

Supporting Information:

CH-01 is a hypoxia-activated prodrug that sensitizes cells to hypoxia/reoxygenation through inhibition of Chk1 and Aurora A

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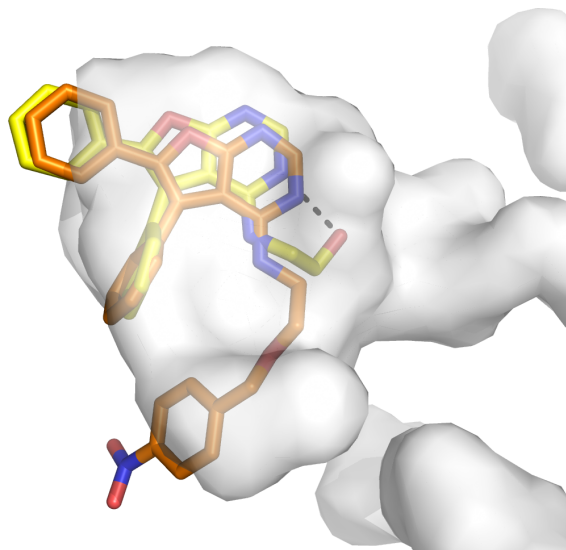
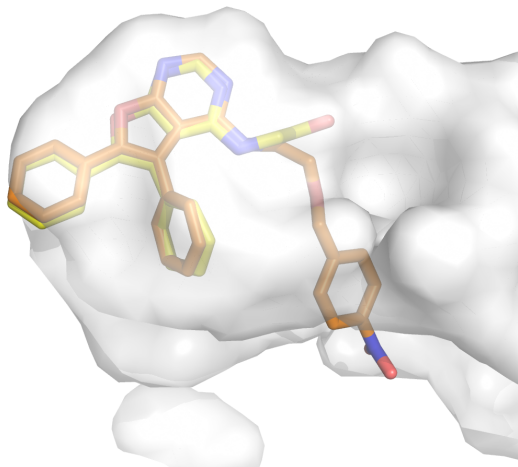


Figure S1. A representative image of compound **1** (carbon = orange) docked to the X-ray crystal structure of compound **6** (carbon = yellow) bound to Chk1 (PDB ID, 2BRB). The surface of the binding site is shown. Docking indicated that the binding of the 4-nitrobenzyl derivative **1** to Chk1 was less favorable than binding of **6**.

A



B

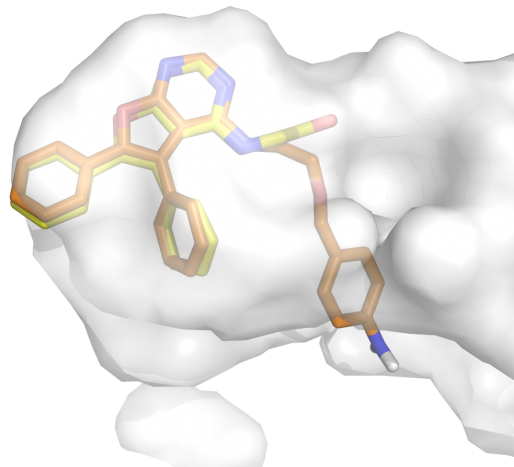


Figure S2. (A). A representative image of compound **1** (carbon = orange) docked to the X-ray crystal structure of compound **6** (carbon = yellow) bound to Aurora A (PDB ID, 3K5U). Docking indicated that the 4-nitrobenzyl group of **1** can potentially be accommodated in the ATP-binding site. (B). A representative image of amine **5** (carbon = orange) docked to the X-ray crystal structure of compound **6** (carbon = yellow) bound to Aurora A (PDB ID, 3K5U). Docking indicated that the 4-aminobenzyl group of **5** can potentially be accommodated in the ATP-binding site. The surface of the binding site is shown.

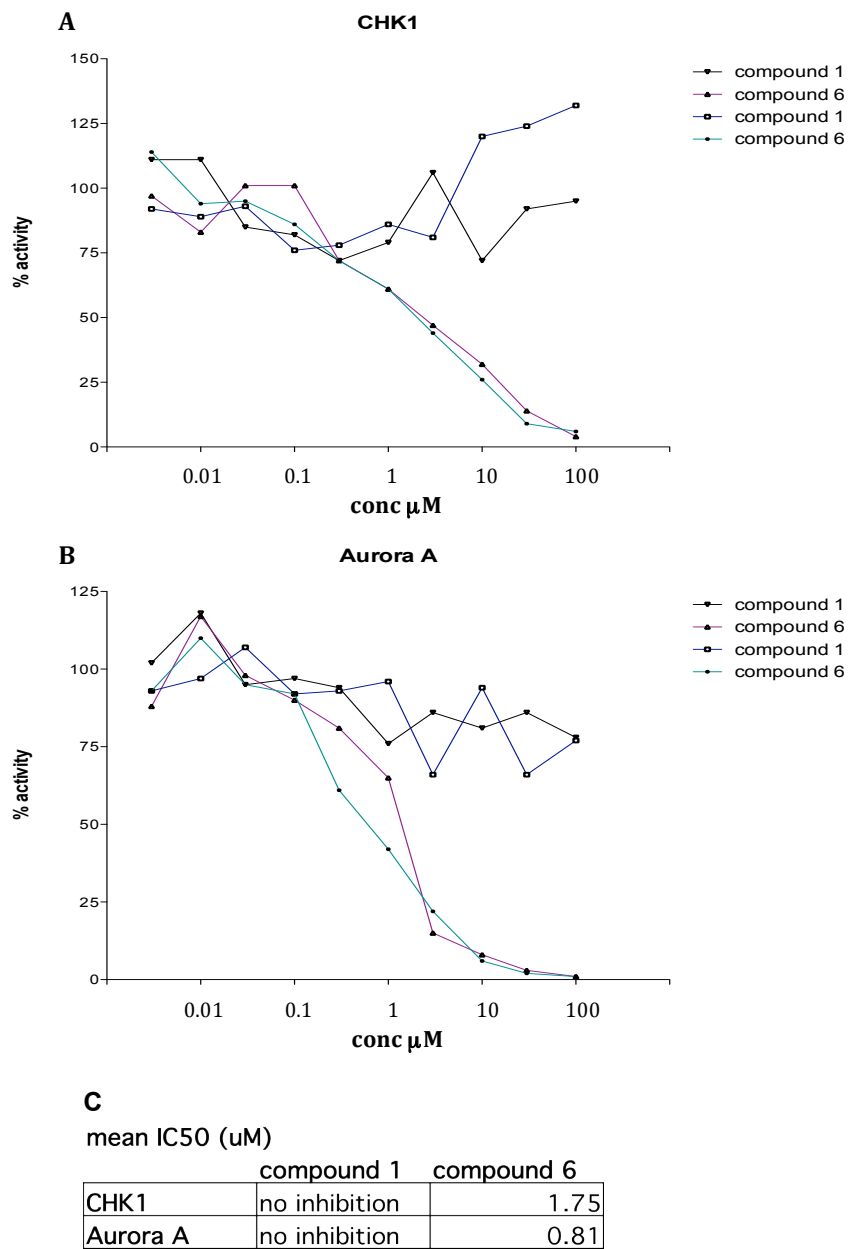
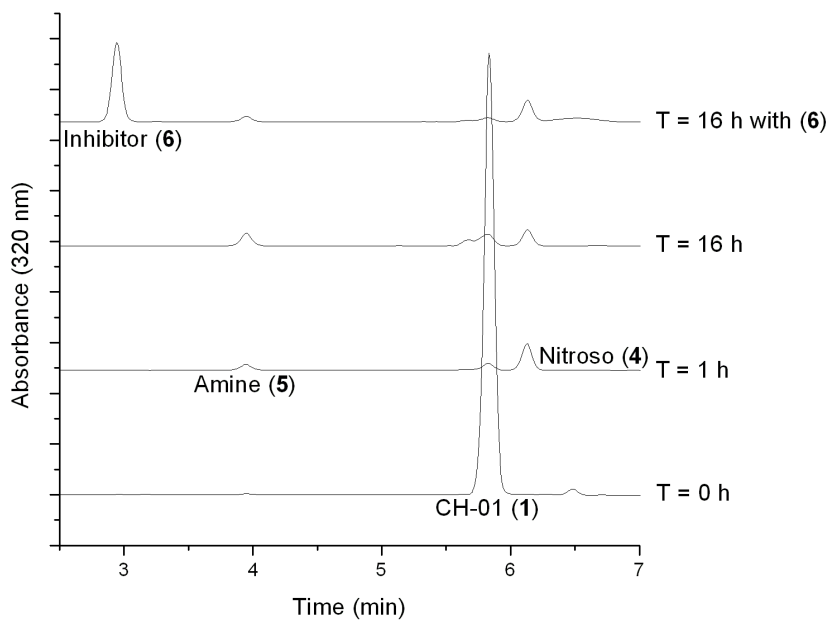


Figure S3. The IC₅₀ values of compounds **1** and **6** were obtained for Chk1 and Aurora kinase A using a radioactive (³³P-ATP) filter-binding assay. (A) Compound **1** shows no inhibition of Chk1 whereas compound **6** has an IC₅₀ value of 1.75 μM. (B) Compound **1** shows no inhibition of Aurora kinase A whereas compound **6** has an IC₅₀ value of 0.81 μM. This analysis was carried out by The International Centre for Kinase profiling, University of Dundee (<http://www.kinase-screen.mrc.ac.uk/>). (C) Mean IC₅₀ values.

A



B

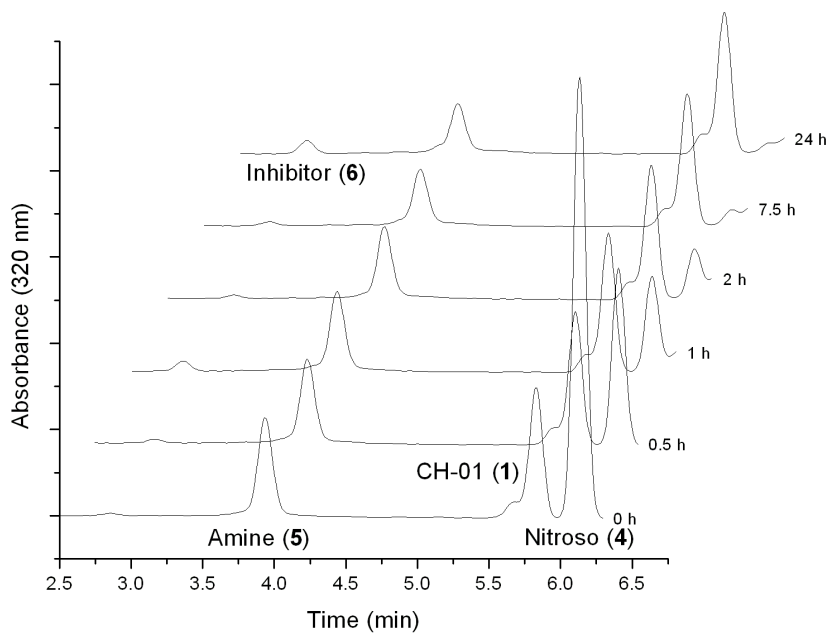


Figure S4. (A) Reduction of **1** using zinc powder and 10% ammonium chloride in DMF was monitored at 0, 1 and 16 h. The new peaks were identified as the corresponding amine and nitroso reduction products. An injection that contained the active inhibitor **6** as an internal control showed that none of the peaks correlated to **6**, confirming that release of **6** was not observed under these conditions. (B) The T = 1 h aliquot from the zinc reduction was injected into potassium phosphate buffer pH 7.4 and incubated at 37 °C. HPLC analysis of the formed precipitate reveals loss of the nitroso **4** and amine **5**.

Docking with AutoDock Vina (version 1.1.2):

Preparation of the receptors:

The receptors (PDB IDs: 2BRB and 3K5U) were initially prepared by adding polar hydrogens at pH 7.4 using Protonate3D in Molecular Operating Environment (MOE) 2011.10. Gasteiger charges were then assigned using AutoDock's graphical user interface AutoDockTools (ADT; version 1.5.4). Following the addition of Gasteiger charges, ADT was implemented to build a "united atom model" of the receptor by merging non-polar hydrogens and adding their partial charges to their parent carbon atoms.

Preparation of the ligands:

The ligands were prepared by adding polar hydrogens at pH 7.4 using Protonate3D in MOE. MOE then assigned Gasteiger charges, before performing an energy minimisation using an MMFF94x forcefield with a 0.05 gradient. AutoDock's graphical user interface AutoDockTools (ADT; version 1.5.4) was subsequently used to build a "united atom model" of the ligands, by merging non-polar hydrogens and adding their partial charges to their parent carbon atoms.

Individual docking with AutoDock Vina (version 1.1.2):

The search space for docking into the receptor 2BRB was defined as a cubic box with dimensions 18.75 Å × 18.75 Å × 18.75 Å and center x = 17.238, y = -2.877, z = 10.364. The exhaustiveness and number of modes were both set to 20 and all other parameters were kept at their default values.

The search space for docking into the receptor 3K5U was defined as a cubic box with dimensions 18.75 Å × 18.75 Å × 18.75 Å and center x = -5.496, y = -31.095, z = 6.03. The exhaustiveness and number of modes were both set to 20 and all other parameters were kept at their default values.

General Experimental

¹H NMR spectra were recorded on Bruker DPX400 or DQX400 (400 MHz) or Bruker AVII 500 (500 MHz) using deuteriochloroform (unless indicated otherwise) as a reference for internal deuterium lock. The chemical shift data for each signal are given as δH in units of parts per million (ppm) relative to tetramethylsilane (TMS) where δH (TMS) = 0.00 ppm. The multiplicity of each signal is indicated by: s (singlet); br s (broad singlet); d (doublet); t (triplet); q (quartet); dd (doublet of doublets); combinations thereof; or m (multiplet). The number of protons (n) for a given resonance signal is indicated by nH. Coupling constants (*J*) are quoted in Hz and are recorded to the nearest 0.1 Hz. Identical proton coupling constants (*J*) are averaged in each spectrum and reported to the nearest 0.1 Hz. The coupling constants are determined by analysis using Bruker TopSpin software.

¹³C NMR spectra were recorded on Bruker DQX400 (100 MHz) or Bruker AVII 500 (126 MHz) spectrometers with broadband proton decoupling and internal deuterium lock. The chemical shift data for each signal are given as δC in units of parts per million (ppm) relative to tetramethylsilane (TMS) where δC (TMS) = 0.00 ppm.

Mass spectra were acquired on a VG platform spectrometer. Electrospray ionization spectra were obtained on Micromass LCT Premier and Bruker MicroTOF spectrometers, operating in positive or negative mode, from solutions of methanol. m/z values are reported in Daltons and followed by their percentage abundance in parentheses.

Melting points were determined using a Kofler hot stage microscope and are uncorrected.

Microanalyses were obtained at the Elemental Analysis Service, London Metropolitan University, London.

Infrared Spectra were obtained from neat samples, either as solids or liquids, using a diamond ATR module. The spectra were recorded on a Bruker Tensor 27 spectrometer. Absorption maxima are reported in wavenumbers (cm^{-1}).

Analytical thin layer chromatography (TLC) was carried out on Merck silica gel 60 F₂₅₄ aluminum-supported thin layer chromatography sheets. Visualization was by absorption of UV light (λ_{max} 254 nm), or thermal development after dipping in one of: **a** ethanolic solution of phosphomolybdic acid (PMA), ninhydrin or 4-anisaldehyde; **b** aqueous solution of cerium-ammonium-molybdate and sulfuric acid or potassium permanganate, potassium carbonate and sodium hydroxide.

Flash Column chromatography was carried out on VWR Prolabo silica gel 60 (240-400 mesh), eluting with solvents as supplied, under a positive pressure of compressed air (unless otherwise stated).

Anhydrous solvents Anhydrous CH_2Cl_2 , Et_2O , toluene, and THF were obtained using a Dalek drying column, where solvent was dried by passage

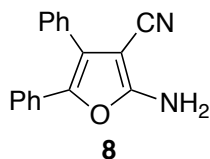
through filter columns and dispensed under an atmosphere of N₂ gas, before being stored over activated 3 Å molecular sieves under a N₂ atmosphere. Dry *N,N*-dimethylformamide was purchased from SigmaAldrich UK in SureSeal™ bottles and used without further purification as was dry 1,4-dioxane which was purchased from Fluka UK. All other solvents were used as supplied (analytical or HPLC grade) without purification.

Chemicals were purchased from Acros UK, Sigma Aldrich UK, Alfa Aesar UK, Fisher UK, Fluka UK, Fluorochem, Merck or TCI-Europe. All solvents and reagents were purified, when necessary, by standard techniques. Where appropriate and if not stated otherwise, all non-aqueous reactions were performed in a flame-dried flask under an inert atmosphere of nitrogen or argon.

In vacuo refers to the use of a rotary evaporator attached to a diaphragm pump. Brine refers to a saturated aqueous solution of sodium chloride. Petroleum ether refers to the fraction boiling between 30–40 °C unless otherwise stated.

Experimental Procedures

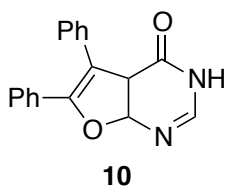
2-Amino-4,5-diphenylfuran-3-carbonitrile (8)



Benzoin (**7**, 5.00 g, 23.5 mmol, 1.0 eq), malononitrile (2.02 g, 30.6 mmol, 1.3 eq) and diethylamine (0.95 mL, 9.19 mmol, 0.4 eq) were added to dry dioxane (25 mL) stirring over 4 Å molecular sieves. The reaction vessel was covered with foil and heated under reflux overnight under an atmosphere of argon. The

reaction solution was filtered and the filtrate concentrated *in vacuo*. The resulting residue was crystallized from dioxane to yield (**8**) (4.60 g, 75%) as a beige solid: R_f 0.3 (1:4 ethyl acetate/petroleum ether); mp 204–207 °C (dioxane) (lit. 204–206 °C [dioxane]);² ν_{\max} (neat) cm^{-1} : 3457, 2215; ^1H NMR (500 MHz; DMSO- D_6): δ 7.72 (s, 2H), 7.49–7.39 (m, 5H), 7.29–7.17 (m, 5H, Ar CH); ^{13}C NMR (126 MHz; DMSO- D_6): δ 163.5, 136.6, 131.2, 129.4, 129.0, 128.9, 128.6, 128.3, 126.9, 124.2, 121.8, 115.5, 69.2; (ESI⁺) m/z 543.18 ([2M + Na]⁺, 100%). These data are in good agreement with literature values.¹

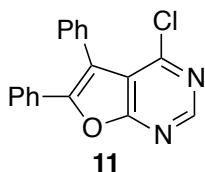
5,6-Diphenylfuro[2,3-d]pyrimidin-4(3H)-one (**10**)



Formic acid (13.85 mL, 367.1 mmol) was added to acetic anhydride (13.85 mL, 146.8 mmol) stirring over 4 Å molecular sieves. The reaction solution was stirred at 60 °C for 2 h, generating acetic formic anhydride as a colorless liquid. Compound **8** (3.00 g, 11.5 mmol, 1.0 eq) was added to the acetic formic anhydride and the resulting solution heated to 85 °C for 6 h. The solution was filtered and adjusted to pH 6 using aqueous NaOH (2 M). The filtrate was extracted with CH_2Cl_2 (50 mL) and the organic layer washed with distilled water (3 × 50 mL). The organic layer was dried over MgSO_4 , filtered and concentrated *in vacuo*. The resulting residue took the form of a brown solid (2.77 g), containing **9** { m/z (ES⁺) 311 ([M+Na]⁺, 100%)}, and was used without further purification.

The brown solid (3.79 g) containing **9** was heated to 220 °C for 30 min. The resulting solid was crystallized from methanol to yield **10** (1.70 g, 51% over two steps) as a brown solid: R_f 0.2 (3:7 ethyl acetate:petroleum ether); mp 258–260 °C (MeOH) (lit. > 240 °C [MeOH]);² ν_{\max} (neat) cm^{-1} : 3443, 1675; ^1H NMR (500 MHz; DMSO- D_6): δ 12.65 (s, 1H), 8.17 (s, 1H), 7.45–7.33 (m, 10H); ^{13}C NMR (126 MHz; DMSO- D_6): δ 164.6, 159.0, 148.0, 146.8, 131.7, 131.0, 129.9, 129.62, 129.55, 129.2, 128.8, 127.2, 119.5, 108.9; (ESI⁺) m/z 599.18 ([2M + Na]⁺, 100%). These data are in good agreement with literature values.²

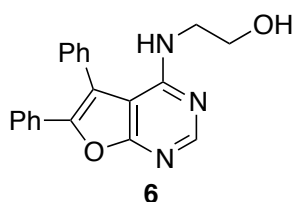
4-Chloro-5,6-diphenylfuro[2,3-d]pyrimidine (**11**)



Phosphorus(V) oxychloride (9.93 mL, 106.53 mmol, 30.7 eq) was added to dry **10** (1.00 g, 3.47 mmol, 1.0 eq). The reaction solution was stirred at 55 °C under argon for 2 h, then quenched by pouring over crushed ice and adjusted to pH 7 using a saturated aqueous solution of Na_2CO_3 . The product was extracted with CH_2Cl_2 (100 mL) and washed with distilled water (3 × 50 mL). The organic layer was dried over MgSO_4 , filtered and concentrated *in vacuo*. The resulting residue was adsorbed onto silica gel and purified using silica gel column chromatography (elution: 1:9:90 triethylamine:ethyl acetate:petroleum ether) to give **11** (0.86 g, 81%) as a pale yellow solid: R_f 0.9 (1:4 ethyl acetate:petroleum ether); mp 103–105 °C (ethyl acetate/petroleum ether) (lit. 121–122 °C);² ^1H NMR (500 MHz;

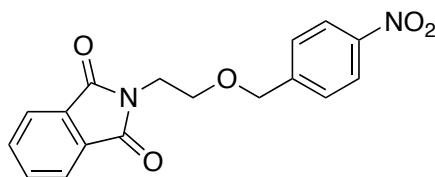
DMSO-D₆): δ 8.86 (s, 1H), 7.53–7.40 (m, 10H); ¹³C NMR (126 MHz; DMSO-D₆): δ 165.3, 153.0, 151.7, 150.9, 130.5, 130.1, 129.9, 129.0, 128.8, 128.0, 126.9, 118.4, 115.0; *m/z* (ES⁺) 307 ([M+H]⁺, 100%). These data are in good agreement with literature values.²

2-(5,6-Diphenylfuro[2,3-d]pyrimidin-4-ylamino)ethanol (**6**)



Chloride **11** (0.70 g, 2.29 mmol, 1.0 eq) and ethanolamine (0.30 mL, 5.03 mmol, 2.2 eq) were added to ethanol (7 mL). The reaction was heated under reflux for 6 h and then concentrated *in vacuo*. The resulting residue was adsorbed onto Celite[®] and purified by silica gel column chromatography, eluting with Et₃N, MeOH, ethyl acetate and petroleum ether (1:4:30:65) to afford **6** (0.72 g, 95%) as a beige solid: *R_f* 0.3 (1:4:30:65 Et₃N:MeOH:ethyl acetate:petroleum ether); mp 154–157 °C (ethyl acetate/petroleum ether) (lit. 163–165 °C);² ¹H NMR (500 MHz; CDCl₃): δ 8.32 (s, 1H, 3.56), 7.52–7.22 (m, 10H), 5.16 (t, *J* = 5.2, 1H), 4.18 (br s, 1H), 3.72 (t, *J* = 4.2, 2H), 3.56 (dt, *J* = 5.2, 4.6, 2H); ¹³C NMR (126 MHz; CDCl₃): δ 164.7, 157.9, 153.6, 147.0, 132.1, 129.72, 129.70, 129.3, 129.0, 128.5, 128.5, 126.4, 114.9, 103.3, 62.4, 44.2; *m/z* (ES⁺) 333 (100%), 332 ([M+H]⁺, 98%), 370 ([M+K]⁺, 78%). These data are in good agreement with literature values.²

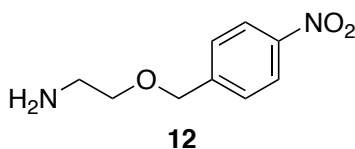
2-(2-([4-Nitrobenzyl]oxy)ethyl)isoindoline-1,3-dione



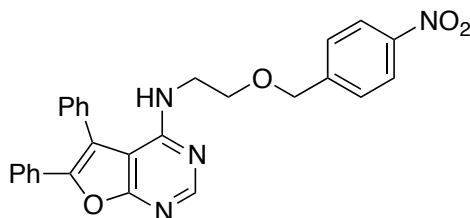
Sodium hydride (0.63 g 60% dispersion in mineral oil, 15.69 mmol, 1.5 eq) was added to a stirred solution of *N*-(2-hydroxyethyl)phthalimide (2.00 g, 10.46 mmol, 1.0 eq) in dry THF (20 mL). The reaction mixture was stirred for 15 min under argon at ambient temperature and then 4-nitrobenzyl bromide (3.39 g, 15.69 mmol, 1.5 eq) and TBAI (0.39 g, 1.05 mmol, 0.1 eq) were added. The reaction mixture was stirred at ambient temperature for a further 24 h, quenched by the addition of methanol (20 mL) and the solvents removed *in vacuo*. The residue was dissolved in CH₂Cl₂ (50 mL), washed with distilled water (3 × 50 mL) and the organic layer dried over MgSO₄, filtered and concentrated *in vacuo*. The resulting residue was purified by silica gel column chromatography, eluting with ethyl acetate and petroleum ether (20:80) to afford 2-(2-([4-nitrobenzyl]oxy)ethyl)isoindoline-1,3-dione (0.78 g, 23%) as a yellow solid: R_f 0.2 (1:4 ethyl acetate:petroleum ether); mp 142–143 °C (ethyl acetate/petroleum ether); ν_{max} (neat) cm⁻¹: 1739, 1515, 1366; ¹H NMR (500 MHz; CDCl₃): δ 8.12 (d, *J* = 8.7, 2H), 7.87–7.85 (m, 2H), 7.74–7.73 (m, 2H), 7.43 (d, *J* = 8.7, 2H), 4.63 (s, 2H), 3.98 (t, *J* = 5.6, 2H), 3.80 (t, *J* = 5.6); ¹³C NMR (126 MHz; CDCl₃): δ 168.2, 147.3, 145.5, 134.1, 132.0, 127.6, 123.6, 123.3, 71.4, 67.7, 37.4; HRMS *m/z* (ESI⁺) found [M+Na]⁺ 349.0791, C₁₇H₁₄N₂O₅Na requires [M+Na]⁺ 349.0795; *m/z*

(ES⁺) 349 ([M+Na]⁺, 100%), 675 ([2M+Na]⁺, 42%). Anal. Calcd for C₁₇H₁₄N₂O₅: C, 62.6; H, 4.3; N, 8.6. Found C, 62.5; H, 4.4; N, 8.5.

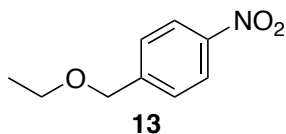
2-([4-Nitrobenzyl]oxy)ethanamine (12)



Hydrazine monohydrate (1.98 g, 1.92 mL, 39.6 mmol, 43.1 eq) was added to a solution of 2-(2-([4-nitrobenzyl]oxy)ethyl)isoindoline-1,3-dione (0.30 g, 0.92 mmol, 1.0 eq.) in ethanol (20 mL). The reaction mixture was stirred at ambient temperature for three days under argon then filtered, washed with ethanol and the filtrate concentrated *in vacuo*. The resulting residue was dissolved in CH₂Cl₂ (20 mL) and washed with aqueous HCl (2 M, 2 × 20 mL). The aqueous phases were combined, washed with diethyl ether (2 × 20 mL) and then adjusted to pH > 11 using NaOH pellets. The organic material was extracted from this mixture with CH₂Cl₂ (20 mL), the combined CH₂Cl₂ layers dried over MgSO₄, filtered and the solvent was removed *in vacuo* to yield 2-([4-nitrobenzyl]oxy)ethanamine (**12**) (0.14 g, 80%) as an orange liquid: *R*_f 0.2 (2:3, ethyl acetate:petroleum ether); *v*_{max} (CHCl₃) cm⁻¹: 1518, 1345; ¹H NMR (400 MHz; CDCl₃): δ 8.11 (d, *J* = 8.6, 2H), 7.44 (d, *J* = 8.6, 2H), 4.57 (s, 2H), 3.50 (t, *J* = 5.2, 2H), 2.86 (t, *J* = 5.2, 2H); ¹³C NMR (100 MHz; CDCl₃): δ 147.2, 146.1, 127.6, 123.5, 73.1, 71.7, 41.8; HRMS *m/z* (ESI⁺) found [M+H]⁺ 197.0929, C₉H₁₃N₂O₃ requires [M+H]⁺ 197.0921; *m/z* (ES⁺) 197 ([M+H]⁺, 100%).

N-(2-([4-Nitrobenzyl]oxy)ethyl)-5,6-diphenylfuro[2,3-*d*]pyrimidin-4-amine (**1**)**1**

A solution of **11** (0.05 g, 0.17 mmol, 1.0 eq), amine **12** (0.07 g, 0.38 mmol, 2.2 eq) and Et₃N (0.05 mL, 0.38 mmol, 2.2 eq) in DMF (3 mL) was stirred at 80 °C under nitrogen for 6 h and then concentrated *in vacuo*. The resulting residue was dissolved in CH₂Cl₂ (20 mL) and washed with distilled water (3 × 20 mL). The organic layer was dried over MgSO₄, filtered and concentrated *in vacuo* and the resulting residue purified by silica gel chromatography, eluting with Et₃N, MeOH, ethyl acetate and petroleum ether (1:4:20:75) to afford *N*-(2-([4-nitrobenzyl]oxy)ethyl)-5,6-diphenylfuro[2,3-*d*]pyrimidin-4-amine (**1**) (0.1 g, 93%) as a yellow solid: R_f 0.2 (1:1, ethyl acetate:petroleum ether); mp 127–129 °C (ethyl acetate); ν_{max} (neat) cm⁻¹: 3426, 1520, 1340; ¹H NMR (400 MHz; CDCl₃): δ 8.42 (s, 1H), 8.22 (d, *J* = 8.7, 2H), 7.52–7.27 (m, 10H), 7.35 (d, *J* = 8.7, 2H), 5.12 (t, *J* = 5.2, 1H), 4.51 (s, 2H), 3.74 (dt, *J* = 5.2, 5.2, 2H), 3.61 (t, *J* = 5.2, 2H); ¹³C NMR (126 MHz; CDCl₃): δ 165.0, 157.4, 154.0, 147.4, 146.9, 145.4, 132.4, 129.7, 129.5, 129.4, 128.7, 128.5, 127.5, 126.4, 123.6, 114.8, 103.3, 71.7, 69.3, 40.4; HRMS *m/z* (ESI⁻) [Found: [M-H]⁻ 465.1575, C₂₇H₂₁N₄O₄ requires [M-H]⁻ 465.1568]; *m/z* (ES⁻) 465 ([M-H]⁻, 100%); Anal. Calcd for C₂₇H₂₂N₄O₄: C, 69.5; H, 4.8; N, 12.0. Found C, 69.4; H, 4.6; N, 11.9.

1-(Ethoxymethyl)-4-nitrobenzene (13)

Anhydrous ethanol (6 mL) was added to dry THF (30 mL) and the solution cooled with an ice-water bath. Sodium hydride was then added in portions and the resulting mixture left to stir at the ambient temperature for 10 min. This mixture was syringed drop wise into an ice-cooled solution of 4-nitrobenzylbromide (1.0 g, 4.62 mmol) in dry THF (20 mL) and the reaction left stirring at the ambient temperature for an additional 2 h. A saturated aqueous solutions of NH_4Cl was added, the mixture concentrated *in vacuo* and then re-dissolved in EtOAc. This solution washed with brine, dried over anhydrous MgSO_4 , filtered and the solvent removed *in vacuo*. Purification by column chromatography eluting with petroleum ether and ethyl acetate (gradient: 10:0 to 9:1) gave 1-(ethoxymethyl)-4-nitrobenzene (**13**) as a yellow oil (161 mg, 19%): R_f 0.4 (1:10 ethyl acetate:petroleum ether); $^1\text{H NMR}$ (400 MHz, CDCl_3): δ 8.21 (d, $J = 8.1$, 2H), 7.51 (d, $J = 8.1$, 2H), 4.61 (s, 2H), 3.60 (q, $J = 7.0$), (t, $J = 7.0$); m/z (FI^+) 181.07 ($[\text{M}]^+$, 100%). These data are in good agreement with literature values.³

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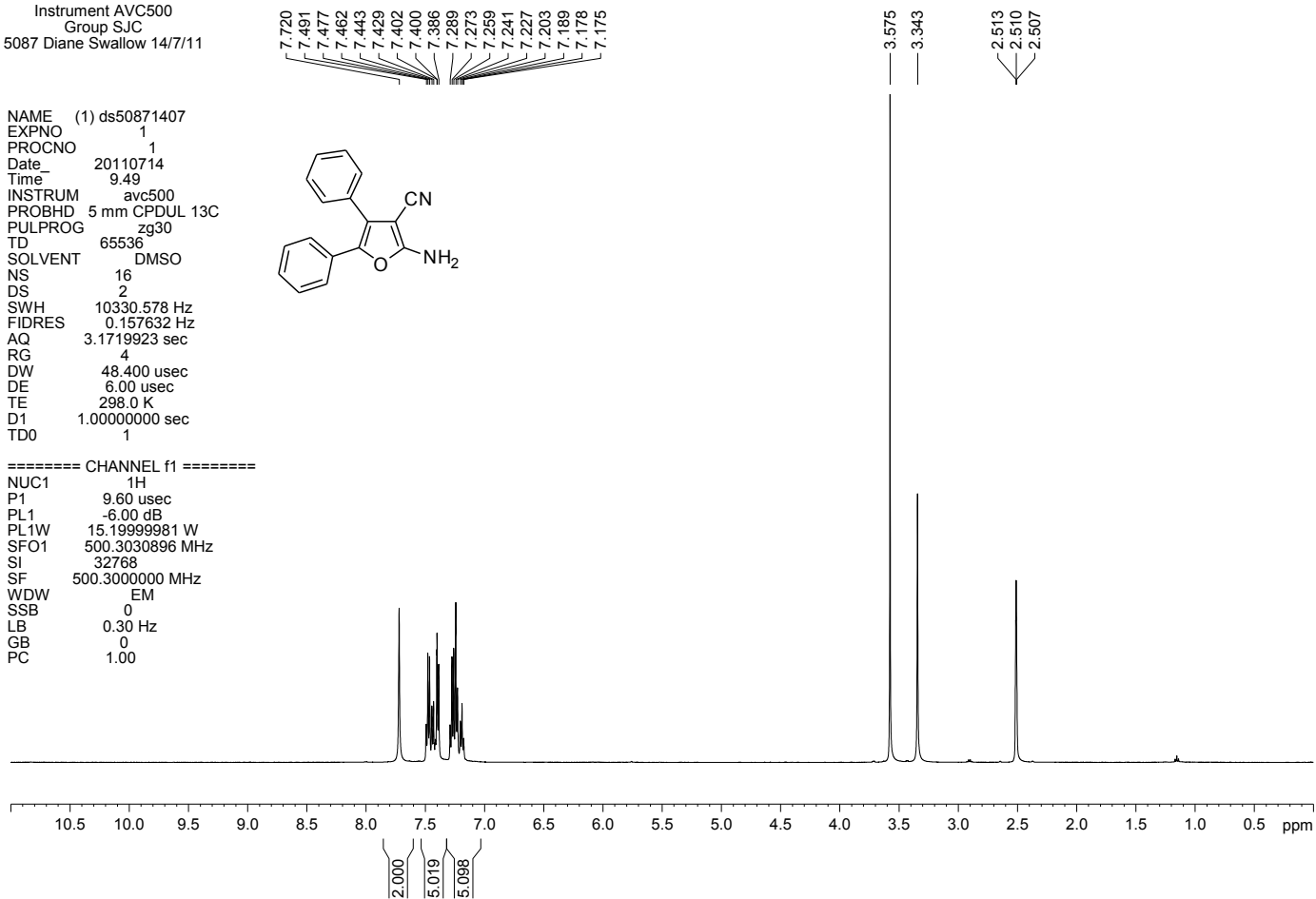
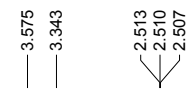
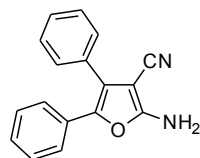
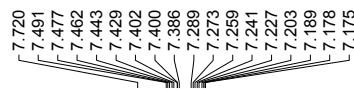
¹H and ¹³C NMR Spectra

2-Amino-4,5-diphenylfuran-3-carbonitrile (**8**) – ¹H NMR spectrum

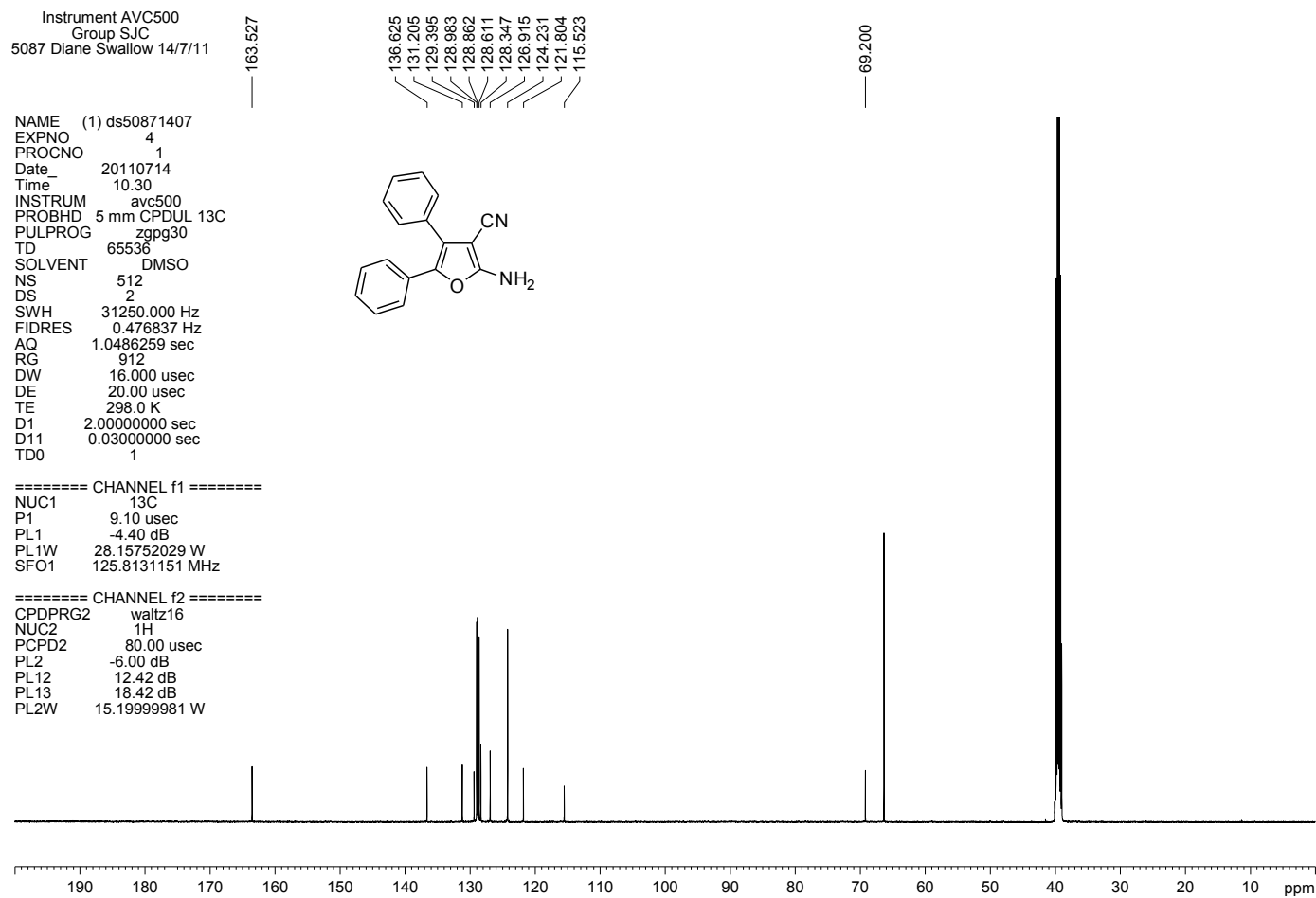
Instrument AVC500
Group SJC
5087 Diane Swallow 14/7/11

NAME (1) ds50871407
EXPNO 1
PROCNO 1
Date_ 20110714
Time_ 9.49
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 4
DW 48.400 usec
DE 6.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 9.60 usec
PL1 -6.00 dB
PL1W 15.19999981 W
SFO1 500.3030896 MHz
SI 32768
SF 500.3000000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



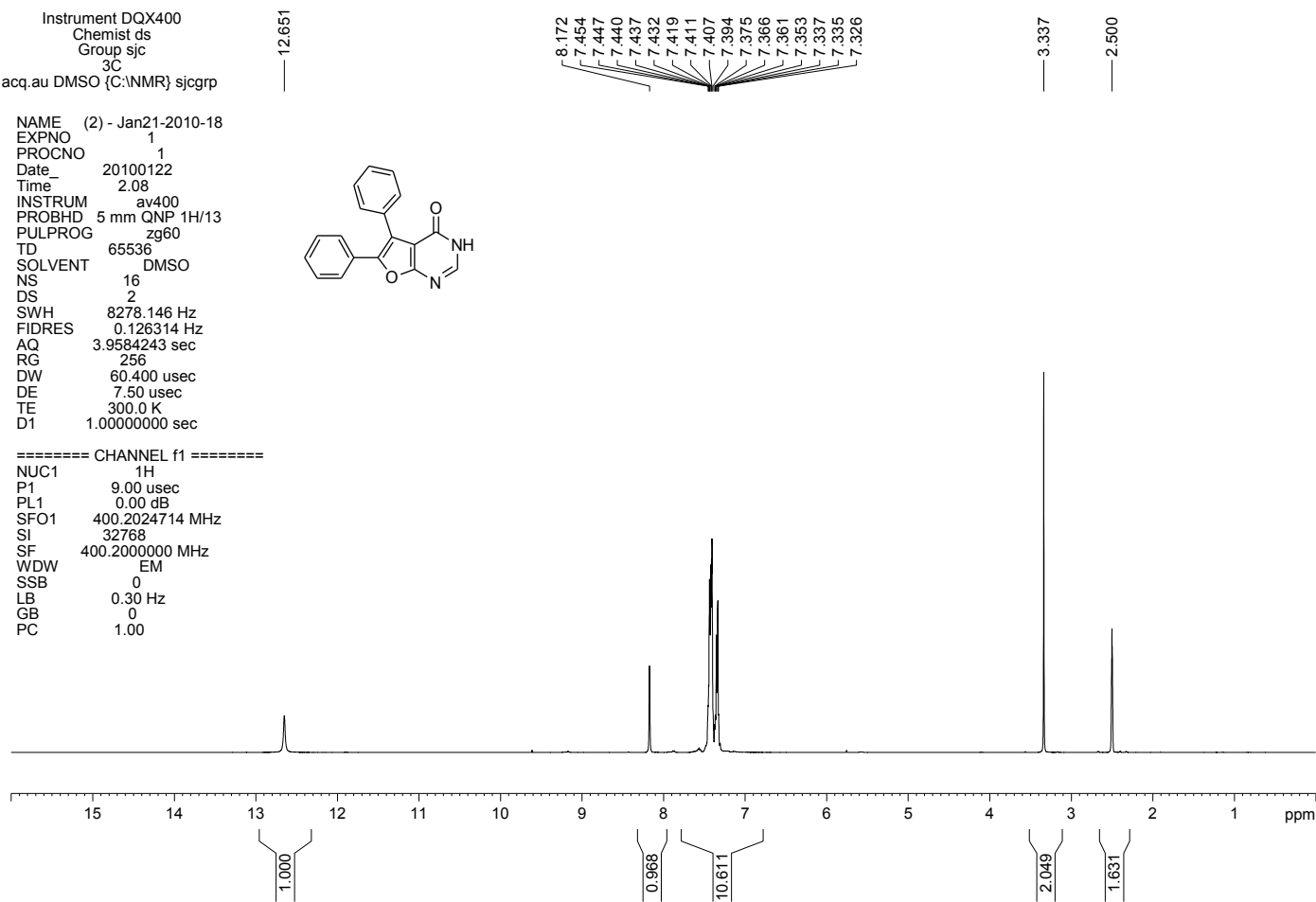
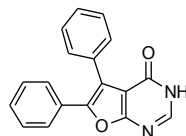
2-Amino-4,5-diphenylfuran-3-carbonitrile (**8**) – ¹³C NMR spectrum



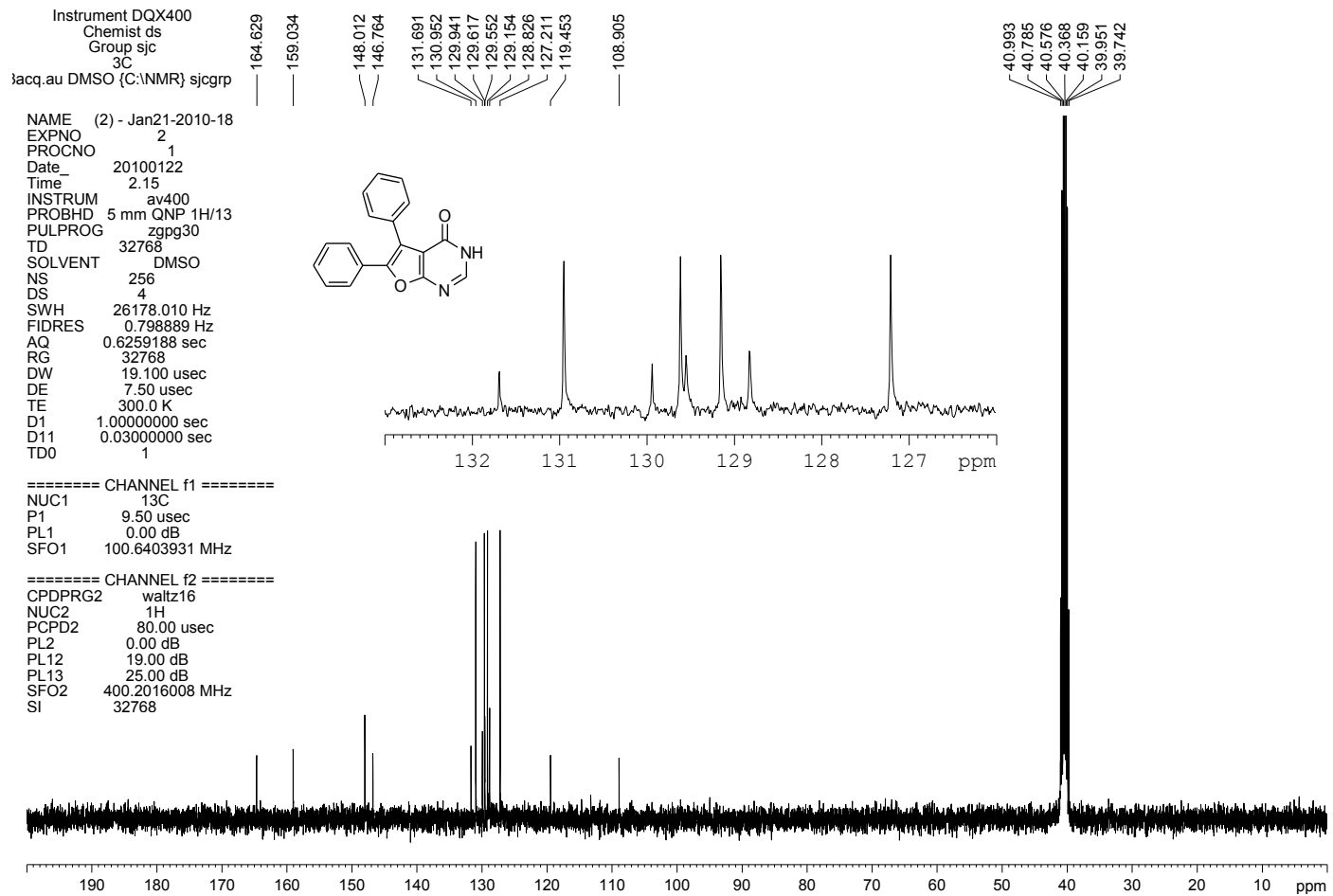
5,6-Diphenylfuro[2,3-d]pyrimidin-4(3H)-one (**10**) – ¹H NMR spectrum

Instrument DQX400
 Chemist ds
 Group sjc
 3C
 acq.au DMSO {C:\NMR} sjcgrp

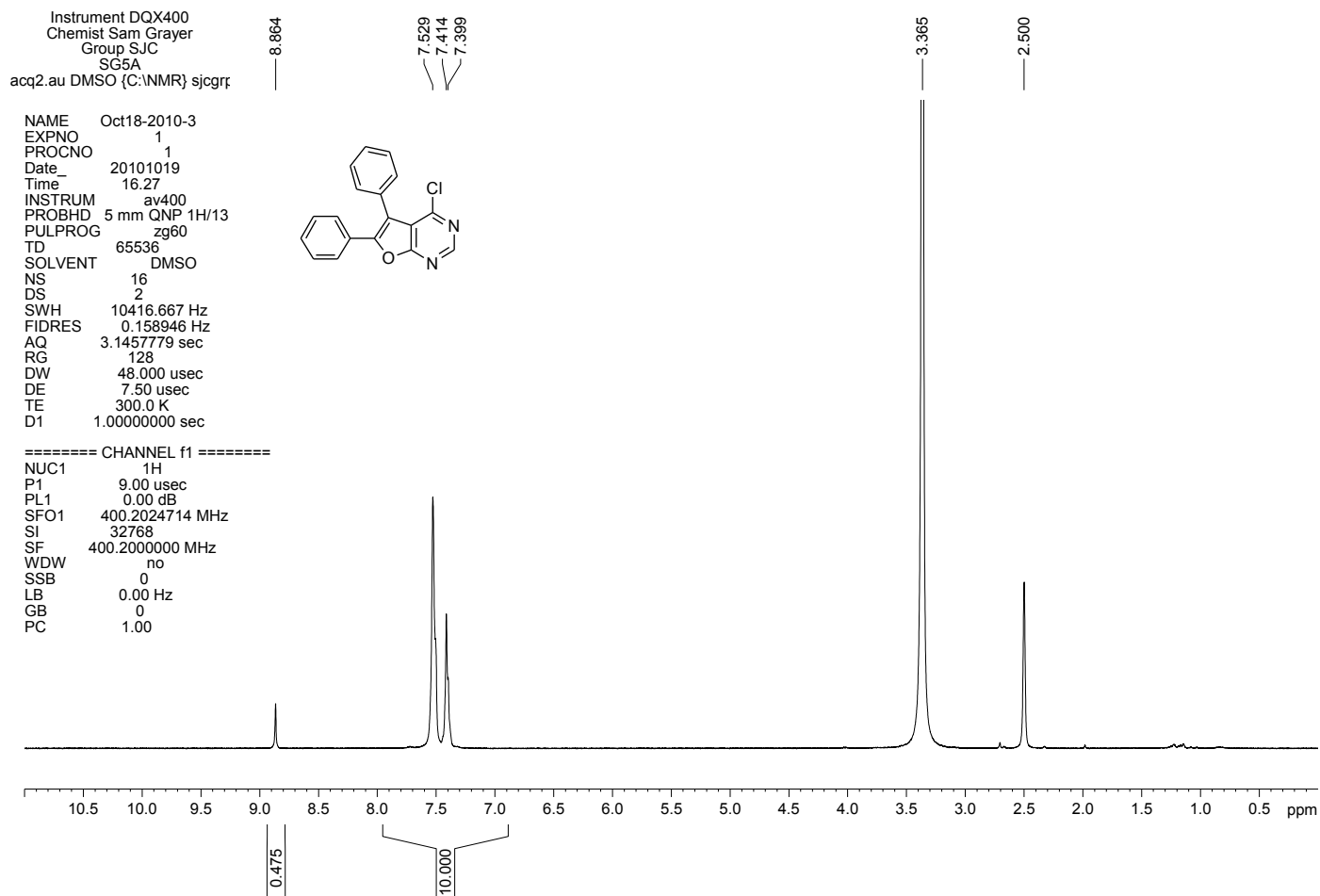
NAME (2) - Jan21-2010-18
 EXPNO 1
 PROCNO 1
 Date_ 20100122
 Time 2.08
 INSTRUM av400
 PROBHD 5 mm QNP 1H/13
 PULPROG zg60
 TD 65536
 SOLVENT DMSO
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 256
 DW 60.400 usec
 DE 7.50 usec
 TE 300.0 K
 D1 1.0000000 sec



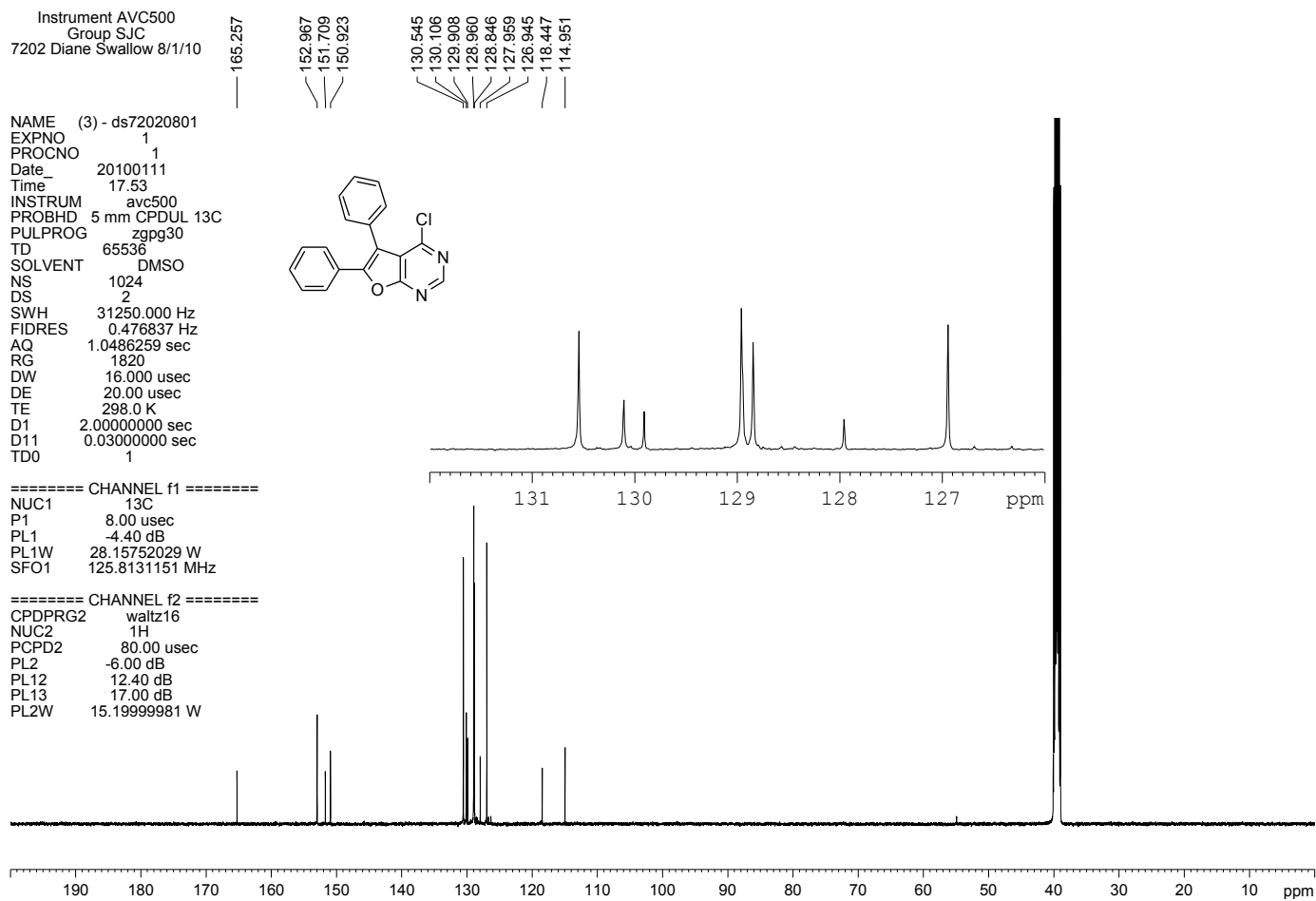
5,6-Diphenylfuro[2,3-d]pyrimidin-4(3H)-one (**10**) – ¹³C NMR spectrum



4-Chloro-5,6-diphenylfuro[2,3-d] pyrimidine (**11**) – ¹H NMR spectrum



4-Chloro-5,6-diphenylfuro[2,3-d] pyrimidine (**11**) – ¹³C NMR spectrum



2-(5,6-Diphenylfuro[2,3-d]pyrimidin-4-ylamino)ethanol (**6**) – ¹H NMR spectrum

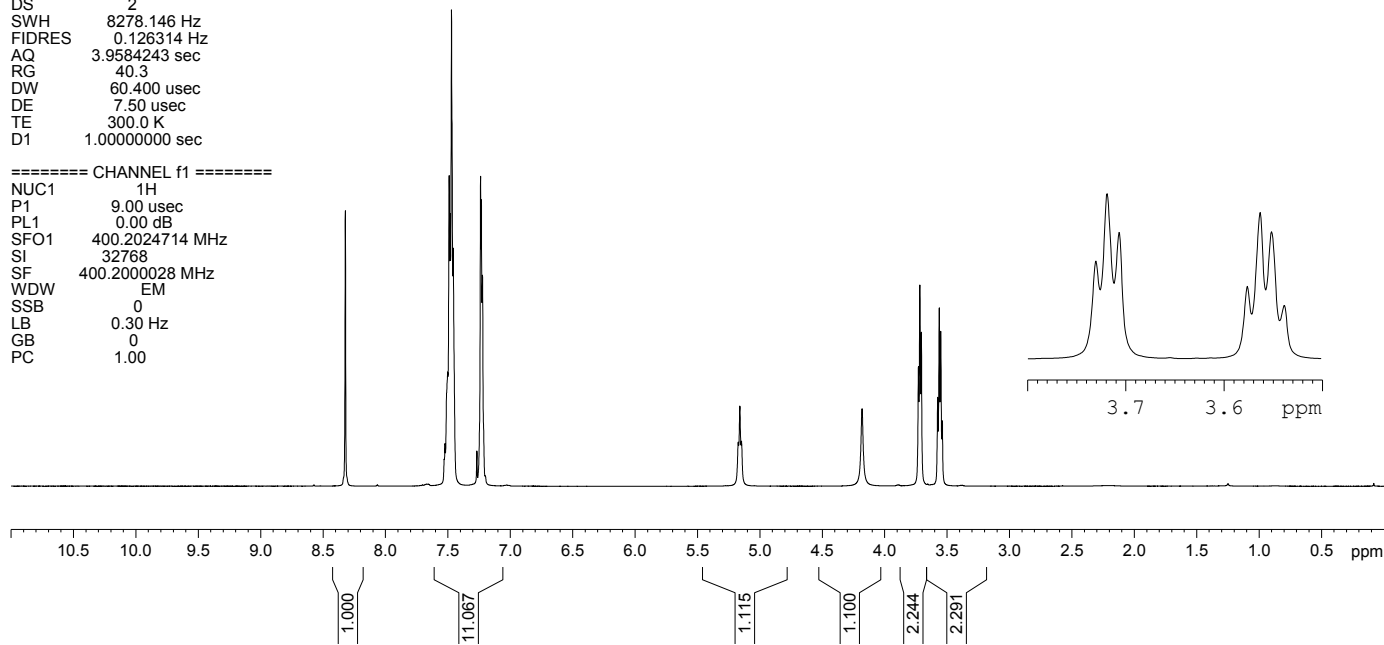
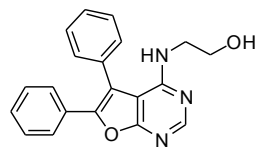
Instrument DQX400
 Chemist ds
 Group sjc
 ds-3Y
 xq.au CDCI3 [E:\NMR2012} sjcg

NAME (4) - Aug07-2012-7
 EXPNO 1
 PROCNO 1
 Date_ 20120808
 Time 1.09
 INSTRUM av400
 PROBHD 5 mm QNP 1H/13
 PULPROG zg60
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 40.3
 DW 60.400 usec
 DE 7.50 usec
 TE 300.0 K
 D1 1.0000000 sec

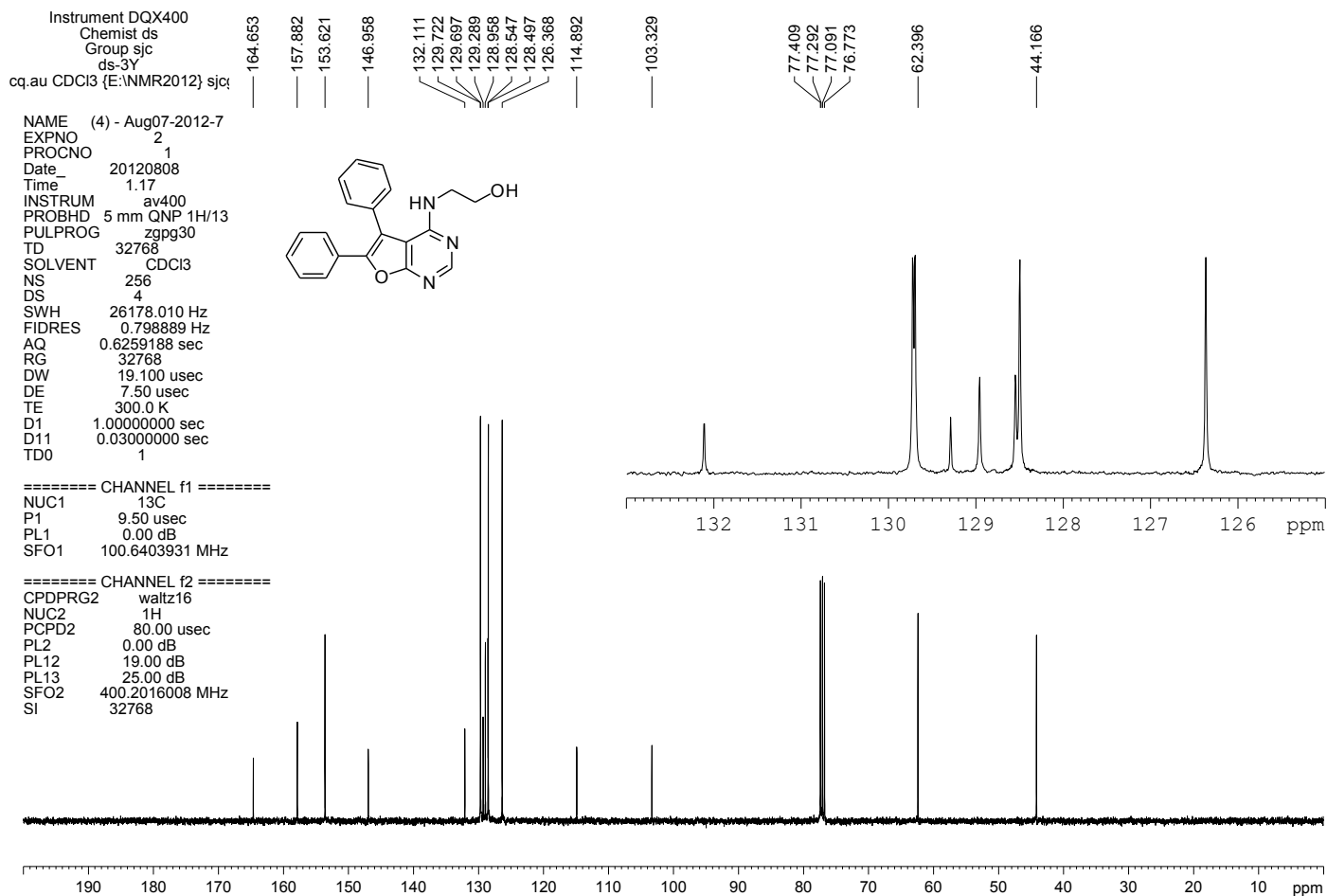
===== CHANNEL f1 =====

NUC1 1H
 P1 9.00 usec
 PL1 0.00 dB
 SFO1 400.2024714 MHz
 SI 32768
 SF 400.2000028 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

8.322
 7.524
 7.503
 7.491
 7.482
 7.472
 7.458
 7.239
 7.232
 7.224
 5.174
 5.161
 5.148
 4.182
 3.730
 3.719
 3.707
 3.576
 3.563
 3.552
 3.539



2-(5,6-Diphenylfuro[2,3-d]pyrimidin-4-ylamino)ethanol (**6**) – ¹³C NMR spectrum



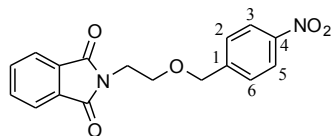
2-(2-(4-Nitrobenzyl)oxy)ethylisoindoline-1,3-dione – ¹H NMR spectrum

Instrument AVC500
Group SJC
3841 Sam Grayer 28/4/11

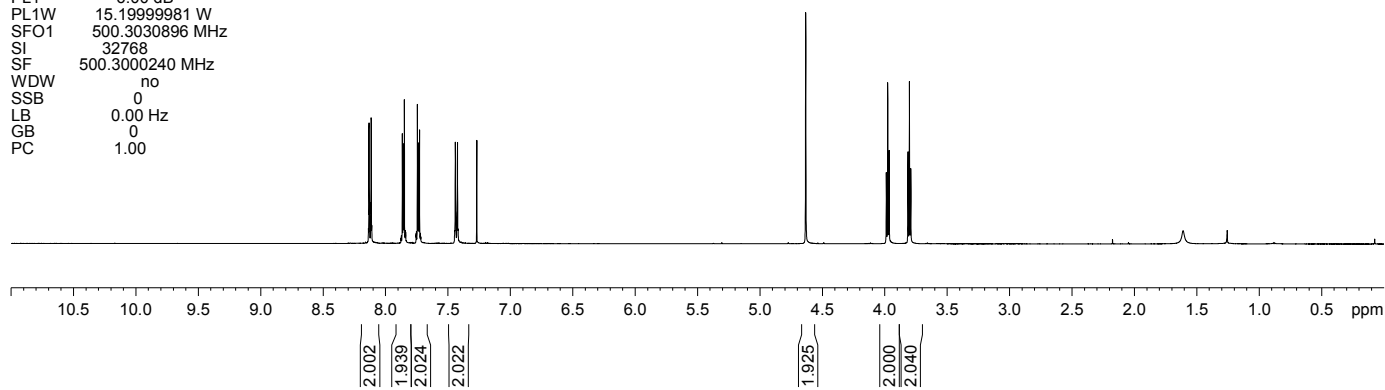
8.132
8.114
7.865
7.859
7.854
7.848
7.744
7.738
7.733
7.727
7.440
7.423
7.268

4.633
3.989
3.978
3.966
3.815
3.804
3.793

NAME sg38412804
EXPNO 1
PROCNO 1
Date_ 20110501
Time 21.53
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 10330.578 Hz
FIDRES 0.157632 Hz
AQ 3.1719923 sec
RG 4
DW 48.400 usec
DE 6.00 usec
TE 298.0 K
D1 1.0000000 sec
TD0 1



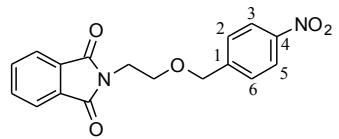
===== CHANNEL f1 =====
NUC1 1H
P1 9.60 usec
PL1 -6.00 dB
PL1W 15.19999981 W
SFO1 500.3030896 MHz
SI 32768
SF 500.3000240 MHz
WDW no
SSB 0
LB 0.00 Hz
GB 0
PC 1.00



2-(2-([4-Nitrobenzyl]oxy)ethyl)isoindoline-1,3-dione – ¹³C NMR spectrum

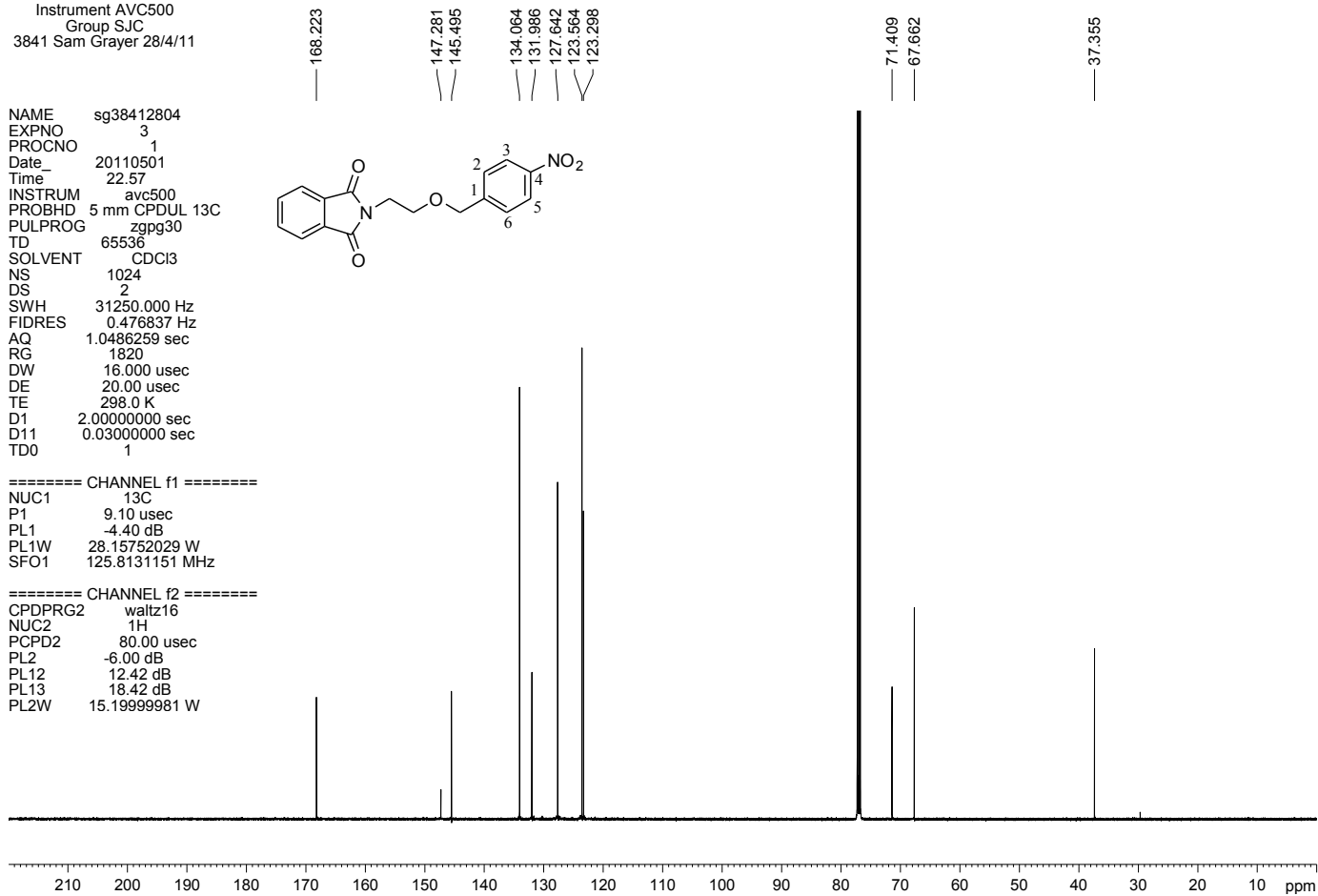
Instrument AVC500
Group SJC
3841 Sam Grayer 28/4/11

NAME sg38412804
EXPNO 3
PROCNO 1
Date_ 20110501
Time 22.57
INSTRUM avc500
PROBHD 5 mm CPDUL 13C
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1024
DS 2
SWH 31250.000 Hz
FIDRES 0.476837 Hz
AQ 1.0486259 sec
RG 1820
DW 16.000 usec
DE 20.00 usec
TE 298.0 K
D1 2.0000000 sec
D11 0.03000000 sec
TD0 1

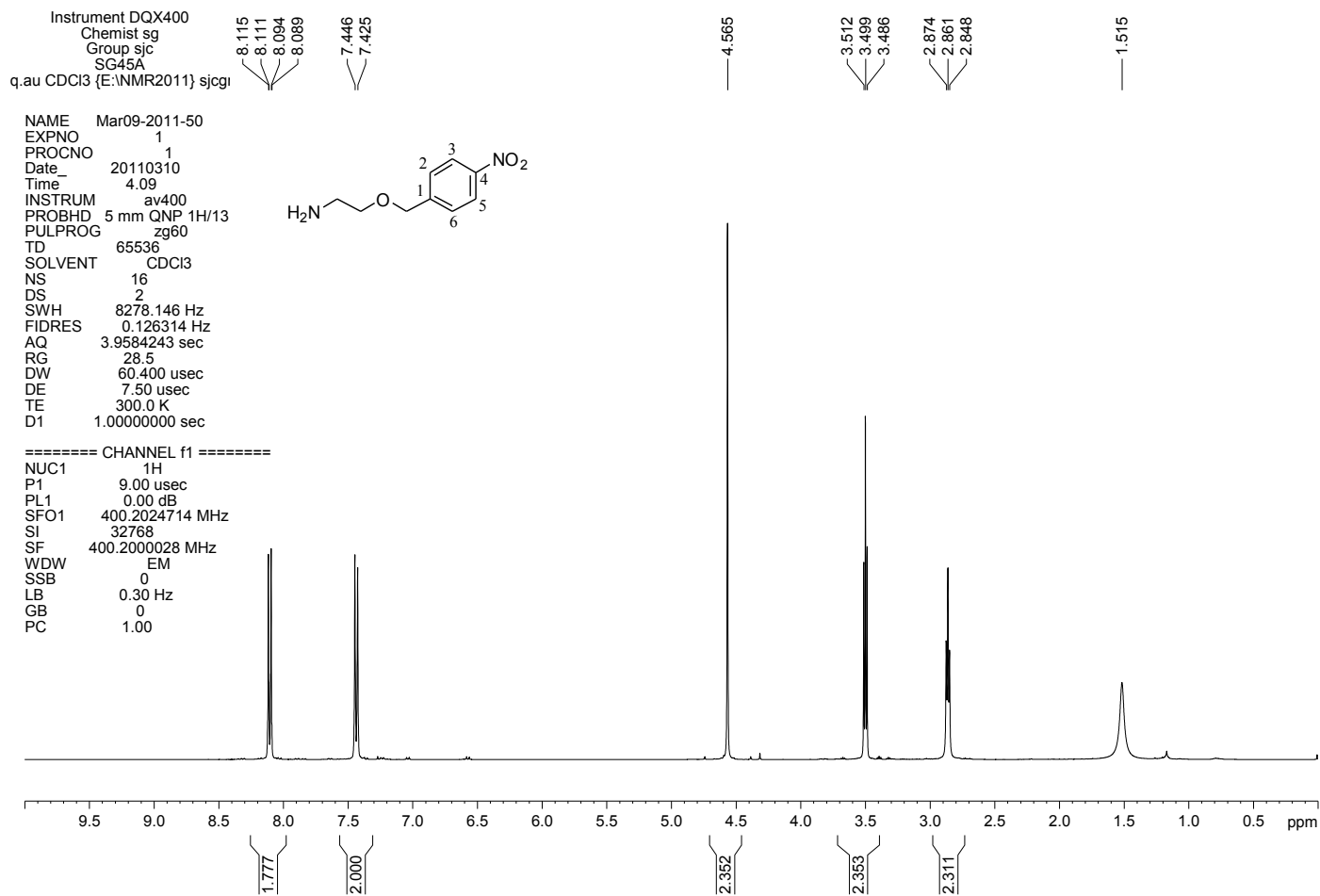


==== CHANNEL f1 =====
NUC1 13C
P1 9.10 usec
PL1 -4.40 dB
PL1W 28.15752029 W
SFO1 125.8131151 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -6.00 dB
PL12 12.42 dB
PL13 18.42 dB
PL2W 15.19999981 W



2-([4-Nitrobenzyl]oxy)ethanamine (**12**) – ¹H NMR spectrum



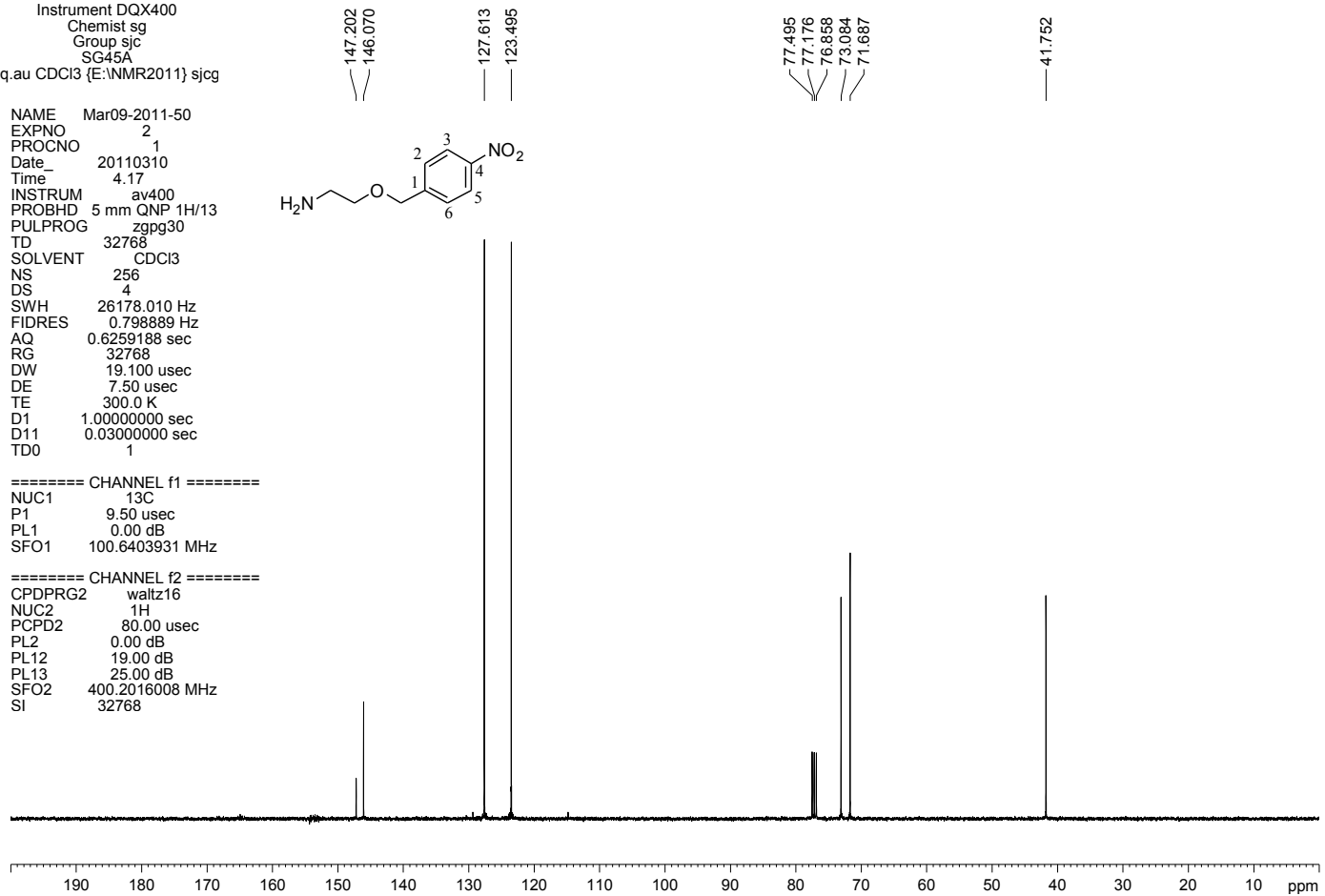
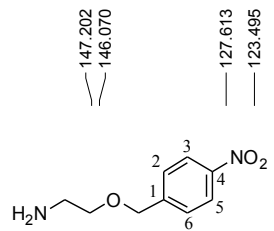
2-([4-Nitrobenzyl]oxy)ethanamine (**12**) – ¹³C NMR spectrum

Instrument DQX400
 Chemist sg
 Group sjc
 SG45A
 xq.au CDCl3 [E:\NMR2011] sjcg

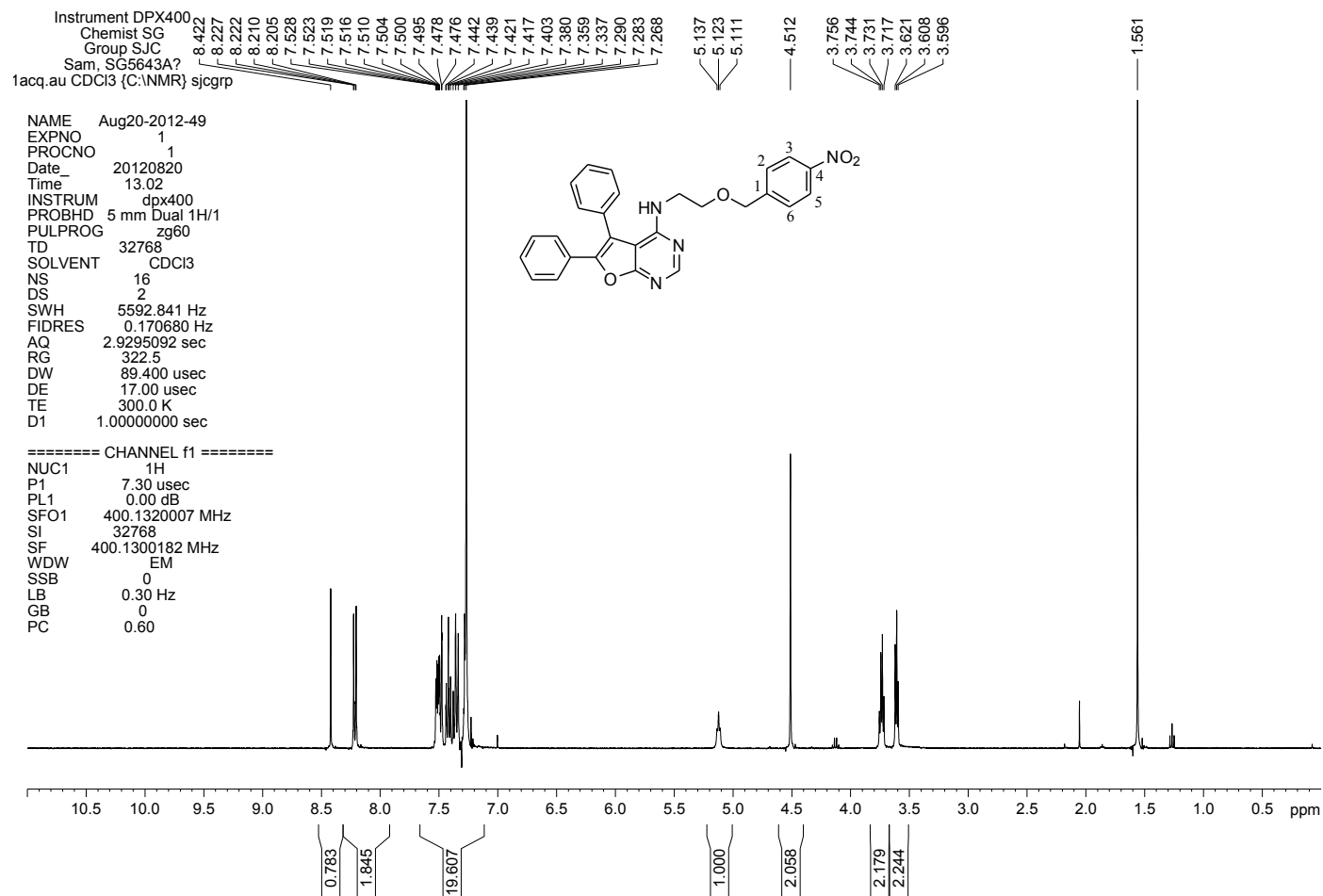
NAME Mar09-2011-50
 EXPNO 2
 PROCNO 1
 Date_ 20110310
 Time 4.17
 INSTRUM av400
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 256
 DS 4
 SWH 26178.010 Hz
 FIDRES 0.798889 Hz
 AQ 0.6259188 sec
 RG 32768
 DW 19.100 usec
 DE 7.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.50 usec
 PL1 0.00 dB
 SFO1 100.6403931 MHz

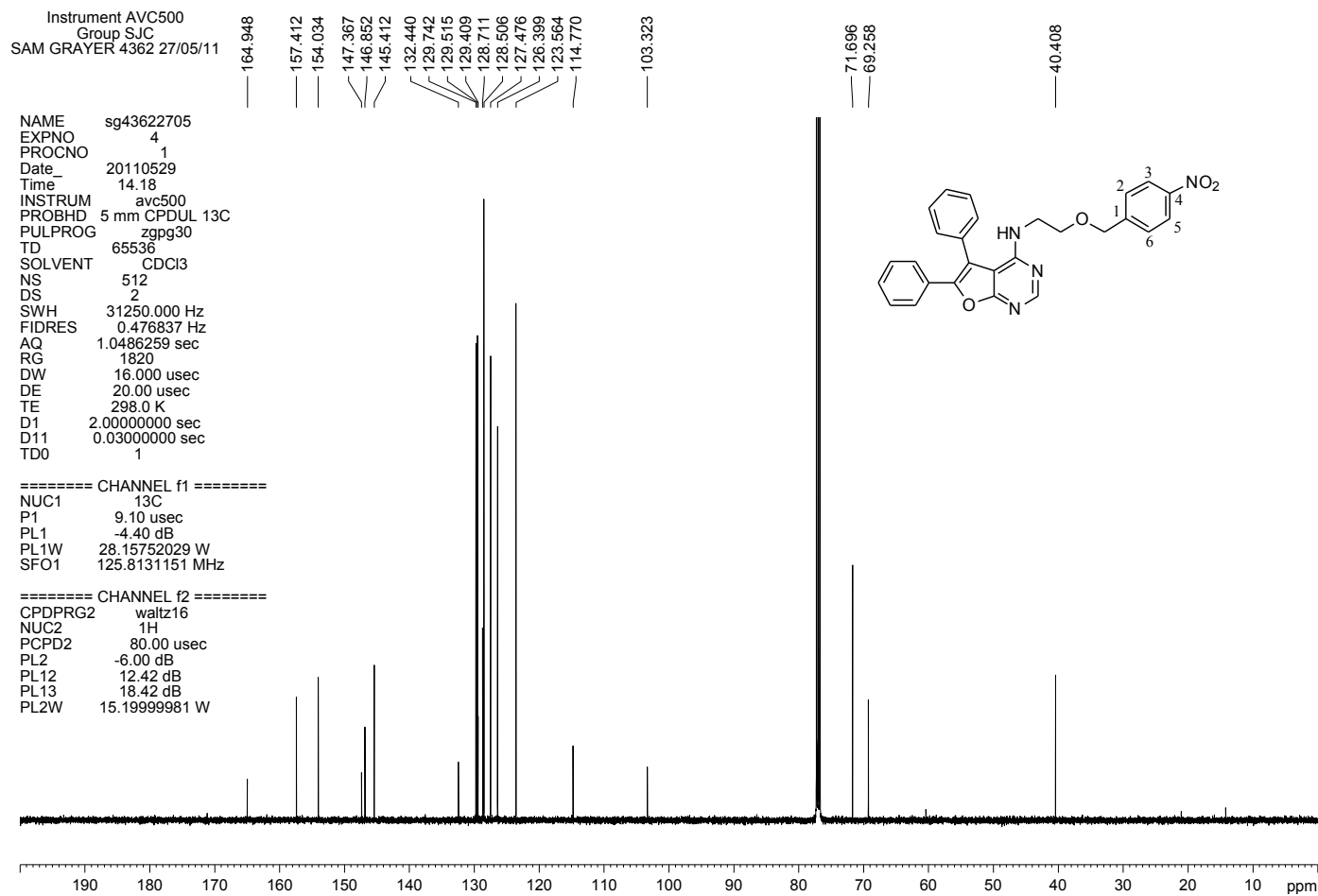
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0.00 dB
 PL12 19.00 dB
 PL13 25.00 dB
 SFO2 400.2016008 MHz
 SI 32768



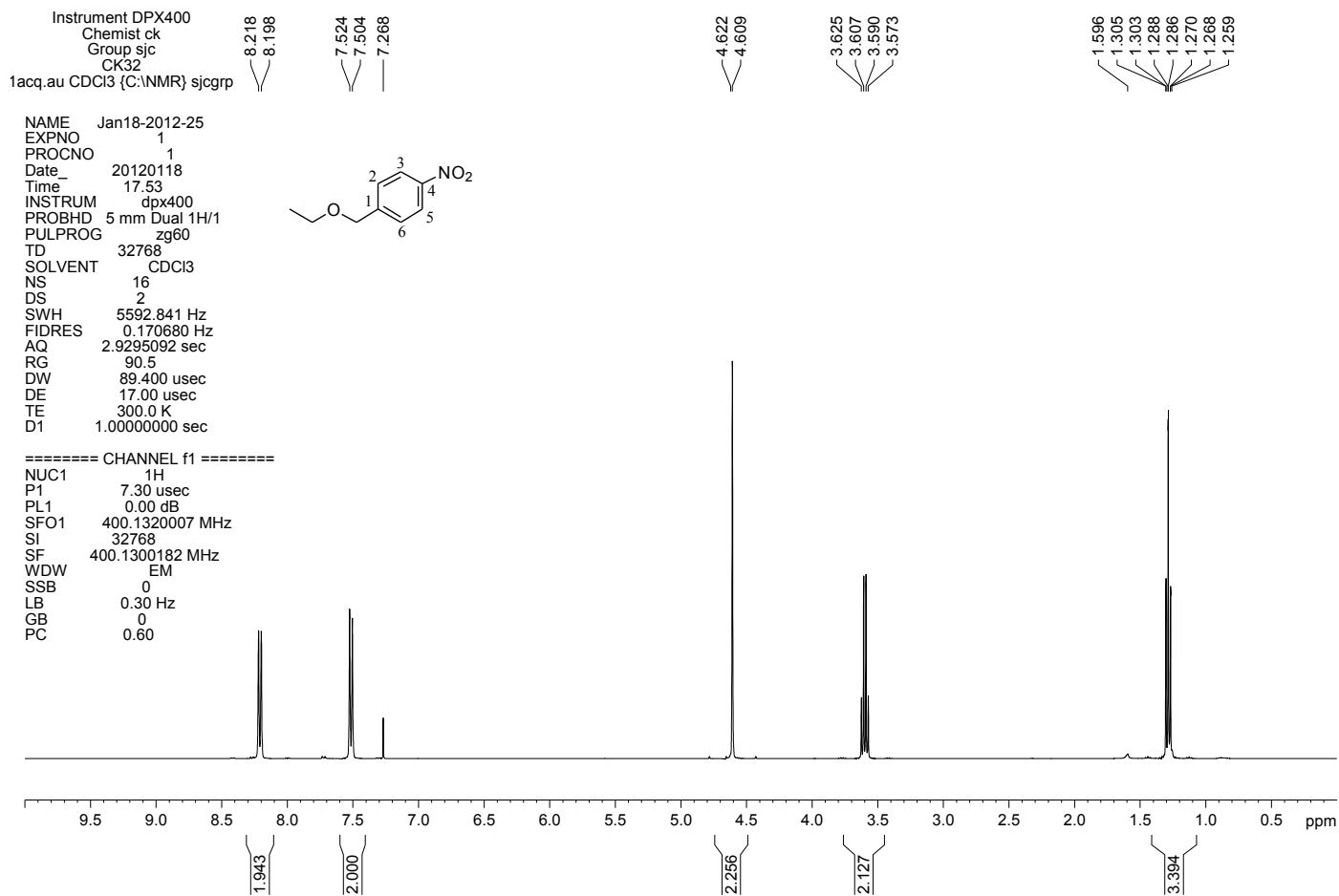
N-(2-([4-Nitrobenzyl]oxy)ethyl)-5,6-diphenylfuro[2,3-*d*]pyrimidin-4-amine (1) – ¹H NMR spectrum



N-(2-([4-Nitrobenzyl]oxy)ethyl)-5,6-diphenylfuro[2,3-*d*]pyrimidin-4-amine (1) – ¹³C NMR spectrum



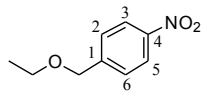
1-(Ethoxymethyl)-4-nitrobenzene (**13**) – ¹H NMR spectrum



1-(Ethoxymethyl)-4-nitrobenzene (13) – ¹³C NMR spectrum

Instrument DQX400
 Chemist ck
 Group sjc
 CK32
 xq.au CDCI3 [E:\NMR2012] sjcg

NAME Jan19-2012-39
 EXPNO 1
 PROCNO 1
 Date_ 20120119
 Time 14.55
 INSTRUM av400
 PROBHD 5 mm QNP 1H/13
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 256
 DS 4
 SWH 26178.010 Hz
 FIDRES 0.798889 Hz
 AQ 0.6259188 sec
 RG 32768
 DW 19.100 usec
 DE 7.50 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1



147.291
 146.348

127.642
 123.595

77.328
 77.010
 76.693
 71.391
 66.449

15.143

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.50 usec
 PL1 0.00 dB
 SFO1 100.6403931 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0.00 dB
 PL12 19.00 dB
 PL13 25.00 dB
 SFO2 400.2016008 MHz
 SI 32768

