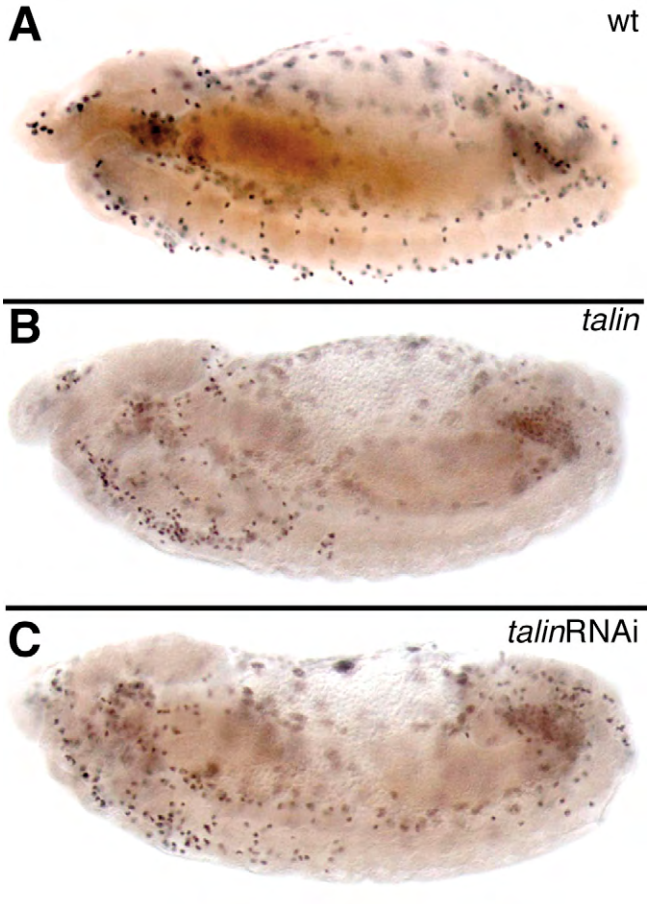

Fig. S1. Talin is required for proper haemocyte migration. (A-C) Lateral view of stage 13 embryos stained with an anti-Srp antibody to label the haemocytes. (A) Wild-type embryo. (B,C) Elimination of both maternal and zygotic Talin (B) or expression of UAS-*talin* RNAi in haemocytes (C) phenocopy the haemocyte migration defects observed in *mys* mutant embryos.

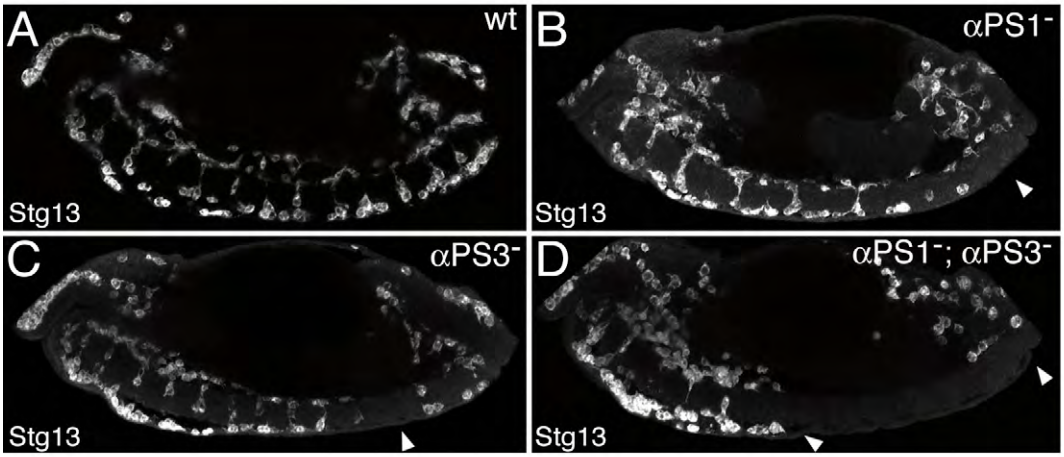

Fig. S2. PS1 and PS3 integrins act redundantly to regulate haemocyte migration. (A-D) Lateral view of stage 13 embryos carrying the combination UAS-CD2/*srph-GAL4* and stained with an anti-CD2 antibody. (A) Wild-type embryo. (B,C) Elimination of either aPS1 (B) or aPS3 (C) causes a small delay in haemocyte migration. (D) However, elimination of both phenocopies loss of bPS function.

Movie 1. Actin and microtubule dynamics in WT haemocytes. Live imaging of WT haemocytes expressing Clip170-GFP, to label microtubules (MTs), and mCherry-Moesin, to label F-actin, migrating randomly in stage 15 embryos. In WT haemocytes, microtubules bundle into an arm and there is a close co-ordination of MT arm disassembly and lamellipodial retraction. Confocal stills were acquired at 30 second time intervals and the movie displayed at 7 frames/second for 30 minutes. Scale bar: 10 mm.

Movie 2. Disrupted migration and microtubule dynamics in stage 15 *mys* mutant haemocytes. *mys* mutant haemocytes expressing Clip170-GFP and mCherry-Moesin. In *mys* mutant haemocytes, microtubules polarised and initially formed an arm, but this structure was not maintained and rapidly collapsed within persisting lamellipodia. In addition, *mys* mutant haemocytes exhibit little migration and remain in close contact throughout imaging. Confocal stills were acquired at 30 second time intervals and the movie displayed at 7 frames/second for 30 minutes. Scale bar: 10 mm.