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Supplemental Information

Insulin/IGF-1 Controls Epidermal Morphogenesis

via Regulation of FoxO-Mediated p63 Inhibition

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Inventory of supplementary data:

- Supplementary Figure S1 belongs to Figure 1
- Supplementary Figure S1 legend
- Supplementary Figure S2 belongs to Figure 3
- Supplementary Figure S2 legend
- Supplementary Figure S3 belongs to Figure 4
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- Supplementary Figure S5 belongs to Figure 6
- Supplementary Figure S5 legend
- Supplementary Figure S6 belongs to Figure 7
- Supplementary Figure S6 legend
- Supplementary experimental procedures

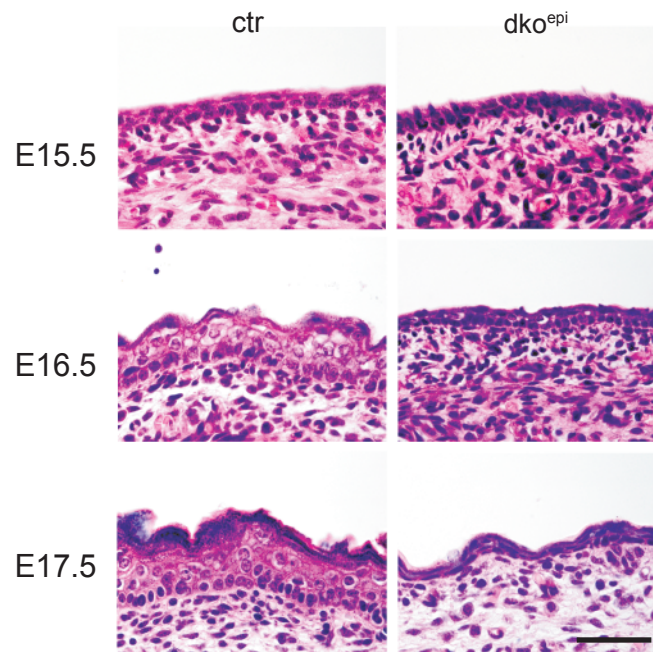
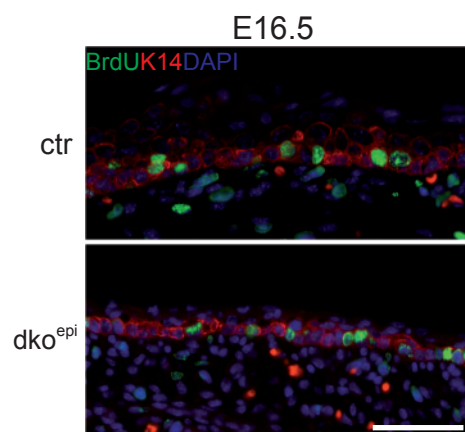
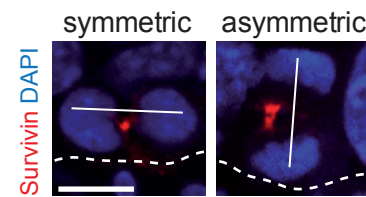
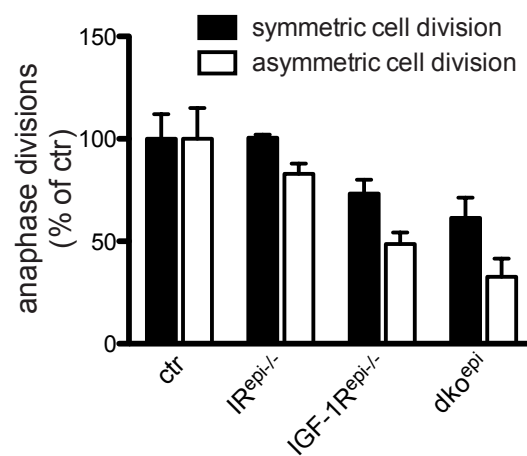
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Figure S1 C.Guenschmann et al.

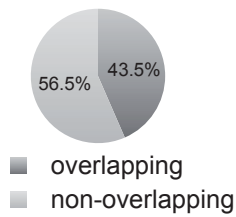
Figure S1 related to Figure 1.

(A) Hematoxylin/Eosin staining of E15.5, E16.5 and E17.5 control and dko^{epi} reveals a reduced number of suprabasal layers in the epidermis in dko^{epi} mice. Scale bar is 100 μm .

(B) Immunofluorescence analysis of keratin 14 (K14, red), BrdU (green) and DAPI-stained nuclei (blue) in E16.5 control and dko^{epi} mice injected with BrdU 1h before sacrificing.

(C) Representative axes of division using the anaphase and telophase marker survivin (red). Dashed line represents the basement membrane. Scale bar is 10 μm .

(D) Quantification of symmetric and asymmetric divisions in control (N=349 divisions), $IR^{epi/-}$ (N=186), $IGF-1R^{epi/-}$ (N=119) and DKO^{epi} (N=118) E16.5 epidermis reveals an increasing loss of asymmetric cell divisions (ACD). Symmetric and asymmetric divisions were set to 100% Quantification was based using DAPI staining to mark anaphase dividing cells and collagen IV as a basement membrane markers with spindles parallel to the basement membrane scored as symmetric and spindles perpendicular scored as asymmetric. (N=5 control embryos, N=3 $IR^{epi/-}$ and N=4 $IGF-1R^{epi/-}$ embryos and dko^{epi} E16.5 embryos, mean \pm SD).

Ap63^{-/-}E18.5 skin vs. p63 kd keratinocytes**B**Common GO terms in arrays of p63^{-/-} E18.5 skin and p63 kd keratinocytes

| GO Term | PValue | Fold Enrichment |
|----------------------------------|----------|-----------------|
| epidermis development | 3,23E-12 | 3,076759789 |
| ectoderm development | 4,81E-12 | 2,963378356 |
| response to organic substance | 1,21E-10 | 1,848474039 |
| response to endogenous stimulus | 9,47E-10 | 2,125876414 |
| regulation of cell proliferation | 2,84E-09 | 1,738414892 |

Unique GO terms in p63^{-/-} E18.5 skin array

| GO Term | PValue | Fold Enrichment |
|-------------------------------|----------|-----------------|
| chemical homeostasis | 4,15E-14 | 2,060255643 |
| response to wounding | 4,85E-13 | 1,990284697 |
| cellular chemical homeostasis | 3,21E-11 | 2,094117647 |
| homeostatic process | 6,52E-11 | 1,7249413 |
| muscle organ development | 9,56E-11 | 2,499646657 |

Unique GO terms in p63 kd keratinocytes array

| GO Term | PValue | Fold Enrichment |
|---------------------------------------|----------|-----------------|
| regulation of programmed cell death | 1,24E-11 | 1,512121392 |
| regulation of apoptosis | 1,37E-11 | 1,51365262 |
| regulation of cell death | 1,80E-11 | 1,506555301 |
| protein localization | 7,50E-10 | 1,44139005 |
| establishment of protein localization | 2,93E-08 | 1,427116006 |

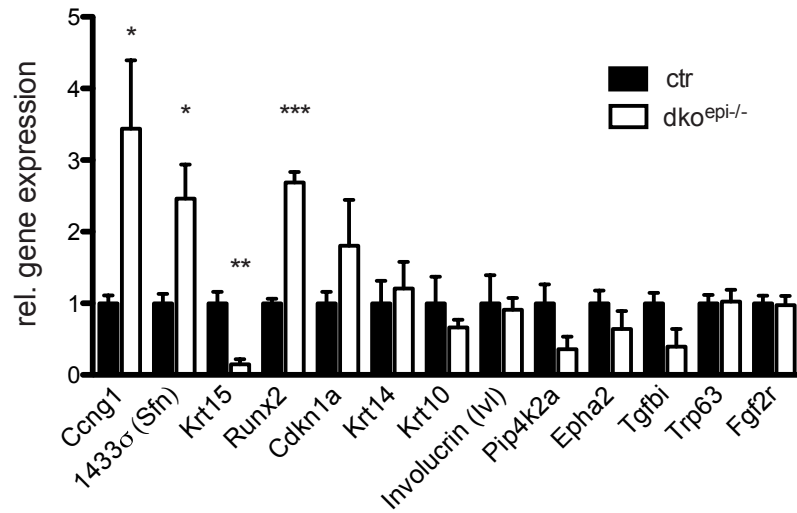
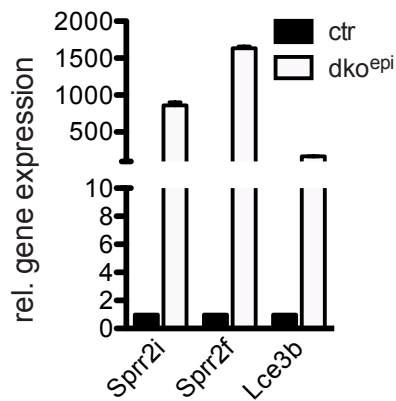
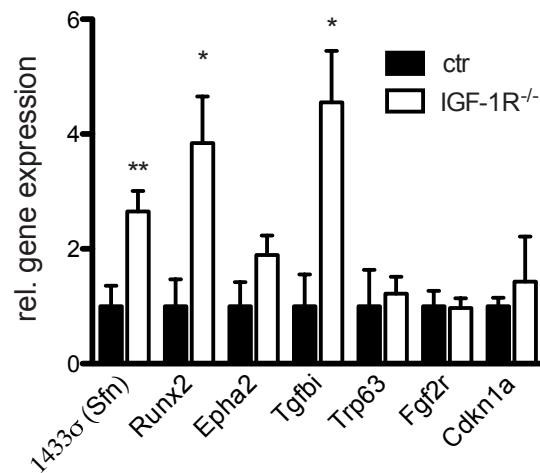
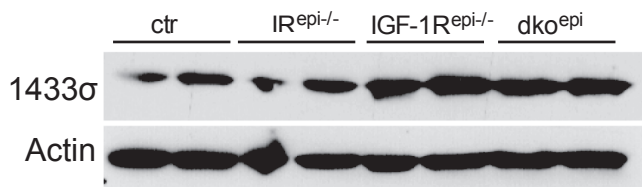
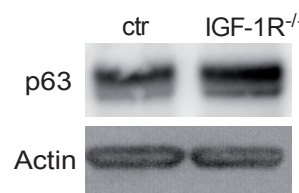
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Figure S2 related to Figure 3.

(A) Pie chart showing the percentage of overlapping and non-overlapping genes of the p63^{-/-} E18.5 skin global gene set with the p63 kd keratinocytes global gene expression set.

(B) GO terms for the overlapping genes from (A) and the unique genes for either the p63^{-/-} E18.5 skin or the p63 kd keratinocyte gene sets.

(C and D) Quantitative real time PCR analysis showing relative gene expression of p63 regulated genes (C) and selected EDC genes (D) in control (ctr) and dko^{epi} newborn epidermis (N=3 newborn epidermis/genotype, mean ± SEM).

(E) Quantitative Real time PCR showing relative gene expression of p63 regulated genes in control (set to 1) and IGF-1R^{-/-} primary cultured keratinocytes (mean of three independently isolated primary keratinocytes/genotype ± SEM).

(F) Increased 14-3-3σ protein expression in newborn epidermis of IR^{epi-/-}, IGF-1R^{epi-/-} and dko^{epi} mice compared to control (ctr). Actin was a loading control.

(G) Western blot analysis for p63 in keratinocytes lysate reveals no change of protein expression upon loss of IGF-1R.

Values in (C), (D) and (E) are means ± SEM and statistical significance was tested by student's T-test and indicated as *P<0.05, **P<0.01, ***p<0.001.

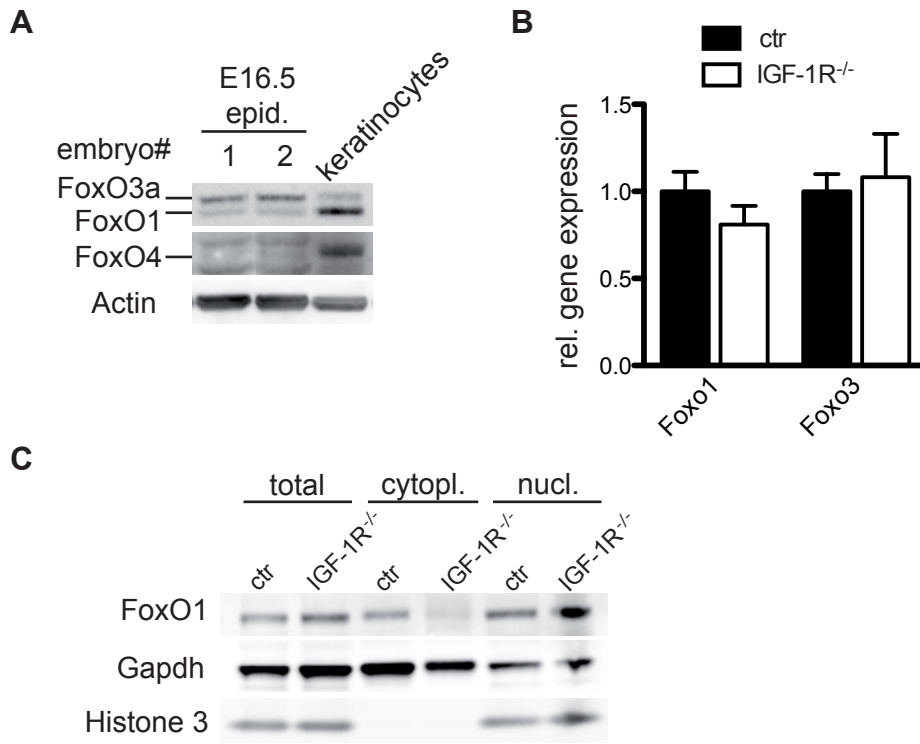


Figure S3 C.Guenschmann et al.

Figure S3 related to Figure 4.

(A) Western blot analysis for FoxO1, 3 and 4 protein in E16.5 epidermis and in primary mouse keratinocytes.

(B) Quantitative real time PCR analysis of E16.5 epidermis reveals no obvious changes in either FoxO1 or Foxo3 expression in IGF-1R^{epi-/-}. Control is set to 1 (mean of N=3 embryos/ genotype \pm SEM). Statistical significance was tested by student's T-test.

(C) Fractionation of keratinocytes shows more FoxO1 in the nucleus upon loss of IGF-1R.

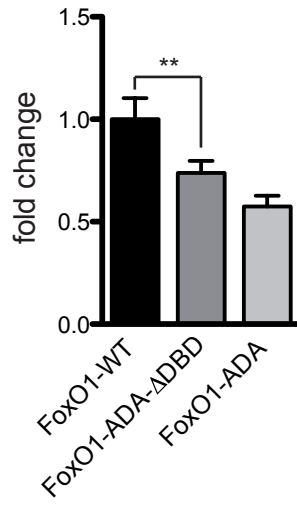
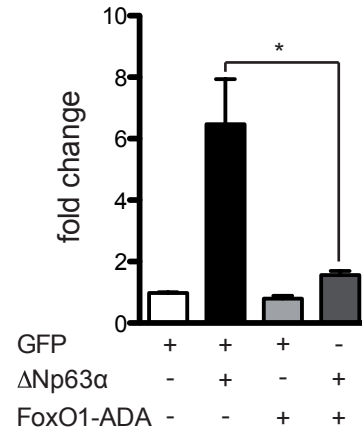
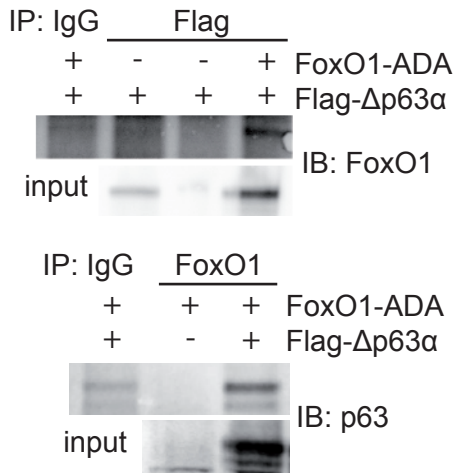
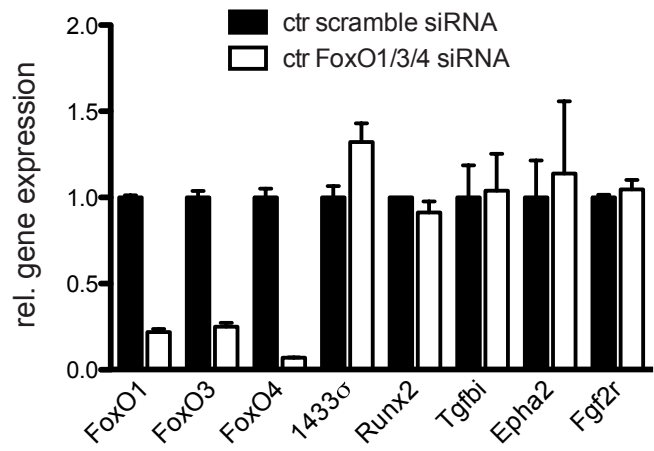
A**B****C****D****Figure S4** C.Guenschmann et al.

Figure S4 related to Figure 5.

(A) Luciferase reporter assay showing that both FoxO1-ADA and the DNA binding deficient Foxo1 mutant (FoxO1-ADA- Δ DBD) repress the activity of the p63-transactivated reporter (BDS-2,3x) in keratinocytes (mean of N=4 independent experiments \pm SEM).

(B) Luciferase reporter assay in CHO cells using the p63-transactivated (BDS-2 (3x)) reporter and co-transfected with GFP and FoxO1-ADA or Δ Np63 α showing that p63 expression is required for FoxO-mediated regulation of the p63 reporter (mean of N=4 independent experiments \pm SEM).

(C) Immunoprecipitation analysis of CHO cells transfected with FoxO1-ADA and Flag- Δ Np63 α shows that FoxO-ADA can immunoprecipitate p63 and, vice versa, Flag-P63 co-precipitates FoxO1-ADA. Shown is one representative experiment of three independent experiments.

(D) Real time PCR analysis on control primary mouse keratinocytes transiently transfected with either scramble or combined knockdown of FoxO1/3/4 using smart pool siRNAs to each of these FoxOs. FoxO1/3/4 knockdown did not alter p63 target expression in control keratinocytes compared to scrambled siRNAs (mean of N=4 independent experiments \pm SEM).

Values in (A), (B) and (D) are means \pm SEM. Statistical significance in (D) and (E) was tested by student's T-test and is indicated as *P<0.05, **P<0.01.

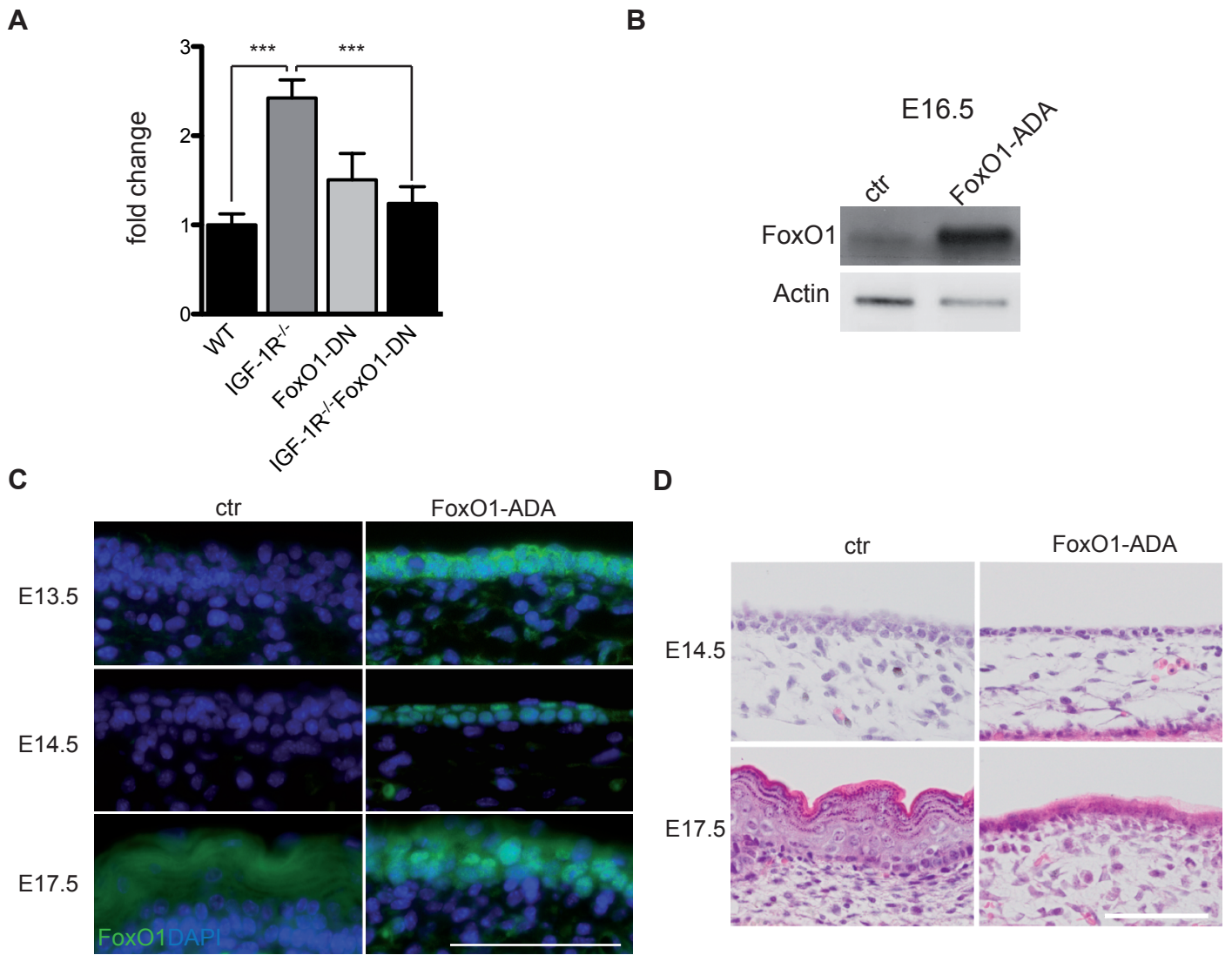


Figure S5 C.Guenschmann et al.

Figure S5 related to Figure 6.

(A) Luciferase reporter assays using the FoxO luciferase reporter (6xDBE) in primary keratinocytes isolated from control, IGF-1R^{epi-/-}, FoxO-DN^{epi} and IGF-1R^{epi-/-} mice show that expression of FoxO-DN represses FoxO transcriptional activity in IGF-1R^{-/-} keratinocytes (N=4 independent experiments, mean \pm SEM). Statistical significance was tested by student's T-test and is indicated as ***P<0.001.

(B) Western blot analysis of FoxO1-ADA on protein lysates of E16.5 control and FoxO1-ADA epidermis.

(C) Immunofluorescence analysis for FoxO1 (green) on skin sections from indicated embryonic stages in control and FoxO1-ADA mice. Nuclei are counterstained with DAPI (blue). Scale bar 50 μ m.

(D) H&E staining on paraffin sections of FoxO1-ADA embryos reveals an hypomorphic epidermis at E14.5. Scale bar is 50 μ m.

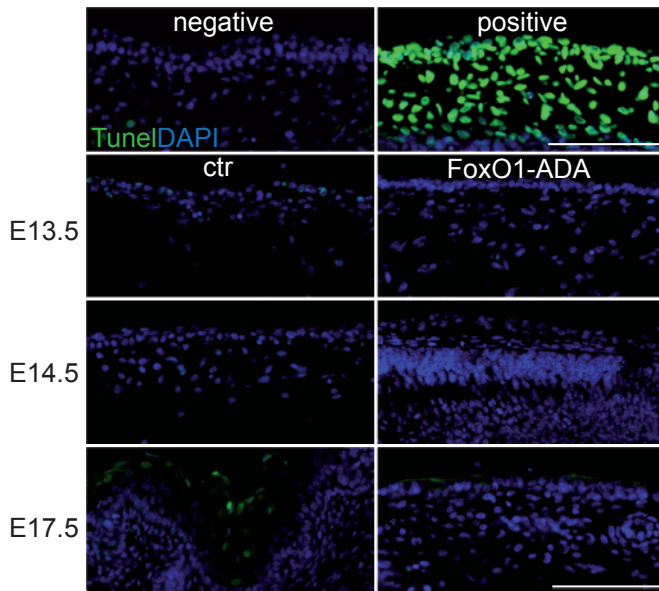
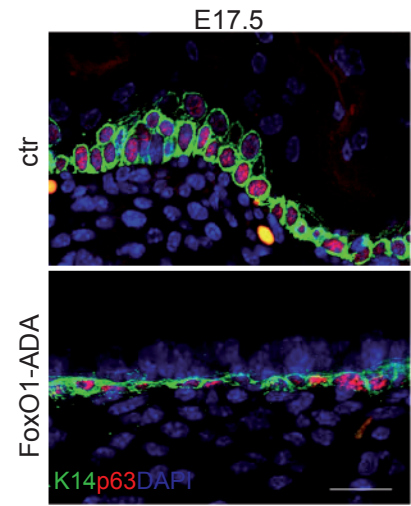
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Figure S6 C.Guenschmann et al.

Figure S6 related to Figure 7.

(A) Apoptosis is not increased in FoxO1-ADA epidermis compared to control at indicated embryonic stages as shown by TUNEL staining (green). Scale bar is 50 μm . Nuclei were counterstained by DAPI (Blue).

(B) Immunofluorescence analysis for p63 (red) and K14 (green) in epidermis of E17.5 FoxO1-ADA mice. Nuclei were counterstained with DAPI (Blue) Scale bar is 10 μm .

Supplementary experimental procedures

Cellular fractionation of primary keratinocytes

Detergent-free lysis of primary keratinocytes was performed as previously described (Maher et al., J. Cell Biol. 186:219). Nuclei were extracted using high salt buffer (20 mM HEPES, pH 7.9, 25% glycerol, 1.5 mM MgCl₂, 1.2 M KCl, 0.2 mM EDTA, 0.5 mM DTT, 0.2 mM PMSF).

Taqman probes used for realtime qPCR

| Gene Symbol | Probe number |
|--------------------|---------------------|
| Foxo1 | Mm00490672_m1 |
| Foxo3 | Mm01185733_m1 |
| Foxo4 | Mm00840140_g1 |
| Foxo6 | Mm00809934_s1 |
| Hprt | Mm00446986_m1 |
| 18s | Mm03928990_g1 |
| Runx2 | Mm00501584_s1 |
| Sfn | Mm01180869_s1 |
| Trp63 | Mm01144752_m1 |
| Krt15 | Mm00492972_m1 |
| Krt14 | Mm00516876_m1 |
| Krt10 | Mm03009921_m1 |
| Ccng1 | Mm00438084_m1 |
| Fgfr2 | Mm01269930_m1 |
| Sprr2f | Mm00448855_s1 |
| Sprr2i | Mm007268832_s1 |
| Lce3b | Mm01333146_g1 |
| Cdkn1a | Mm01303209_m1 |
| Tgfb1 | Mm00493634_m1 |
| Pip4k2a | Mm00435721_m1 |
| Epha2 | Mm00438726_m1 |
| Jag1 | Mm00496902_m1 |

Primary antibodies used for IF, WB or IP

| Antigen | Host | Manufacturer |
|-------------------------|-------------|--|
| Actin (C4) | mouse | MP Biomedicals, Santa Ana, CA |
| Akt1 (C73H10) | rabbit | Cell Signaling Technology, Danvers, MA |
| phospho-Akt (S473)(D9E) | rabbit | Cell Signaling Technology, Danvers, MA |
| BrdU (B44) | mouse | BD Bioscience, Franklin Lakes, NJ |
| FoxO1 (C29H4) | rabbit | Cell Signaling Technology, Danvers, MA |
| FoxO3a (75D8) | rabbit | Cell Signaling Technology, Danvers, MA |
| FoxO4 | rabbit | Santa Cruz Biotechnology, Santa Cruz, CA |
| phospho-FoxO1 (S256) | rabbit | Cell Signaling Technology, Danvers, MA |
| Flag M2 | mouse | Sigma-Aldrich, St Louis, MO |
| GFP | chicken | Abcam, Cambridge, MA |
| IGF-1R β (C20) | rabbit | Santa Cruz Biotechnology, Santa Cruz, CA |
| Keratin 10 | rabbit | Covance, Princeton, NJ |
| Keratin 14 | rabbit | Covance, Princeton, NJ |
| Loricrin | rabbit | Covance, Princeton, NJ |
| p63 α (H129) | rabbit | Santa Cruz Biotechnology, Santa Cruz, CA |
| p63 (4A4) | mouse | Santa Cruz Biotechnology, Santa Cruz, CA |
| Survivin (71G4B7) | rabbit | Cell Signaling Technology, Danvers, MA |
| 14-3-3 σ (N14) | goat | Santa Cruz Biotechnology, Santa Cruz, CA |

List of primers used for Chromatin immunoprecipitation assays

| Gene Symbol | Sense/ Antisense Primer Sequence |
|---|--|
| 1433sigma (Sfn) | CGATGTGGAGAACCAGAGAG CCAATATGTTTGTGGACACCT |
| Cdkn1a | CATGTTTCAGCCCTGGAATTG GTAGTTGGGTATCATCAGGTCTC |
| Runx2 | GACTGTCAGGAGCTGGGAAG GGCCATATAGCCTTGCATCA |
| Fgfr2 (Ferone et al., Embo Mol Med, 2011) | AATGAGCGCGCAAGTTAGAAC GCCGCGCCGAGATGT |
| negative control (Ferone et al., Embo Mol Med, 2011) | ACTCTGACGGATGGCTCTTCA AGGCAGACTTGTGTGGAGATGA |