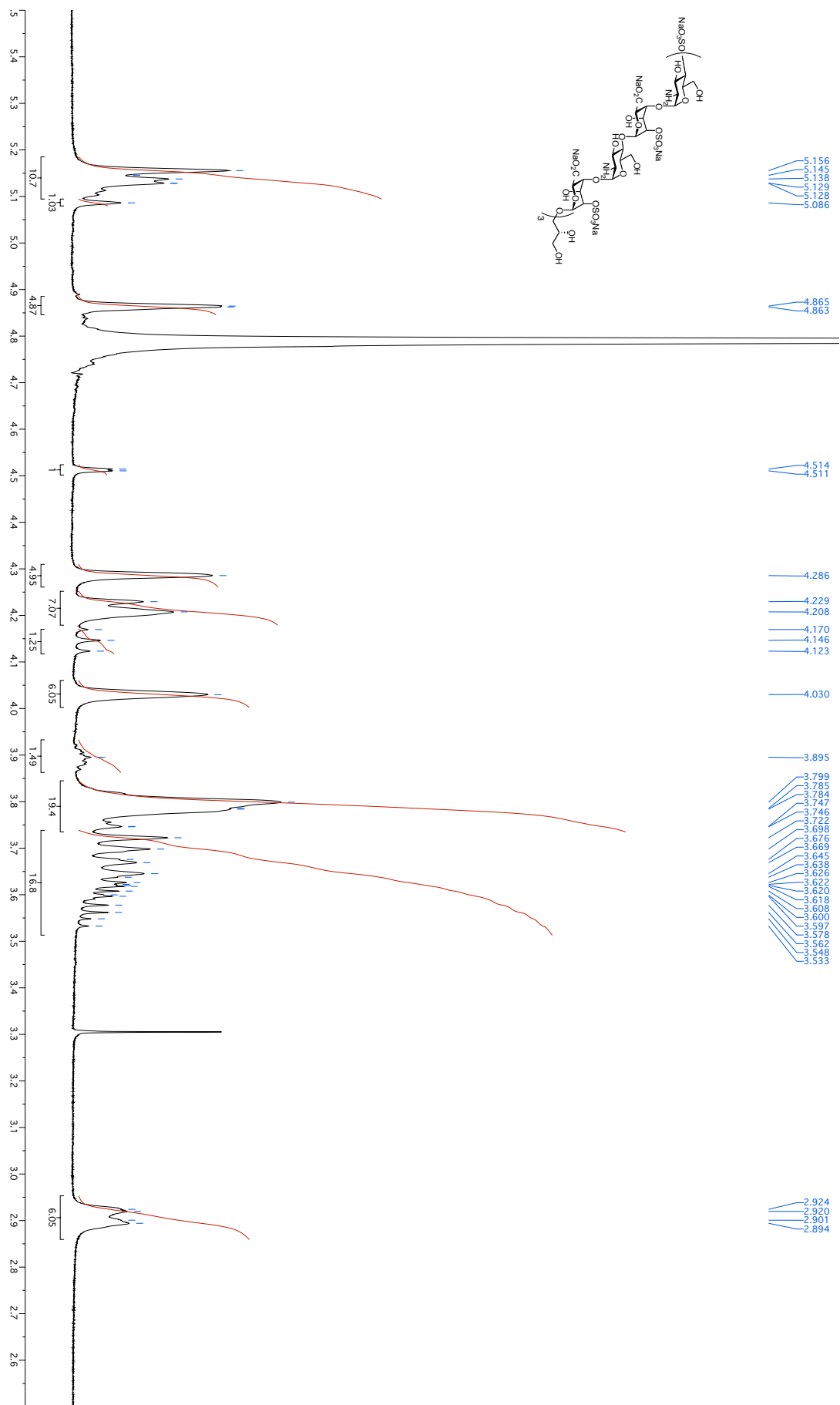
Supplementary Figure S54. MALDI-TOF MS spectrum for 13



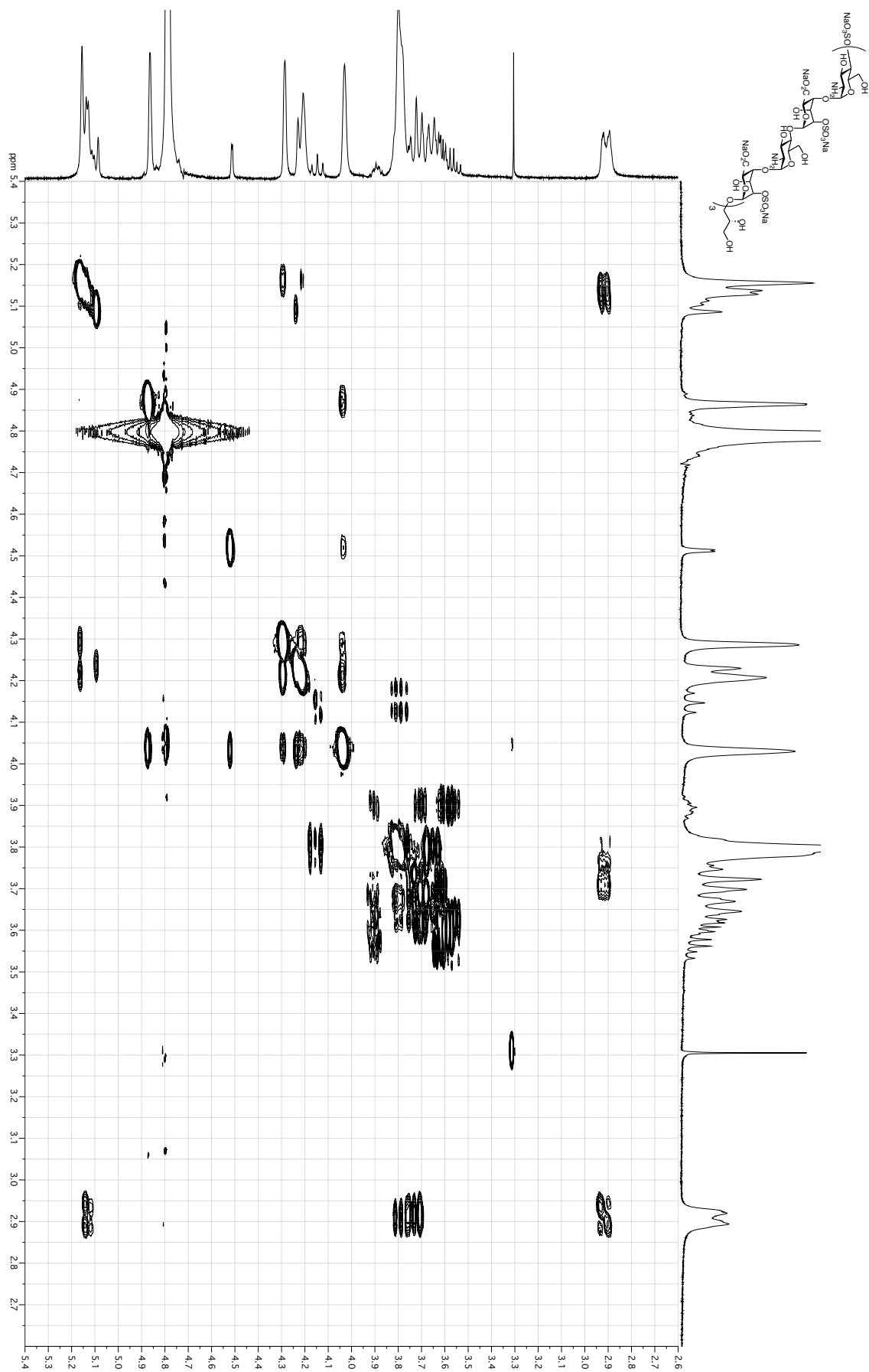
Supplementary Figure S55. ¹H NMR (400 MHz; CDCl₃/CD₃OD 10:1) spectrum for B



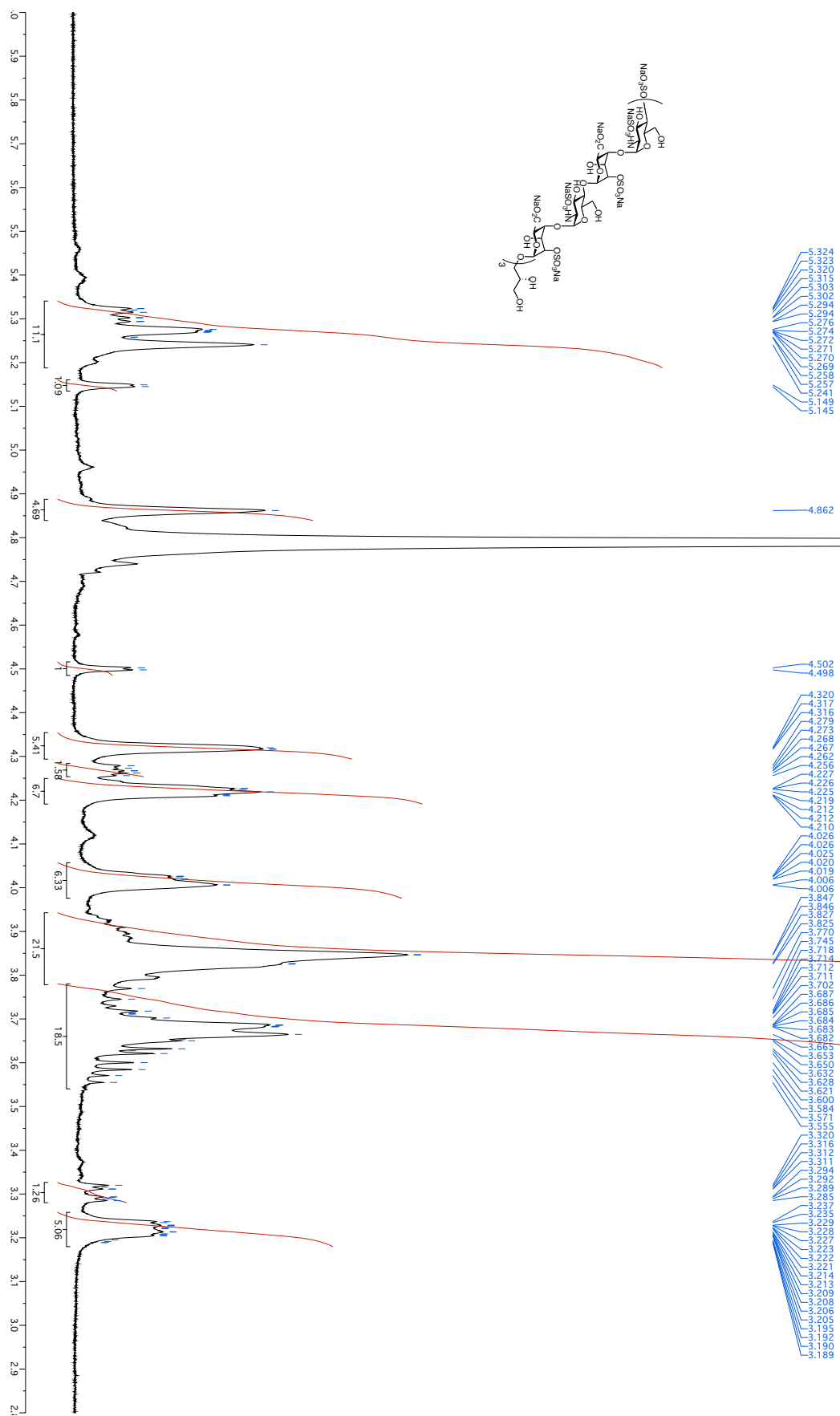
Supplementary Figure S56. COSY NMR spectrum for **B**



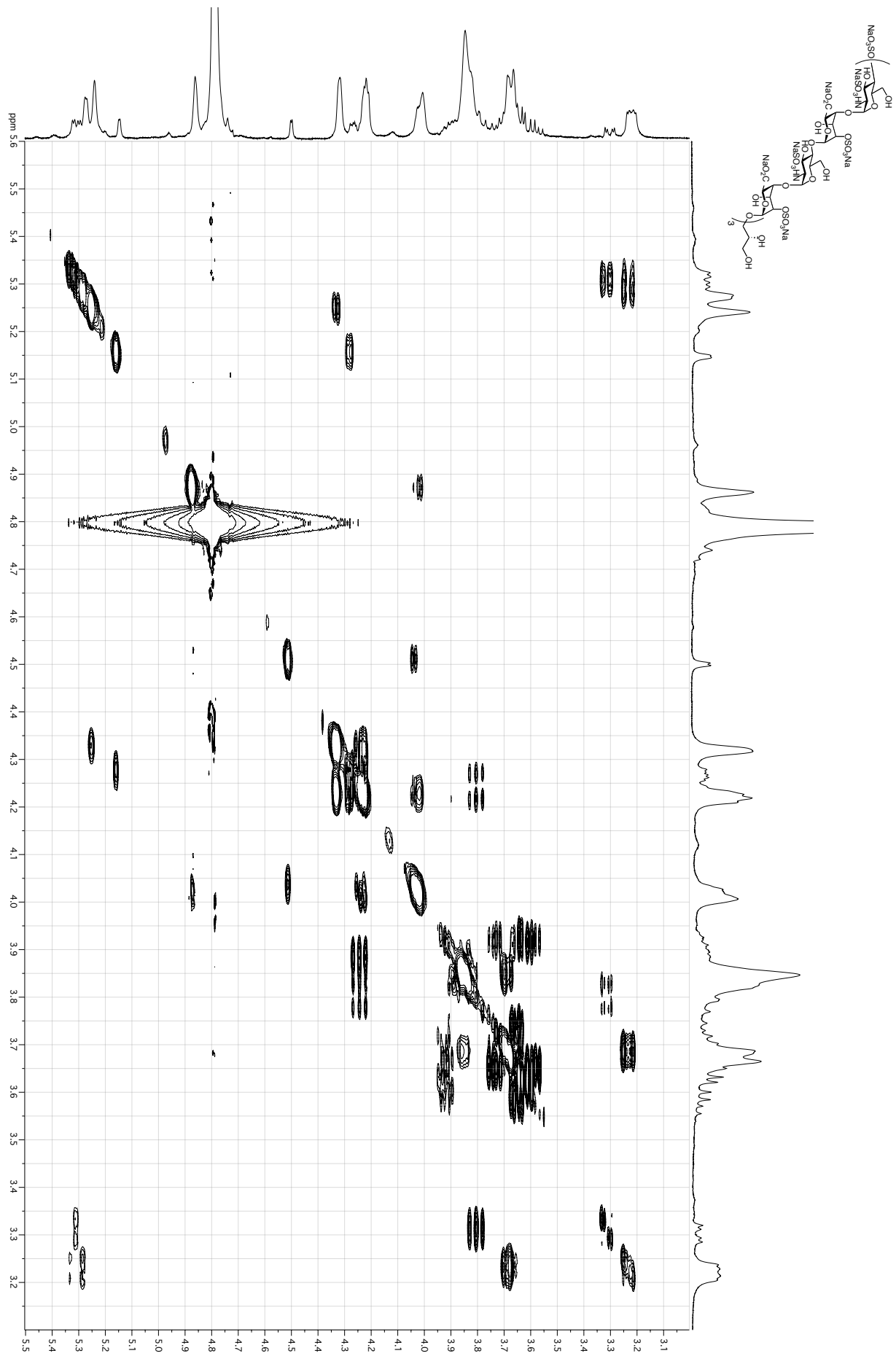
Supplementary Figure S57. ¹H NMR (400 MHz; D₂O) spectrum for dodecasaccharide NH₂ intermediate C



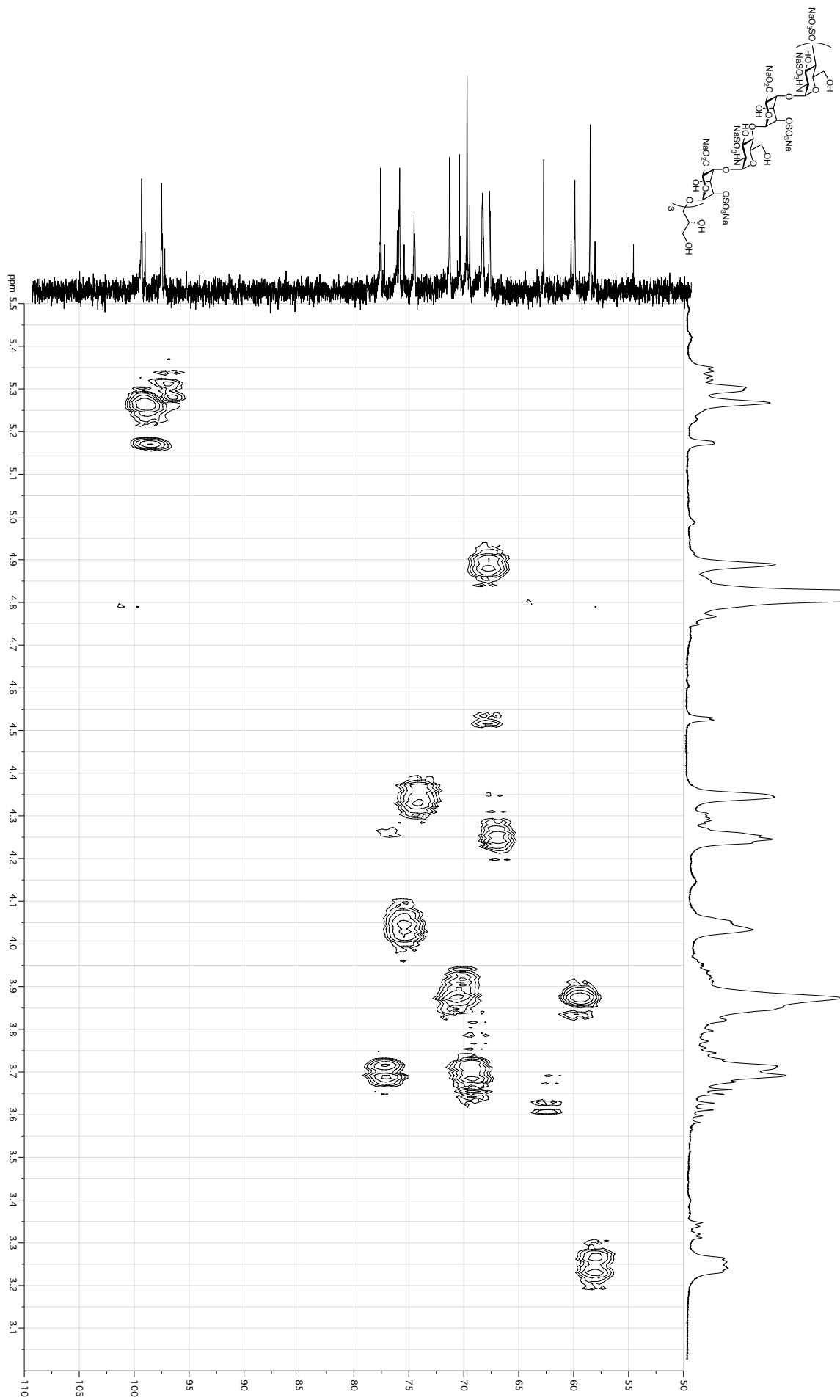
Supplementary Figure S58. COSY NMR spectrum for dodecasaccharide NH₂ intermediate C



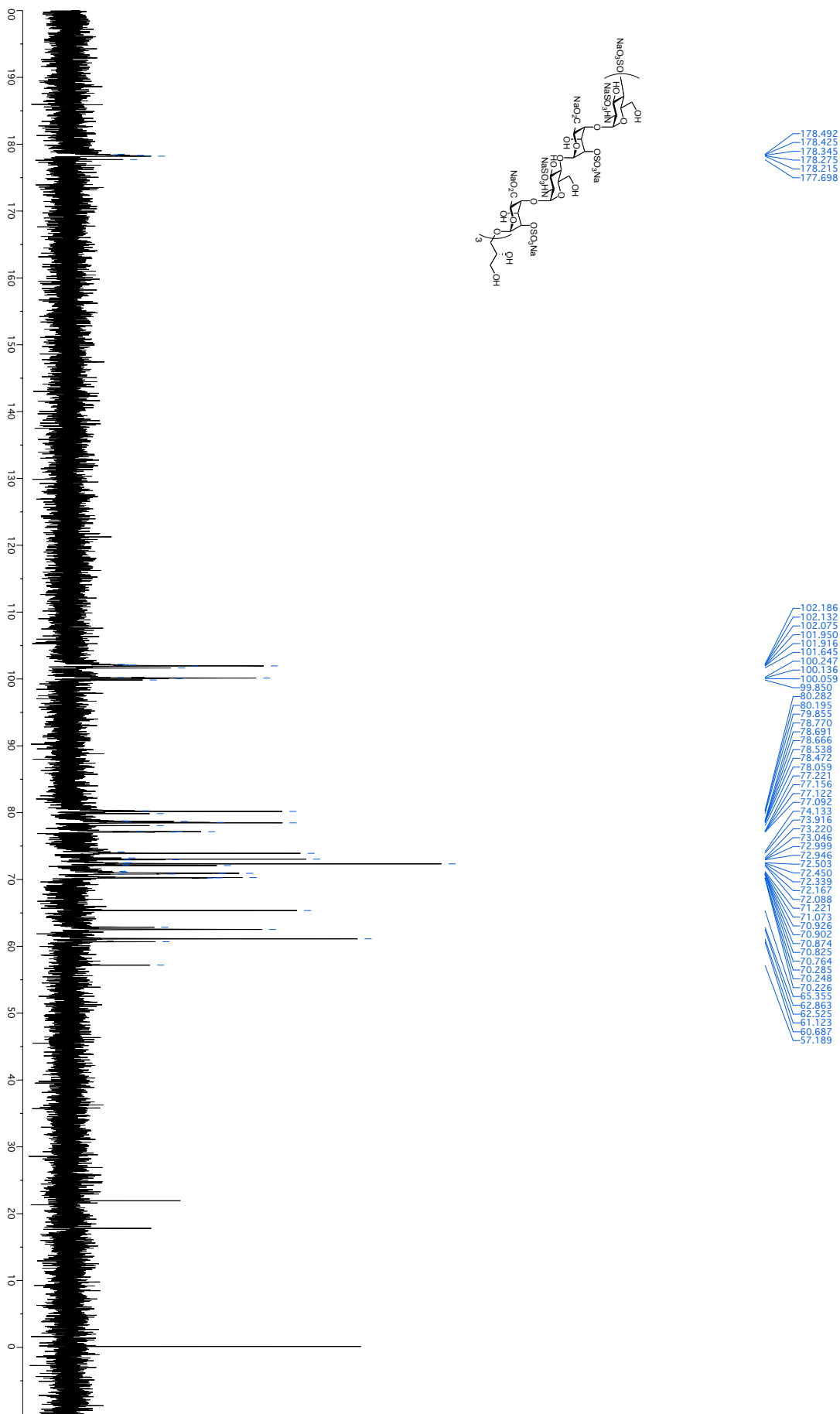
Supplementary Figure S59. ^1H NMR (400 MHz; D_2O) spectrum for 14



Supplementary Figure S60. COSY NMR spectrum for 14



Supplementary Figure S61. HMBC NMR spectrum for 14



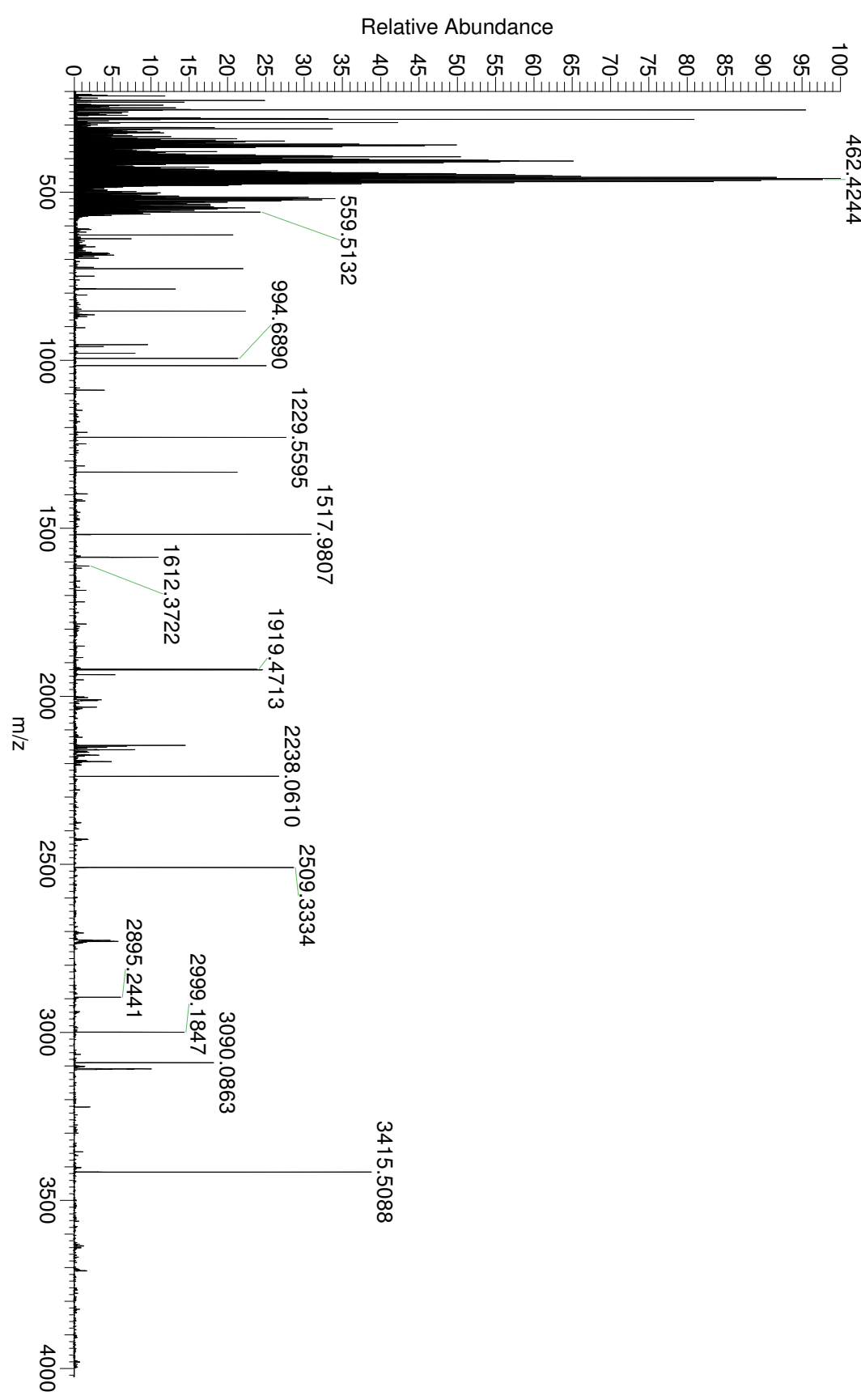
Supplementary Figure S62. ^{13}C NMR (100 MHz; D_2O) spectrum for 14

SU1311 MW=3571?
(H₂O)/MeOH

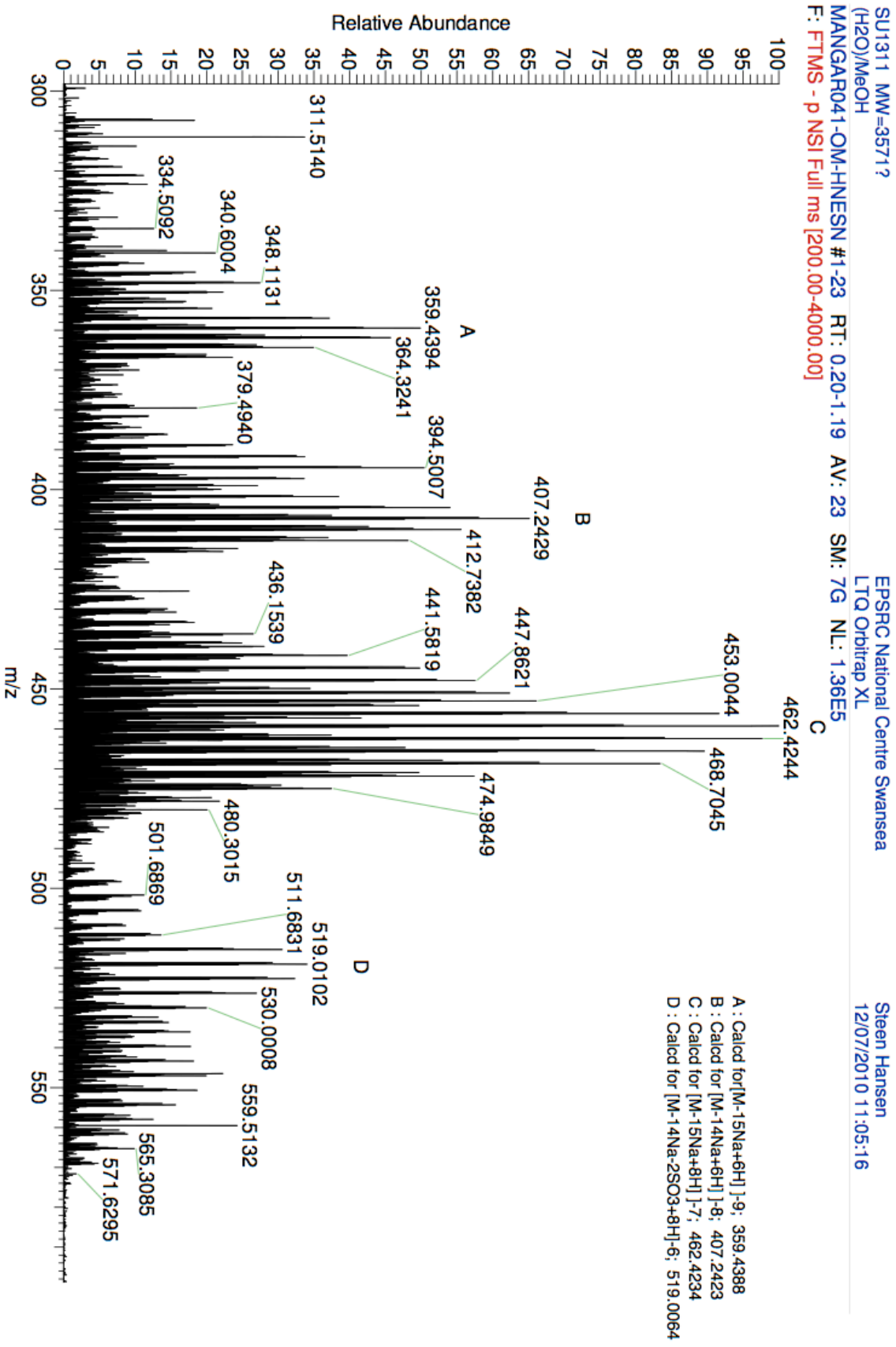
EPSRC National Centre Swansea
LTQ Orbitrap XL

Steen Hansen
12/07/2010 11:05:16

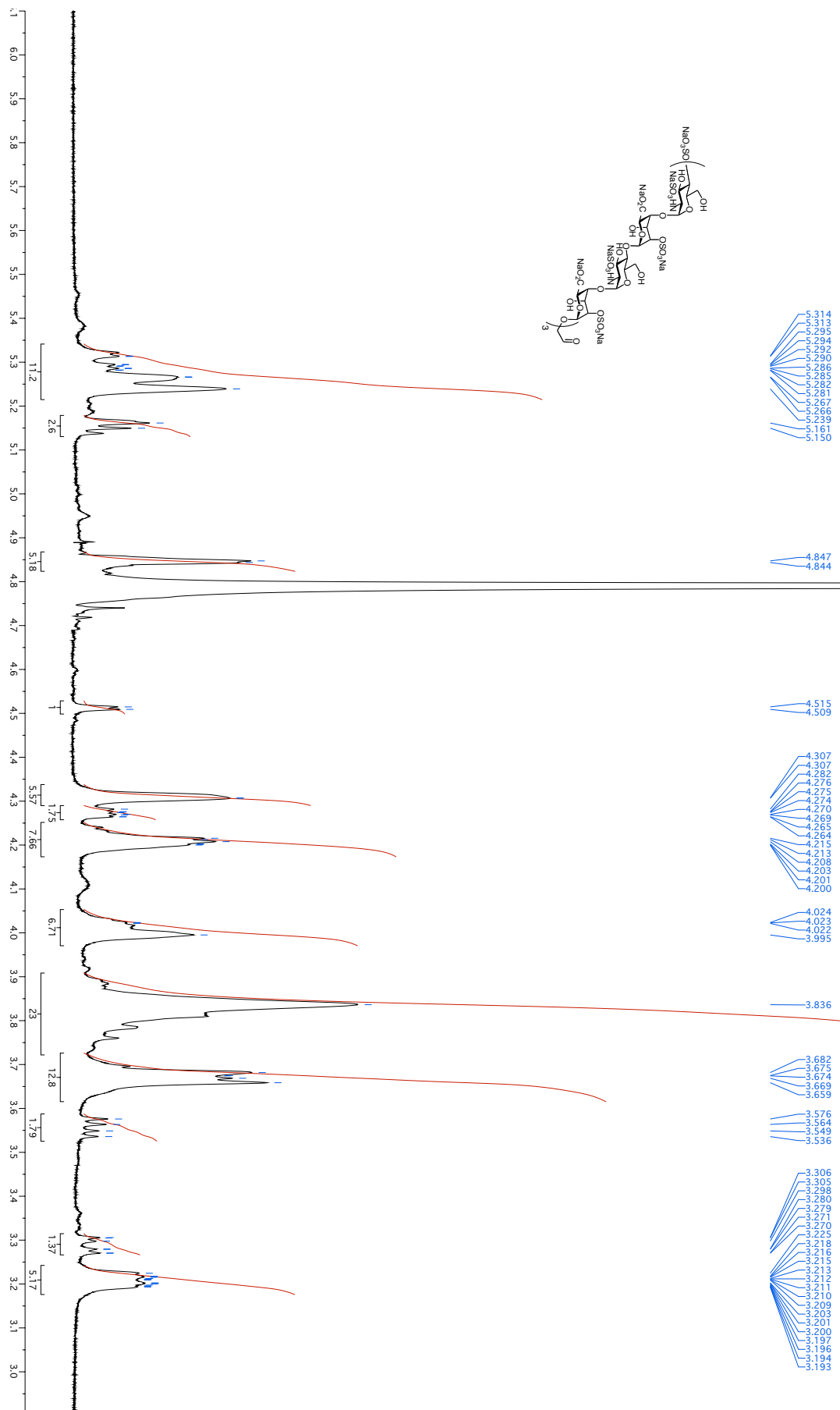
MANGAR041-OM-HNESN #1-23 RT: 0.20-1.19 AV: 23 SM: 7G NL: 1.36E5
F: FTMS - p NSI Full ms [200.00-4000.00]



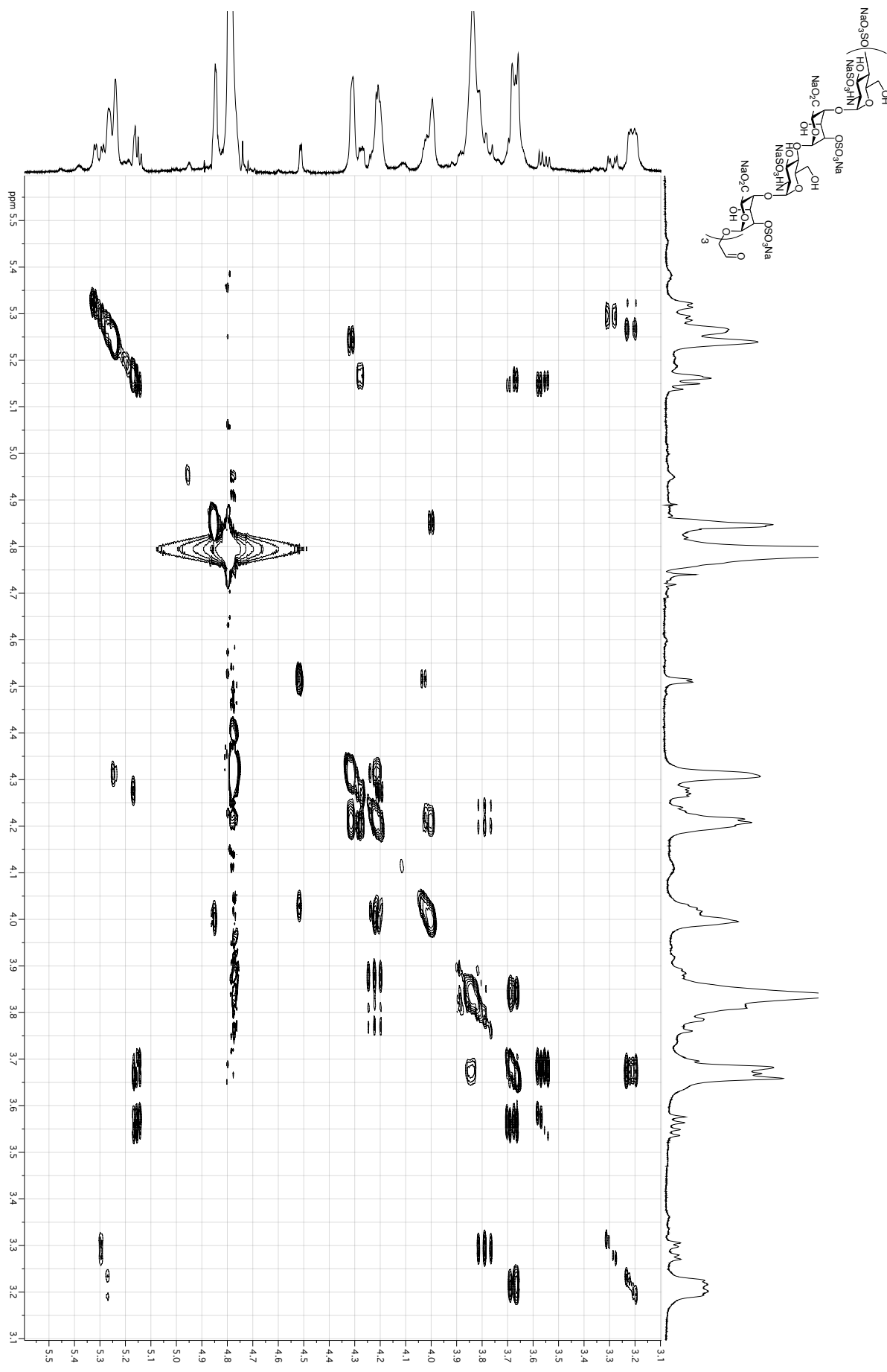
Supplementary Figure S63. FT MS negative mode spectrum for 14



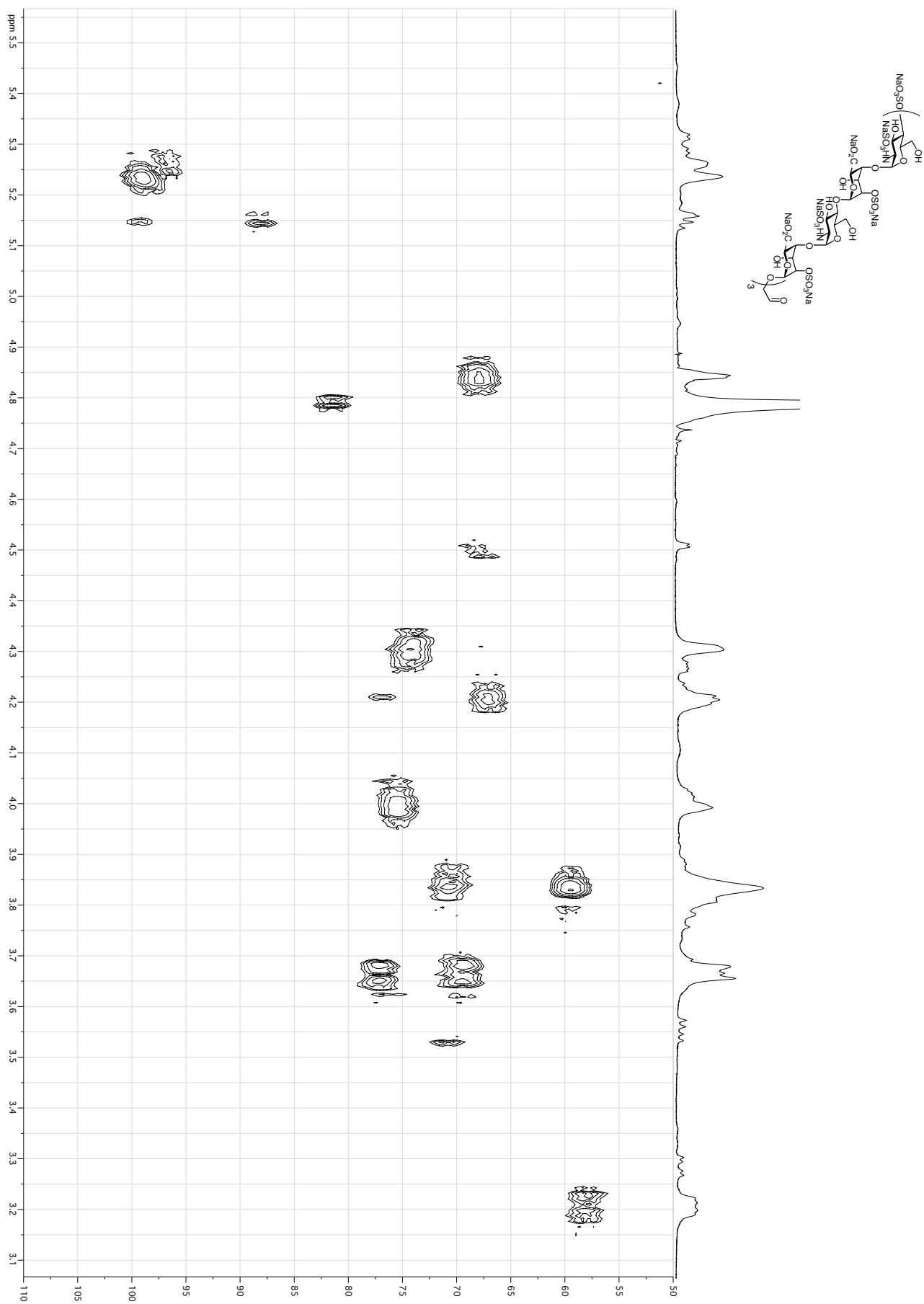
Supplementary Figure S64. FT MS negative mode assigned spectrum for 14



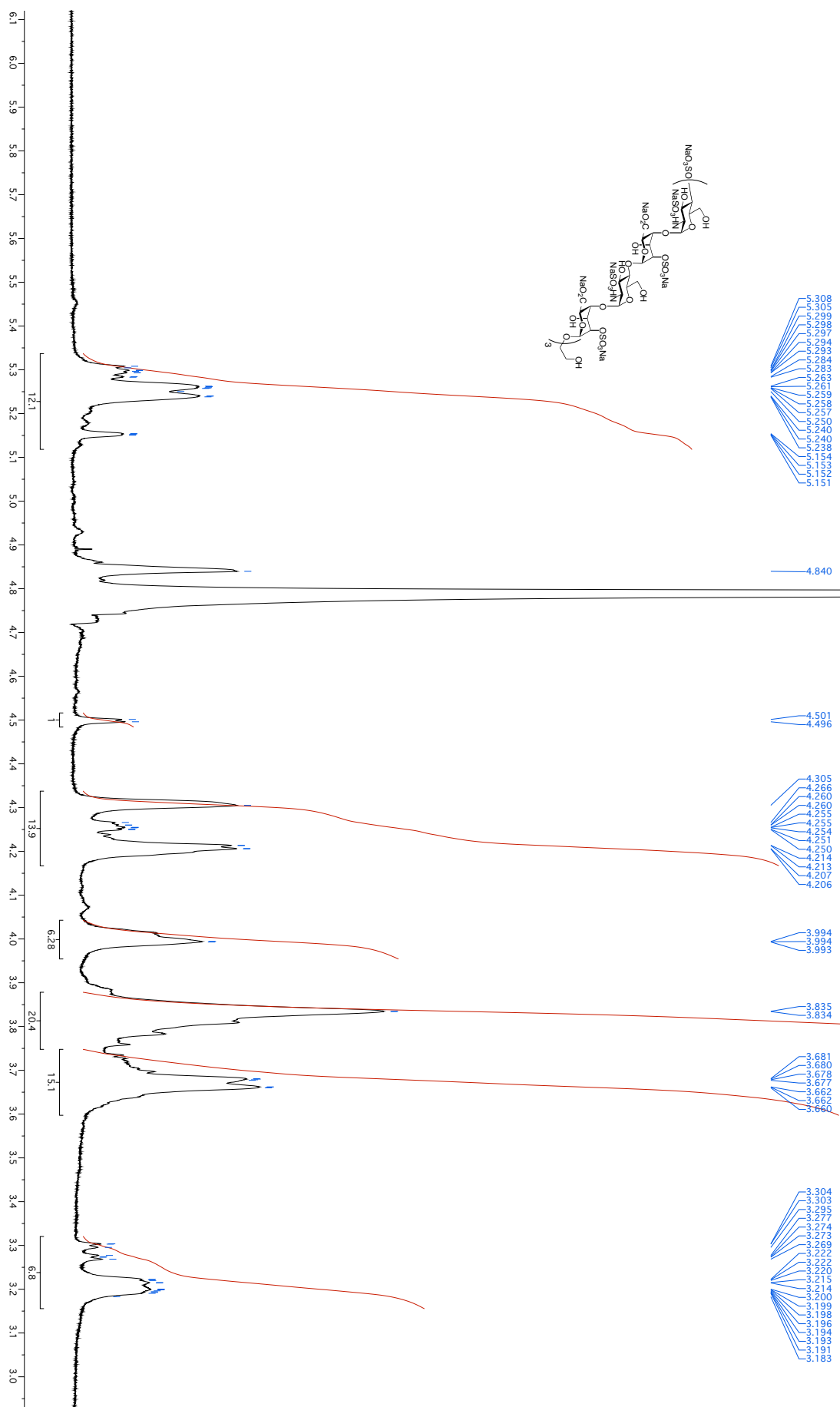
Supplementary Figure S65. ^1H NMR (400 MHz; D_2O) spectrum for 15



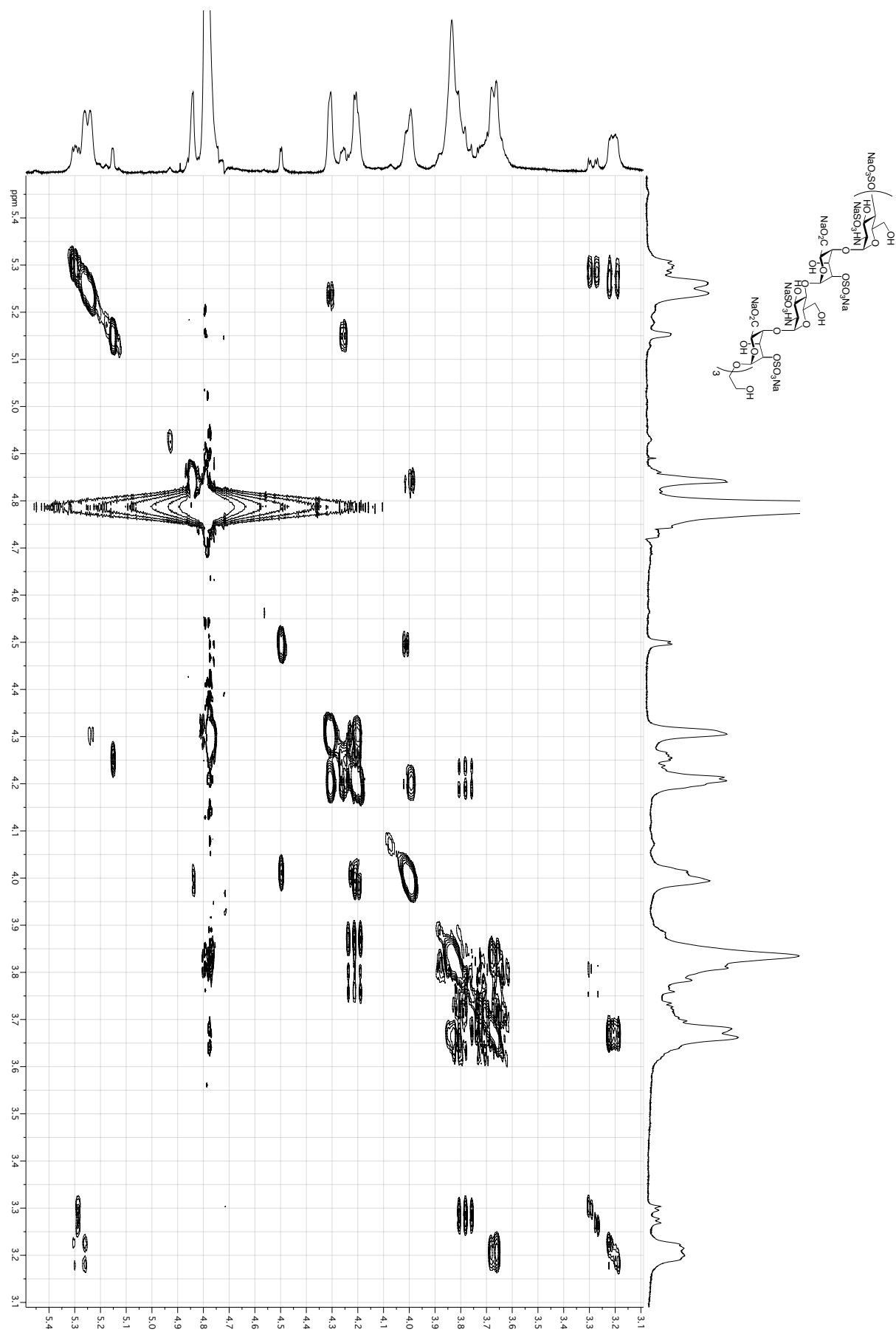
Supplementary Figure S66. COSY NMR spectrum for 15



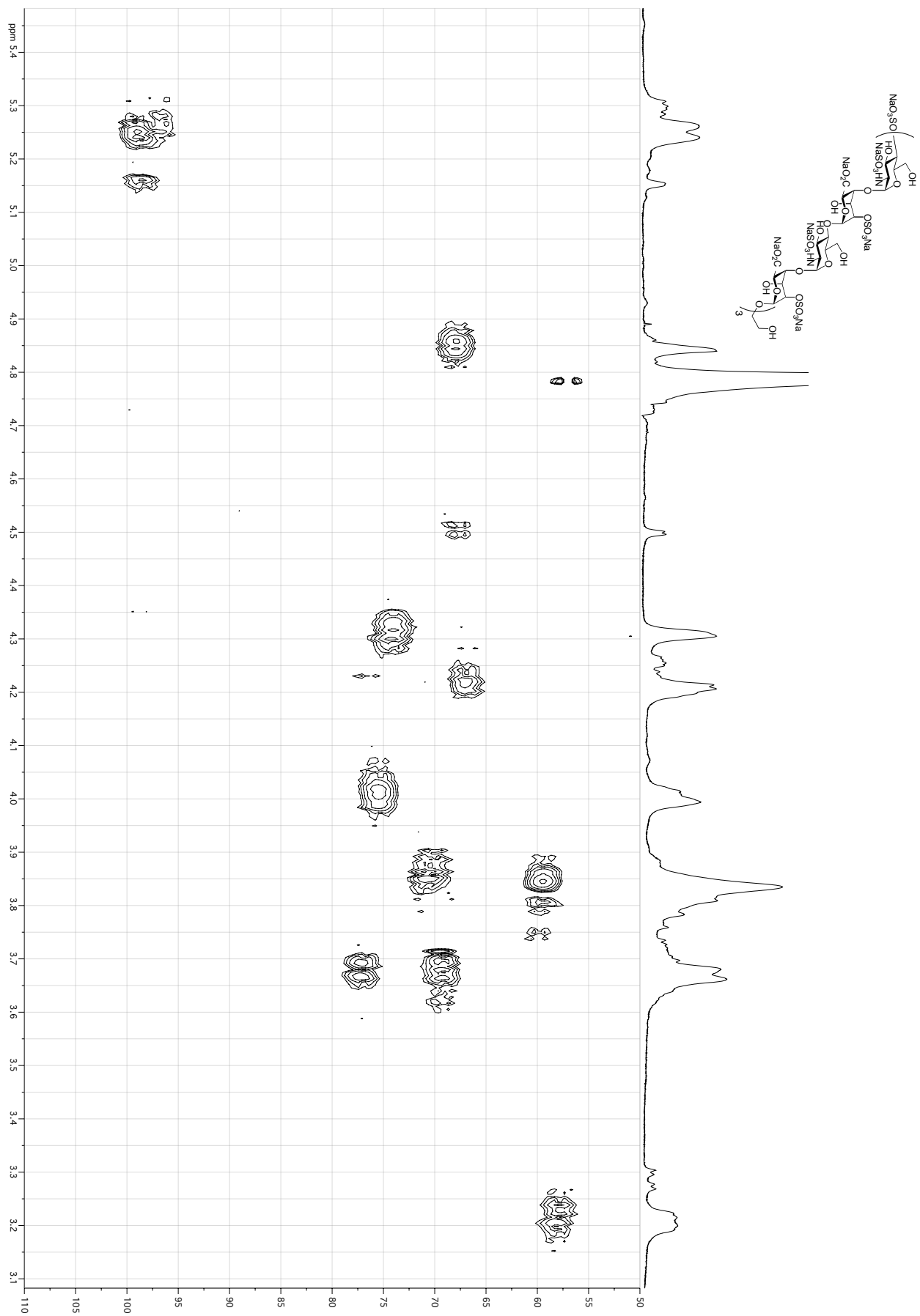
Supplementary Figure S67. HMPC NMR spectrum for 15



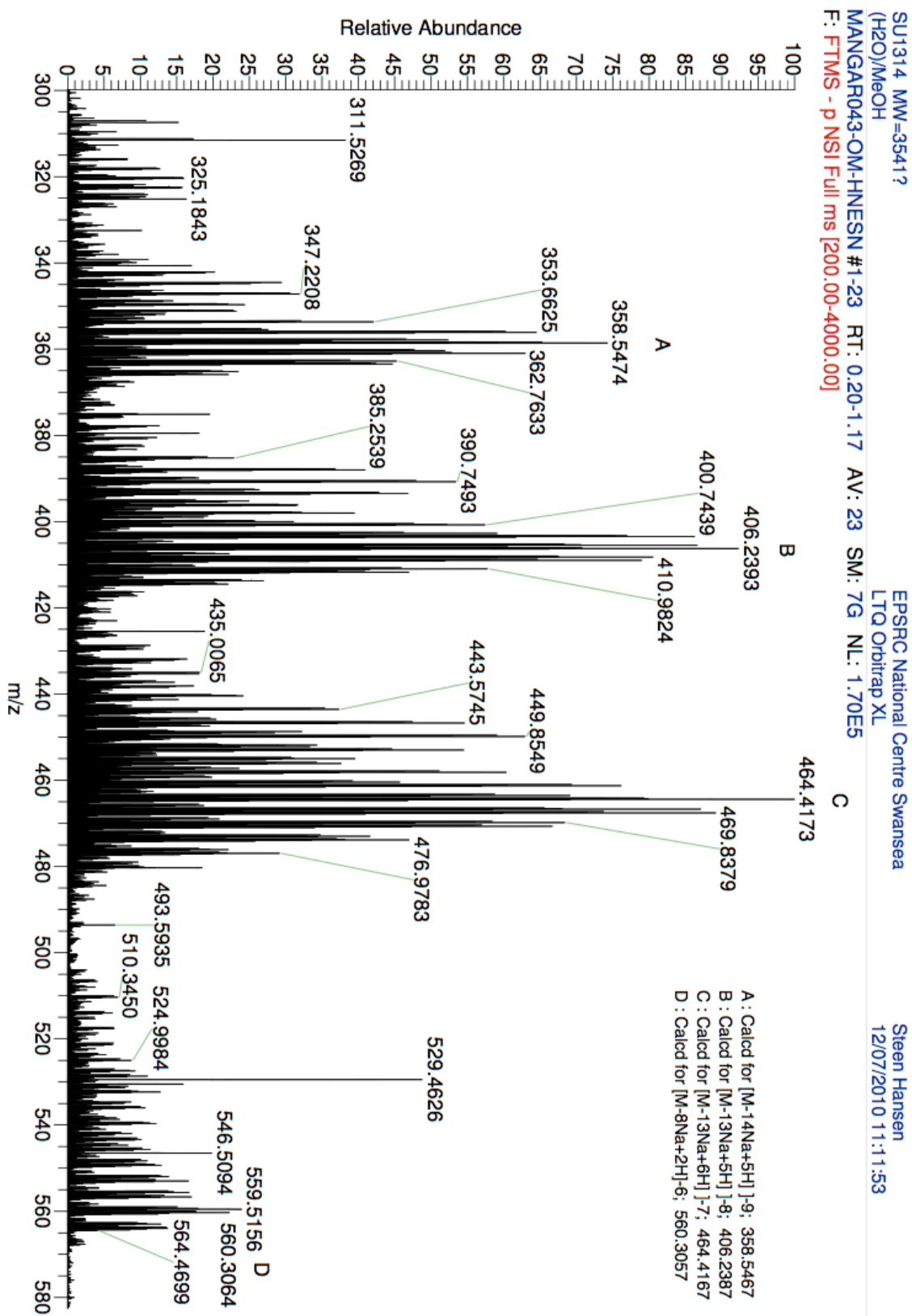
Supplementary Figure S68. ¹H NMR (400 MHz; D₂O) spectrum for 16



Supplementary Figure S69. COSY NMR spectrum for 16



Supplementary Figure S70. HMQC NMR spectrum for 16



Supplementary Figure S71. FT MS negative mode assigned spectrum for 16

Supplementary methods

General methods

All the chemicals used were purchased from commercial sources without further purification. All reactions were monitored by TLC on Merck silica gel plates ⁶⁰F₂₅₄. Silica gel 60 (particle size 0.035-0.070 mm) was used for column chromatography. ¹H NMR spectra were recorded at 500 or 400 MHz and ¹³C spectra at 100 MHz respectively on Bruker DPX spectrometers. Mass spectra (MS) were recorded using a Micromass Platform II spectrometer using an electro spray ionization source or via the EPSRC National Mass Spectrometry Service (Swansea). Isotope patterns for compounds with mass > 1000 are included in the SI. Optical rotations were obtained using an AA-1000 polarimeter. Elemental analyses were performed by Micro Analytical Laboratory, School of Chemistry, The University of Manchester.

NMR Data were reprocessed using iNMR 4 from Nucleomatica.

Synthesis of Oligosaccharides

Methyl 2-azido-3,6-di-*O*-benzyl-2-deoxy-4-*O*-*p*-methoxybenzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl-L-idopyranose uronate) (3)

To **2** (1.85 g, 1.88 mmol) was added acetone 50 mL and cooled to 0 °C in an ice-bath. *N*-Bromosuccinimide (335 mg, 1.88 mmol) was then added and the mixture stirred for 45 min. The reaction was quenched by addition of saturated NaHCO₃ solution (5 mL), the acetone evaporated and DCM (200 mL) and water (200 mL) added. The organic phase was separated, dried (MgSO₄), filtered and evaporated. The crude product was purified using flash column chromatography (EtOAc/hexane 1:2) to yield 1.45 g (87%) of **3** as a white foam ($\alpha/\beta \sim 1:1$). *R*_f 0.17 (EtOAc/Hexane 1:2). ¹H NMR (400 MHz; CDCl₃) δ 8.11-8.07 (m, 2H, Bz), 7.40-7.17 (m, 16H, Ph), 7.11-7.07 (m, 2H, Bz), 7.02 (d, *J* = 8.8 Hz, 2H, PMB), 6.84 (d, *J* = 8.8 Hz, 2H, PMB), 5.46-5.43 (m, 1H, H-1 α), 5.22 (dd, *J* = 11.6 Hz, 2 Hz, 1H, H-1 β), 5.08-5.06 (m, 2H, H-2 α , H-2 β), 4.91-4.90 (m, 1H, H-5 α), 4.90-4.75 (m, 2H, CH₂Ph), 4.69 (d, *J* = 3.6 Hz, 1H, H'-1), 4.66 (d, *J* = 3.6 Hz, 1H, H'-1), 4.62-4.35 (m, 4H, CH₂Ph, CH₂PMP), 4.60-4.59 (m, 1H, H-5 β), 4.33 (t, *J* = 2.8 Hz, 1H, H-3 β), 4.30 (dt, *J* = 2.8 Hz, 1.2 Hz, 1H, H-3 α), 4.20 (d, *J* = 9.2 Hz, 1H, OH α), 4.06-3.59 (m, 14H, H-4, H'-4, H'-5, H'-6, CH₂Ph), 3.82 (s, 3H, PhOCH₃), 3.81 (s, 3H, PhOCH₃), 3.74 (s, 3H, COOCH₃), 3.73 (s, 3H, COOCH₃), 3.41-3.36 (m, 2H, H'-3), 3.25-3.21 (m, 2H, H'-2). ¹³C NMR (100 MHz; CDCl₃) δ 169.5, 168.7, 165.9, 165.7, 159.2, 137.9, 137.9, 137.8, 137.0, 136.5, 133.4, 133.3, 130.4, 130.0, 129.5, 129.4, 129.0, 128.9, 128.8, 128.7, 128.6, 128.5, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 127.8, 127.7, 127.7, 127.6, 113.6, 100.4, 93.8, 92.6, 80.2, 80.0, 77.4, 77.3, 77.3, 76.2, 75.9, 74.5, 74.5, 74.0, 73.7, 73.6, 73.4, 73.0, 71.8, 71.7, 68.6, 67.6, 67.5, 67.0, 63.7, 63.7, 56.3, 55.3, 52.5, 52.4. HRMS (FT MS): *m/z*: calcd for C₄₉H₅₅N₄O₁₃ [*M*+NH₄]⁺: 907.3760; found: 907.3766.

Methyl 2-azido-3,6-di-*O*-benzyl-2-deoxy-4-*O*-*p*-methoxybenzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl-1-trichloroacetimidate- α/β -L-idopyranuronate) (4)

To **3** (505 mg, 0.57 mmol) was added dry DCM (10 mL), CCl₃CN (0.40 mL, 3.99 mmol), 1,8-diazabicyclo[5.4.0]undec-7-ene (DBU) (10 μ L, 0.07 mmol) and the mixture stirred under N₂ for 1 h. The solution was evaporated and the crude product was purified using flash column

chromatography (EtOAc/hexane 1:3 with 1% NEt₃), yielding 535 mg (91%) of **4** as an oil ($\alpha/\beta \sim 3:1$). α -**4**: *R_f* 0.30 (EtOAc/Hexane 1:3). ¹H NMR (400 MHz; CDCl₃) δ 8.70 (s, 1H, C=NH), 8.15-8.12 (m, 2H, Bz), 7.44-7.11 (m, 18H, Ph), 7.07 (d, *J* = 8.6 Hz, 2H, PMB), 6.87 (d, *J* = 8.6 Hz, 2H, PMB), 6.57-6.56 (m, 1H, H-1), 5.36-5.35 (m, 1H, H-2), 5.03 (d, *J* = 2.6 Hz, 1H, H-5), 4.96-4.77 (m, 2H, CH₂Ph), 4.81 (d, *J* = 3.5 Hz, 1H, H[']-1), 4.63-4.40 (m, 4H, CH₂Ph, CH₂PMP), 4.27-4.26 (m, 1H, H-3), 4.20-4.19 (m, 1H, H-4), 4.13-4.11 (m, 1H, CH₂Ph), 3.93-3.81 (m, 4H, H[']-5, H[']-6a, H[']-6b, CH₂Ph), 3.83 (s, 3H, PhOCH₃), 3.75 (s, 3H, COOCH₃), 3.71-3.66 (m, 1H, H[']-4), 3.50 (t, *J* = 10.0 Hz, 1H, H[']-3), 3.25 (dd, *J* = 10.3 Hz, *J* = 3.5 Hz, 1H, H[']-2). ¹³C NMR (100 MHz; CDCl₃) δ 168.7, 165.5, 160.2, 159.3, 137.9, 137.8, 137.3, 133.5, 130.5, 130.1, 129.5, 129.3, 128.9, 128.4, 128.4, 128.2, 128.0, 127.9, 127.8, 127.7, 127.7, 113.7, 100.3, 96.1, 90.8, 80.1, 77.4, 77.1, 76.7, 75.8, 74.6, 74.5, 73.6, 72.5, 72.2, 71.8, 69.0, 67.7, 65.7, 63.7, 55.4, 52.5.

β -**4**: *R_f* 0.16 (EtOAc/Hexane 1:3). ¹H NMR (400 MHz; CDCl₃) δ 8.68 (s, 1H, C=NH), 8.14-8.12 (m, 2H, Bz), 7.43-7.12 (m, 18H, Ph), 7.04 (d, *J* = 8.6 Hz, 2H, PMB), 6.85 (d, *J* = 8.6 Hz, 2H, PMB), 6.31 (d, *J* = 1.9 Hz, 1H, H-1), 5.48 (dd, *J* = 3.0 Hz, *J* = 1.9 Hz, 1H, H-2), 4.92-4.79 (m, 2H, CH₂Ph), 4.62-4.37 (m, 5H, H-3, CH₂Ph, CH₂PMP), 4.13-4.11 (m, 1H, CH₂Ph), 4.04-4.03 (m, 1H, H-4), 3.88-3.61 (m, 5H, H[']-4, H[']-5, H[']-6a, H[']-6b, CH₂Ph), 3.82 (s, 3H, PhOCH₃), 3.75 (s, 3H, COOCH₃), 3.48 (t, *J* = 10.2 Hz, 1H, H[']-3), 3.28 (dd, *J* = 10.2 Hz, *J* = 3.5 Hz, 1H, H[']-2). ¹³C NMR (100 MHz; CDCl₃) δ 167.9, 166.3, 163.6, 160.4, 159.3, 137.9, 137.8, 136.9, 133.3, 130.4, 130.1, 129.6, 128.8, 128.7, 128.5, 128.4, 128.4, 128.3, 128.3, 128.2, 127.9, 127.8, 127.7, 113.7, 100.5, 95.2, 90.5, 80.2, 77.4, 77.1, 76.8, 76.7, 76.2, 74.7, 74.6, 74.5, 73.8, 73.6, 73.3, 71.8, 67.7, 66.5, 63.8, 55.4, 52.5. HRMS (FT MS): *m/z*: calcd for C₅₁H₅₁Cl₃N₄O₁₃ [M+NH₄]⁺: 1052.2844; found: 1052.2842.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy-4-*O*-*p*-methoxybenzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(phenyl 2-*O*-benzoyl-3-*O*-benzyl-1-thio- β -L-idopyranoside)] uronate (β -6)

To **5** (1.05 g, 1.21 mmol) and **4** (1.43 g, 1.38 mmol) was added dry DCM (20 mL), the mixture cooled to -25 °C and TMSOTf (2 μ L, 0.01 mmol) added and the mixture stirred under a N₂ atmosphere for 30 min. More TMSOTf (2 μ L, 0.01 mmol) was then added, the mixture left another 30 min. and then quenched by addition of a few drops of NEt₃. The solution was evaporated and the crude product was purified using flash column chromatography (EtOAc/hexane 1:3). A second column (DCM/EtOAc 20:1) removed remaining Cl₃CONH₂. This yielded 1.80 g (85%) of the product β -**6** as a foam. *R_f* 0.16 (EtOAc/Hexane 1:3). [α]_D²⁰ = +21.1 (*c* = 0.85, CH₂Cl₂). ¹H NMR (400 MHz; CDCl₃) δ 8.23-8.21 (m, 2H, Bz), 7.96-7.94 (m, 2H, Bz), 7.56-7.53 (m, 2H, Bz), 7.41-7.21 (m, 35H, Ph), 7.09-7.05 (m, 4H, Bz, PMB), 6.84 (d, *J* = 8.4 Hz, 2H, PMB), 5.50 (d, *J* = 4.0 Hz, 1H, H[']-1), 5.26 (d, *J* = 2.0 Hz, 1H, H-1), 5.23-5.22 (m, 1H, H-2), 5.19 (t, *J* = 4.4 Hz, 1H, H[']-2), 4.96 (d, 1H, *J* = 3.6 Hz, H[']-1), 4.88-4.74 (m, 4H, CH₂Ph, CH₂PMP), 4.65-4.42 (m, 12H, H[']-1, H-4, H-5, H[']-5, 4xCH₂Ph), 4.62-4.35 (m, 4H, CH₂Ph, CH₂PMP), 4.60-4.59 (m, 1H, H-5 β), 4.33 (t, *J* = 2.8 Hz, 1H, H-3 β), 4.30-4.21 (m, 4H, H-3, H[']-3, CH₂Ph), 4.04 (t, *J* = 4.4 Hz, 1H, H[']-4), 3.93-3.37 (m, 10H, H[']-3, H[']-4, H[']-5, H[']-6, H[']-3, H[']-4, H[']-5, H[']-6), 3.81 (s, 3H, PhOCH₃), 3.48 (s, 3H, COOCH₃), 3.43 (s, 3H, COOCH₃), 3.27-3.21 (m, 2H, H[']-2, H[']-2). ¹³C NMR (100 MHz; CDCl₃) δ 169.2, 168.6, 166.4, 165.2, 159.3, 138.0, 137.9, 137.9, 137.4, 137.0, 134.8, 133.4, 133.3, 131.5, 130.4, 130.1, 129.9, 129.7, 129.5, 129.4, 129.0, 128.7, 128.6, 128.5, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 128.1, 128.0, 127.9, 127.8, 127.8, 127.7, 127.7, 127.2, 113.7, 99.7, 99.0, 98.3, 86.0, 79.9, 78.7, 77.6, 77.5, 77.4, 77.2, 76.8, 75.7, 75.5, 75.4, 75.1, 74.9,

74.6, 74.4, 73.9, 73.6, 72.9, 72.4, 71.7, 71.4, 70.2, 70.0, 69.7, 67.7, 67.6, 63.8, 63.5, 55.4, 52.1, 51.9. HRMS (FT MS): m/z : calcd for $C_{96}H_{100}N_7O_{23}S$ [$M+NH_4$] $^+$: 1750.6586; found: 1750.6578.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(phenyl 2-*O*-benzoyl-3-*O*-benzyl-1-thio- β -L-idopyranoside)] uronate (β -7)

To β -6 (952 mg, 0.55 mmol) was added CH_3CN (33 mL) and water (3 mL) followed by ammonium cerium(IV) nitrate (99.99+%, 751 mg, 1.37 mmol). The orange solution was stirred for 90 min and was then extracted with DCM (200 mL, 50 mL) and water (200 mL), dried ($MgSO_4$), filtered and evaporated. The crude product was purified using flash column chromatography (EtOAc/hexane 1:3) yielding β -7 (596 mg, 67%) as a white foam and recovered starting material (259 mg, 27%). R_f 0.20 (EtOAc/Hexane 1:2). $[\alpha]_D^{20} = +10.4$ ($c = 0.12$, CH_2Cl_2). 1H NMR (400 MHz; $CDCl_3$) δ 8.21-8.18 (m, 2H, Bz), 7.97-7.94 (m, 2H, Bz), 7.53-7.50 (m, 2H, Bz), 7.42-7.05 (m, 35H, Ph), 5.55 (d, $J = 4.8$ Hz, 1H, $H''-1$), 5.23 (d, $J = 1.6$ Hz, 1H, H-1), 5.21-5.18 (m, 2H, H-2, $H''-2$), 4.95 (d, 1H, $J = 3.6$ Hz, $H'-1$), 4.86-4.72 (m, 4H, $2 \times CH_2Ph$), 4.59-4.46 (m, 9H, $H'''-1$, $H'''-5$, H-5, $3 \times CH_2Ph$), 4.26-4.20 (m, 2H, H-3, $H''-3$), 4.26-3.34 (m, 13H, H-4, $H''-4$, $H'-3$, $H'-4$, $H'-5$, $H'-6$, $H'''-3$, $H'''-5$, $H'''-6$, CH_2Ph), 3.43 (s, 3H, $COOCH_3$), 3.41 (s, 3H, $COOCH_3$), 3.23-3.17 (m, 3H, $H'-2$, $H'''-2$, $H'''-4$). ^{13}C NMR (100 MHz; $CDCl_3$) δ 169.3, 168.6, 166.4, 165.2, 138.0, 138.0, 137.9, 137.6, 137.4, 136.9, 134.7, 133.4, 133.3, 131.4, 131.1, 130.1, 130.0, 129.9, 129.9, 129.6, 129.4, 129.0, 128.9, 128.8, 128.6, 128.6, 128.6, 128.6, 128.5, 128.5, 128.4, 128.3, 128.3, 128.3, 128.1, 128.0, 128.0, 127.9, 127.8, 127.8, 127.7, 127.7, 127.6, 127.2, 99.3, 99.0, 98.1, 85.9, 79.2, 78.5, 77.4, 77.1, 77.1, 76.8, 75.8, 75.7, 75.6, 75.0, 75.0, 74.8, 74.5, 74.1, 73.8, 73.6, 72.8, 72.5, 72.3, 71.3, 70.8, 70.5, 70.2, 70.0, 69.4, 67.4, 63.6, 62.7, 52.1, 51.9. HRMS (FT MS): m/z : calcd for $C_{88}H_{92}N_7O_{22}S$ [$M+NH_4$] $^+$: 1630.6011; found: 1630.6009.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy-4-*O*-trichloroacetyl- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(phenyl 2-*O*-benzoyl-3-*O*-benzyl-1-thio-L-idopyranoside)] uronate (α -8 and β -8)

The tetrasaccharide α -7 (596 mg, 0.37 mmol) was dissolved in dry DCM (5 mL), cooled to 0 °C in an icebath, then dry pyridine (0.15 mL, 1.85 mmol) and trichloroacetyl chloride (0.10 mL, 0.93 mmol) was added. The solution was stirred for 1 h. The solution was extracted with DCM (50 mL) and water (50 mL), dried ($MgSO_4$), filtered and evaporated. The crude product was purified using flash column chromatography (EtOAc/hexane 1:3). This yielded α -8 (610 mg, 94%) as a white foam. The same procedure was used for converting β -7 into β -8.

α -8: R_f 0.23 (EtOAc/Hexane 1:3). $[\alpha]_D^{20} = -18.3$ ($c = 0.84$, CH_2Cl_2). 1H NMR (500 MHz; $CDCl_3$) δ 8.13-8.11 (m, 2H, Bz), 8.02-8.00 (m, 2H, Bz), 7.53-7.12 (m, 35H, Ph), 5.78-5.77 (m, 1H, H-1), 5.55 (d, $J = 3.8$ Hz, 1H, $H''-1$), 5.37-5.38 (m, 1H, H-2), 5.31-5.27 (m, 2H, $H'''-4$, H-5), 5.17 (t, 1H, $J = 4.4$ Hz, $H''-2$), 4.97-4.75 (m, 4H, $2 \times CH_2Ph$), 4.90 (d, $J = 3.6$ Hz, 1H, $H'-1$ or $H'''-1$), 4.69 (d, $J = 3.6$ Hz, 1H, $H'-1$ or $H'''-1$), 4.64 (d, $J = 4.1$ Hz, 1H, $H''-5$), 4.53-4.25 (m, 6H, $3 \times CH_2Ph$), 4.21 (t, $J = 5.2$ Hz, 1H, $H''-3$), 4.19-4.18 (m, 1H, H-3), 4.02-4.01 (m, 1H, H-4), 3.99-3.41 (m, 13H, $H''-4$, $H'-3$, $H'-4$, $H'-5$, $H'-6$, $H'''-3$, $H'''-4$, $H'''-5$, $H'''-6$, CH_2Ph), 3.55 (s, 3H, $COOCH_3$), 3.45 (s, 3H, $COOCH_3$), 3.37 (dd, 1H, $J = 10.1$ Hz, 3.4 Hz $H'-2$ or $H'''-2$), 3.27 (dd, 1H, $J = 10.3$ Hz, 3.6 Hz $H'-2$ or $H'''-2$). ^{13}C NMR (100 MHz; $CDCl_3$) δ 169.3, 169.2, 165.6, 165.3, 160.2, 138.0, 137.8, 137.3, 137.1, 136.8, 135.4, 133.6, 133.5, 131.3, 130.0, 129.9, 129.6, 129.5, 129.1, 128.8, 128.6,

128.6, 128.5, 128.5, 128.4, 128.3, 128.2, 128.2, 128.1, 128.0, 128.0, 127.9, 127.7, 127.6, 127.4, 127.2, 99.8, 99.2, 98.5, 89.7, 86.8, 78.6, 77.6, 77.5, 77.2, 76.8, 76.5, 76.0, 75.9, 74.9, 74.8, 74.8, 74.1, 73.9, 73.8, 73.6, 72.8, 71.9, 71.4, 70.5, 69.6, 69.3, 69.2, 68.5, 67.7, 67.2, 63.8, 63.1, 60.5, 52.1, 51.9. ES MS: m/z: calcd for C₉₀H₈₇Cl₃N₆O₂₃SNa [M+Na]⁺: 1779.5; found: 1779.5. Elemental analysis calcd (%) for C₉₀H₈₇Cl₃N₆O₂₃S: C 61.45, H 4.98, N 4.78; found C 61.71, H 5.05, N 4.69. **β-8**: *R_f* 0.18 (EtOAc/Hexane 1:3). [α]_D²⁰ = +32.9 (*c* = 0.61, CH₂Cl₂). ¹H NMR (400 MHz; CDCl₃) δ 8.24-8.21 (m, 2H, Bz), 8.03-8.00 (m, 2H, Bz), 7.57-7.54 (m, 2H, Bz), 7.45-7.07 (m, 35H, Ph), 5.56 (d, *J* = 3.8 Hz, 1H, H^{''-1}), 5.32 (t, *J* = 9.4 Hz, 1H, H^{'''-4}), 5.27 (d, *J* = 1.9 Hz, 1H, H-1), 5.24-5.23 (m, 1H, H-2), 5.18 (t, *J* = 4.4 Hz, H^{''-2}), 4.90-4.75 (m, 5H, H^{'-1}, 2xCH₂Ph), 4.64 (d, *J* = 4.4 Hz, H-5 or H-5^{''}), 4.55-4.45 (m, 6H, H^{'''-1}, H^{''-5} or H-5, 2xCH₂Ph), 4.33-4.18 (m, 4H, H-3, H^{''-3}, CH₂Ph), 4.01-3.90 (m, 4H, H-4, H^{''-4}, CH₂Ph), 3.74-3.37 (m, 10H, H^{'-2}, H^{'-3}, H^{'-4}, H^{'-5}, H^{'-6}, H^{'''-3}, H^{'''-5}, H^{'''-6}), 3.50 (s, 3H, COOCH₃), 3.39 (s, 3H, COOCH₃), 3.25 (dd, *J* = 10.3 Hz, *J* = 3.6 Hz, 1H, H^{'''-2}). ¹³C NMR (100 MHz; CDCl₃) δ 169.2, 168.6, 166.4, 165.3, 160.2, 138.0, 137.8, 137.3, 137.3, 137.0, 136.8, 134.7, 133.4, 133.4, 131.5, 130.1, 129.9, 129.7, 129.5, 129.0, 128.8, 128.7, 128.7, 128.7, 128.6, 128.6, 128.5, 128.5, 128.4, 128.3, 128.3, 128.2, 128.1, 128.0, 128.0, 127.9, 127.9, 127.7, 127.7, 127.6, 127.5, 127.4, 127.1, 99.3, 99.2, 98.3, 89.7, 85.9, 78.5, 77.5, 77.4, 76.1, 75.7, 75.3, 74.9, 74.8, 74.6, 74.2, 73.8, 73.8, 73.6, 72.9, 72.5, 71.3, 70.2, 70.0, 69.4, 69.1, 67.8, 67.2, 63.9, 63.1, 52.1, 51.9.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)- 2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-((*S*)-2,3-bis(benzyloxy)propyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranoside)] uronate (9**)**

Tetrasaccharide **α-7** (145 mg, 0.082 mmol) was dissolved in dry DCM (2 mL) under N₂. (*S*)-2,3-Bis(benzyloxy)propanol (44 mg, 0.163 mmol) was added and the clear solution cooled to 0 °C. Freshly activated 4Å powdered molecular sieves (137 mg) were added, after 10 min. NIS (50 mg, 2.22 mmol) and after another 10 min. AgOTf (catalytic amount). The suspension changed colour from pale yellow to deep red and was stirred for a further 20 min. The reaction was quenched into NaHCO₃ (250 mg) and Na₂S₂O₃ (250 mg) in water (5 mL). After stirring for 10 mins and the reaction colour changing to pale yellow the suspension was filtered through a short pad of Celite® washing with water and DCM. The layers were separated and the aqueous extracted with DCM (10 mL). The organics were combined, dried (MgSO₄) and solvent removed *in vacuo* to reveal the crude product as a pale yellow oil. This was purified by flash column chromatography (EtOAc/Hexane 3:5) to give **9** (130 mg, 82%) as a white foam. *R_f* 0.22 (EtOAc/Hexane 1:2). [α]_D²⁰ = +0.7 (*c* = 0.37, CH₂Cl₂). ¹H NMR (500 MHz; CDCl₃) δ 8.11-8.09 (m, 2H, Bz), 8.00-7.98 (m, 2H, Bz), 7.54-7.14 (m, 40H, Ph), 5.55 (d, *J* = 4.7 Hz, H^{''-1}), 5.21 (t, 1H, *J* = 5.4 Hz, H^{''-2}), 5.14-5.13 (m, 1H, H-1), 5.12-5.11 (m, H-2), 4.94 (d, *J* = 3.6 Hz, 1H, H^{'-1}), 4.87-4.41 (m, 19H, H^{'''-1}, H-5, H^{''-5}, 8xCH₂Ph), 4.21 (t, *J* = 6.0 Hz, 1H, H^{''-3}), 4.10-4.09 (m, 1H, H-3), 4.04-4.01 (m, 1H, H^{''-4}), 3.99-3.98 (m, 1H, H-4), 3.96-3.42 (m, 15H, H^{'-3}, H^{'-4}, H^{'-5}, H^{'-6}, H^{'''-3}, H^{'''-4}, H^{'''-5}, H^{'''-6}, CH₂CHOBNCH₂OBN), 3.46 (s, 3H, COOCH₃), 3.44 (s, 3H, COOCH₃), 3.23 (dd, 1H, *J* = 10.2 Hz, 3.6 Hz H^{'-2} or H^{'''-2}), 3.19 (dd, 1H, *J* = 10.3 Hz, 3.6 Hz H^{'-2} or H^{'''-2}). ¹³C NMR (100 MHz; CDCl₃) δ 169.6, 169.5, 169.4, 165.8, 165.4, 138.6, 138.3, 138.2, 138.1, 138.0, 137.7, 137.7, 137.5, 133.6, 130.1, 129.8, 129.6, 128.9, 128.7, 128.7, 128.6, 128.6, 128.5, 128.5, 128.4, 128.4, 128.3, 128.2, 128.1, 128.1, 128.0, 127.9, 127.8, 127.8, 127.7, 127.4, 99.4, 99.3, 98.4, 79.4, 78.6, 77.5, 76.8, 76.0, 75.9, 75.8, 75.1, 74.9, 74.5, 74.2, 73.9, 73.8, 73.5, 72.8, 72.5, 71.5, 71.2, 70.7, 70.5, 69.8, 69.6, 68.6, 67.8, 67.6, 67.5, 63.7, 62.9, 60.6, 52.1, 52.0. HRMS (FT MS): m/z: calcd for

C₉₉H₁₀₆N₇O₂₅ [*M*+NH₄⁺]: 1792.7233; found: 1792.7212. Elemental analysis calcd (%) for C₉₉H₁₀₂N₆O₂₅: C 66.96, H 5.79, N 4.73; found C 66.71, H 5.69, N 4.68.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy-4-*O*-*p*-methoxybenzyl- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-((*S*)-2,3-bis(benzyloxy)propyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranoside)] uronate (10)

Tetrasaccharide **9** (55 mg, 0.031 mmol) and donor β -**6** (65 mg, 0.037 mmol) was dissolved in dry DCM (1 mL) under N₂. Freshly activated 4Å powdered molecular sieves (100 mg) were added and the solution cooled to 0 °C in an ice-bath. After 10 min. NIS (13 mg, 0.046 mmol) was added, and after another 10 min. AgOTf (catalytic amount) was added. The suspension changed colour from pale yellow to deep red and was stirred for a further 35 min. The reaction was quenched by pouring into a separating funnel containing a mixture of DCM (20 mL), saturated aqueous NaHCO₃ (20 mL) and Na₂S₂O₃ (2 mL, 10% aqueous). After shaking until the iodine colour was removed the suspension was filtered through a short pad of Celite® washing with water and DCM. The layers were separated and the aqueous extracted with DCM (10 mL). The organic layers were combined, dried (MgSO₄) and solvent removed *in vacuo*. The crude product was purified by silica gel flash column chromatography (EtOAc/hexane 3:7) to give **10** (85 mg, 81%) as a white foam. *R*_f 0.24 (EtOAc/Hexane 1:2). [α]_D²⁰ = +9.1 (*c* = 0.78, CH₂Cl₂). ¹H NMR (400 MHz; CDCl₃) δ 8.10-8.08 (m, 2H, Bz), 7.95-7.91 (m, 6H, Bz), 7.55-7.00 (m, 84H, Ph, PMB), 6.83 (d, 1H, *J* = 8.8 Hz, PMB), 5.55-5.51 (m, 3H, H^{''-1}, H^{'''-1}, H^{''''-1}), 5.19-5.15 (m, 3H, H^{''-2}, H^{'''-2}, H^{''''-2}), 5.11-5.10 (m, 2H, H-1, H-2), 4.97 (d, 1H, *J* = 3.5 Hz, H^{''-1} or H^{'''-1} or H^{''''-1}), 4.93 (d, 1H, *J* = 3.5 Hz, H^{''-1} or H^{'''-1} or H^{''''-1}), 4.89 (d, 1H, *J* = 3.8 Hz, H^{''-1} or H^{'''-1} or H^{''''-1}), 4.88-4.29 (m, 33H, H^{''''''-1}, H-5, H^{''-5}, H^{'''-5}, H^{''''-5}, 14xCH₂Ph), 4.23-4.07 (m, 4H, H-3, H^{''-3}, H^{'''-3}, H^{''''-3}), 4.05-3.19 (m, 32H, H^{''-2}, H^{'''-2}, H^{''''-2}, H^{''''''-2}, H^{''-3}, H^{'''-3}, H^{''''-3}, H^{''''''-3}, H-4, H^{''-4}, H^{'''-4}, H^{''''-4}, H^{''''''-4}, H^{''-4}, H^{'''-4}, H^{''''-4}, H^{''''''-4}, H^{''-5}, H^{'''-5}, H^{''''-5}, H^{''''''-5}, H^{''-6}, H^{'''-6}, H^{''''-6}, H^{''''''-6}, CH₂CHOBnCH₂OBN), 3.81 (s, 3H, PhOCH₃), 3.46 (s, 3H, COOCH₃), 3.41 (s, 3H, COOCH₃), 3.30 (s, 3H, COOCH₃), 3.23 (s, 3H, COOCH₃). HRMS (FTMS) : *m/z*: calcd for C₁₈₉H₂₀₀N₁₄O₄₈ [*M*+2NH₄]²⁺: 1717.6848; found: 1717.6827.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy-4-*O*-trichloroacetyl- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranosyl uronate)-(1 \rightarrow 4)-2-azido-3,6-di-*O*-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-((*S*)-2,3-bis(benzyloxy)propyl 2-*O*-benzoyl-3-*O*-benzyl- α -L-idopyranoside)] uronate (11)

Tetrasaccharide **9** (68 mg, 0.038 mmol) and donor α -**8** (81 mg, 0.046 mmol) was dissolved in dry DCM (1.5 mL) under N₂. Freshly activated 4Å powdered molecular sieves (111 mg) were added and the solution cooled to 0 °C in an icebath. After 10 min. NIS (17 mg, 0.076 mmol) was added, and after another 10 min. AgOTf (catalytic amount) was added. The suspension changed colour from pale yellow to deep red and was stirred for a further 45 min. The reaction was quenched into a

separating funnel containing a mixture of DCM (30 mL), saturated aqueous NaHCO₃ (25 mL) and Na₂S₂O₃ (2 mL, 10% aqueous). After shaking until the iodine colour was removed the suspension was filtered through a short pad of Celite® washing with water and DCM. The layers were separated and the aqueous extracted with DCM (10 mL). The organic layers were combined, dried (MgSO₄) and solvent removed *in vacuo*. The crude was purified by silica gel flash column chromatography (toluene/acetone 20:1) to give **11** (120 mg, 92%) of as a white foam. *R_f* 0.23 (EtOAc/Hexane 1:2). [α]_D²⁰ = +12.9 (*c* = 0.35, CH₂Cl₂). ¹H NMR (500 MHz; CDCl₃) δ 8.10-8.08 (m, 2H, Bz), 8.02-8.00 (m, 2H, Bz), 7.96-7.93 (m, 4H, Bz), 7.53-7.03 (m, 82H, Ph), 5.57-5.52 (m, 3H, H^{''}-1, H^{''''}-1, H^{''''''}-1), 5.30 (t, 1H, *J* = 9.8 Hz, H^{''''''''}-4), 5.19-5.16 (m, 3H, H^{''}-2, H^{''''}-2, H^{''''''}-2), 5.12-5.11 (m, 2H, H-1, H-2), 4.94-4.90 (m, 3H, H[']-1, H^{''}-1, H^{''''}-1), 4.87-4.24 (m, 33H, H^{''''''''}-1, H-5, H^{''}-5, H^{''''}-5, H^{''''''}-5, 14xCH₂Ph), 4.24-4.09 (m, 4H, H-3, H^{''}-3, H^{''''}-3, H^{''''''}-3), 4.04-3.20 (m, 32H, H[']-2, H^{''}-2, H^{''''}-2, H^{''''''}-2, H^{''''''''}-2, H[']-3, H^{''}-3, H-3^{''''}, H^{''''''}-3, H-4, H^{''}-4, H^{''''}-4, H^{''''''}-4, H[']-4, H^{''}-4, H^{''''}-4, H^{''''''}-4, H[']-5, H^{''}-5, H^{''''}-5, H^{''''''}-5, H[']-6, H^{''}-6, H^{''''}-6, H^{''''''}-6, CH₂CHOBnCH₂OBn), 3.47 (s, 3H, COOCH₃), 3.44 (s, 3H, COOCH₃), 3.33 (s, 3H, COOCH₃), 3.27 (s, 3H, COOCH₃). ¹³C NMR (100 MHz; CDCl₃) δ 169.4, 169.3, 169.2, 165.6, 165.2, 165.1, 160.2, 138.5, 138.1, 138.0, 137.9, 137.8, 137.7, 137.7, 137.5, 137.4, 137.4, 137.2, 137.2, 136.7, 133.6, 133.5, 129.9, 129.9, 129.8, 129.8, 129.6, 129.4, 129.3, 129.2, 128.8, 128.7, 128.7, 128.5, 128.4, 128.4, 128.4, 128.3, 128.3, 128.2, 128.1, 128.1, 128.0, 128.0, 127.9, 127.9, 127.8, 127.7, 127.7, 127.6, 127.5, 127.3, 127.3, 127.1, 99.3, 99.3, 99.2, 99.1, 99.1, 98.3, 98.0, 98.0, 89.7, 78.3, 78.2, 78.1, 77.5, 77.4, 77.2, 77.0, 77.0, 76.8, 76.7, 76.7, 76.7, 76.6, 76.6, 76.5, 76.5, 75.9, 75.8, 75.7, 75.7, 75.6, 75.4, 75.4, 75.3, 74.9, 74.8, 74.8, 74.6, 74.4, 74.2, 74.1, 74.1, 73.8, 73.7, 73.6, 73.6, 73.5, 73.3, 73.2, 72.6, 72.4, 71.3, 71.2, 71.1, 70.5, 70.5, 70.4, 69.6, 69.2, 68.4, 68.3, 67.5, 67.3, 67.2, 67.2, 63.4, 63.4, 63.2, 63.2, 63.1, 60.4, 53.5, 51.9, 51.7, 51.6. MS (MALDI-TOF): *m/z*: calcd for C₁₈₃H₁₈₃Cl₃N₁₂NaO₄₈ [M+Na]⁺: 3444.1; found: 3444.1. Elemental analysis calcd (%) for C₁₈₃H₁₈₃Cl₃N₁₂O₄₈: C 64.18, H 5.39, N 4.91; found C 64.10, H 5.32, N 4.89.

Methyl [2-azido-3,6-di-*O*-benzyl-2-deoxy-α-D-glucopyranosyl-(1→4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl-α-L-idopyranosyl uronate)-(1→4)-2-azido-3,6-di-*O*-benzyl-2-deoxy-α-D-glucopyranosyl-(1→4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl-α-L-idopyranosyl uronate)-(1→4)-2-azido-3,6-di-*O*-benzyl-2-deoxy-α-D-glucopyranosyl-(1→4)-(methyl 2-*O*-benzoyl-3-*O*-benzyl-α-L-idopyranosyl uronate)-(1→4)-2-azido-3,6-di-*O*-benzyl-2-deoxy-α-D-glucopyranosyl-(1→4)-((*S*)-2,3-bis(benzyloxy)propyl 2-*O*-benzoyl-3-*O*-benzyl-α-L-idopyranoside)] uronate (12**)**

The octasaccharide **11** (346 mg, 0.10 mmol) was dissolved in a mixture of MeOH/pyridine (5 mL/2 mL) and heated to 50 °C for 4 h. The solvents were evaporated and co-evaporated with toluene (2x20 mL). The crude product was purified using flash column chromatography (EtOAc/hexane 1:2 and 3:5). This yielded **12** (300 mg, 91%) as a white foam, along with recovered starting material (20 mg, 5%). *R_f* 0.10 (EtOAc/Hexane 1:2). [α]_D²⁰ = +9.8 (*c* = 0.32, CH₂Cl₂). ¹H NMR (500 MHz; CDCl₃) δ 8.10-8.08 (m, 2H, Bz), 8.00-7.93 (m, 6H, Bz), 7.54-7.03 (m, 82H, Ph), 5.57-5.52 (m, 3H, H^{''}-1, H^{''''}-1, H^{''''''}-1), 5.22-5.15 (m, 3H, H^{''}-2, H^{''''}-2, H^{''''''}-2), 5.12-5.11 (m, 2H, H-1, H-2), 4.97 (d, 1H, *J* = 3.5 Hz, H[']-1 or H^{''}-1 or H^{''''}-1), 4.93 (d, 1H, *J* = 3.6 Hz, H[']-1 or H^{''}-1 or H^{''''}-1), 4.90 (d, 1H, *J* = 3.6 Hz, H[']-1 or H^{''}-1 or H^{''''}-1), 4.86-4.34 (m, 33H, H^{''''''''}-1, H-5, H^{''}-5, H^{''''}-5, H^{''''''}-5, 14xCH₂Ph), 4.24-4.08 (m, 4H, H-3, H^{''}-3, H^{''''}-3, H^{''''''}-3), 4.05-3.20 (m, 32H, H[']-2, H^{''}-2, H^{''''}-2, H^{''''''}-2, H[']-3, H^{''}-3, H-3^{''''}, H^{''''''}-3, H-4, H^{''}-4, H^{''''}-4, H^{''''''}-4, H[']-4, H^{''}-4, H^{''''}-4, H^{''''''}-4, H[']-5, H^{''}-5, H^{''''}-5, H^{''''''}-5, H[']-6, H^{''}-6, H^{''''}-6, H^{''''''}-6, CH₂CHOBnCH₂OBn), 3.48 (s, 3H, COOCH₃), 3.44 (s, 3H, COOCH₃), 3.32 (s, 3H, COOCH₃), 3.27

