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Figure S1. Effectors that have undergone more than 8 divisions (D>8) have less potential to transit to memory. CFSE-labeled naïve DO11.10 T cells were transferred into Rag2^{-/-} mice and the hosts were immunized with OVAp in CFA. Eight days later the CD4⁺CFSE⁺ (D2-8) and CD4⁺CFSE⁻ (CFSE⁻) LN effectors were sorted as indicated in (A) and transferred into Rag2^{-/-} parking hosts. Seven weeks later, the SP cells were harvested and the CD4⁺KJ1-26⁺ cells were analyzed for IFN γ production (B). The results show that highly activated (D>8) effectors are less likely to transit to memory.

Figure S2. Re-expression of IL-7R during transition to memory may be dependent on IL-2 signaling. BALB/c mice recipient of CFSE-labeled naïve T cells were immunized with 125µg OVA/CFA and D2-8 effector T cells were isolated and transferred into Rag2^{-/-} mice to generate CFA discontinued animals (OVAp+CFA discontinued). Rag2^{-/-} mice, recipient of CFSE-labeled naïve T cells, were immunized with 125µg OVA/CFA and used as CFA continued (OVA+CFA continued) mice. (A) Shows IL-7R expression on KJ1.26⁺CD4⁺ splenic T cells harvested after 2, 4 and 8 weeks of parking. (B) Shows IL-2R (CD25) and IL-7R (CD127) expression on naïve T cells and on D2-8 effector T cells 3 and 5 days after immunization with OVAp+CFA. The cells were gated on KJ1.26 and CD4 and analyzed for CD25 and CD127 expression in comparison with isotype control (dotted lines) by flow cytometry. The results indicate that IL-2R (CD25) expression precedes IL-7R re-expression, perhaps suggesting that signaling through IL-2R is required for IL-7R expression and transition to memory .

Figure S3. Effectors primed in Rag2^{-/-} and wild type (WT) BALB/c hosts show similar responses to antigen-free adjuvant. CFSE-labeled naïve DO11.10 T cells were transferred into Rag2^{-/-} or WT BALB/c mice and the hosts were immunized with OVAp in CFA. Eight days later the CFSE⁺CD4⁺ LN D2-8 effectors were sorted from each group and transferred into Rag2^{-/-}

hosts recipient of effectors generated in Rag2^{-/-} (Rag2^{-/-} \rightarrow Rag2^{-/-}) or WT (WT \rightarrow Rag2^{-/-}) mice. Next, each group was divided into 2 subgroups, one of which was given CFA (D2-8/CFA) and the other was not (D2-8). After 7 weeks of parking, the SP cells were harvested and the percentage and absolute number of surviving CD4⁺KJ1-26⁺ cells were determined by flow cytometry and their IFNy responses were measured by ELISA.

Figure S4. CpG increases the percentage of IL-2-producing IFN γ^+ KJ1.26⁺ T cells. Late (D6-8) effectors were isolated from BALB/c mice 3 days after immunization with OVAp/CFA and transferred into Rag2^{-/-} mice. On day 1, 4, 7 and 10 after cell transfer, the hosts were given CpG TLR9 ligand (50µg/injection) i.p. or left untreated. The splenic cells were harvested after 4 months of parking and analyzed for IL-2 and IFN γ production by flow cytometry. (A) Shows the gating strategy for CD3 and KJ1.26 (upper panel) or CD44, KJ1.26 and CD3 (lower panel), followed by analysis of IL-2 and IFN γ production by CD3⁺KJ1.26⁺ and CD44⁺CD3⁺KJ1.26⁺ T cells from the 4-month parking experiments. (B) Shows compiled results for the percentage of KJ1.26⁺ T cells producing IFN γ (KJ1.26⁺IFN γ^+), IL-2 (KJ1.26⁺IL2⁺) or both IFN γ and IL-2 (KJ1.26⁺IFN γ^+). Each bar represents the mean ± SD of four independent experiments. The results indicate that CpG treatment increases the frequency of cytokine-producing memory cells.