#### **SUPPLEMENT**

#### NATURE METHODS LipidBlast - in-silico tandem mass spectrometry database for lipid identification

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**Supplementary Figure 1:** Compound structure examples from all 26 lipid classes covered in LipidBlast. File type is MS PowerPoint.

**Supplementary Figure 2**: Investigation of fragmentations and rearrangements of different phospholipid classes from authentic reference compounds using electrospray ion trap tandem mass spectrometry. File type is MS PowerPoint

**Supplementary Figure 3**: Custom modeling of m/z fragments and abundances shown for phosphatidylcholine. The candidate PC 36:2 cannot be identified with the standard LipidBlast libraries. A custom created library allows the subsequent identification of MS/MS spectra obtained from a quadrupole time-of-flight (Q-TOF) instruments. File type is MS PowerPoint.

**Supplementary Figure 4:** Detailed search results of all 134 MS/MS spectra from forty different platforms containing screenshots of pictures from NIST MS Search GUI program. All raw mass spectral files and search statistics are supplied together with the LipidBlast software. File type is MS PowerPoint.

**Supplementary Figure 5:** Validation of LipidBlast with MS/MS spectra obtained from a quadrupole time-of-flight (Q-TOF) instrument. All raw mass spectral files are supplied with the LipidBlast software. File type is MS PowerPoint.

**Supplementary Note 1:** Complete literature reference collection used for LipidBlast development, covering ~300 external literature references. References were used to develop or validate known fragmentations and to manually extract high-resolution tandem mass spectra. File type is MS DOC.

**Supplementary Table 1**: Detailed statistics of the LipidBlast MS/MS libraries with detailed lipid compound numbers, MS/MS spectra numbers, covered adduct types ([M+H]<sup>+</sup>; [M+Na]<sup>+</sup>; [M+N4]<sup>+</sup>; [M+H4]<sup>+</sup>; [M-H]<sup>-</sup>; [M-2H]<sup>(2-)</sup>; [M+NH4-CO]<sup>+</sup>; [M+2Na-H]<sup>+</sup>; [M]<sup>+</sup>; [M-H+Na]<sup>+</sup>; [M+Li]<sup>+</sup>) and histogram statistics of accurate mass values and peak numbers in library. File type is XLS and stored externally.

**Supplementary Table 2**: Complete table of mass spectrometry platforms that can be used with the LipidBlast libraries. Forty different mass spectrometer types from seven major vendors can be used with LipidBlast. The table shows cross-platform independence and the ability of the LipidBlast libraries to identify compounds from different MS platforms. File type is XLS and stored externally.

#### **Online Supplement LipidBlast:**

The LipidBlast software itself and all MS/MS in-silico spectra, all references spectra, compound structure examples as well as all statistical evaluations can be found under:

#### http://fiehnlab.ucdavis.edu/projects/LipidBlast

Questions regarding LipidBlast and software should be directed to: Dr. Tobias Kind (tkind@ucdavis.edu) http://fiehnlab.ucdavis.edu/staff/kind/

Davis, May15 2013





### LipidBlast - In silico created MS/MS libraries for lipid profiling

Supplement of covered structures The structure drawing files (\*.mrv) MarvinSketch can be found under:

http://fiehnlab.ucdavis.edu/projects/LipidBlast

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#### Covered structures and MS/MS spectra in LipidBlast

Num	Lipid class	PIC	Short Name	Number compounds	Number MS/MS spectra	Number MS/MS libraries	
1	Phosphatidylcholines	ok	PC	5,476	10,952	2	
2	Lysophosphatidylcholines	ok	lysoPC	80	160	2	
3	Plasmenylphosphatidylcholines	ok	plasmenyl-PC	222	444	2	
4	Phosphatidylethanolamines	ok	PE	5,476	16,428	3	
5	Lysophosphatidylethanolamines	ok	lysoPE	80	240	3	
6	Plasmenylphosphatidylethanolamines	ok	plasmenyl-PE	222	666	3	
7	Phosphatidylserines	ok	PS	5,123	15,369	3	
8	Sphingomyelines	ok	SM	168	336	2	
9	Phosphatidic acids	ok	PA	5,476	16,428	3	
10	Phosphatidylinositols	ok	PI	5,476	5,476	1	
11	Phosphatidylglycerols	ok	PG	5,476	5,476	1	
12	Cardiolipins	ok	CL	25,426	50,852	2	
13	Ceramide-1-phosphates	ok	CerP	168	336	2	
14	Sulfatides	ok	ST	168	168	1	
15	Gangliosides	ok	[glycan]-Cer	880	880	1	
16	Monoacylglycerols	ok	MG	74	148	2	
17	Diacylglycerols	ok	DG	1,764	3,528	2	
18	Triacylglycerols	ok	TG	2,640	7,920	3	
19	Monogalactosyldiacylglycerols	ok	MGDG	5,476	21,904	4	
20	Digalactosyldiacylglycerols	ok	DGDG	5,476	10,952	2	
21	Sulfoquinovosyldiacylglycerols	ok	SQDG	5,476	5,476	1	
22	Diacylated phosphatidylinositol monomannoside	ok	Ac2PIM1	144	144	1	
23	Diacylated phosphatidylinositol dimannoside	ok	Ac2PIM2	144	144	1	
24	Triacylated phosphatidylinositol dimannoside	ok	Ac3PIM2	1,728	1,728	1	
25	Tetraacylated phosphatidylinositol dimannoside	ok	Ac4PIM2	20,736	20,736	1	
26	Diphosphorylated hexaacyl Lipid A	ok	LipidA-PP	15,625	15,625	1	
Total	All libraries			119,200	212,516	50	



Mass

Abbreviation TG(16:0/18:1(11E)/20:0) Systematic Name 1-hexadecanoyI-2-(11E-octadecenoyI)-3-eicosanoyI-sn-glycerol Formula C57H108O6 888.81



Abbreviation DG(16:0/18:1(11E)/0:0) Systematic Name 1-hexadecanoyl-2-(11E-octadecenoyl)-sn-glycerol Formula C37H70O5 594.52 Mass

Abbreviation MG(16:0/0:0/0:0) Systematic Name 1-hexadecanoyl-sn-glycerol Formula C19H38O4 Mass 330.28

Abbreviation PA(16:0/18:1(11E)) Systematic Name 1-hexadecanoyI-2-(11E-octadecenoyI)-sn-glycero-3-phosphate Formula C37H71O8P Mass 674.49

Abbreviation PE(16:0/18:1(11E)) Systematic Name 1-hexadecanoyl-2-(11E-octadecenoyl)-sn-glycero-3phosphoethanolamine Formula C39H76NO8P Mass 717.53

Abbreviation PG(16:0/18:1(11E)) Systematic Name 1-hexadecanoyl-2-(11E-octadecenoyl)-sn-glycero-3-phospho-(1'-snglycerol) Formula C40H77O10P Mass 748.53



Abbreviation PI(16:0/18:1(11E)) Systematic Name 1-hexadecanoyI-2-(11E-octadecenoyI)-sn-glycero-3-phospho-(1'-myoinositol) Formula C43H81O13P Mass 836.54

Abbreviation PC(16:0/18:1(11E)) Systematic Name 1-hexadecanoyl-2-(11E-octadecenoyl)-sn-glycero-3-phosphocholine C42H82NO8P Formula Mass 759.58





AbbreviationPS(16:0/18:1(11E))Systematic Name1-hexadecanoyl-2-(11E-octadecenoyl)-sn-glycero-3-phosphoserineFormulaC40H76NO10PMass761.52

AbbreviationCL(1'-[14:0/16:0],3'-[14:0/18:1(11E)])Systematic Name1'-[1-tetradecanoyl-2-hexadecanoyl-sn-glycero-3-phospho],3'-[1-<br/>tetradecanoyl-2-(11E-octadecenoyl)-sn-glycero-3-phospho]-sn-glycerolFormulaC71H136017P2Mass1322.93





Common Name	PC(16:0/0:0)[U] (lysoPC)
Systematic Name	1-hexadecanoyI-sn-glycero-3-phosphocholine
Exact Mass	495.33
Formula	C24H50NO7P

LM ID	LMGP01030007 (plasmenyI-PC)
Common Name	PC(P-16:0/18:1(9Z))[U]
Exact Mass	743.58
Formula	C42H82NO7P



LM IDLMGP02050003 (lysoPE)Common NamePE(14:0/0:0)Exact Mass425.25FormulaC19H40NO7P



LM ID	LMGP02030004 (plasmenyl-PE)
Common Name	PE(P-18:0/18:1(9Z))
Exact Mass	729.57
Formula	C41H80NO7P





LM ID	LMSP03010003
Common Name	SM(d18:1/16:0)
Systematic Name	N-(hexadecanoyl)-sphing-4-enine-1-phosphocholine
Synonyms	C16 Sphingomyelin
Exact Mass	702.57
Formula	C39H79N2O6P

LM ID	LMSP02050002 (Ceramide-phosphate)
Common Name	CerP(d18:1/16:0)
Systematic Name	N-(hexadecanoyl)-sphing-4-enine-1-phosphate
Synonyms	C16 CerP
Exact Mass	617.48
Formula	C34H68NO6P



LM ID	LMSP06020002
Common Name	C16 Sulfatide
Systematic Name	(3'-sulfo)Galß-Cer(d18:1/16:0)
Synonyms	C16 Sulfatide
Exact Mass	779.52
Formula	C40H77NO11S



LM ID	LMSP0601AJ01 (GM3 ganglioside)				
Common Name	-				
Systematic Name	NeuAca2-3Galß1-4Glcß-Cer(d18:1/16:0)				
Synonyms	-				
Exact Mass	1152.71				
Formula	C57H104N2O21				



 LM ID
 LMGL05010024

 Common Name
 MGDG(18:2(9Z,12Z)/18:3(9Z,12Z,15Z))

 Systematic Name
 1-(9Z,12Z-octadecadienoyl)-2-(9Z,12Z,15Z-octadecatrienoyl)-3-O-ß-D-galactosyl-sn-glycerol

 Synonyms
 Monogalactosyldiacylglycerol(18:2(9Z,12Z)/18:3(9Z,12Z,15Z))

 Exact Mass
 776.54

 Formula
 C45H76010



### LM ID LMGL05010010 Common Name DGDG(18:5(3Z,6Z,9Z,12Z,15Z)/18:4(6Z,9Z,12Z,15Z)) Systematic Name 1-(3Z,6Z,9Z,12Z,15Z-octadecapentaenoyI)-2-(6Z,9Z,12Z,15Z-octadecatetraenoyI) 3-O-(6'-O-a-D-galactosyl-&D-galactosyl)-sn-glycerol 1 Synonyms Digalactosyldiacylglycerol(18:5(3Z,6Z,9Z,12Z,15Z)/18:4(6Z,9Z,12Z,15Z)) Exact Mass 930.53 Formula C51H78015



LM ID	LMGL05010007
Common Name	SQDG(16:0/16:1(11Z))
Systematic Name	1-hexadecanoyl-2-(11Z-hexadecenoyl)-3-(6'-sulfo-a-D-quinovosyl)-sn-glycerol
Synonyms	sulfoquinovosyldiacylglycerols; SQDG(16:0/16:1)
Exact Mass	791.50
Formula	C41H75O12S



Diacylated phosphatidylinositol monomannoside A Ac2PIM1(16:0/methyl-18:0) C50H95O18P 1014.625603

Ac2PIM1

Ac2PIM2



Diacylated phosphatidylinositol dimannoside Ac2PIM2(16:0/methyl-18:0) C56H105O23P Mass: 1176.678426



Triacylated phosphatidylinositol dimannoside Ac3PIM2(16:0/methyl-18:0/16:0) C72H135O24P 1414.908092

Tetraacylated phosphatidylinositol dimannoside C88H165O25P 1653.137757 Ac4PIM2(16:0/16:0/16:0/methyl-18:0)

Diphosphorylated hexaacyl Lipid A 1825.250692 C96H182N2O25P2 LipidA-PP [14/14/14/14/3O-(14)/3O-(14)]





### LipidBlast - In silico created MS/MS libraries for lipid profiling

Supplement of fragmentation modeling from standard reference compounds. Raw MS/MS Spectra can be found under:

http://fiehnlab.ucdavis.edu/projects/LipidBlast

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### Phospholipids ID **Collection A** Dr. KH LIU **UC Davis Metabolomics** LipidBlast

### **Fragmentation Pattern of Phospholipid on LTQ MS**

PL	(+)	(+)		(-)	(-)	(-) FFA	Formed Adduct
PC	-18 (H+)	-59 (both)	-183 (both)	-74 (Ac <sup>-</sup> )		No	H⁺, Na⁺ Ac⁻
PE	-43 (Na+)	-141 (both)				Yes	H+, Na+ -H+
PS	-87	-185 (both)		-87 (-H+)		Yes	H+, Na+ -H+
SM	-18 (H+)	-59 (Na+)	-183 (Na+)	-74 (Ac <sup>-</sup> )	-145 (Ac <sup>-</sup> )	No	H⁺, Na⁺ Ac⁻
PI	No signal			-162 (-H+)	241*	Yes	-H+
PG	No signal			-74 (-H+)	227*	Yes	-H+
PA	-98 (Na+)	-120 (Na⁺)				Yes	Na⁺ -H⁺

(Na<sup>+</sup>), (H<sup>+</sup>), (Ac<sup>-</sup>): Adduct Ion, Both: (Na<sup>+</sup>) or (H<sup>+</sup>) \* Characteristic Fragment Ion

## Phosphatidylcholine ID





#### *m/z* 782.6 (PC 34:1, Sodium Adduct (16:0, 18:1))

#### - TriMe Ammonium Ion 🤒 PC\_Palmitoyl\_Oleoyl\_Pos\_MSMS\_01 #249-289 RT: 0.52-0.98 AV: 9 NL: 2.90E3 T: Average spectrum MS2 782.60 (249-289) -59.0 +Ņ< O -P OH -PC 2500 2000 -58.9 te 1500-1500-1000 500 -182.7 -H2O -285.1 -60.0 -257.1 -245.0 -205.0 -18.0 -387.1 -341.2 -315.2 -539.4 -499.3 -469.2 -441.3 -402.1 -164.1 -118.0 -82.3 -61.1 58 9.7 0 250 350 400 500 550 600 650 700 750 300 450 m/z PC\_Palmitoyl\_Oleoyl\_Pos\_MSMS\_01 #248-288 RT: 0.51-0.96 AV: 9 NL: 8.79E1 0 T: Average spectrum MS2 760.60 (248-288) 701.4 -16:0-H<sub>2</sub>O 701.3 80-70--18 496.3 60 <del>-</del> 478.4 Aise atu -18:1-H<sub>2</sub>O 577.5 742.5 -16:0 30-478.1 522.3 700.4 20-742.2 522.5 577.1 10-741.5 743.0 577.8 601.3 645.3 660.5 688.5 281.0 536.6 576.4 227.1 250.9 401.1 416.2 343.1 356.1 445.1 311.3 0-250 300 350 400 450 500 550 600 650 700 750 200

Pos. Mode

#### *m/z* 782.6 (PC 34:1, Sodium Adduct (16:0, 18:1))





Pos. Mode

*m/z* 818.6 (PC 34:1, Acetate Adduct (16:0, 18:1))





#### *m/z* 818.6 (PC 34:1, Acetate Adduct (16:0, 18:1))



Neg. Mode

#### PC – Bovine brain



090325\_PCs\_Pos\_CE25\_MSMS\_01 #45-214 RT: 0.09-0.43 AV: 170 NL: 5.20E3 T: ITMS + p ESI Full ms[300.00-1100.00]





# Sphingomyelin ID





*m/z* 729.6 (SM 36:1 (18:1))



Pos. Mode



*m/z* 787.6 (SM 36:1, Acetate Adduct (18:1))







Neg. Mode

#### *m/z* 787.6 (SM 36:1, Acetate Adduct (18:1))



## Phosphatidylserine ID





#### *m/z* 762.6 (PS 34:1 (16:0, 18:1))

#### Pos. Mode



#### *m/z* 784.5 (PS 34:1, Sodium Adduct (16:0, 18:1))



Pos. Mode

#### *m/z* 760.6 (PS 34:1 (16:0, 18:1))

Neg. Mode





#### *m/z* 760.6 (PS 34:1 (16:0, 18:1))

#### Neg. Mode



### Phosphatidylethanolamine ID

#### *m/z* 718.6 (PE 34:1 (16:0, 18:1))





#### *m/z* 718.6 (PE 34:1 (16:0, 18:1))



*m/z* 740.6 (PE 34:1, sodium adduct (16:0, 18:1))


*m/z* 716.7 (PE 34:1 (16:0, 18:1))





#### *m/z* 716.7 (PE 34:1 (16:0, 18:1))



### Phosphatidylinositol ID





*m/z* 861.6 (PI 36:2 (18:1, 18:1))



Exact Mass: 241.01

Neg. Mode

#### *m/z* 883.1 ?? (PI 36:1 (18:1, 18:1))





### Phosphatidic acid ID







Neg. Mode

#### *m/z* 697.5 (PA 34:1, Sodium Adduct (16:0, 18:1))



Pos. Mode

## Phosphatidylglycerol ID

*m/z* 747.7 (PG 34:1 (16:0, 18:1))

### Neg. Mode









Neg. Mode

*m/z* 749.7 (PG 34:1 (16:0, 18:1)) M+H





Pos. Mode

# Phospholipids ID Collection B

### Dr. KH LIU UC Davis Metabolomics LipidBlast

#### DLOPC (18:1, 18:1, diether form, *m/z* 758.8 [M+H]+)



#### DLOPC (18:1, 18:1, diether form, *m/z* 780.8 [M+Na]+)



#### LPC (14:0, *m/z* 468.6 [M+H]+)

090623\_LPL3Mix\_2\_5ug\_ml\_Pos\_CE35\_01 #38-101\_RT: 0.15-0.97\_AV: 3\_NL: 1.22E2

#### T: Average spectrum MS2 468.37 (38-101)

#### Positive mode 🧟



090623\_LPL3Mix\_2\_5ug\_ml\_Pos\_CE35\_01 #54 RT: 0.35 AV: 1 NL: 5.21E1 T: ITMS + p ESI d Full ms2 490.35@cid35.00 [125.00-505.00]



#### LPC 16:0 (m/z 496.4, [M+H]+)

090708\_LPCs\_EY\_Pos\_CE25\_02 #56-117 RT: 0.11-0.86 AV: 4 NL: 1.39E4 T: Average spectrum MS2 496.46 (56-117)

#### Positive mode 🧕



EY\_Pos\_CE25\_02 #56-117 RT: 0.11-0.86 AV: 4 NL: 1.96E2 ctrum MS2 496.46 (56-117)



#### Plasmalogen PC (P-18:0, 18:1, *m/z* 772.7 [M+H]+)



#### Plasmalogen PE (P-18:0, 18:1, *m/z* 730.7 [M+H]+)



#### Plasmalogen PE (P-18:0, 18:1, *m/z* 752.7 [M+Na]+)



- Positive mode
- Characteristic ion of plasmalogen PE
  - P-20:0: m/z 442
  - P-20:1: m/z 440
  - P-18:0: m/z 414
  - P-18:1: m/z 412
  - P-16:0: m/z 386
  - P-16:1: m/z 384

#### Plasmalogen PE (P-18:0, 18:1, *m/z* 728.6 [M-H]-)



#### LPE (18:0, *m/z* 482.4 [M+H]+)



LPE (18:0, *m/z* 504.4 [M+Na]+)



#### LPE (16:0, *m/z* 452.3 [M-H]-)



#### LPE (18:0, *m/z* 480.5 [M-H]-)



All LPE: m/z 196 and m/z 214

#### LPE (16:0, *m/z* 454.4 [M+H]+)



LPE (16:0, *m/z* 476.4 [M+Na]+)



#### DMPS (14:0, 14:0, *m/z* 680.6 [M+H]+)



#### DMPS (14:0, 14:0, *m/z* 702.6 [M+Na]+)



#### DMPS (14:0, 14:0, *m/z* 678.6 [M-H]-)





POPS (16:0, 18:1, *m/z* 760.7 [M-H]-)





#### SM 38:0 (d18:0, 20:0 (N-Acyl), m/z 761.7, [M+H]+)



#### SM 38:0 (d18:0, 20:0 (N-Acyl), m/z 783.6, [M+Na]+)



#### SM 36:1 (d18:0, 18:1 (N-Acyl), m/z 731.6, [M+H]+)



0 090708\_SMs\_BB\_Pos\_CE35\_01 #65-122 RT: 0.23-0.98 AV: 2 NL: 7.97E3 - Choline T: Average spectrum MS2 753.57 (65-122) 694.45 100-80 Relative Abundance 60-40-- PC 20-570.55 723.00 736.00 233,27 258,82 283.09 309.27 339.91 379.36 411.09 452.82 481.45 507.55 548.64 612,45 628.55 650.45 692.09 O٠ 200 250 300 350 400 450 500 550 600 650 700 750 m/z



SM 34:1 (d18:1, 16:0)

TG 54:0 (17:0, 17:0, 17:0, m/z 866.8, [M+NH<sub>4</sub>]+)



#### TG 48:0 (16:0, 16:0, 16:0, m/z 824.7, [M+NH<sub>4</sub>]+)



#### DG 32.0 (16:0, 16:0, m/z 586.6, [M+NH<sub>4</sub>]+)

090814\_DG\_DP\_Pos\_CE35\_MSMS\_400\_1000\_01#64-125\_RT: 0.11-0.83\_AV: 4\_NL: 1.66E3 T: Average spectrum MS2 586.57 (64-125)





#### Cardiolipin 14:0 (CL 14:0, m/z 620, [M-2H]2-)



Exact Mass: 227.2 0






- Negative mode
- Characteristic ion
  - m/z 1448: [M-H]-
  - m/z 831.5: loss of diacyl group
  - m/z 751.6: loss of diacyl + 80
  - m/z 695.6: loss of diacyl + 136
  - m/z 433.3: acyl chain information
  - m/z 415.3: acyl chain information

### ✓ Positive mode: No signal

### Cardiolipin 18:2 (CL 18:2, m/z 724, [M-2H]2-)



m/z

Negative mode

Relative Abundance

- Characteristic ion
  - m/z 724: [M-2H]2-
  - m/z 695.4
  - m/z 593.1: loss of monoacyl group
  - m/z 415.5: loss of diacyl group
  - m/z 279.6: 18:2

# Li Adduct

By adding LiOH in reconstitution solvent (Final concentration of Li: 2 mM)

### PLPC (16:0, 18:1, m/z 760.6, [M+H]+)



### PLPC (16:0, 18:1, m/z 766.6, [M+Li]+)



### Identified Lithium Adduct of Phosphatidylcholine

- Lithium adduct: Acyl chain information
- Sodium adduct: no acyl chain information

	PL	Acyl chain	[MH]+	[M+Li]+	PL	Acyl chain	[MH]+	[M+Li]+
F	PC 34:2	16:0, 18:2	758.7	764.7	PC 36:2	18:0, 18:2	786.7	792.7
F	PC 34:1	16:0, 18:1	760.7	766.7	PC 36:1	18:0, 18:1	788.7	794.7
F	PC 34:0	16:0, 18:0	762.7	768.7	PC 38:7	16:1, 22:6	804.6	810.6
F	PC 36:6	16:1, 20:5	778.7	784.7	PC 38:6	16:0, 22:6	806.6	812.6
F	PC 36:5	16:0, 20:5	780.7	786.7	PC 38:5	18:1, 20:4	808.6	814.6
F	PC 36:4	18:2, 18:2	782.7	788.7	PC 38:4	18:0, 20:4	<mark>810</mark> .6	816.6
F	PC 36:3	18:1, 18:2	784.7	790.7	PC 38:3	18:0, 20:3	<b>812</b> .6	818.6

## Identification of major PL and NL from standard blood

m/z	PL/TG	m/z	PL/TG	m/z	PL/TG	m/z	PL/TG	m/z	PL/TG
468.5	LPC 14:0	614.5	DG 34:0 NH4+	780.7	PC 34:2 Na	850.6	TG 50:1 NH4+	904.6	TG 54:2 NH4+
480.5	LPC P-16:0	638.5	DG 36:2 NH4+	782.7	PC 34:1 Na	852.6	PC 39:1 Na	906.5	TG 54:1 NH4+
482.5	LPC 15:0	640.5	DG 36:1 NH4+	784.7	PC 34:0 Na	854.6	PC 39:0 Na	908.6	TG 54:0 NH4+
494.7	LPC 16:1	642.5	DG 36:0 NH4+	786.7	PC 36:2	856.6	PC 40:6 Na	916.5	TG 56:10 NH4+
496.6	LPC 16:0	666.5	DG 38:2 NH4+	788.7	PC 36:1	858.6	PC 40:5 Na	918.5	TG 56:9 NH4+
508.7	LPC P-18:0	668.5	DG 38:1 NH4+	794.6	PC 35:2 Na	860.6	PC 40:4 Na	920.6	TG 56:8 NH4+
510.6	LPC 17:0	669.4	SM 30:1 Na	796.6	PC 35:1 Na	862.6	PC 40:3 Na	922.6	TG 56:7 NH4+
518.5	LPC 18:3	670.5	DG 38:0 NH4+	798.6	PC 35:0 Na	864.6	PC 40:2 Na	924.6	TG 56:6 NH4+
520.6	LPC 18:2	671.6	SM 30:1 Na	800.7	PC 36:6 Na	866.6	PC 40:1 Na	926.6	TG 56:5 NH4+
522.6	LPC 18:1	675.6	SM 32:1	802.7	PC 36:5 Na	868.6	PC 40:0 Na	928.6	TG 56:4 NH4+
524.6	LPC 18:0	677.5	SM 32:0	804.7	PC 36:4 Na	870.6	TG 52:5 NH4 <sup>+</sup>	930.6	TG 56:3 NH4+
526.6	LPE 22:6	697.4	SM 32:1 Na	806.6	PC 36:3 Na	872.7	TG 52:4 NH4+	932.6	TG 56:2 NH4+
536.7	LPC P-20:0	699.4	SM 32:0 Na	808.7	PC 36:2 Na	874.6	TG 52:3 NH4+	934.6	TG 56:1 NH4+
538.6	LPC 19:0	703.7	SM 34:1	810.7	PC 36:1 Na	876.6	TG 52:2 NH4+	936.6	TG 56:0 NH4+
542.5	LPC 20:5	705.8	SM 34:0	812.7	PC 36:0 Na	878.6	TG 52:1 NH4+	938.6	TG 58:12 NH4+
544.6	LPC 20:4	725.5	SM 34:1 Na	814.7	PC 37:6 Na	880.6	TG 52:0 NH4+	940.6	TG 58:11 NH4+
546.6	LPC 20:3	727.5	SM 34:0 Na	820.6	PC 37:3 Na	886.6	TG 53:4 NH4+	942.6	TG 58:10 NH4+
548.6	LPC 20:2	753.3	SM 36:1 Na	822.6	PC 37:2 Na	888.5	TG 53:3 NH4+	946.6	TG 58:9 NH4+
550.9	LPC 20:1	755.1	SM 36:0 Na	824.6	PC 38:8 Na	890.6	TG 54:9 NH4+	952.6	TG 58:6 NH4+
552.9	LPC 20:0	756.5	PC 34:3	826.6	PC 38:7 Na	892.6	TG 54:8 NH4+	954.6	TG 58:5 NH4+
568.6	LPC 22:6	758.7	PC 34:2	832.6	PC 38:4 Na	894.6	TG 54:7 NH4+	956.6	TG 58:4 NH4+
570.6	LPC 22:5	760.7	PC 34:1	834.7	PC 38:3 Na	896.6	TG 54:6 NH4+	958.6	TG 58:3 NH4+
578.6	LPC 22:1	762.7	PC 34:0,	836.7	PC 38:2 Na	898.7	TG 54:5 NH4+	960.6	TG 58:2 NH4+
610.5	DG 34:2 NH4+	768.7	PE 38:4	846.6	TG 50:3 NH4+	900.7	TG 54:4 NH4+	986.6	TG 60:3 NH4+
612.5	DG 34:1 NH4+	774.6	PE 32:1	848.7	TG 50:2 NH4+	902.7	TG 54:3 NH4+		

Reconstitution solvent: Chloroform/Methanol (1/9) containing 7.5 mM ammonium acetate





### LipidBlast - In silico created MS/MS libraries for lipid profiling

### Custom modeling for other instruments

http://fiehnlab.ucdavis.edu/projects/LipidBlast

Tobias Kind, Oliver Fiehn FiehnLab – Metabolomics UC Davis Genome Center, Davis, USA Supplement: Custom creation of LipidBlast libraries New PC library for QTOF Time to create new library <10 min using EXCEL templates.

### Q-TOF spectrum of PC 36:2; [M+H]+; 786.5987 can not be found in standard LipidBlast library, due to missing fragments in the spectrum. A custom library can be created to model specific m/z values (next slide).

Source MS/MS data Agilent 6530 QqTOF; [M+H]+;

Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Sandra K, Pereira Ados S, Vanhoenacker G, David F, Sandra P.; Journal of Chromatography A, 1217 (2010) 4087-4099

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#   Src.   Name     22   A   Agilent 6410 triple quadrupole MS; [M-H]-;     23   A   Agilent 6520 Q-TOF; [M-H]-;     24   A   Agilent 6530 QqTOF; [M+H]+;     25   A   Agilent 6530 QqTOF; [M+H]+;     26   A   Agilent 6530 QqTOF; [M+H]+;     27   A   Agilent 6530 QqTOF; [M+H]+; PC/PE mix     27   A   Agilent 6530 QqTOF; [M+H]+; TAG     28   A   Agilent 6530 QqTOF; [M+N]+; TAG     29   A   Agilent LC/MSD 1100 Ion Trap; [M-H];     30   A   Agilent Ion Trap SL; [M+Na]+; <b>V</b> Image: A   Spec List	100-   786.5987     50-   ?     20   330   440   550   660   770     (Text File) Agilent 6530 QqTOF; [M+H]+;   .   .   .   .     Plot/Text of Search Spectrum A   Plot of Search Spectrum A   Plot/Text of Spec List /   .   .
lipidblast-neg; lipidblast-pos; 212516 total spectra     #   Library     Score   Dot Product   Prob. (%)     Rev-Dot   Name	
Names (Structures / Hit List	Plot/Text of Hit / Plot of Hit /
Lib, Search Other Search Names Compare Librarian	

Peptide

Peptide

### The custom LipidBlast library correctly identifies PC 36:2 from MS/MS data of a Q-TOF mass spectrometer.

### For QTOFs at selected voltage only precursor and product ion m/z 184 are observed.

Source MS/MS data:

Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Sandra K, Pereira Ados S, Vanhoenacker G, David F, Sandra P.; Journal of Chromatography A, 1217 (2010) 4087-4099

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60	<b> </b> 🖉   🖬	🛛 🛛 1. Agile	nt 6530	QqTOF; [M	+H]+;	- <b>B</b>	<b>L</b> . 🔎	0												
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23	A A	gilent 6520 Q-	TOF; [M-I	H]-;	•								786.5987	2 largest peak	36:2 ; [M+ <mark>8:</mark>	H]+; 786.598	37; Comprei	nensive bloc	ia piasma i	.piao
24		gilent 6530 Qa gilent 6530 Qa	TUF; [M TOF: [M	+HCUUJ-; +H1+:										184.0728 99	9.00   78	6.5987 800.	00			
26	A A	gilent 6530 Qo	TOF; [M·	+H]+; PC/PE r	nix				50-		4			no synonyms:						
27		gilent 6530 Qo ailant 6520 Qo	TOF; [M-	+H]+; PE+PC	MIX															
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30	A A	gilent Ion Trap	SL; [M+I	Na]+;				-	ľ	220 330	440 550	660	770			_				
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1	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+	Differe	Agilent 6530 Qo	(TOF; [M+H]+; it A Side by Side	λ Subtra	Head to	Tail MF=7 RMF=7	37	I▼PC 36:	2; [M+H]+; I	GPCho(18:1	(11E)/18:1	(11) 1 P
2	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+.		nce V Head to Ta		V_ocona							73711 0.6	11
3	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+.						Name: PC 36:	2; [M+H]+	; GPCho(18:1	/(11E)/18:1	(11E))		-
	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+.	1004					Comment: Par	2009 <u>06</u> ent=786.6	0126 Mz_exa	act=786.60	126 ; PC 36:	2; [M+H]+,	;
C C	custompo	+npos.msp +hpos.msp	7	737 737	0.61	737	PC 36(2) PC 36(2)	[M+H]+. [M+H]+						7 largest peak	<u>s:</u> 00.001 =	-	0.001.500	05010,000		
7	custompo	+hpos.msp	7	737	0.61	737	PC 36:2:	[M+H]+						786.60126 2	99.0015 00.0017	04.34554 2L 68.59070 1	70.00   522 10.00	.30610-200	.001-603.5	).
8	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+						7 m/z Values a	and Intens	ities:				
9	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+	50-	1 1				184.07387 99	19.00 frag	ment C5H15 H1.op1-H20-J	NO4P II MuHlaní	2.420		
10	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+.						522.35610 20	0.00 [M+	H]-sn1    [M+l	H]-sn2	5120		
11	custompo	+hpos.msp	7	737	0.61	737	PC 36:2;	[M+H]+.			504.34554	727	7.52776	603.53522 20	0.00 (M+	H]-C5H14NC	14P (-183)			
12	custompo	+hpos.msp	7	/37	0.61	737	PC 36:2;	[M+H]+.						768.59070 10	0.00 [M+ .00 [M+	nj-congn (-: H1-H20 (-18)	10)			
13	custompo	+npos.msp +bpos.msp	7	737	0.61	737	PC 36:2; PC 36:2;	[™I+FI]+. [M+H]+ ▼I	ــلە 🛛	220 220	440 550		770	786.60126 20	0.00 (M+	H])				Ţ
	custompt	прозтвр	r	1.51	0.01	rur	r C 30.2,	Po *0 (*. ≦	foustom	220 330 ne+hnos msn) PC	36:2: [M+H]+: GP	000 Cho(18:1)	11E1/18-10							۲
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Peptide

Peptide

Supplement: Custom creation of LipidBlast libraries New formate adduct library [M+HCOO]-Time to create new library <10 min using EXCEL templates.

### Q-TOF spectrum of PC 36:2 ; [M+HCOO]-; m/z 830.5966 is falsely identified as PE (with low hit score). A custom library can be created to model specific m/z values (next slide).

Source MS/MS data Agilent 6530 QqTOF; [M+H]+;

Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Sandra K, Pereira Ados S, Vanhoenacker G, David F, Sandra P.; Journal of Chromatography A, 1217 (2010) 4087-4099



#### Q-TOF spectrum of PC 36:2; [M+HCOO]-; m/z 830.5966 is correctly identified using the custom format pc-form-neg library.

Source MS/MS data Agilent 6530 QqTOF; [M+H]+;

Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Sandra K, Pereira Ados S, Vanhoenacker G, David F, Sandra P.; Journal of Chromatography A, 1217 (2010) 4087-4099

NIST MS Search 2.0 - [Peptide, Presearch Default - 394 spectra]	
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#     Src.     Name       22     A     Agilent 6410 triple quadrupole MS; [M-H]-;       23     A     Agilent 6520 Q-T0F; [M-H]-;       24     A     Agilent 6530 QqT0F; [M+HC00]-;       25     A     Agilent 6530 QqT0F; [M+H]+;       26     A     Agilent 6530 QqT0F; [M+H]+;       27     A     Agilent 6530 QqT0F; [M+H]+;       28     A     Agilent 6530 QqT0F; [M+H]+;       29     A     Agilent 6530 QqT0F; [M+N]+;       30     A     Agilent Ic/MSD 1100 Ion Trap ; [M+N]+;	100-   770.5758     50-   ?     50-   ?     830.5966   830.5966     300   400   500   600   700     300   400   500   600   700   800     (Text File) Agilent 6530 QqTOF; [M+HCOO];   Intersities:   279.2344   200.00   283.2645   100.00   770.5758   999.00   830.5966   25     Synonyms:   no synonyms: <t< td=""></t<>
Names Structures Spec List	Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /
lipidblast-neg; lipidblast-pos; custompc+hpos.msp; custompc+napos.msp; msmslib; pc-ac-neg.msp; pc-form-neg.msp; 235420 total spectra 1000- 100- 10- 1-	100- 770.5758   50- 279.2344   0- 4   279.23226 830.59110
	- 100- 770.56997
#     Library     Score     Dot Product     Prob. (%)     Rev-Dot     Name       1     pc-form-neg.msp     683     991     7.54     992     PC 36:2; [M+HCC       2     pc-form-neg.msp     683     991     7.54     992     PC 36:2; [M+HCC	280     320     360     400     440     480     520     560     600     640     680     720     760     800     840       Agilent 6530 QqT0F; [M+HC00];     I     Head to Tail MF=683 RMF=991     I ▼PC 36:2; [M+HC00]; GPCho(18:0/18:2/18:2/18:2/18:2/18:2/18:2/18:2/18:2
3     pc-form-neg.msp     683     991     7.54     992     PC 36.2; [M+HCl       4     pc-form-neg.msp     683     991     7.54     992     PC 36.2; [M+HCl       5     pc-form-neg.msp     683     991     7.54     992     PC 36.2; [M+HCl       5     pc-form-neg.msp     683     991     7.54     992     PC 36.2; [M+HCl	Name:     PC 36:2; [M+HC00]; GPCho(18:0/18:2(2E,4E))       100-     MW:     830 ID#; 2563 DB; pc-form-neg.msp       Comment:     Parent=830.59110 Mz_exact=830.59110 ; PC 36:2; [M+HC00];       4 largest peaks:     Parent=830.59110 Mz_exact=830.59110 ; PC 36:2; [M+HC00];
6     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU       7     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU       8     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU       9     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU       9     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU       10     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU       11     pc-form-heg.msp     683     991     7.54     992     PC 36.2; [M+HCU	50-     ?     279.23226 100.00   283.26354 100.00   830.5911       4 m/2 Values and Intensities;     279.23226 100.00 FA sn2       283.26354 100.00 FA sn1     279.23226 100.00 FA sn1       770.56997 999.00 [M-CH3]- (-15)     830.59110 100.00 [M+HC00]- (M+44.9976)
12   pc-form-neg.msp   683   991   7.54   992   PC 36:2; [M+HCl     13   pc-form-neg.msp   83   838   0.07   951   PC 36:2; [M+HCl     14   pc-form-neg.msp   83   838   0.07   951   PC 36:2; [M+HCl     ▲   ▲   ▲   ▲   ▲   ▲   ▲     Names   Structures   ✓   Hit List   ➡	O 300 400 500 600 700 800 (pc-form-neg.msp) PC 36:2; [M+HCDO]-; GPCho(18:0/18:2(2E,4E) Plot/Text of Hit / Plot of Hit /
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	Peptide Peptide





### LipidBlast - In silico created MS/MS libraries for lipid profiling

Supplement data mass spectral search All raw files and software can be found under:

http://fiehnlab.ucdavis.edu/projects/LipidBlast

Tobias Kind, Kwang-Hyeon Liu, Do Yup Lee, Oliver Fiehn FiehnLab – Metabolomics UC Davis Genome Center, Davis, USA The MS/MS search results are shown here for peer review and proof of concept. The complete set of the electronic 134 spectra (\*.MSP) is found in the supplement section.

All machine readable spectra and all supplement data can be found at: <u>http://fiehnlab.ucdavis.edu/projects/LipidBlast</u>

For questions please contact the curator: Dr. Tobias Kind <a href="http://fiehnlab.ucdavis.edu/staff/kind/">http://fiehnlab.ucdavis.edu/staff/kind/</a>

### NIST MS Search GUI – Search parameters

NIST MSPepSearch mass spectral library search program Written by Dmitrii V. Tchekhovskoi and Stephen E. Stein Mass Spectrometry Data Center National Institute of Standards and Technology (NIST)

Visit http://peptide.nist.gov or http://chemdata.nist.gov for more information, software, or MS libraries.

#### About NIST MS Search 2.0

The NIST Mass Spectral Search Program for the NIST/EPA/NIH Mass Spectral Library Version 2.0 f, build Dec 3 2009 X

Software by S. Stein, Y. Mirokhin, D. Tchekhovskoi, and G. Mallard. Data Evaluation by A. Mikaia, V. Zaikin, J. Little, Damo Zhu, E. White and D. Sparkman.

NIST uses its best effort to deliver a high quality copy of the database and to ensure that the data shown are accurate. However, NIST makes no warranties to that effect,



### This is a special compiled version

NIST <u>MS Search 2.0</u> The built date must be minimum May 2010 Version 2.0g 2011 produces slightly different hit scores.

### The latest NIST.EXE is available from

http://peptide.nist.gov/ Search scores can change slightly with new versions. NIST MS Search is an independent product from LipidBlast.



**4000 Q-Trap mass spectrometer; [M-H]-**Ceramide-phosphate CerP(d18:1/12:0); [M-H]-; Prec. m/z: 874.8; LipidMaps http://www.lipidmaps.org/data/standards/standards.php?lipidclass=LMSP

SNIST MS Search 2.0 - [Peptide, Presearch Default - 39 spectra]	
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å 🖻 🖻 🚑 🛄 📴 📲 🖂 #/z ← 💡	
🚳 🎾 🚔 📮 1. ABI 4000 Q-Trap; [M-H]-; CerP(d18:1/ 💌 🛞 🖳 😥 🔍	
#     Src.     Name       1     A     ABI 4000 Q-Trap; [M-H]; CerP(d18:1/12:0)       2     A     ABI 4000 Q-Trap; [M-H]; CerP(d18:1/12:0)       3     A     ABI QTRAP 4000; [M-H]; Lipid A (diPP-14-hexaacyl)       4     A     ABI 4000 Q-Trap; [M-H]; Pl 36:1       5     A     ABI 4000 Q-Trap; [M-H]; GM2(d18:1/C18:0)       7     A     ABI-API 4000 QTrap; [M-H]; GM2(d18:1/C18:0)       8     A     ABI 4700 MALDI-TOF; [M-H]; GM2(d20:1/C18:0)       8     A     ABI 4700 MALDI-TOF; [M-H]; GM2(d18:1/N124:1)       10     A     ABI 4700 MALDI-TOF; [M-H]; Sulfatide(d18:1/N124:1)       11     A     ABI 4700 MALDI-TOF/TOF; [M-H]; Lipid A       10     A     ABI 4700 MALDI-TOF/TOF; [M-H]; Sulfatide(d18:1/N124:1)       11     A     ABI API 2000 triple quadrupole; [M-H]; CL 76:12       4     ABI 000 CTADEX D- actional DOF/TOF: M-H]; Sulfatide(d18:1/N124:1)       11     A     ABI API 2000 triple quadrupole; [M-H]; CL 76:12       14     ABI 000 CTADEX D- actional DOF/TOF/TOF/TOF/TOF/TOF/TOF/TOF/TOF/TOF/T	100-   97.0     100-   378.2     0   378.2     0   378.2     0   378.2     0   370.0     280   560     (Text File) ABI 4000 Q-Trap;     Values and Intensities:     97.0
10-	50-
# Library Score Dot Product Prob. (%) Rev-Dot Name	ABI 4000 Q-Trap; [M-H]; CerP(d1   Head to Tail MF=149 RMF=761   ▼CerP 30:1; [M-H]; CerP(d18:1(4E)
1 lipidblast-neg 149 761 63.8 762 CerP 30:1; [M-H]-; CerP(d18:1(4E)/12:0)	Difference A Head to Tail Side by Side A Subtraction / 149 761R 63.8P
2 lipidblast-neg 55 517 5.22 649 CerP 30:1; [M-H]-; CerP(d14:1(4E)/16:0)	Name: CerP 30:1; [M-H]; CerP(d18:1(4E)/12:0)
3 lipidblastineg 55 517 5.22 649 CerP 30:1; [M-H]-; CerP(d16:1(4E)/14:0)	100- <u>MW:</u> 560 <u>ID#</u> : 22881 <u>DB</u> : lipidblast-neg
4 Custompo+n 13 261 1.23 422 FC 16.0, [M+Na]+, GFCh0(12:0) 5 custompo+n 19 261 1.29 422 FC 18:0: [M+Na]+; GFCh0(12:0/6:0)	5 largest peaks:
6 linidblast-ons 5 91 0.80 150 PC 18:0: [M+Na]+; GPCbo(6:0/12:0)	78,95851 999.00   96,96908 999.00   360.2
7 lipidblast-pos 5 91 0.80 150 PC 18:0; [M+Na]+; GPCho(12:0/6:0)	360.23049 542.39745 542.39745 542.39745 78 95951 999 00 ion PD2 (79 95951)
8 custompc+h 1 24 0.68 122 PC 20:3; [M+H]+; GPCho(2:0/18:3(6Z,9Z,12Z))	0.4 100 200 300 400 500 96.96908 999.00 ion H2P04- (96.96908)
	(lipidblast-neg) CerP 30:1; [M-H]-; CerP(d18:1(4E)/12:0)
Names Structures Hit List	Plot/Text of Hit Plot of Hit
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide //

#### 4000 Q-Trap mass spectrometer; M+H

Ceramide-phosphate CerP(d18:1/12:0); M+H; Prec. m/z: 874.8; LipidMaps http://www.lipidmaps.org/data/standards/standards.php?lipidclass=LMSP



#### ABI QTRAP 4000; Lipid A (diPP-14-hexaacyl); [M-H]-;

Accurate Mass should be 1824.24287; ELECTROPHORESIS, Volume 29, Issue 10 (p 2171-2181) ; Characterization of intact lipopolysaccharides

#### 0.8 precursor SET



Comment Curator: Precursor Search set to 0.8 Da (accurate mass not reported in publication)

#### ABI 4000 Q-Trap mass spectrometer; [M-H]-

PI(18:0/18:1) (putative); [M-H]-; Prec. m/z: 863.6; Applied Biosystems/MDS Sciex; Metabolomic identification of potential phospholipid biomarkers for chronic glomerulonephritis by using high performance liquid chromatography-mass spectrometry; Lewen Jia, Chang Wang, Sumin Zhao, Xin Lu and Guowang Xu

📕 NIST MS Search 2.0 - [Peptide, Presearch Default - 65 spectra]	
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🚳 🗫 🗃 📮 1. ABI 4000 Q-Trap; [M-H]-; PI 36:1 💽 🛞 🖳 😥 🍭 🚳	
#     Src.     Name: ABI 4000 Q-Trap; [M-H]; PI 36:1	
1 A ABI 4000 Q-Trap; [M-H]; CerP(d18:1/12:0)	usti
2 A ABI 4000 Q-1 rap ; [M+H]+; CerP(d18:1/12:0) 3 A ABI QTRAP 4000; [M-H]-; Lipid A (diPP-14-bexaacvi) 410.1 863.6 <u>8 largest peaks:</u>	10
4     A     ABI 4000 Q-Trap; [M-H]; PI 36:1     281.3     570.00     863.6     300.00     410.1     200.00     303.1       4     A     ABI 4000 Q-Trap; [M-H]; PI 36:1     340     680     403.4     50.00     591.0     50.00     297.0     10.00	"코
5   A   ABI 4000 Q-Trap; [M+NH4]+; TG 52:3     6   A   ABI-API 4000 QTrap; [M-H]; GM2[d18:1/C18:0]     Interference of Second List (Control	<u> </u>
7 A ABI-API 4000 QTrap; [M-H]-; GM2(d20:1/C18:0)	
8 A ABI 4/00 MALDI-TUF/TUF/ [M-H]; AC2PIM I[16:0/methyl-18:0] 9 A ABI 4700 MALDI TUF/TUF/ [M-H]: Lipid A 100	
10 A ABI 4800 MALDI-TOF/TOF; [M-H]-; Sulfatide(d18:1/N24:1)	
Names / Structures / Spec List /10.1	63.6
linidblast-pos: custompo+boos msp: custompo+boos msp: pc-ac-ped msp: pc-form-ped msp: 234420 total spectral	
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10-	
	ð I
H Library Score Dot Product Prob. (%) Rev-Dot Name ABI 4000 Q-Trap; (M-H); PI 36:1 Head to Tail MF=143 RMF=577 ▼PI 36:1; (M-H); GPIns(18:0/	18:10 4000
1     Ipidplast-neg     143     577     5.18     639     P13617 (M-H); GPIns(18:0/18:1(11E))     143 0776 0.18     143 0776 0.18       2     Inidblast-neg     143     577     5.18     639     P13617 (M-H); GPIns(18:0/18:1(11E))     143 0776 0.18     143 0776 0.18	
3 lipidblast-neg 143 577 5.18 639 PI 36:1; [M-H]; GPIns(18:0/18:1(13Z)) 597.30413 <u>Name:</u> PI 36:1; [M-H]; GPIns(18:0/18:1(11E))	-
4 lipidblast-neg 143 577 5.18 639 PI 36:1; [M-H]-; GPIns(18:0/18:1(17Z))	545
5 lipidblast-neg 143 577 5.18 639 PI 36:1; (M-H)-; GPIns(18:0/18:1(4E)) 50- 417.24074 281.24790 999.00   283.26354 999.00   597	7.31
7 lipidplastneg 143 577 5.18 639 PI 36:1; [M-H]; defins[16.0/16:1[62]])	1.3
8 lipidblast-neg 143 577 5.18 639 PI 36:1; [M-H]-; GPIns(18:0/18:1(9E))	-
(lipidblast-neg) PI 36:1; (M-H]-; GPIns(18:0/18:1(11E))	
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#### 4000 Q-Trap mass spectrometer; M+NH4

TAG(16:0/18:1/18:2); M+NH4; Prec. m/z: 874.8; Detection of the abundance of diacylglycerolnext term and triacylglycerol molecular species in cells using neutral loss mass spectrometry; doi:10.1016/j.ab.2007.03.012; Robert C. Murphy, Corresponding Author Contact Information, E-mail The Corresponding Author, Patrick F. James, Andrew M. McAnoy, Jessica Krank, Eva Duchoslav and Robert M. Barkley



Name: ABI-API 4000 QTrap; [M-H]-; GM2(d18:1/C18:0) <u>MW:</u> 1383 <u>ID#:</u> 276 <u>DB:</u> Text File <u>Comment:</u> GM2(d18:1/C18:0); [M-H]-; Prec. m/z: 1383.0; Imaging MALDI Mass Spectrometry Using an Oscillating Capillary Nebulizer Matrix Coating System and Its Application to Analysis of Lipids in Brain from a Mouse Model of Tay-Sachs/Sandhoff Disease; Anal. Chem., 2008, 80 (8), pp 2780-2788; http://pubs.acs.org/doi/full/10.1021/ac702350g 10 largest peaks:



#### Name: ABI-API 4000 QTrap; [M-H]-; GM2(d20:1/C18:0)

MW: 1411 ID#: 4241 DB: Text File

Comment: GM2(d20:1/C18:0); [M-H]-; Prec. m/z: 1411; Imaging MALDI Mass Spectrometry of Sphingolipids Using an Oscillating Capillary Nebulizer Matrix Application System Series: Methods in Molecular Biology | Volume: 656 | Year: 2010 | Page ; 10.1007/978-1-60761-746-4\_7



<u>Name:</u> ABI 4700 MALDI-TOF/TOF; [M-H]-; Ac2PIM1(16:0/methyl-18:0) <u>MW:</u> 1013 <u>ID#:</u> 4240 <u>DB:</u> Text File <u>Comment:</u> Ac2PIM1(16:0/methyl-18:0); [M-H]-; Prec. m/z:1013.63; Synthesis and Structure of Phosphatidylinositol Dimannoside; http://pubs.acs.org/doi/full/10.1021/jo0625599; Supplement

塔 NIST MS Search 2.0 - [Peptide, Presearch Default - 22 spectra]	
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X 🖻 🖻 🚑 🔠 🌇 🚮 🚔 🗆 #/z ← 🎗	
🚳 🎥 🚔 🗐 1. ABI 4700 MALDI-TOF/TOF; [M-H]-; / 💌 🛞 🖳 🔍 🥘	
# Src. Name	100 391.2299 Name: ABI 4700 MALDI-TOF/TOF; [M-H]; Ac
4 A ABI 4000 Q-Trap; [M-H]-; PI 36:1 5 A ABI 4000 Q-Trap; [M+NH4]+; TG 52:3	255.2307 Comment: Ac2PIM1(16:0/methyl-18:0); [M-H]-;
6 A ABI-API 4000 QTrap; [M-H]; GM2(d18:1/C18:0)	1013.075 <u>101argest peaks:</u> 50 1 715.3599 <b>?</b> 391.2299 999.00 255.2307 800.00 2
8 A ABI-API 4000 QTrap; [M-H]-; GDTa 8 A ABI-API 4000 QTrap; [M-H]-; GM2(d20:1/C18:0)	403.0551 550.00   78.9465 400.00   4
9 A ABI 4700 MALDI-TOF/TOF; [M-H]; Ac2PIM1(16:0/methyl-18:0)	
	210 420 630 840 237.2791 /30.001 331.2239 999.001 4 ▼
Names Structures Spec List	Plot/Text of Search Spectrum Plot of Search Spectrum P
lipidblast-neg; 134202 total spectra	201 2200
	100-255,2307
	152,9803
14	50- 715.3599 🦅
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# Library Score Dot Product Prob. (%) Rev-Dot Name	
1 lipidblast-neg 483 603 26.1 807 Ac2PIM2 35:0; [M-H]-; A	50 255 2226 291 2251 4 <b>9</b>
2 lipidblast-neg 483 603 26.1 807 Ac2PIM2 35:0; [M-H]-; A	1013.6177
3 lipidblast-neg 480 599 23.1 802 Ac2PIM2 350; [M-H]-; A 4 lipidblast-neg 480 599 23.1 802 Ac2PIM2 350; [M-H]-; A	100-
5 lipidblast-neg 187 272 0.23 707 DGDG 41:2; [M-H]-; DGI	
6 lipidblast-neg 187 272 0.23 707 DGDG 41:2; [M-H]-; DGI	90 180 270 360 450 540 630 720 810 900 990
8 lipidblast-neg 187 272 0.23 707 DGDG 41:2; [M-H]-; DGL 8 lipidblast-neg 187 272 0.23 707 DGDG 41:2; [M-H]-; DGL	Difference Head to Tail Side by Side Subtraction 483 603R 26.1P
9 lipidblast-neg 187 272 0.23 707 DGDG 41:2; [M-H]-; DGI	Name: Ac2PIM2 35:0: [M-H]-: Ac2PIM1(methyl-18:0/16:1
10 lipidblast-neg 187 272 0.23 707 DGDG 41:2; [M-H]-; DG[	100- /15.330/8 MW: 1013 ID#: 135 DB: lipidblast-neg
11 lipidblast-neg 160 236 0.06 707 DGDG 41:2; [M-H]-; DGL 12 lipidblast-neg 160 236 0.06 707 DGDG 41:2; [M-H]-; DGL	1012 C177 9largest peaks:
13 lipidblast-neg 58 90 0.00 236 SQDG 48:2; [M-H]-; SQC	
14 lipidblast-neg 58 90 0.00 236 SQDG 48:2; [M-H]-; SQD	9 m/z Values and Intensities:
15 iipiadiast-neg 58 90 0.00 236 SQDG 48:2; [M-H]-; SQL 16 ligidblast-neg 58 90 0.00 236 SQDG 48:2: [M-H]-: SQC	270 540 810
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Name: ABI 4700 MALDI TOF/TOF; [M-H]-; Lipid A <u>MW</u>: 1796 <u>ID#</u>: 4239 <u>DB</u>: Text File <u>Comment</u>: Lipid A from E coli F583; [M-H]-; Prec. m/z: 1796.521; C94H178N2O25P2; LipidA-PP-[14('O-14)/14('O-12)/14/14]; Determination of pyrophosphorylated forms of lipid A in Gram-negative bacteria using a multivaried mass spectrometric approach; DOI: 10.1073/pnas.0800445105



#### Name: ABI 4800 MALDI-TOF/TOF; [M-H]-; Sulfatide(d18:1/N24:1)

<u>MW:</u> 888 <u>ID#:</u> 272 <u>DB:</u> Text File

Comment: Sulfatide(d18:1/N24:1); [M-H]-; Prec. m/z:888.67; Selective desorption/ionization of sulfatides by MALDI-MS facilitated using 9- aminoacridine as matrix;Hua Cheng, Gang Sun, Kui Yang, Richard W. Gross, and Xianlin Han;Journal of Lipid Research, Vol. 51, 1599-1609, June 2010

9 largest peaks:



#### API 2000 triple quadrupole; [M-H]-;

Cardiolipin CL 76:12 (MIX from rat heart); [M-H]-; Prec. m/z: 1495.97;

Journal of Lipid ResearchVolume 46, 2005;

Quantitation of cardiolipin molecular species in spontaneously hypertensive heart failure rats using electrospray ionization mass spectrometry



<u>Name:</u> ABI-QSTAR-XL-Quadrupol-TOF; [M+Na]+ ; PC 34:0 <u>MW:</u> N/A <u>ID#:</u> 4236 <u>DB:</u> Text File <u>Comment:</u> PC 34:0; [M+Na]+ ; Prec. m/z: 784.55; Application of electrospray ionization mass spectrometry to characterize glycerophospholipids in Francisella tularensis subsp. novicida;X. Wang et al. / International Journal of Mass Spectrometry 293 (2010) 4550

🖷 NIST MS Search 2.0 - [Peptide, Presearch Default - 327 spectra]	
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🝈 🎾 🖆 🚔   1. ABI-QSTAR-XL-Quadrupol-TOF; [M+N 🛛 🛞 崖 😥 🔍	
# Src. Name	1146.98
12 A ABI API 2000 triple quadrupole; [M-H]-; CL 76:12	100- House Spectrometry 293 (2010) 4550
13 A ABI-QSTAR-XL-Quadrupol-TOF; [M+Na]+; PC 34:0	601.49
15 A ABI Sciex API III QQQ; [M-H]; PE MIX	50- ?? 725.48
16 A ABI Sciex API III QQQ; [M-H]-; PE 40:6	
17 A ABI Sciex API III QQQ; [M+H]+; PE MIX	
	(Text File) ARI-QSTAR-XI -Quadrupol-TOF: [M+Na]+ : P
Names Structures Spec List	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
lipidblast-neg; custompc+hpos.msp; custompc+napos.msp; lipidblast-pos; pc-ac-neg.msp;	
pc-torm-neg.msp; 234420 total spectra	100-146.98
	601.49
	50- 725.48 204.55
	123,40 784,55
# Library Score Dot Product Prob. (%) Rev-Dot Name	455.25399
1 lipidblast-pos 231 580 4.96 943 PC 34:0; [M+Na]+; GPC)	50-
2 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC	601.51717
3 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC	100
4 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC	725.50971
5 lipidblast-pos 221 567 3.50 922 PL 34(0) [M+Na]+) GPCF	180 240 300 360 420 480 540 600 660 720 780
0 iipidulast-pos 221 367 3.30 322 FC 34.0; [M+Na]+; GPCF 7 iipidulast-pos 221 567 3.50 922 PC 34.0; [M+Na]+; GPCF	ABI-QSTAR-XL-Quadrupol-TOF; I Head to Tail MF=231 RMF=580 TC 34:0; (M+Na)+; GPCho(17:0/
8 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC	Uttrerence A Head to Tail A Side by Side A Subtraction / 231 580R 4.96P
9 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC	725 50971 Name: PC 34:0; [M+Na]+; GPCho(17:0/17:0)
10 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC}	100- <u>MW:</u> 784 <u>ID#:</u> 44465 <u>DB:</u> lipidblast-pos <u>Comment: Parent-784 58321 Mar. eu-ty-704 59321 - PC</u>
11 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPCł	601.51717 5 largest peaks:
12 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC	50-
13 lipidblast-pos 221 567 3.50 922 PC 34:0; [M+Na]+; GPC}	455,25399 455,25399 40.00 [M+NaL59.on2
14 lipidbiast-pos 221 567 3,50 922 PC 34;0; [M+Na]+; GPC	0 180 360 540 720 514.32749 20.00 [M+Na]-sn1-H20    [M+Na]-sn2-H20 ↓
13 IIUIUUIas(0005 221 307 3.00 322 FC 34.0. IM+NaI+: 0FC →	(lipidblast-pos) PC 34:0: [M+Na]+: GPCho(17:0
Names Structures Hit List	Plot/Text of Hit / Plot of Hit /
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For Help, press F1	Peptide Peptide //

Name: ABI QSTAR-XL QTOF; [M-H]-; Sulfatide (d18:1/C22:0) <u>MW:</u> 862 <u>ID#:</u> 269 <u>DB:</u> Text File <u>Comment:</u> Sulfatide (d18:1/C22:0); [M-H]-; Prec. m/z: 862.5729; M. Cameron Sullards, Jeremy C. Allegood, and Alfred H. Merrill, Jr.; Identification and Structure Determination of Sulfatides using Chip-based NanoESI with Hybrid Q-TOF MS and MS/MS; <u>5 largest peaks:</u>



<u>Name:</u> ABI Sciex API III QQQ; [M-H]-; PE MIX <u>MW:</u> 750 <u>ID#:</u> 4234 <u>DB:</u> Text File <u>Comment:</u> PE MIX; alkeny/acyl PE(18:0/20:4) and PE(16:0/22:4); [M-H]-; Prec. m/z:750.3; JL Kerwin, AR Tuininga, and LH Ericsson; Identification of molecular species of glycerophospholipids and sphingomyelin using electrospray mass spectrometry; J. Lipid Res., Jun 1994; 35: 1102 - 1114.

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#     Src.     Name       12     A     ABI API 2000 triple quadrupole; [M-H]; CL 76:12       13     A     ABI-QSTAR-XL-Quadrupol-TOF; [M+Na]+; PC 34:0       14     A     ABI QSTAR-XL QTOF; [M-H]; Sulfatide (d18:1/C22:0)       15     A     ABI Sciex API III QQQ; [M-H]; PE MIX       16     A     ABI Sciex API III QQQ; [M-H]; PE MIX       17     A     ABI Sciex API III QQQ; [M-H]; PE MIX       18     A ABI Sciex API III QQQ; [M-H]+; PE MIX       19     A     ABI Sciex API III QQQ; [M-H]+; PE MIX       18     A BI Sciex API III QQQ; [M-H]+; PE MIX       19     A     ABI Sciex API III QQQ; [M-H]+; PE MIX       18     A     ABI Sciex API III QQQ; [M-H]+; PE MIX       19     A     ABI Sciex API III QQQ; [M-H]+; PE MIX       19     A     ABI Sciex API III QQQ; [M-H]+; PE MIX       19     A     ABI Sciex API III QQQ; [M-H]+; PE MIX       19     A     ABI Sciex API III QQQ; [M-H]+; PE MIX	Name: ABI Sciex API III QQQ; [M-H]; PE MIX       100-     750.3       50-     331.5       70-     31/2 See See See See See See See See See Se
lipidblast-neg; custompc-hpos.msp; custompc-+napos.msp; lipidblast-pos; pc-ac-neg.msp; pc-form-neg.msp; 234420 total spectra 1 1 1 1000 900 800 700 600 500 400 300 200 100 0	100- 50- 3331.5 <b>?</b>
.(%)     Rev-Dot     Name       1.5     877     plasmenyl-PE 38:4; [M-H]; PE(P-16:0/22:4(7Z,10Z,13Z,16Z))       38     877     plasmenyl-PE 38:4; [M-H]; PE(P-18:0/20:4(5E,8E,11E,14E))       38     877     plasmenyl-PE 38:4; [M-H]; PE(P-18:0/20:4(5E,8E,11E,14E))       38     877     plasmenyl-PE 38:4; [M-H]; PE(P-18:0/20:4(5E,8E,211Z,14Z))       38     877     plasmenyl-PE 38:4; [M-H]; PE(P-18:0/20:4(5E,12E,12E,13E,16E))       39     632     PE 37:5; [M-H]; GPEtn(15:1(3Z)/22:4(7Z,10Z,13Z,16Z)/15:1(3Z))       36     632     PE 37:5; [M-H]; GPEtn(22:4(7Z,10Z,13Z,16Z)/15:1(3Z))       36     632     PE 37:5; [M-H]; GPEtn(22:4(7Z,10Z,13Z,16Z)/15:1(3Z))       36     632     PE 37:5; [M-H]; GPEtn(22:4(7Z,10Z,13Z,16Z)/15:1(3Z))       36     632     PE 37:5; [M-H]; GPEtn(12:1(3Z)/20:4(5E,8E,11E,14E))       37     632     PE 37:5; [M-H]; GPEtn(12:1(3Z)/20:4(5E,7Z,11Z,1Z))	436.28295     ?       100-     331.26354       320     360     400     440     480     520     560     600     640     680     720     760       I_ABI Sciex API III QQQ: [M-H]: PI Head to Tail MF=161 RMF=630     (▼plasmenyl-PE 38:4: [M-H]: PE[P-I     Difference     Head to Tail      Subtraction     161 630R 19.5P
26   632   PE 37:5; [M-H]; GPEth(17:1(§2)/20:4(52,82,112,142))     26   632   PE 37:5; [M-H]; GPEth(17:1(§2)/20:4(7E,10E,13E,16E))     26   632   PE 37:5; [M-H]; GPEth(20:4(5E,8E,11E,14E)/17:1(§2))     26   632   PE 37:5; [M-H]; GPEth(20:4(5E,8E,11E,14E)/17:1(§2))     26   632   PE 37:5; [M-H]; GPEth(20:4(5E,8E,11E,14E)/17:1(§2))     26   632   PE 37:5; [M-H]; GPEth(20:4(5E,82,112,142)/17:1(§2))     26   632   PE 37:5; [M-H]; GPEth(20:4(5E,82,112,142)/17:1(§2))     35   452   CL 76:9; [M-2H](2-); CL(18:1/20:3/18:1/20:4)     35   452   CL 76:9; [M-2H](2-); CL(18:1/20:2/16:1/20:2/20:4)     35   452   CL 76:9; [M-2H](2-); CL(10:1/20:2/20:4)     35   452   CL 76:9; [M-2H](2-); CL(10:2/20:4)     4   Names   Structures	Name:     plasmenyl-PE     38:4;     (M-H);     PE(P-16:0/22:4(72,1C \ MW, 750 ID:ft;       100-1331.26354     MW;     750 ID:ft;     123275 DB;     (piddbast-neg)       50-1436.28295     331.26354     MW;     750 ID:ft;     123275 DB;     (piddbast-neg)       331.26354     999.001     436.28295     250.001     418.27239     331.26354     999.001     436.28295     250.001     418.27239     31.26354     999.001     436.28295     250.001     418.27239     31.26354     999.001     436.28295     250.001     418.27239     31.26354     999.001     436.28295     250.001     418.27239     31.26354     999.001     418.27239     31.26354     999.001     418.27239     31.26354     999.001     418.27239     31.26354     31.26354     399.001     418.27239     31.26354
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Name: ABI Sciex API III QQQ; [M-H]-; PE 40:6 <u>MW:</u> 790 <u>ID#:</u> 4233 <u>DB:</u> Text File <u>Comment:</u> PE 40:6; PE(18:0/22:6); [M-H]-; Prec. m/z:790.4; JL Kerwin, AR Tuininga, and LH Ericsson; Identification of molecular species of glycerophospholipids and sphingomyelin using electrospray mass spectrometry; J. Lipid Res., Jun 1994; 35: 1102 - 1114.

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# Src. Name	100 283.4 Name: ABI Sciex API III QQQ; [M-H]-; PE 40:6
12 A ABI API 2000 triple quadrupole; [M-H]-; CL 76:12	Comment: PE 40:6; PE(18:0/22:6); [M-H]-; Prec
14 A ABI QSTAR XL QTOF; (M-H); Sulfatide (d18:1/C22:0)	50 327.5 790.4 31argest peaks:
15 A ABI Sciex API III QQQ; [M-H]; PE MIX	30- 203.4 333.00 327.3 330.00 1 730.4 300.0
16 A ABI Sciex API III UUU; [M-H]; PE 40:6	
	330 440 550 660 770 <u>no sunnyums</u>
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1000 900 800 700 600 500 400 300 200 100 0	
# Library Score Dot Product Prob. (%) Rev-Dot Name	
1 lipidblast-neg 257 881 40.2 885 PE 40:6; [M-H]-; GPEtn[	50 480.30917 50 7
2 lipidblast-neg 257 881 40.2 885 PE 40:6; [M-H]-; GPE tn(;	*
3 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	100
4 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	
5 lipidblast-neg 50 534 0.40 536 0L 82(11) [M-2H][2-); 0L 6 lipidblast-neg 50 534 0.40 536 0L 92(11, [M-2H][2-); 0L	300 350 400 450 500 550 600 650 700 750 800
7 lipidblast-neg 50 534 0.40 536 CL 82.11, [M-2H][21], CL	ABI Sciex API III QQQ; [M-H]; PI Head to Tail MF=257 RMF=881 ▼PE 40:6; [M-H]; GPEth(18:0/22:1
8 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	Difference A Head to Tail A Side by Side A Subtraction / 257 881R 40.2P
9 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	Name: PE 40:6; [M-H]-; GPEtn(18:0/22:6(4Z,7Z,10Z,132
10 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	100- <u>MW:</u> 790 <u>D#:</u> 108478 <u>DB:</u> lipidblast-neg
11 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	6 largest peaks:
12 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	
13 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL 14 lipidblast-neg 50 534 0.40 536 CL 82:11; [M-2H](2-); CL	400.30317 505.25733 50.001 6 m/z Values and Intensities:
14 iipidbiast-neg 50 554 0.40 556 CL 82:11; [M-2H][2-]; CL 15 linidbiast-neg 25 361 0.12 452 CL 82:11 (M-2H)[2.)· CL ▼	300 450 600 750 283.26354 999.00 sn1 FA
	(lipidblast-neg) PE 40:6; [M-H]-; GPEtn(18:0/2
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	Peptide Peptide ///

Name: ABI Sciex API III QQQ; [M+H]+; PE MIX <u>MW</u>: 752 <u>ID#</u>: 4232 <u>DB</u>: Text File <u>Comment</u>: PE MIX; alkeny/acyl PE(18:0/20:4) and PE(16:0/22:4) alkenyl-acyl PE; [M+H]+; Prec. m/z:752.4; JL Kerwin, AR Tuininga, and LH Ericsson; Identification of molecular species of glycerophospholipids and sphingomyelin using electrospray mass spectrometry; J. Lipid Res., Jun 1994; 35: 1102 - 1114.

not found



<u>Name:</u> ABI Sciex API III QQQ; [M+H]+; PE 40:6 <u>MW:</u> 792 <u>ID#:</u> 4231 <u>DB:</u> Text File <u>Comment:</u> PE 40:6; PE(18:0/22:6); [M+H]+; Prec. m/z:792.2; JL Kerwin, AR Tuininga, and LH Ericsson; Identification of molecular species of glycerophospholipids and sphingomyelin using electrospray mass spectrometry; J. Lipid Res., Jun 1994; 35: 1102 - 1114.



Name: ABI-QSTAR-Pulsar-Quadrupol-TOF MALDI; [M-H]-; PI 38:4 <u>MW:</u> 885 <u>ID#:</u> 4230 <u>DB</u>: Text File <u>Comment:</u> PI 38:4; PI(18:0/20:4);Elevation of sulfatides in ovarian cancer: An integrated transcriptomic and lipidomic analysis including tissue-imaging mass spectrometry;;doi:10.1186/1476-4598-9-186



Name: ABI-QSTAR-Pulsar-Quadrupol-TOF PSer; [M-H]-; PS 40:6 <u>MW</u>: 834 <u>ID#</u>: 263 <u>DB</u>: Text File <u>Comment</u>: 834.75; PSer(18:0/22:6);MALDI-MS DIRECT TISSUE ANALYSIS OF PROTEINS:IMPROVING SIGNAL SENSITIVITY USING ORGANIC TREATMENTS;R. Lemaire, M. Wisztorski, A. Desmons, J.C. Tabet, R. Day, M.Salzet, I. Fournier;DOI : 10.1021/ac060565z <u>8 largest peaks</u>:



#### Name: ABI-QSTAR-Pulsar-Quadrupol-TOF; [M-H]-; SQDG 34:2 MW: 817 ID#: 262 DB: Text File

Comment: SQDG(18:2/16:0); [M-H]-; Prec. m/z: 817.51; SQDG(18:2/16:0); Characterization by high-performance liquid chromatography/electrospray ionization quadrupole time-of-flight mass spectrometry of the lipid fraction of Spirulina platensis pressurized ethanol extract; Rapid Commun. Mass Spectrom. 2007; 21: 1729-1738;Miguel Herrero, Maria J. Vicente, Alejandro Cifuentes and Elena Ibanez

8 largest peaks:


Name: ABI-QSTAR-Pulsar-Quadrupol-TOF MALDI; [M-H]-; ST 34:1 <u>MW:</u> 778 <u>ID#:</u> 4227 <u>DB:</u> Text File <u>Comment:</u> ST 34:1; ST(d18:1/16:0);Elevation of sulfatides in ovarian cancer: An integrated transcriptomic and lipidomic analysis including tissue-imaging mass spectrometry;;doi:10.1186/1476-4598-9-186



Name: Agilent 6410 triple quadrupole MS; [M-H]-; PI3P 37:1 <u>MW:</u> 957 <u>ID#:</u> 12671 <u>DB:</u> Text File <u>Comment:</u> PI3P 37:1; [M-H]-; m/z 957.6; STRESS-INDUCED SYNTHESIS OF PHOSPHATIDYLINOSITOL 3-PHOSPHATE IN MYCOBACTERIA; http://www.jbc.org/cgi/doi/10.1074/jbc.M110.119263 9 largest peaks:

PI3P species (Phosphatidylinositol 3-Phosphates) not in LipidBlast – very low hit score (24) therefore false ID

🖷 NIST MS Search 2.0 - [Peptide, Presearch Default - 22 spectra]	
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🔟 🏷 🗁 📮   1. Agilent 6410 triple quadrupole MS; [M 📉 🕅 🛓 🗮 😥 🗮 🥨	
# Src. Name	659.0 Name: Agilent 6410 triple quadrupole MS; [M-H
1 A Agilent 6410 triple quadrupole MS; [M-H]-; PI3P 37:1	100- 253.3 <u>MW:</u> 957 <u>ID#:</u> 1257 <u>IDB:</u> Text File Comment: PI3P 37:1: [M-H]-: m/z 957 6: STBE
2 A Agilent 5410 triple quadrupole MS; [M-H]-; PI 34:1	9 largest peaks:
4 A Agilent 6530 QcT OF; [M+HCDO]; PC 36:2	
5 A Agilent 6530 QqTOF; [M+H]+; PC 36:2	100.1 200.0 200.0 200.0 417.2 200.0
6 A Agilent 6530 QqTOF; [M+H]+; PC/PE mix	
7 A Aailant 6530 DaTOF: IM+HI+: PE+PC MIX	340 510 680 850 <u>417 2 250 001 595 0 400 001 659 0 999 0</u>
Names Structures Spec List	Plot/Text of Search Spectrum A Plot of Search Spectrum A Plot/Text of Spec List
lipidblast-neg; 134202 total spectra	
100-1	100 659.0
	253.3
	321.1 595.0 🤊
1-	504 595.0 <b>°</b>
1000 900 800 700 600 500 400 300 200 100 0	957.6
1 lipidblast-neg 24 61 5.17 236 SQD6 44:2; [M-H]-; SQL 2 Faidblast-neg 24 61 5.17 236 SQD6 44:2; [M-H]-; SQL	50-
2 lipidblast-neg 24 61 5.17 236 5QDG 44:2; [M-H]-; 5QL 2 lipidblast-neg 24 61 5.17 236 5QDG 44:2; [M-H]-; 5QL	
3 lipidblastmeg 24 61 5.17 236 5QDG 44:2; [M-H]-; 5QL 4 lipidblastmeg 24 61 5.17 236 SODG 44:2; [M-H]-; 5QL	100
5 lipidblast-neg 24 61 517 236 SQDG 44:2; [M-H]-; SQD	
6 lipidblast-neg 24 61 517 236 SQDG 44:2: [M-H]; SQC	
7 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-: SQE	Agilent 6410 triple quadrupole MSI Head to Tail MF=24 RMF=61 I▼SQDG 44:2; [M-H]-; SQDG[18:1[]
8 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQD	Dimenence A Head to fail A side by side A subtraction 24 61k 6.17P
9 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQD	Name: SQDG 44:2; [M-H]-; SQDG(18:1(11E)/26:1(5Z))
10 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQE	100-223.00030 <u>MW:</u> 957 <u>ID#:</u> 130332 <u>DB:</u> lipidblast-neg
11 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQE	5 largest peaks:
12 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQC	
13 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQC	393.37302 5 m/z Values and Intensities:
14 lipidblast-neg 24 61 5.17 236 SQDG 44:2; [M-H]-; SQD	QL 220,00000 00000 00000 000000 000000 000000
15 lipidblast-neo 24 61 5. 7 236 SODG 44:2: IM-H1: SOL	(ipidblact.neg) SODG 44/2 (M.H1: SODG(19)
Names Structures Hit List	Plot/Text of Hit / Plot of Hit /
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	Peptide Peptide

Name: Agilent 6410 triple quadrupole MS; [M-H]-; PI 34:1 <u>MW:</u> 835 <u>ID#:</u> 12670 <u>DB:</u> Text File <u>Comment:</u> PI (16:0/18:1); [M-H]-; m/z 835.6; STRESS-INDUCED SYNTHESIS OF PHOSPHATIDYLINOSITOL 3-PHOSPHATE IN MYCOBACTERIA; http://www.jbc.org/cgi/doi/10.1074/jbc.M110.119263 <u>8 largest peaks:</u>



Name: Agilent 6520 Q-TOF; [M-H]-; PI 35:0 <u>MW:</u> 851 <u>ID#:</u> 25 <u>DB:</u> Spec. List <u>Comment:</u> PI (16:0/19:0); [M-H]-; 851.5650; Comprehensive LC-MS Profiling of Mycobacterium tuberculosis Complex Lipids; Mark J. Sartain; COSMOS 2009 <u>6 largest peaks:</u>



Name: Agilent 6530 QqTOF; [M+HCOO]-; PC 36:2 <u>MW</u>: 830 <u>ID#</u>: 12668 <u>DB</u>: Text File <u>Comment</u>: PC 36:2 ; [M+HCOO]-; 830.5966; Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Journal of Chromatography A, 1217 (2010) 4087-4099 <u>4 largest peaks</u>:



<u>Name:</u> Agilent 6530 QqTOF; [M+H]+; PC 36:2 <u>MW:</u> 786 <u>ID#:</u> 27 <u>DB:</u> Spec. List <u>Comment:</u> PC 36:2 ; [M+H]+; 786.5987; Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Journal of Chromatography A, 1217 (2010) 4087-4099 <u>2 largest peaks:</u>

RIST MS Search 2.0 - [Peptide, Presearch Default - 179 spectra]	
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🚳 🍉 🗃 📮 1. Agilent 6530 QqTOF; [M+H]+; PC 36:: 💌 🛞 🖳 🔍 🍭 🚳	
# Src. Name	Name: Agilent 6530 QqTOF; [M+H]+; PC 36:2
20 L ABI-QSTAR-Pulsar-Quadrupol-TOF PSer; [M-H]-; PS 40:6	100- 786,5987 <u>MW:</u> 786 <u>ID#:</u> 27 <u>DB:</u> Spec. List <u>Comment:</u> PC 36:2 ; [M+H]+; 786,5987; Comprehensive blood plasma lipidon
21 L ABI-QSTAR-Pulsar-Quadrupol-TOF; [M-H]; SQDG 34:2 22 L ABI-QSTAR-Pulsar-Quadrupol-TOF MALDI; [M-H]; ST 34:1;	2 largest peaks: 104.0729.099.001.795.5997.900.001
23 L Agilent 6410 triple quadrupole MS; [M-H]; PI3P 37:1	360 720 2 m/z Values and Intensities:
24 L Agilent 6410 triple quadrupole MS; [M-H]; PI 34:1 25 L Agilent 6520 Q-TDE: [M-H]-: PI 35:0	(Spec. List) Agilent 6530 Qq1
26 L Agilent 6530 QqTOF; [M+HCOO]-; PC 36:2	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
27 L Agilent 6530 QqTOF; (M+H)+; PC 36;2 28 L Agilent 6530 QqTOF; (M+H)+; PC/PE mix	100
29 L Agilent 6530 QqT0F; [M+H]+; PE+PC MIX	786,5987
30 L Agilent 6530 QqTOF; [M+NH4]+; TG 54:3	
	. 50-
Names Structures Spec List	
acylooa+mol+hpos; acylooa+mol-hneg; mainlib; acyloarnitine+custom+mol+hpos; acyloarnitine+mol+hpos; acyloarnitine+mol+napos; custompo+hpos msp; custompo+napos msp; hilio-urine; linidblast-neg; po-ac-neg msp; po-form-neg msp;	
respectdbinmdlformat; 365886 total spectra	504.34554 603.53522 727.52776
1000-1	50-
10-	
1-	100
# Library Score Dot Product Prob (%) Rev-Dot Name	200 250 300 350 400 450 500 500 600 650 700 750 800
1 custompc+h 7 737 0.61 737 PC 36:2; [M+H]+; GPCho(18:1(11E)/18:1(11E))	Difference Head to Tail Side by Side A Subtraction 7 737R 0.61P
2 custompc+h 7 737 0.61 737 PC 36:2; [M+H]+; GPCho(18:1(11E)/18:1(11Z))	Name: PC 36/2: [M+H]+: GPCbo(18:1(11E)/18:1(1
3 custompc+h 7 737 0.61 737 PC 36:2; [M+H]+; GPCho(18:1(11E)/18:1(13Z))	100- <u>MW:</u> 786 <u>ID#</u> , 2569 <u>DB</u> : custompc+hpos.msp
4 custompc+h 7 737 0.61 737 PC 36:2; [M+H]+; GPCho[18:1(11E]/18:1(17Z)]	Comment: Parent=786.60126 Mz_exact=786.6012 7 Jaroest peaks:
6 custompc+n 7 737 0.61 737 PC 36.2; [M+H]+; GPCho(18:1(1E)/18:1(4E)]	50-
7 custompc+h 7 737 0.61 737 PC 36:2; [M+H]+; GPCho(18:1(11E)/18:1(7Z))	004.34554 727.52776 786.60126 200.00 768.59070 10.00
8 custompc+h 7 737 0.61 737 PC 36:2; [M+H]+; GPCho(18:1(11E)/18:1(9E))	280 420 560 700 184.07387 999.00 fragment C5H15N04P
	(custompc+hpos.msp) PC 36:2; [M+H]+; GPCho(18:1(1
Names A outocures Hit List	Plot/Text of Hit Plot of Hit
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	Peptide Peptide

Name: Agilent 6530 QqTOF; [M+H]+; PC/PE mix <u>MW:</u> 744 <u>ID#:</u> 28 <u>DB</u>: Spec. List <u>Comment:</u> PC 36:2 + PE 36:2 mix ; [M+H]+; 744.5864; Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Journal of Chromatography A, 1217 (2010) 4087-4099 <u>3 largest peaks</u>:

MIX of different compounds (here annotated as PC36:2

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🔞 🖕 🗃 📮 1. Agilent 6530 QqTOF; [M+H]+; PC/PE 🗾 🛞 🖳 😥 🍭 🔍	
	Name: Apilent 6530 QpTOF: [M+H]+: PC/PE mix
20 L ABI-QSTAR-Pulsar-Quadrupol-TOF PSer; [M-H]-; PS 40:6	100- 744 E02 0 26 20 26 20 26 20 26 20 26 20 26 20 26 20 26 20 26 26 26 26 26 26 26 26 26 26 26 26 26
21 L ABI-QSTAR-Pulsar-Quadrupol-TOF; [M-H]; SQDG 34:2	3 largest peaks:
22 L ABI-USTAH-Pulsan-Quadrupol-FOF MALDI; [M-H]; ST 34:1; 23 L Agilent 6410 triple guadrupole MS; [M-H]; PI3P 37:1	0 1 1 184.0723 999.00   744.5864 500.00   603.5311 200.00   330 660 2 m (2) (1) (1) (2) (2) (2) (2) (2) (2) (2) (2) (2) (2
24 L Agilent 6410 triple quadrupole MS; [M-H]-; PI 34:1	(Spec. List) Agilent 6530 Dg1
25 L Agilent 6520 Q-TOF; [M-H]; PI 35:0	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
27 L Agilent 6530 QqT0F; [M+H]+; PC 36:2	
28 L Agilent 6530 QqTDF; [M+H]+; PC/PE mix	100-
29 L Agilent 6530 QqT UF; (M+H)+; PE+PC MIX 30 L Agilent 6530 QqT OF; (M+NH4)+; TG 54:3	
	50 744.5864
Names Structures Spec List	603 5311
aculona+mol+bnos: aculona+mol-bneg: mainlib: aculorarritine+custom+mol+bnos: aculorarritine+mol+bnos:	
acylcamitine+mple; bybos unto ming, maining; byboan and book action in the report option main poor, acylcamitine+mple; pc-ac-neg.msp; pc-form-neg.msp;	
respectdbinmdlformat; 365886 total spectra	352.18913 561.48829 632.46557 744.55433
	50-
1-	100-
# Library Score Dot Product Prob. (%) Rev-Dot Name	[ Agilent 6530 QqT0F; [M+H]+; PC.[ Head to Tail MF=19 RMF=621 [▼PC 33;2; [M+H]+; GPCho(7:0/26;2
1 custompc+h 19 621 1.72 676 PC 33:2; [M+H]+; GPCho(7:0/26:2(5E,9Z))	Difference Head to Tail Side by Side Subtraction / 19 621R 1.72P
2 custompc+h 19 621 1.72 676 PC 33:2; [M+H]+; GPCho(7:0/26:2(5Z,9E))	Name: PC 33:2; [M+H]+; GPCho(7:0/26:2(5E,9Z))
3 custompc+h 19 621 1.72 676 PU 33:2; [M+H]+; GPCho[7:0/26:2(52,52]]	100- <u>MW:</u> 744 <u>ID#:</u> 1648 <u>DB:</u> custompc+hpos.msp
4 custompc+h 19 621 1.72 676 PC 33.2; [vi+h]+; GPCho[1::0/22.2[132,162]]	Comment: Parent=744.00403 Mz_exact=744.0042     Jargest peaks:
6 custompc+h 19 621 1.72 676 PC 33:2; [M+H]+; GPCho(13:0/20:2[5Z.8Z])	50- 252 19912 551 49929 744 5542 999.00 352,18913 200.00 370.11
7 custompc+h 19 621 1.72 676 PC 33:2; [M+H]+; GPCho(15:0/18:2(2E,4E))	302.10313 301.40023 744.3343 632.46557 200.00 685.48083 200.00 744.5
8 custompc+h 19 621 1.72 676 PC 33:2; [M+H]+; GPCho(15:0/18:2(6Z,9Z))	240 360 480 600 720 184.07387 999.00 fragment C5H15N04P
	(custompc+hpos.msp) PC 33:2; [M+H]+; GPCho(7:0/2€ ▲
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	Peptide Peptide //

Name: Agilent 6530 QqTOF; [M+H]+; PE+PC MIX <u>MW:</u> 744 <u>ID#:</u> 29 <u>DB</u>: Spec. List <u>Comment:</u> PE 36:2 + PC18:1/P-16:0; [M+H]+; 744.5538; Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Journal of Chromatography A, 1217 (2010) 4087-4099 <u>3 largest peaks:</u>



<u>Name:</u> Agilent 6530 QqTOF; [M+NH4]+; TG 54:3 <u>MW:</u> 902 <u>ID#:</u> 30 <u>DB:</u> Spec. List <u>Comment:</u> TG 54:3 (TG(18:1/18:1); [M+H]+; 902.8133; Comprehensive blood plasma lipidomics by liquid chromatography/quadrupole time-of-flight mass spectrometry; Journal of Chromatography A, 1217 (2010) 4087-4099 <u>1 largest peaks:</u>

TAG M+H adduct not in library

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× 🖻 🖻 🗇 🗮 🎬 🜉 📲 🖽 🕫 🔶 👘	
© ▶ ≥ 🚔 1. Agilent 6530 QqTOF; [M+NH4]+; TG ! 🗹 🔞 🖳 🔍	
#       Src.       Name         20       L       ABI-QSTAR-Pulsar-Quadrupol-TOF PSer; [M-H]; PS 40:6         21       L       ABI-QSTAR-Pulsar-Quadrupol-TOF; [M-H]; SQDG 34:2         22       L       ABI-QSTAR-Pulsar-Quadrupol-TOF MALD); [M-H]; ST 34:1;         23       L       Agilent 6410 triple quadrupole MS; [M-H]; PI3P 37:1         24       L       Agilent 6410 triple quadrupole MS; [M-H]; PI 35:0         26       L       Agilent 6530 QqTOF; [M+H]+; PI 35:0         26       L       Agilent 6530 QqTOF; [M+H]+; PC 36:2         28       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         29       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         30       L       Agilent 6530 QqTOF; [M+H]+; PC 7PE mix         310       L       Agilent 653	100-       Name: Agilent 6530 QqT0F; [M+NH4]+; TG 54:3         100-       MW: 902 [DH: 30 DB: Spec. List         Comment: TG 54:3 (TG(18:1/18:1/18:1); [M+H]+; 902.8133; Comprehensive         1 largest peaks:         603.5315         603.5315         900         1 m/z Values and Intensities:         Plot/Text of Search Spectrum         Plot/Text of Search Spectrum
acylcarnitme+mol+napos; custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; respectdbinmdlformat; 365886 total spectra	Difference A Head to Tail A Side by Side A Subtraction
Names Structures Hit List Lib. Search Other Search Names Compare Librarian MSMS	Plot/Text of Hit / Plot of Hit /
	Peptide Peptide //

<u>Name:</u> Agilent LC/MSD 1100 Ion Trap ; [M-H]-; PE 34:1 <u>MW:</u> 716 <u>ID#:</u> 31 <u>DB:</u> Spec. List <u>Comment:</u> PE 34:1; [M-H]-;716.5 ;Use of electrospray ionization mass spectrometry for profiling of crude oil effects on the phospholipid molecular species of two marine bacteria; DOI: 10.1002/rcm.2231 <u>5 largest peaks:</u>

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🔞 🍉 🚔 📮 1. Agilent LC/MSD 1100 Ion Trap ; [M-H 🗸 🛞 🐘 🔎 📵 🚳	
	Name Asland CAVED 1100 Jan Tree (M H1) DE 24-1
	100- 281.2 MW: 716 ID#: 31 DB: Spec. List
28 L Agilent 6530 QqTOF; [M+H]+; PC/PE mix	Comment: PE 34:1; [M-H]-;716.5 ;Use of electrospray ionization mass spectro
29 L Agilent 6530 QqTOF; (M+H)+; PE+PC MIX	
31 L Agilent LC/MSD 1100 Ion Trap ; (M-H); PE 34:1	Shee List Anilent LC/MSD
32 L Agilent Ion Trap SL; [M+Na]+; PC 34:1	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
34 L Agilent Ion Trap XCT ESI; [M-H]-; PI 38:4	201.2
35 L Agilent Ion Trap XCT ESI; [M+H]+; PE 34:2	100-
37 L Agilent Ion Trap XCT ESI; [M-H]; PI 38:4	
100 I A THURD 1100 MAR THAT IS NO. 101.101.00 22.0	50- 255.2
Names Spec List	7105
acylcoa+mol+hpos; acylcoa+mol-hneg; mainlib; acylcarnitine+custom+mol+hpos; acylcarnitine+mol+hpos;	
acylcarnitine+mol+napos; custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; respect/dbimmdformat; 365896; total spectra	
1004	452.27790
	50-
	100- 255.23226
	150 200 250 300 350 400 450 500 550 600 650 700
# Library Score Dot Product Prob. (%) Rev-Dot Name	Agilent LC/MSD 1100 Ion Trap : [/ Head to Tail MF=236 RMF=851 / PE 34:1; [M-H]-; GPEtn(16:0/18:1]
1 lipidblast-neg 236 851 5.38 881 PE 34:1; [M-H]-; GPEtn(16:0/18:1(11E))	Unterence A Head to Tail A side by side A subtraction 236 851R 5.38P
2 lipidplast-neg 236 651 5.38 661 PE 34.1; [M-H]-; GPEtn(16:0/18:1(12)) 3 lipidplast-neg 236 851 5.38 881 PE 34:1: [M-H]-; GPEtn(16:0/18:1(137))	Name: PE 34:1; [M-H]-; GPEtn(16:0/18:1(11E))
4 lipidblast-neg 236 851 5.38 881 PE 34:1; [M-H]-; GPEtn(16:0/18:1(17Z))	Tuu- <u>Comment:</u> Parent=716.52306 Mz_exact=716.5230
5 lipidblast-neg 236 851 5.38 881 PE 34:1; [M-H]-; GPEtn(16:0/18:1(4E))	50-
6 lipidblast-neg 236 851 5.38 881 PE 34:1; [M-H]-; GPEtn(16:0/18:1(62))	452.27790 460.28298 50.00
7 lipidplast-neg 236 851 5.38 881 PE 34:1; [M-H]; GPEth(16:0/18:1(72)) 8 linidplast-neg 236 851 5.38 881 PE 34:1: [M-H]; GPEth(16:0/18:1(9E)) =1	6 m/z Values and Intensities:
	(inidblast-neg) PE 3411 [M-H]- GPE In(16:0/18:1(11E))
Names Structures / Hit List	Plot/Text of Hit / Plot of Hit /
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide //

<u>Name:</u> Agilent Ion Trap SL; [M+Na]+; PC 34:1 <u>MW:</u> 782 <u>ID#:</u> 32 <u>DB:</u> Spec. List <u>Comment:</u> PC (18:1/16:0); [M+Na]+; LC/MS Analysis of Bronchoalveolar Lavage Fluid Phospholipids as Biomarkers for Chronic Lung Inflammation; 5989-1491EN; Barroso, Bischoff <u>6 largest peaks:</u>



Name: Agilent Ion Trap XCT; [M+H]+; NA <u>MW:</u> 808 <u>ID#:</u> 33 <u>DB</u>: Spec. List <u>Comment:</u> PC (18:1/20:4) putative; [M+H]+; Alterations in phospholipid and fatty acid lipid profiles in primary neocortical cells during oxidant-induced cell injury; http://dx.doi.org/10.1016/j.cbi.2008.05.028 <u>3 largest peaks</u>:



<u>Name:</u> Agilent Ion Trap XCT ESI; [M-H]-; PI 38:4 <u>MW:</u> 885 <u>ID#:</u> 34 <u>DB:</u> Spec. List <u>Comment:</u> PI 38:4; Prec: 750.6; Analysis of phospholipid species in rat peritoneal surface layer by liquid chromatography/electrospray ionization ion-trap mass spectrometry ;http://dx.doi.org/10.1016/j.bbalip.2006.03.022 <u>5 largest peaks:</u>

NIST MS Search 2.0 - [Peptide, Presearch Default - 10 spectra]	
File Search View Tools Options Window Help	_ l=l
# Src. Name	100 303.3 Name: Agilent Ion Trap XCT ESI; [M-H]; PI 38:4
27 L Agilent 6530 QqT0F; [M+H]+; PC 36:2 28 L Agilent 6530 QgT0F; [M+H]+; PC/PE mix	Comment: PI 38:4; Prec: 750.6; Analysis of phospholipid species in rat peritc
29 L Agilent 6530 QqTOF; [M+H]+; PE+PC MIX	
30 L Agilent 6530 QqT0F; [M+NH4]+; TG 54:3 31 L Agilent LC/MSD 1100 lon Tran : [M-H]+; PE 34:1	380 760 5 m/z Values and Intensities:
32 L Agilent Ion Trap SL; [M+Na]+; PC 34:1	Spec. List) Aglient Ion Trap?
33 L Agilent Ion Trap XCT; [M+H]+; NA 24 L Agilent Ion Trap XCT ESI: [M H] - PI 29:4	
35 L Agilent Ion Trap XCT ESI; [M+H]+; PE 34:2	100- 303.3
36 L Agilent Ion Trap XCT ESI; [M-H]-; PI 38:4	
20 L Agrent tion Trap Act Est (M-H): P1 36.4	50
Names / Structures / Spec List	259.3 331 3 484.3
aculcoa+mol+boos: aculcoa+mol-boos: mainlib: aculcarritine+custom+mol+boos: aculcarritine+mol+boos:	436.3
acylcarnitine+mol+napos; custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp;	
respectdbinmdiformat; 365886 total spectra 10-	
	50- 419.25640
	100-283.26354 599.31979
	300 360 420 480 540 600 660 720 780 840 900
# Library Score Dot Product Prob. (%) Rev-Dot Name	Agilent Ion Trap XCT ESI; [M-H];: Head to Tail MF=37 RMF=331 🔽 PI 38:4; [M-H]-: GPIns(18:0/20:4(
1 lipidblast-neg 37 331 12.2 438 PI 38:4; [M-H]-; GPIns(18:0/20:4(5E,8E,11E,14E	Difference A Head to Tail Side by Side A Subtraction / 37 331R 12.2P
2 lipidblast-neg 37 331 12.2 438 PI 38:4; [M-H]-; GPIns(18:0/20:4(5Z,8Z,11Z,14Z)	1283 26354 599 31979 Name: PI 38:4; [M-H]; GPIns(18:0/20:4(5E,8E,111
4 linidblast-neg 37 331 12.2 436 PI 36.4, [M-H]-; GPIns(16.0/20.4(7E, 10E, 13E, 16	100-203.2034 333.1313 MW: 885 ID#: 119419 DB: lipidblast-neg
5 lipidblast-neg 37 331 12.2 438 Pl 38:4; [M-H]; GPIns(20:4(5Z,8Z,11Z,14Z)/18:(	8 largest peaks:
6 lipidblast-neg 37 331 12.2 438 PI 38:4; [M-H]-; GPIns(20:4(7E,10E,13E,16E)/18	
7 lipidblast-neg 24 239 7.89 707 MGDG 44:6; [M-H]-; MGDG(22:2[132,162]/22:4]	0 8 m/z Values and Intensities:
0 inprovidescriteg 24 233 7.03 707 MidDid 44.0, [m+n]+, MidDid (22.4(72,102,132,16 €	280 420 560 700 840 283.20394 533.00 sni FA ▼
Names Structures Hit List	Plot/Text of Hit / Plot of Hit /
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide

Name: Agilent Ion Trap XCT ESI; [M+H]+; PE 34:2 <u>MW:</u> 714 <u>ID#:</u> 35 <u>DB:</u> Spec. List <u>Comment:</u> PE 34:2; PE(16:0/18:2); Analysis of phospholipid species in rat peritoneal surface layer by liquid chromatography/electrospray ionization ion-trap mass spectrometry ;http://dx.doi.org/10.1016/j.bbalip.2006.03.022 4 largest peaks:



Name: Agilent Ion Trap XCT ESI; [M-H]-; PI 38:4 <u>MW:</u> 885 <u>ID#:</u> 36 <u>DB:</u> Spec. List <u>Comment:</u> PI 38:4; PI(18:0/20:4); Prec: 885.6; Analysis of phospholipid species in rat peritoneal surface layer by liquid chromatography/electrospray ionization ion-trap mass spectrometry ;http://dx.doi.org/10.1016/j.bbalip.2006.03.022 8 largest peaks:



Name: Agilent Ion Trap XCT ESI; [M-H]-; PS 38:4 <u>MW:</u> 810 <u>ID#:</u> 134 <u>DB:</u> Text File <u>Comment:</u> PS 38:4; PS(18:0/20:4); Prec:810.5; Analysis of phospholipid species in rat peritoneal surface layer by liquid chromatography/electrospray ionization ion-trap mass spectrometry ;http://dx.doi.org/10.1016/j.bbalip.2006.03.022 <u>6 largest peaks:</u>



Name: Agilent MSD 1100 single quadrupole MS; [M+H]+; PE 32:0 <u>MW:</u> 692 <u>ID#:</u> 38 <u>DB:</u> Spec. List <u>Comment:</u> PE (16:0/16:0); [M+H]+; Quantitation and characterization of phospholipids in pharmaceutical formulations by liquid chromatography-mass spectrometry; http://dx.doi.org/10.1016/S0021-9673(00)00148-5 7 largest peaks:

Hit score lower than 20, or 100 in dot product, non-hit, also obtained from single quad, no-MSMS



Name: Agilent QTOF; [M+NH4]+; TG <u>MW:</u> 896 <u>ID#:</u> 39 <u>DB:</u> Spec. List <u>Comment:</u> TAG LLL 878; +NH4=896; No accurate precursor assigned, 0.8 Da error;Maximizing Detection of Complex Hydrophobic Lipids: Optimization Efficiency and Nano-Chromatography; ASMS 2009 David A. Weil1, Michael Woodman and Carol Ball, Agilent Technologies Inc.,Schaumburg, IL1 and Raleigh, NC2 <u>3 largest peaks:</u>

no precursor assigned, hit score = 0

Efe         Search         Yew         Total         Search         Yew         Ye	🖷 NIST MS Search 2.0 - [Peptide, Presearch Default - 2 spectra]	
Image: Appler Line: Image: Example 520 0p10F; [M+NH4]+: T6       Image: Appler Line: Appler 1010F; [M+NH4]-: T6       Image: Appler Line: Appler 1010F; [M-NH4]-: T6       Image: Appler Line: Apple: T010F; [M-NH4]-: T6       Image: Apple	I File Search View Iools Options Window Help	
Image: Source         Totagient QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+NH4]+: T6           Image: Applent QTDF: [M+NH4]+: T6         Image: Applent QTDF: [M+1]+: N4         Image: Applent QTDF: [M+1]+: N4           Image: Applent QTDF: [M+1]+: P8 344         Image: Applent QTDF: [M+1]+: P8 344         Image: Applent QTDF: [M+1]+: P8 344           Image: Applent QTDF: [M+1]+: P8 344         Image: Applent QTDF: [M+1]+: P8 344         Image: Applent QTDF: [M+1]+: P8 344           Image: Image: QTDF: Image: Applent QTDF: [M+1]+: P8 344         Image: QTDF: QTDF: [M+1]+: P8 344         Image: QTDF: QTDF: [M+1]+: P8 344           Image: QTDF: Image: QTDF: Image: QTDF: QTDF: P8 344         Image: QTDF: QTDF: QTDF: QTDF: QTDF: QTDF: QTDF: QTDF: QT	1 🗈 🖻 ở 🏭 🕎 📲 🖽 🚧 🔶 🦻	
#         Sice         Name         Sign: Aglent IC/Si 00 qTOF; [M+NH4+; T6 54:3           31         L         Aglent IC/Si 00 qTOF; [M+NH4+; T6 54:3         1 <t< th=""><th>© &gt; ≥ = 1. Agilent QTOF; [M+NH4]+; TG S</th><th></th></t<>	© > ≥ = 1. Agilent QTOF; [M+NH4]+; TG S	
37       L       Aglert Ion Trap XCT ESI; [M+H]: P1 38.4         38       L       Aglert Ion Trap XCT ESI; [M+H]: P1 38.4         38       L       Aglert Ion Trap XCT ESI; [M+H]: P1 38.4         38       L       Aglert Ion Trap XCT ESI; [M+H]: P1 38.4         39       L       Aglert ND TO Signed WS; [M+H]: P1 38.4         30       L       Aglert ND TO Signed WS; [M+H]: P1 38.4         31       L       Butter Existic 2000 in the ESI FICR APEXAg. [M+H]: P1 38.4         41       L       Butter Existic 2000 in the ESI FICR APEXAg. [M+H]: PC MX         41       L       Butter Existic 2000 in the ESI FICR APEXAg. [M+H]: PC MX         41       Dutter Existic 2000 in the ESI FICR APEXAg. [M+H]: PC MX         41       Dutter Existic 2000 in the ESI FICR APEXAg. [M+H]: PC MX         41       Dutter Existic 2000 in the ESI FICR APEXAg. [M+H]: PC MX         41       Dutter Existic 2000 in the ESI FICR APEXAg. [M+H]: PC MX         420 stand Spectra       Spec List         430 stand Spectra       Spec List         430 stand Spectra       Spec List         441 spectra       Spec List         533499 total spectra       Spec List         1       Ipidblast-reg       70         500       70       50.0         700 <th>#         Src.         Name           31         L         Agilent 6530 QqTOF; [M+NH4]+; TG 54:3           32         L         Agilent LC/MSD 1100 Ion Trap; [M+H]; PE 34:1           33         L         Agilent Ion Trap SL; [M+Na]+; PC 34:1           34         L         Agilent Ion Trap XCT; [M+H]+; NA           35         L         Agilent Ion Trap XCT ESI; [M+H]+; PI 38:4           36         L         Agilent Ion Trap XCT ESI; [M+H]+; PI 34:2</th> <th>100-       599.5         100-       599.5         100-       599.5         100-       380 [D#; 39 DB; Spec. List         100-       100-</th>	#         Src.         Name           31         L         Agilent 6530 QqTOF; [M+NH4]+; TG 54:3           32         L         Agilent LC/MSD 1100 Ion Trap; [M+H]; PE 34:1           33         L         Agilent Ion Trap SL; [M+Na]+; PC 34:1           34         L         Agilent Ion Trap XCT; [M+H]+; NA           35         L         Agilent Ion Trap XCT ESI; [M+H]+; PI 38:4           36         L         Agilent Ion Trap XCT ESI; [M+H]+; PI 34:2	100-       599.5         100-       599.5         100-       599.5         100-       380 [D#; 39 DB; Spec. List         100-       100-
Names         Spec List           acylcoarmol+hpos; acylcoarmol+hops; acylcoarmo	37       L       Agilent Ion Trap XCT ESI; [M-H]; PI 38:4         38       L       Agilent Ion Trap XCT ESI; [M-H]; PI 38:4         39       L       Agilent MSD 1100 single quadrupole MS; [M+H]+; PE 32:0         40       L       Agilent QTOF; [M+NH4]+; TG         41       L       Bruker DESI FTICR APEX-Q, [M+H]+; PC MIX         41       L       Bruker DESI FTICR APEX-Q, [M+H]+; PC MIX	100-     599.5       50-     50-
#       Library       Score       Dot Product       Prob. (%)       Rev-Dot       Name         1       lipidblast-neg       0       70       50.0       707       MGDG 44:1; [M-H]; MGDG(22:0/22:1(132))         2       lipidblast-neg       0       70       50.0       707       MGDG 44:1; [M-H]; MGDG(22:0/22:1(132))         37.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)       Name; MGDG 44:1; [M-H]; MGDG(22:0/22:1(132)/22:0)         100       337.31046       Name; MGDG 4	Names (Structures / Spec List acylcoa+mol+hpos; acylcoa+mol-hneg; mainlib; acylcarnitine+custom+mol+hpos; acylcarnitine+mol+hpos; acylcarnitine+mol+napos; custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; 359499 total spectra	0 <u>337.27</u> 50
	#         Library         Score         Dot Product         Prob. (%)         Rev-Dot         Name           1         lipidblast-neg         0         70         50.0         707         MGDG 44:1; [M-H]-; MGDG(22:0/22:1(13Z))           2         lipidblast-neg         0         70         50.0         707         MGDG 44:1; [M-H]-; MGDG(22:1(13Z)/22:0)	337.31046         300       360       420       480       540       600       660       720       780       840       900         ▲ Agilent OTDF: [M+NH4]+: TG       Head to Tail MF=0 RMF=70       ▼MGDG 44:1: [M+H]: MGDG[22:07         Difference       Head to Tail       Side by Side       Subtraction       0       708       600         100-       337.31046       Name: MGDG 44:1; [M+H]: MGDG[22:07/22:1[132 ▲         100-       337.31046       Name: MGDG 44:1; [M-H]: MGDG[22:07/22:1[132 ▲         50-       2 largest peaks:       337.31046       99.00         339.32610       99.00
Image: Structures     0     337.31046     999.00     sn2 FA       280     420     500     700     840     339.32610     999.00     sn1 FA       Init List	Names <u>Structures</u> Hit List     Hit List     Lib. Search Other Search Names Compare Librarian MSMS	0         280         420         560         700         840         337.31046         999.00         sn2 FA           (lipidblast-neg) MGDG         44:1; [M-H]; MGDG(22:0/22:1(1)         Image: Constraint of the state

Name: Bruker DESI FTICR APEX-Q, [M+H]+; PC MIX <u>MW</u>: 758 <u>ID#</u>: 40 <u>DB</u>: Spec. List <u>Comment:</u> PC MIX PC (34:2); 758.5694; [M+H]+; Characterization of DESI-FTICR mass spectrometry from ECD to accurate mass tissue analysis <u>9 largest peaks</u>:

PC -MIX



Name: Bruker Esquire 3000 ion trap; ESI, [M-H]-; LipidA-PP 56:26:0 <u>MW</u>: 1796 <u>ID#</u>: 41 <u>DB</u>: Spec. List <u>Comment:</u> LipidA-PP 56:26:0; LipidA-PP [14/14/14/3O-(12)/3O-(14)]; [M+H]+; Prec. m/z: 1796.2;Gram-negative bacterial lipid A analysis by negative electrospray Gram-negative bacterial lipid A analysis by negative electrospray Gram-negative bacterial lipid A analysis by negative electrospray; International Journal of Mass Spectrometry 249250 (2006) 7792 <u>10 largest peaks</u>:



Name: Bruker Esquire 3000 ion trap; ESI, [M-H]-; LipidA-PP 56:26:0 <u>MW:</u> 1796 <u>ID#:</u> 42 <u>DB:</u> Spec. List <u>Comment:</u> LipidA-PP 56:26:0; LipidA-PP [14/14/14/3O-(12)/3O-(14)]; [M+H]+; Prec. m/z: 1796.2;Gram-negative bacterial lipid A analysis by negative electrospray Gram-negative

bacterial lipid A analysis by negative electrospray Gram-negative bacterial lipid A analysis by negative electrospray; International Journal of Mass Spectrometry 249250 (2006) 7792

10 largest peaks:

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X 🖻 💼 🚭 🏭 📲 🖽 🚧 🔶 🖌	
🔞 🍉 😅 🚔 1. Bruker Esquire 3000 ion trap; ESI, [M 🔽 😥 🖳 🔍 🍭	
#         Src.         Name           36         L         Agilent Ion Trap XCT ESI; [M+H]+; PE 34:2           37         L         Agilent Ion Trap XCT ESI; [M-H]; PI 38:4	Name:         Bruker Esquire 3000 ion trap; ESI, [M-H]-; LipidA-PP 56:26:0           100-         1035.6         1470           1035.6         1470         201/201/201/201/201/201/201/201/201/201/
38         L         Agilent Ion Trap XCT ESI; [M-H]-; PI 38:4           39         L         Agilent MSD 1100 single quadrupole MS; [M+H]+; PE 32:0           40         L         Agilent QTOF; [M+NH4]+; TG           41         L         Bruker DESI FTICR APEX-Q, [M+H]+; PC MIX	0         1         1088.2         999.00         1470         400.00         1035.6         333.00         1243.8         333.00         101;           1410         999.6         180.00         1225.8         100.00         1261.8         100.00         1454.0         50.00         1495.           Ispec. List) Bruker Esquire 3:         Image: Construction of Search Sections         Image: Construction of Sections         Image: Constructing : Constructions         Image: Construction
42         L         Bruker Esquire 3000 ion trap; ESI, [M-H]-; LipidA-PP 56:26:0           43         L         Bruker Esquire 3000 ion trap; ESI, [M-H]-; LipidA-PP 56:26:0           44         L         Bruker Esquire ion trap; ESI MS/MS, [M+H]+; PC 34:1           45         L         Bruker Esquire ion trap; ESI MS/MS, [M-H]-; DGDG 36:8	100- 1698.2
46 L Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; NA  46 L Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; NA  47 Names Structures  Spec List	50- 1035.6 1243.8 1470
acylcoa+mol+hpos; acylcoa+mol-hneg; mainlib; acylcarnitine+custom+mol+hpos; acylcarnitine+mol+hpos; acylcarnitine+mol+napos; custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; 359499 tota 1000-	
100- 10- 1-	50- 1698.23467
#         Library         Score         Dot Product         Prob. (%)         Rev-Dot         Name           1         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/10/18/30-(12)]	Image: Provide and to Tail         Figure 3000 ion trap;         ESI.         Head to Tail         MF=167 RMF=436         Image: Provide Arrow 100 rmin and 1
2         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/10/18/30-[14           3         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/12/16/30-[12           4         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/12/16/30-[12           4         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/12/16/30-[12	100- 1552.00785 Name: LipidA-PP 56:26:0; LipidA-PP [14/14/14/14] MW: 1796 [D#: 87941 DB: lipidblast-neg Comment: Parent=1796 21157 Mz exact=1796 21
5         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/14/14/30-(12           6         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/14/14/30-(12           7         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/14/14/30-(14           7         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/14/14/30-(14           7         lipidblast-neg         167         436         0.99         592         LipidA-PP 56:26:0; LipidA-PP [14/14/14/14/14/14/14/14/14/14/14/14/14/1	1698.23467         9 largest peaks:           50-         1552.00785         999.00   1698.23467         600.00   175           111         11         1454.03095         250.00   1568.00277         250.00   155
r         inploudastrieg         for         436         0.33         332         LipidA-PP (14/14/16/12/30-(12/3	0 9 m/2 Values and Intensities: 1080 1260 1440 1620 1800 (lipidblast-neg) LipidA-PP 56:26:0; LipidA-PP [14/14/14]
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide //

## Name: Bruker Esquire ion trap; ESI MS/MS, [M+H]+; PC 34:1 <u>MW</u>: 760 <u>ID#</u>: 43 <u>DB</u>: Spec. List <u>Comment</u>: PC 34:1; PC(16:0/18:1); [M+H]+; Prec. m/z: 760.7; Developmental profiling by mass spectrometry of phosphocholine containing phospholipids in the rat nervous system reveals temporo-spatial gradients; DOI: 10.1111/j.1471-4159.2010.06836.x 5 largest peaks:



<u>Name:</u> Bruker Esquire ion trap; ESI MS/MS, [M-H]-; DGDG 36:8 <u>MW:</u> 931 <u>ID#:</u> 44 <u>DB:</u> Spec. List <u>Comment:</u> DGDG(18:4/18:4); DGDG 36:8; [M-H]-; Prec. m/z: 931.8; Chimica e biologia a confronto: pigmenti e altri metaboliti secondari prodotti da dinoflagellati del Lago di Tovel; Studi Trent. Sci. Nat., Acta Biol., 81 (2004), Suppl. 2: 413-426;Rita FRASSANITO, Ines MANCINI & Graziano GUELLA <u>5 largest peaks:</u>

🕌 NIST MS Search 2.0 - [Peptide, Presearch Default - 4 spectra]	
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	[
#         Src.         Name         931.8         Name: Bruker Esquire ion trap; E           100         931.8         MW/ 931 ID#: 44 DB: Spec List	SI MS/MS, [M-H]-; DGDG 36:8
36 L Aglient Ion Trap XCT ESI; [M+H]+; PE 34:2 37 L Aglient Ion Trap XCT ESI; [M+H]+; PE 34:2 Comment: DGDG(18:4/18:4); DC	GDG 36:8; [M-H]-; Prec. m/z: 931.8; Chimic.
38 L Agilent Ion Trap XCT ESI; [M-H]; PI 38:4	
39 L Agilent MSD 1100 single quadrupole MS; [M+H]+; PE 32:0 390 780 5 m/z Values and Intensities:	
41 L Bruker DESI FTICR APEX-Q, [M+H]+; PC MIX	Plot/Text of Spec List /
42 L Bruker Esquire 3000 ion trap: ESI, [M-H]; LipidA-PP 56:26:0	
43 L Bruker Esquire Jobo for trap; ESI MS/MS, [M+H]+; PC 34:1	673.7 931.8
45 L Bruker Esquire ion trap; ESI MS/MS, [M-H]; DGDG 36:8	
40 L Brutes Esquite for day, Estimotives, Internative, NA 47 L Brutes Esquite for the for the DALUE ACCE 2000	
Names / Structures / Spec List 275.5	
custompc+hpos msp: custompc+napos msp: hilic-urine: lipidblast-neg: pc-ac-neg msp: pc-form-neg msp: lipidblast-pos: 235370	
total spectra	· · · · · · · · · · · · · · · · · · ·
10-	
# Library Score Dot Product Prob. (%) Rev-Dot Name	F=381 ITDGDG 36:8; [M-H]-; DGDG(18:4(6
1 lipidblast-neg 12 381 25.0 1e+3 DGDG 36:8; [M-H]-; DGDG(18:4(6Z,9Z,12Z,15Z)/1{ Difference Head to Tail Side by Side Subtraction /	12 381R 25.0P
2 lipidblast-neg 12 381 25.0 1e+3 DGDG 36.8; [M-H]-; DGDG(18:4(62,92,122,152)/18	DGDG 36:8; [M-H]-; DGDG(18:4(6Z,9Z,12Z
4 linidblast-neg 12 381 25.0 1e+3 DGDG 36.8; (M-H]; DGDG(18:4(3E,11E,13E,15E)/ 100-	031 <u>ID#:</u> 76774 <u>DB:</u> lipidblast-neg ent: Parent=931 54192 Mz_exact=931 5415
	ist peaks:
2/5.2 1 m/z	Values and Intensities:
	0098 999.00 sn1 FA    sn2 FA
300 450 600 750 900 3000	
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For Help, press F1 Peptide	Peptide

<u>Name:</u> Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; NA <u>MW:</u> 374 <u>ID#:</u> 45 <u>DB:</u> Spec. List <u>Comment:</u> MG(18:1); MG(16:0); [M+NH4]+; Prec. m/z: 374.3; Identification of molecular species of simple lipids by normal phase liquid chromatographypositive electrospray tandem mass spectrometry, and application of developed methods in comprehensive analysis of low erucic acid rapeseed oil lipids <u>4 largest peaks:</u>



Name: Bruker Esquire ion trap; ESI MS/MS, [M-H]-; MGDG 38:9 <u>MW:</u> 795 <u>ID#:</u> 46 <u>DB:</u> Spec. List <u>Comment:</u> MGDG(20:5/18:4); MGDG 38:9; [M-H]-; Prec. m/z: 795.5; Chimica e biologia a confronto: pigmenti e altri metaboliti secondari prodotti da dinoflagellati del Lago di Tovel; Studi Trent. Sci. Nat., Acta Biol., 81 (2004), Suppl. 2: 413-426;Rita FRASSANITO, Ines MANCINI & Graziano GUELLA 6 largest peaks:



## <u>Name:</u> Bruker Esquire ion trap;ESI MS/MS, [M+Na]+; MGDG 38:9 <u>MW:</u> 819 <u>ID#:</u> 47 <u>DB:</u> Spec. List <u>Comment:</u> MGDG(20:5/18:4); MGDG 38:9; [M+Na]+; Prec. m/z: 819.7; Chimica e biologia a confronto: pigmenti e altri metaboliti secondari prodotti da dinoflagellati del Lago di Tovel; Studi Trent. Sci. Nat., Acta Biol., 81 (2004), Suppl. 2: 413-426;Rita FRASSANITO, Ines MANCINI & Graziano GUELLA

3 largest peaks:

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🔞 🍃 🚔 🚔   1. Bruker Esquire ion trap;ESI MS/MS, 💽 🛞 🖺 🛞 🔍	
	Name: Bruker Esquire ion tran: ESLMS/MS_IM+Nal+: MGDG-38:9
39 L Agilent MSD 1100 single quadrupole MS; [M+H]+; PE 32:0	100- 517.5 MW: 819 ID#: 47 DB: Spec. List
40 L Agilent QTOF; [M+NH4]+; TG 41 D D D S S S S S S S S S S S S S S S S	3 largest peaks:
41 L Bruker DEST FTICH APEX-Q, [M+H]+; PC MIX 42 L Bruker Esquire 3000 ion trap; ESI, [M-H]-; LipidA-PP 56:26:0	517.5 999.00   819.7 600.00   543.5 250.00   540 720 3m/z Values and Intensities:
43 L Bruker Esquire 3000 ion trap; ESI, [M-H]; LipidA-PP 56:26:0	(Spec. List) Bruker Esquire io
44 L Bruker Esquire ion trap; ESI MS/MS, [M+H]+; PC 34:1 45 L Bruker Esquire ion trap: ESI MS/MS. [M-H]-: DGDG 36:8	Plot/Text of Search Spectrum Plot of Search Spectrum A Plot/Text of Spec List
46 L Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; NA	1 517.5
47 L Bruker Esquire ion trap; ESI MS/MS, [M-H]+; MGDG 38;9 48 L Bruker Esquire ion tran: ESI MS/MS, [M+Na]+; MGDG 38;9	100-
49 L Bruker microTOF qQ-TOF; NA	819.7
	50-
Names Spec List	543.5
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	
100-	
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# Library Score Dot Product Prob. (%) Rev-Dot Name	ST0 540 570 600 630 660 690 720 750 780 810
1 lipidblast-pos 131 944 22.4 949 MGDG 38:9; [M+Na]+; MGDG(18:4(6Z,9Z,12Z,1	Difference A Head to Tail Side by Side A Subtraction / 131 944R 22.4P
2 lipidblast-pos 131 944 22.4 949 MGDG 38:9; [M+Na]+; MGDG(18:4(9E,11E,13E,	Name: MGDG 38:9; [M+Na]+; MGDG(18:4(6Z.9Z,
3 lipidblast-pos 131 944 22.4 949 MGDG 38:9; [M+Na]+; MGDG(20:5(5Z,8Z,11Z,1	100- 100- <u>MW:</u> 819 <u>ID#</u> , 12656 <u>DB</u> ; lipidblast-pos <u>Converse</u> Baset-919 50227 Ma. susat-919 5022
<ul> <li>4 ipidulas:rpos</li> <li>131</li> <li>544</li> <li>544</li> <li>545</li> <li>644</li> <li>644</li> <li>745</li> <li>746</li> <li>747</li> <li>744</li> <li>744</li> <li>744</li> <li>745</li> <li>744</li> <li>745</li> <li>744</li> <li>745</li> <li>744</li> <li>744</li> <li>745</li> <li>744</li> <li>744</li> <li>745</li> <li>744</li> <li>744</li> <li>745</li> <li>745</li> <li>746</li> <l< td=""><td>2 largest peaks:</td></l<></ul>	2 largest peaks:
6 lipidblast-pos 9 513 1.10 577 PA 42:7; [M+Na2:H]+; GPA(22:2(13Z,16Z)/20:5(	504 517.27793 999.00   543.29357 999.00   2 m/z Values and Intensities
7 lipidblast-neg 2 222 0.84 250 PG 39:0; [M-H]-; GPGro(14:0/25:0)	0 517.27793 999.00 [M+Na]-sn2
8 lipidblast-neg 2 222 0.84 250 PG 40:7; [M-H]; GPGro(20:2(112,142)/20:5(52,1	560 630 700 770 543.29357 999.00 [M+Na]-sn1
Names Structures Hit List	[IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII
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or Help, press F1	Peptide Peptide

<u>Name:</u> Bruker microTOF qQ-TOF; NA <u>MW:</u> 744 <u>ID#:</u> 48 <u>DB:</u> Spec. List <u>Comment:</u> PC(16:0/18:1) wrongly assigned?; 744.4; [M+H]+; MS/MS mass spectra of 1palmitoyl-, 2-oleyl-phosphatidycholine; Hyphenated Tools for Lipidomics;Jan Willmann1,Herbert Thiele2, Dieter Leibfritz1;HUPO 2007, Poster M-195 <u>5 largest peaks:</u>

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6 b 2 i i Bruker microTOF qQ-TOF; NA 🕑 🖳 🔎 🖳 🔍	
#       Stc.       Name         45       L       Bruker Esquire ion trap; ESI MS/MS; [M+NH4]+; NA         46       L       Bruker Esquire ion trap; ESI MS/MS; [M+NH4]+; MS/MS mass         47       L       Bruker Esquire ion trap; ESI MS/MS; [M+NH4]+; MS/MS mass         48       L       Bruker microTOF qQ-TOF; NA         49       L       Bruker ultrafeski IMALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0)         50       L       Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0)         51       L       Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0)         52       L       Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SO20         53       L       Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SO20         54       L       Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SO20         54       L       Bruker ultraFlex II MALDI TOF-TOF; [M+Na]+; SO20         53       L       Bruker ultraFlex II MALDI TOF-TOF; [M+Na]+; SO20         54       L       Bruker ultraFlex II MALDI TOF-TOF; [M+Na]+; SO20         55       L       Bruker ultraFlex II MALDI TOF-TOF; MH3]+; SO20         54       L       Bruker ultraFlex II MALDI TOF-TOF; MH4; IV-DC 24:1         56       Bruker ultraFlex II MALDI TOF-TOF; MH4; IV-DC 24:1       IMALDI TOF-TOF; MH4; IV-DC 24:1	× 5 ▼ ▼ ↓
	<u>_</u>
200 320 360 400 440 480 520 560 600 640 680 720	718-1
1       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(11E))         2       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(11Z))         3       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(13Z))         4       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(13Z))         5       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(17Z))         6       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(17Z))         7       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(12Z))         7       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(7Z))         7       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(7Z))         8       lipidblast-neg       86       676       2.37       700       PE 36:1; [M-H]; GPEtn(18:0/18:1(7Z))         8       lipidblast-neg       86       676	77₽ 43 .2 ▼
Peptide Peptide Deptide	

Name: Bruker ultraflex II MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0) <u>MW:</u> 725 <u>ID#:</u> 49 <u>DB:</u> Spec. List <u>Comment:</u> SM 34:1; SM(d18:1/16:0); Prec. m/z: 725.6; [M+Na]+;Comparison of Phospholipid Molecular Species between Terminal and Stem Villi of Human Term Placenta by Imaging Mass Spectrometry;Placenta 31 (2010) 245248; http://dx.doi.org/10.1016/j.placenta.2009.12.026 <u>4 largest peaks:</u>



## <u>Name:</u> Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; DGDG 30:0 <u>MW:</u> 887 <u>ID#:</u> 50 <u>DB:</u> Spec. List <u>Comment:</u> DGDG 30:0; Prec. m/z: 887.7; [M+Na]+; Lipid compositions in Escherichia coli and Bacillus subtilis during growth as determined by MALDI-TOF and TOF/TOF mass spectrometry; <u>6 largest peaks:</u>

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# Src. Name Style UltraFlex II MALDI TOF-TOF; [M+Na]+; DGDG 30:0	
45 L Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; NA 46 L Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; Lipid compositions in E 887.7 Comment: DGDG 30:0; Prec. m/z: 887.7; [M+Na]+; Lipid compositions in E	Es(
47 L Bruker Esquire ion trap; ESI MS/MS, [M+Na]+; MGDG 38:9	15
48 L Bruker microTOF qQ-TOF; NA 49 L Bruker ultraflex II MALDI TOF/TOF: [M+Na]+: SM(d18:1/16:0)	
50 L Bruker UltraFlex II MALDI TOF-TOF: [M+Na]+: DGDG 30:0 Plot/Text of Search Spectrum Plot of Search Spectrum Plot of Search Spectrum Plot of Search Spectrum Plot. Example 2:1	
51 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 31:0	-
53 L Bruker UltraFlex II MALDI TOF-TOF; [M+K]+; PC 36:1 54 J Bruker UltraFlex II MALDI TOF-TOF; [M+K]+; SM(d18:1/16:0)	
55 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; lysoPC 18:0 ↓	77
Names Structures Spec List 323.1 382.2 731.1	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; lipidblast-pos; 235370	
10-	
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	900
	G(15: 9.4P
2 lipidblast-pig 9 131 10.2 438 PI 38:3; [M-H]-; GPIns(18:3(6Z,9Z,12Z)/20:0)	
3 lipidblast-neg 9 131 10.2 438 Pl 38:3; [M-H]-; GPIns(18:3(92,122,152)/20:0) 483.29351 645.34633 MV: 887 ID#: 4747 DB; lipidblast-pos	" 41
4 lipidblast-neg 9 131 10.2 438 PI 38:3; [M-H]-; GPIns(20:0/18:3(6Z,9Z,12Z)) 5 linidblast-neg 9 131 10.2 438 PI 38:3: [M-H]-; GPIns(20:0/18:3(6Z,9Z,12Z)) 2 largest peaks:	/07
483,29351 999,00   645,34633 999,00   2 m/2 (Aluee and Intervitien:	
0 <sup>1</sup> 483.29351 999.00 [M+Na]-sn1-C6H1005 (-162	2)1
390 520 650 780 645.34633 999.00 [M+Na]-sn2	₋่่
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<u>Name:</u> Bruker UltraFlex II MALDI TOF-TOF; DGDG 31:0 <u>MW:</u> 901 <u>ID#:</u> 51 <u>DB:</u> Spec. List <u>Comment:</u> DGDG 31:0; Prec. m/z: 901.7; [M+Na]+; Lipid compositions in Escherichia coli and Bacillus subtilis during growth as determined by MALDI-TOF and TOF/TOF mass spectrometry; <u>8 largest peaks:</u>

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45 L Bruker Esquire ion tran: ESLMS/MS_IMANIA/1+: NA	45.4 901.7 MW: 901 ID#: 51 DB: Spec. List	
46 L Bruker Esquire ion trap; ESI MS/MS, [M-H1]-; MGDG 38:9	Comment: DGDG 31:0; Prec. m/z: 901.7; [M+Na]+; Lipid compositions in Esc	
47 L Bruker Esquire ion trap;ESI MS/MS, [M+Na]+; MGDG 38:9		
49 L Bruker ultraflex II MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0)	680 745.2 300.00 523.4 100.00 405.2 50.00	
50 L Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; DGDG 30:0	arch Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /	
51 L Bruker UltraFlex II MALDI TOF-TOF: DGDG 31:0		
53 L Bruker UltraFlex II MALDI TOF-TOF; [M+K]+; PC 36:1 100-	645.4	
54 L Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0) 55 L Bruker ultrafleXtreme Mál DI TOF/TOF: [M+H]+; lusoPC 18:0	551.4	
	331.4	
Names Structures Spec List 327.1	745.2	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	405.2 523.4	
total spectra		
1 50-		
	483,29357 645,34639	
	400 450 500 550 600 650 700 750 800 850 900	
# Library Score Dot Product Prob. (%) Rev-Dot Name	ex II MALDI TOF-TCI Head to Tail MF=219 RMF=504 (▼DGDG 31:0; [M+Na]+; DGDG(15;	
1 lipidblast-pos 219 504 38.8 697 DGDG 31:0; [M+Na]+; DGDG(15:0/16:0)	ead to Tail Side by Side Subtraction / 219 504R 38.8P	
2 lipidblast-pos 219 504 38.8 697 DGDG 31:0; [M+Na]+; DGDG(16:0/15:0)	83 29357 645 34639	
4 linidblast-pos 125 340 1.76 577 TG 55:5: [M+1 i]+: TG(16:0/16:0/22:5)	<u>MW:</u> 901 ( <u>D#:</u> 4/49 <u>DB:</u> lipidblast-pos Comment: Parent=901 58647 Mz_exact=901 5864	
5 lipidblast-pos 125 340 1.76 577 TG 55:5; [M+Li]+; TG(16:0/17:1/22:4)	4 largest peaks:	
6 lipidblast-pos 125 340 1.76 577 TG 55:5; [M+Li]+; TG(16:0/17:2/22:3)	483.29357 999.00   497.30921 999.00   645.3 4 // 4 // 2 // 2 // 4 // 2 // 2 // 2 /	
7 lipidblast-pos 125 340 1.76 577 TG 55:5; [M+Li]+; TG(16:0/19:0/20:5)	483.29357 999.00 [M+Na]-sn2-C6H1005 (-162)	
8 iipidblast-pos 120 340 1.76 577 1G 54:6; [M+Naj+; 1G(16:0716:1722:5) 390	520 650 780 910 497.30921 999.00 [M+Naj-sn1-C6H1005 (-162)	
Names (Structures / Hit List Plot/Text of Hit .	CDG 31(0; [M+Na]+; DGDG[15(0/16)]     Plot of Hit /	
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Peptide Peptide		

<u>Name:</u> Bruker UltraFlex II MALDI TOF-TOF; DGDG 32:0 <u>MW:</u> 915 <u>ID#:</u> 52 <u>DB:</u> Spec. List <u>Comment:</u> DGDG 32:0; Prec. m/z: 915.7 [M+Na]+; Lipid compositions in Escherichia coli and Bacillus subtilis during growth as determined by MALDI-TOF and TOF/TOF mass spectrometry; <u>5 largest peaks:</u>



Name: Bruker UltraFlex II MALDI TOF-TOF; [M+K]+; PC 36:1 <u>MW:</u> 826 <u>ID#:</u> 53 <u>DB:</u> Spec. List <u>Comment:</u> PC 36:1; Prec. m/z: 826.5723; [M+K]+; Solvent-Free Matrix Dry-Coating for MALDI Imaging of Phospholipids ;Satu M. Puolitaival, Kristin E. Burnum, D. Shannon Cornett, Richard M. Caprioli; doi:10.1016/j.jasms.2008.02.013 <u>4 largest peaks:</u> FALSE Positive ID M+K not in LipidBlast



<u>Name:</u> Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0) <u>MW:</u> 725 <u>ID#:</u> 54 <u>DB:</u> Spec. List <u>Comment:</u> SM(d18:1/16:0); Prec. m/z: 725.5; [M+Na]+; MALDI-based imaging mass spectrometry revealed abnormal distribution of phospholipids in colon cancer liver metastasis; http://dx.doi.org/10.1016/j.jchromb.2007.02.037 <u>4 largest peaks:</u>

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# Src. Name	100 666.5 Name: Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0)
45 L Bruker Esquire ion trap; ESI MS/MS, [M+NH4]+; NA 46 L Bruker Esquire ion trap; ESI MS/MS, [M-H1: MGDG 38:9	542.5 542.5
47 L Bruker Esquire ion trap;ESI MS/MS, [M+Na]+; MGDG 38:9	
48 L Bruker microTOF qQ-TOF; NA 49 L Bruker ultraflev II Mól DI TOF/TOF: [M+Na]+: SM(d19:1/16:0)	330 660 4 m/z Values and Intensities: ▼
50 L Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; DGDG 30:0	Spec. List) Bruker Ultra-lex Plot of Search Spectrum A Plot/Text of Spec List /
51 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 31:0	
53 L Bruker UltraFlex II MALDI TOF-TOF; [M+K]+; PC 36:1	100-
54 L Bruker UltraFlex II MALDI TOF-TOF; (M+Na)+; SM(d18:1/16:0) 55 L Bruker ultrafleXizere MALDI TOF (TOF: (M+L)+; largeC 19:0	
SO E Dicker didate-Arene MALDI TOF/TOF/ [W+H]+, [ssore 16.0	542.5
Names Structures Spec List	184.0
custompc+hoos msp: custompc+hoos msp: hilic-urine: lipidblast-hog: pc-ac-hog msp: pc-form-hog msp: lipidblast-pos: 235370	722.5
total spectra	542.49129
10	
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	100-
# Library Score Dot Product Prob. (%) Rev-Dot Name	ABruker UltraFlex II MALDI TOF-TCI Head to Tail MF=271 RMF=831 ITSM 34:1; (M+Na)+; SM(d14:1(4E))
1 lipidblast-pos 271 831 24.9 879 SM 34:1; [M+Na]+; SM(d14:1(4E)/20:0)	Difference A Head to Tail A Side by Side A Subtraction / 271831R 24.9P
2 lipidblast-pos 2/1 831 24.9 8/9 5M 34:1; [M+Naj+; 5M(d16:U/18:1(32)] 3 lipidblast-pos 271 831 24.9 879 5M 34:1; [M+Naj+; 5M(d16:14/E)/18:0)	666 48383
4 lipidblast-pos 271 831 24.9 879 SM 34:1; [M+Na]+; SM(d18:1(4E)/16:0]	100- Comment: Parent=725.55733 Mz_exact=725.5573
5 lipidblast-neg 33 336 0.24 438 PI 26:0; [M-H]-; GPIns(12:0/14:0)	2 largest peaks:
6 lipidblast-neg 33 336 0.24 438 PI 26:0; [M-H]-; GPIns(14:0/12:0)	2 m/z Values and Intensities:
	0 542.49129 542.49129 20.00 [M+Na]-C5H14N04P (-183) 240 200 730 566 48383 999.00 [M+Na]-C5H14N04P (-183)
	(lipidblast-pos) SM 34:1; [M+Na]+; SM(d14:1(4E)/20:0)
Names Structures Hit List	Plot/Text of Hit Plot of Hit
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	Peptide Peptide ///

2<sup>nd</sup> probability group

Name: Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; lysoPC 18:0 <u>MW:</u> 524 <u>ID#:</u> 55 <u>DB:</u> Spec. List Comment: lysoPC 18:0; Prec. m/7; 524 38 [M+H]+; ultrafleXtreme; Hig

<u>Comment:</u> IysoPC 18:0; Prec. m/z: 524.38 [M+H]+; ultrafleXtreme; High Performance TLC-MALDI; Martin Schürenberg, Beate Fuchs, Annabell Bischoff, Rosemarie Sü, Detlev Suckau, Jürgen Schiller, Gerda Morlock and Ulrike Anders; DGMS2010\_185\_Poster\_TLC-MALDI.pd 6 largest peaks:



<u>Name:</u> Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; PC 34:1 <u>MW:</u> 760 <u>ID#:</u> 56 <u>DB:</u> Spec. List <u>Comment:</u> PC 34:1; PC(16:0/18:1); Prec. m/z: 760.59; [M+H]+; ultrafleXtreme; High Performance TLC-MALDI; Martin Schürenberg, Beate Fuchs, Annabell Bischoff, Rosemarie Sü, Detlev Suckau, Jürgen Schiller, Gerda Morlock and Ulrike Anders; DGMS2010\_185\_Poster\_TLC-MALDI.pd <u>8 largest peaks:</u>

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🚳 🗫 🚔 📮 1. Bruker ultrafleXtreme MALDI TOF/TO 🛛 🛞 🖳 😥 🍭 🚳	
# Src. Name	L 194.06 Name: Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; PC 34:1
49 L Bruker ultraflex II MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0)	100-104.00 MW: 760 ID#: 56 DB: Spec. List Comment: PC 34:1: PC(16:0/18:1): Prec. m/z: 760.59: [M+H]+: ultrafleXtrem
50 L Bruker UltraFlex II MALDI TUF-TUF; (M+Na]+; DGDG 30:0 51 L Bruker UltraFlex II MALDI TOF-TOF: DGDG 31:0	577,33 8 largest peaks:
52 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 32:0	
53 L Bruker UltraFlex II MALDI TOF-TOF; [M+K]+; PC 36:1	(Spec. List) Bruker ultrafleXtr
55 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; lysoPC 18:0	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
56 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; PC 34:1	1 184.06
57 L Bruker ultrafieXtreme MALDI TOF/TOF; [M+Na]+; PC 3b:1 58 L Bruker ultrafieXtreme MALDI TOF/TOF: [M+Na]+; PE 33:1	100-
59 L Bruker ultrafle×treme MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0)	
	50-
Names Structures Spec List	146 97 577 33 760 59
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	
total spectra	200 20400 577 51000 701 51214
100	306.20400 577.31360 701.31214
10	204
1-	
	- 100- 184.07387
# Library Score Dot Product Prob. (%) Rev-Dot Name	■ Bruker ultrafleXtreme MALDI TOF. Head to Tail MF=25 RMF=683 ▼PC 34:1; (M+H)+; GPCho(8:0/26:1
1 custompc+h 25 683 0.93 737 PC 34:1; [M+H]+; GPCho(8:0/26:1(5Z))	Difference A Head to Tail A Side by Side A Subtraction A 25 683R 0.93P
2 custompo+h 25 683 U.93 737 PC 34:1; [M+H]+; GPCho[10:0/24:1[152]]	Name: PC 34:1; [M+H]+; GPCho(8:0/26:1(5Z))
4 custompc+h 25 683 0.93 737 PC 34.1; [M+H]+; GPCho(12.0/22.1(132))	100- 100- <u>MW:</u> 760 <u>IU#:</u> 1918 <u>DB:</u> custompc+hpos.msp <u>Comment:</u> Parent=760.58564 Mz_exact=760.5856
5 custompc+h 25 683 0.93 737 PC 34:1; [M+H]+; GPCho(14:0/20:1(11Z))	9 largest peaks:
6 custompc+h 25 683 0.93 737 PC 34:1; [M+H]+; GPCho(14:0/20:1(13E))	366.20480 577.51960 634.48124 200.00 701.51214 200.00 760.5
7 custompo+h 25 683 0.93 737 PC 34:1; [M+H]+; GPCho(14:0/20:1(13Z))	
6 custompc+n 20 663 0.33 737 PC 34.1; [M+H]+; GPCh0[14.1(32)/20.0] ♥	150 300 450 600 750 184.07387 993.00 tragment LSH ISNU4P ▼
Names Structures / Hit List	Plot/Text of Hit / Plot of Hit /
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide //

Name: Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; PC 36:1 <u>MW:</u> 782 <u>ID#:</u> 57 <u>DB:</u> Spec. List <u>Comment:</u> PC 36:1; PC(16:0/18:1); Prec. m/z: 782.53; [M+Na]+; ultrafleXtreme; BDAL; http://www.bdal.de/uploads/media/ultrafleXtreme-eBook.pdf 10 largest peaks:


## Name: Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; PE 33:1 <u>MW:</u> 726 <u>ID#:</u> 58 <u>DB:</u> Spec. List <u>Comment:</u> PE 33:1; Prec. m/z: 726.5 [M+Na]+; Lipid Compositions in Escherichia coli and Bacillus subtilis During Growth as Determined by MALDI-TOF and TOF/TOF Mass Spectrometry; doi:10.1016/j.ijms.2009.03.005. <u>6 largest peaks:</u>

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X 🗈 🖻 🕌 🞹 🙀 🖶 ⊟ ™/z ← 💡	
🚳 🍃 🚔 🚔 1. Bruker ultrafleXtreme MALDI TOF/TO 🗾 🛞 🖳 😥 🍭 🚳	
# Src. Name	100 164.0
49 L Bruker ultraflex II MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0)     50 L Bruker Ultraflex II MALDI TOF-TOF: (M+Na]+; DGDG 30:0	726.5 Comment: PE 33:1; Prec. m/z: 726.5 [M+Na]+; Lipid Compositions in Escher
51 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 31:0	
52 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 32:0 53 L Bruker UltraFlex II MALDI TOF-TOF; IM+K1+; PC 36:1	360 720 <u>121.0 100.00</u> ▼
54 L Bruker UltraFlex II MALDI TOF-TOF; [M+Na]+; SM(d18:1/16:0)	Plot/Text of Search Spectrum A Plot of Search Spectrum A Plot/Text of Spec List
55 L Bruker ultrafieXtreme MALDI TUF/TUF/ [M+H]+; lysoPU18:0 56 L Bruker ultrafieXtreme MALDI TOF/TOF: [M+H]+; PC 34:1	
57 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; PC 36:1	100-
58 L Bruker ultrafleXtreme MALDI TUF/TUF/ [M+Na]+; PE 33:1 59 L Bruker ultrafleXtreme MALDI TOF/TOF: [M+Na]+; SM(d18:1/16:0)	
	50- 683.5 726.5
Names Structures Spec List	563.5
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	
total spectra	289.08193
	50 585 48587
	30
1-	100
	683.46277
H Libury Com DesDedust Date (%) Des Det Mana	
	Difference λ Head to Tail
2 lipidblast-pos 69 460 1.46 989 PE 33:1; [M+Na]+; GPEtn(9:0/24:1(15Z))	Name: PE 33:1: [MaNala: GPEtn(7:0/26:1(57))
3 lipidblast-pos 69 460 1.46 989 PE 33:1; [M+Na]+; GPEtn(11:0/22:1(13Z))	100- 683.46277 <u>Wallet</u> P 33.1; [WHVary, dr Euly, 0/28,1(32)]
4 lipidblast-pos 69 460 1.46 989 PE 33:1; [M+Na]+; GPEtn(13:0/20:1(11E)) E lipidblast-pos 69 460 1.46 999 PE 33:1; [M+Na]+; GPEtn(13:0/20:1(11Z))	Comment: Parent=726.50497 Mz_exact=726.5049 4 Jargest peaks:
6 lipidblast-pos 69 460 1.46 989 PE 33.1; [M+Na]+; GPEth[13:0/20:1[(12)]	50- 585.48587 585.48587 400.00 289.0
7 lipidblast-pos 69 460 1.46 989 PE 33:1; [M+Na]+; GPEtn(13:0/20:1(13Z))	4 m/z Values and Intensities: 289.08193 289.08193 3.00 [M+Na]-43-SN2-H
8 lipidblast-pos 69 460 1.46 989 PE 33:1; [M+Na]+; GPEtn(14:1(92)/19:0)	140 280 420 560 700 553.36345 3.00 [M+Naj-43-SN1-H
Names A Structures / Hit List	(lipidblast-pos) PE 33:1; [M+Na]+; GPEtn(7:0/26:1(5Z))
13 Cault Other Caulty Caulty Caulty House	
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	Peptide Peptide //

<u>Name:</u> Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0) <u>MW:</u> 725 <u>ID#:</u> 59 <u>DB:</u> Spec. List <u>Comment:</u> SM(d18:1/16:0); Prec. m/z: 725.6 [M+Na]+; ultrafleXtreme; High Performance TLC-MALDI; Martin Schürenberg, Beate Fuchs, Annabell Bischoff, Rosemarie Sü, Detlev Suckau, Jürgen Schiller, Gerda Morlock and Ulrike Anders; DGMS2010\_185\_Poster\_TLC-MALDI.pd <u>10 largest peaks:</u>

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🚳 🍉 🚔 🚎 1. Bruker ultrafleXtreme MALDI TOF/TO 🛛 🛞 🖳 🔎 🍭 🚳	
# Src. Name	666.6 Name: Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0)
49 L Bruker ultraflex II MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0) 50 L Bruker Ultraflex II MALDI TOF-TOF: [M+Na]+: DGDG 30:0	TUU4 MW, 725 IDH, 59 DB; Spec. List Comment: SM(d18:1/16:0); Prec. m/z: 725.6 [M+Na]+; ultrafleXtreme; High
51 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 31:0	
52 L Bruker UltraFlex II MALDI TOF-TOF; DGDG 32:0 53 L Bruker UltraFlex II MALDI TOF-TOF; IM+K1+; PC 36:1	390 198.09 10.00 320.84 10.00 429.62 10.00 502.47 10.00 63
54 L Bruker UltraFlex II MALDI TOF-TOF; (M+Na]+; SM(d18:1/16:0)	Plot/Text of Search Spectrum A Plot of Search Spectrum A Plot/Text of Spec List
55 L Bruker ultrafieXtreme MALDI 10F710F; [M+H]+; [ysoPC 18:0 56 L Bruker ultrafieXtreme MALDI TOF/TOF; [M+H]+; PC 34:1	
57 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; PC 36:1	100-
58 L Bruker ultrafleXtreme MALDI 10F710F7 [M+Na]+; PE 33:1 59 L Bruker ultrafleXtreme MALDI T0F/T0F7 [M+Na]+; SM(d18:1/16:0)	
	50- 542.53
Names Structures Spec List	86,11
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	0 168,99 320,84 429,62 502,47 630,5
100-1	042.40120
	50-
1	100-
# Library Score Dot Product Prob. (%) Rev-Dot Name	A Bruker ultrafleXtreme MALDI TOF./ Head to Tail MF=270 RMF=819 [▼SM 34:1; [M+Na]+; SM(d14:1(4E).
1 lipidblast-pos 270 819 24.5 889 SM 34:1; [M+Na]+; SM(d14:1(4E)/20:0)	Difference Head to Tail Side by Side Subtraction 270 819R 24.5P
2 lipidblast-pos 270 819 24.5 889 SM 34:1; [M+Na]+; SM(d16:0/18:1(92)) -	666 48383 Name: SM 34:1; [M+Na]+; SM(d14:1(4E)/20:0)
4 lipidblast-pos 270 819 24.5 889 SM 34.1; [M+Na]+; SM(d18:1(4E)/16:0)	100- Comment: Parent=725.55733 Mz_exact=725.5573
5 lipidblast-neg 49 315 0.24 438 Pl 26:0; [M-H]-; GPIns(12:0/14:0)	2 largest peaks:
6 lipidblast-neg 49 315 0.24 438 PI 26:0; [M-H]-; GPIns(14:0/12:0)	2 m/z Values and Intensities:
7 iipiapiast-pos 5 42 0.05 175 DG 43:2; [M+Li]+; DG (17:0/26:2/0:0) 8 iipidblast-pos 5 42 0.05 175 DG 43:2; [M+Li]+; DG (17:1/26:1/0:0)	0 542.49129 542.49129 20.00 [M+Na]-C5H14N04P (-183)
	(ipidblast-pos) SM 34:1; [M+Na]+; SM(d14:1(4E)/20:0)
Names Structures / Hit List Plot/Text of Hit Plot of Hit /	
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	Peptide Peptide //

Name: JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]-; DGDG 36:4 <u>MW:</u> 939 <u>ID#:</u> 60 <u>DB:</u> Spec. List <u>Comment:</u> DGDG 36:4; DGDG(18:2/18:2); [M-H]-; Prec. m/z: 939.5; Structural Characterization of Sulfoquinovosyl, Monogalactosyl and Digalactosyl Diacylglycerols by FAB-CID-MS/MS; JOURNAL OF MASS SPECTROMETRY, VOL. 32, 968-977 (1997) <u>8 largest peaks:</u>



<u>Name:</u> JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]-; GM3 <u>MW:</u> 1151 <u>ID#:</u> 61 <u>DB:</u> Spec. List <u>Comment:</u> Ganglioside GM3; [M-H]-; Prec. m/z: 1151.7; Studies of the Chemical Structure of Gangliosides in Deer Antler, Cervus nippon; Chem. Pharm. Bull. 47(1) 123-127 (1999) <u>9 largest peaks:</u>

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🔞 🍃 🖻 🚔 1. JEOL JMS-HX110A/110A 🛛 Tandem M 🔽 🛞 🖺 🖳 🍭 🔍	
# Src Name	Name: JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]-; GM3
55 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; lysoPC 18:0	100- <sup>297,2</sup> 1151.7 <u>MW:</u> 1151 <u>ID#</u> , 61 <u>DB</u> : Spec. List Comment: Ganglioside GM3: IM-HL: Prec. m/z: 1151 7: Studies of the Chem
56 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; PC 34:1 57 L Bruker ultrafleXtreme MALDI TOF/TOF: [M+Na]+; PC 36:1	468.1 860.8 9largest peaks
58 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; PE 33:1	
60 L JEOLJMS-HX110A/110A Tandem MS; FAB; [M-H]; DGDG 36:4	Spec. List) JEOL JMS-HX11
61 L JEOLJMS-HX110A/110A Tandem MS; FAB; [M-H];; GM3	
63 L JEDLJMS-HX110A/110A Tandem MS; FAB; [M-H]; MAD d 30.4	1151.7 1004 1
64 L Kratos MALDI-TOF AXIMA-CFR; [M-H]-; Lipid A 65 L SHIMADZU KBATOS MALDI TBAP TOF: [M-H]-; SDDG 36:5	
	50-
Names Structures Spec List	468.1 536 599.3 698 860.8 1031.7
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	
	536.50395 698.55677
	50-
	_ 1001151 7053
	320 400 480 560 640 720 800 880 960 1040 1120
# Library Score Dot Product Prob. (%) Rev-Dot Name	▲JEOL JMS-HX110A/110A_Tandel Head to Tail MF=116 RMF=374 【▼(glycan) Cer 34:1; GM3(d18:1/16:1
1 lipidblast-neg 116 374 100.0 824 [glycan]-Cer 34:1; GM3(d18:1/16:0)); [M-H]-; NeuAr	Difference A Head to Tail A Side by Side A Subtraction / 116 374R 100.0P
	1151.7053 Name: [glycan]-Cer 34:1; GM3(d18:1/16:0)]; [M-H] ▲ 100 MW: 1151 ID#: 79290 DB: lipidblastneg
	Comment: Parent=1151.70534 Mz_exact=1151.70
	50- 50- 50- 50- 502- 502- 502- 502- 502-
	0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
	380 570 760 950 1140 536.50395 200.00 ion ceramide ▼
Names Structures / Hit List	Plot/Text of Hit / Plot of Hit /
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or Help, press F1	Peptide Peptide

Name: JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]-; MGDG 36:4 <u>MW:</u> 777 <u>ID#:</u> 62 <u>DB</u>: Spec. List <u>Comment:</u> MGDG 36:4; MGDG(18:2/18:2); [M-H]-; Prec. m/z: 777.4; Structural Characterization of Sulfoquinovosyl, Monogalactosyl and Digalactosyl Diacylglycerols by FAB-CID-MS/MS; JOURNAL OF MASS SPECTROMETRY, VOL. 32, 968-977 (1997) <u>4 largest peaks:</u>



 Name:
 JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]-; NA

 <u>MW:</u>
 819 <u>ID#:</u>
 63 <u>DB:</u>
 Spec. List

 <u>Comment:</u>
 SQDG 36:1; wrong; m/z 255 and 281 missing; SQDG(18:1/16:0); [M-H]-; Prec. m/z: 819.6; Structural Characterization of

 Sulfoquinovosyl, Monogalactosyl and Digalactosyl Diacylglycerols by FAB-CID-MS/MS; JOURNAL OF MASS SPECTROMETRY, VOL.

 32, 968-977 (1997)

 <u>6 largest peaks:</u>

SQDG 36:1; wrong; m/z 255 and 281 missing; SQDG(18:1/16:0); [M-H]-; Prec. m/z:

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(1) 1. JEOL JMS-HX110A/110A Tandem MS   (위) 밖 (요) (영) (영)	
	Name: IEOL IMS HV1104 (1104 Tandem MS: EAR-IM H1: NA
Src. Name     Soc. Name	100-148 MW: 819 ID#: 63 DB: Spec. List
56 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; PC 34:1	297 819 <u>Comment:</u> SQDG 36:1; wrong; m/z 255 and 281 missing; SQDG(18:1/16:0);
57 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+Na]+; PC 36:1	
59 L Bruker ultrafieXtreme MALDI TOF/TOF; [M+Na]+; SM(d18:1/16:0)	440 /33 100.00 ·
60 L JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]; DGDG 36:4	Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /
62 L JEOL JMS-HX1104/1104 Tandem MS; FAB; [M-H]; MGDG 36:4	
63 L JEOLJMS-HX110A/110A Tandem MS; FAB; [M-H]-; NA	100-
65 L SHIMADZU KRATOS MALDI TRAP TOF: (M-H1): SQDG 36:5	
	<b>50-</b> 297 819
Names Structures Spec List	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	
total spectra	297.27918
	521.24223
	504
	- 100- 225,00690
	140 210 280 350 420 490 560 630 700 770
# Library Score Dot Product Prob. (%) Rev-Dot Name	AJEOL JMS-HX110A/110A Tander Head to Tail MF=9 RMF=113 ▼SQDG 34:1; [M-H]-; SQDG(15:1[9])
1 lipidblast-neg 36 341 11.4 707 MGDG 39:4; [M-H]-; MGDG(19:0/20:4(5E,8E,11)	Difference A Head to Tail A Side by Side A Subtraction 9 113R 4.38P
2 lipidblast-neg 36 341 11.4 707 MGDG 39.4 (M-H)-; MGDG (19:0/20:4(52,52,11, 3 lipidblast-neg 36 341 11.4 707 MGDG 39.4 (M-H)-; MGDG (19:0/20:4(7E 10E 1	Name: SQDG 34:1; [M-H]-; SQDG(15:1(92)/19:0)
4 lipidblast-neg 36 341 11.4 707 MGDG 39:4; [M-H]-; MGDG[20:4[5E,8E,11E,14E	100- Comment: Parent=819.52923 Mz_exact=819.5292
5 lipidblast-neg 36 341 11.4 707 MGDG 39:4; [M-H]-; MGDG(20:4(5Z,8Z,11Z,14Z-	50 50 50 50 50 50 50 50 50 50 50 50 50 5
6 lipidblast-neg 36 341 11.4 707 MGDG 39:4; [M-H]-; MGDG(20:4(7E,10E,13E,16	521.24223 223.00650 333.00 521.24223 300.00 573.3.
7 lipidblast-neg 26 269 8.03 559 PG 33:0; [M-H]-; GPGro(13:0/20:0) 8 lipidblast-neg 26 269 8.03 559 PG 39:0; [M-H]-; GPGro(20:0/19:0)	225.00690 999.00 fragment C6H907S
	(inidblast-peg) SDDG 3411 (M.H.)- SDDG(15-1(97)/19
Names (Structures / Hit List	Plot/Text of Hit / Plot of Hit /
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide

# <u>Name:</u> Kratos MALDI-TOF AXIMA-CFR; [M-H]-; Lipid A <u>MW:</u> 2048 <u>ID#:</u> 64 <u>DB:</u> Spec. List <u>Comment:</u> Lipid A; The Lipid A 1-Phosphatase of Helicobacter pylori Is Required for Resistance to the Antimicrobial Peptide Polymyxin;JOURNAL OF BACTERIOLOGY, June 2006, p. 45314541 <u>3 largest peaks:</u>

🚝 NIST MS Search 2.0 - [Peptide, Presearch Default - 126 spectra]	
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🔞 ⊳ ਫ਼ 🚔 🛛 1. Kratos MALDI-TOF AXIMA-CFR; [M-H 🗹 🔞 🖳 😥 🍭 🚳	
	Name: Kratos MALDI-TOE AXIMA-CEB · [M-H1-1 linid A
55 L Bruker ultrafleXtreme MALDI TOF/TOF; [M+H]+; lysoPC 18:0	100- 2048.6 <u>MW:</u> 2048 <u>ID#</u> ; 64 <u>DB</u> ; Spec. List
56 L Bruker ultrafleXtreme MALDI TOF/TOF; (M+H)+; PC 34:1	2071.1 2071.1 3 largest peaks:
57 L Bruker ultrafleXtreme MALDI TOF/TOF/ [M+Na]+; PC 36.1	2048.6 999.00   2021.2 700.00   2071.1 300.00   2040 2070   3 m/z Values and Intensities:
59 L Bruker ultrafleXtreme MALDI TOF/TOF; (M+Na]+; SM(d18:1/16:0)	Spec. List) Kratos MALDI-TC
61 L JEOLJMS-HX1104/1104 Tandem MS; FAB; [M-H]; DiaDia 36:4	Plot/Text of Search Spectrum Plot of Search Spectrum A Plot/Text of Spec List
62 L JEOL JMS-HX1104/110A Tandem MS; FAB; [M-H]; MGDG 36:4	2048.6
64 L Kratos MALDI-TOF AXIMA-CFR; [M-H]-; Lipid A	100-
65 L SHIMADZU KRATOS MALDI TRAP TOF; [M-H]-; SQDG 36:5	
	50-
Names Structures Spec List	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370 total spectra	
1000-1	
100-]	50-
10-	2048.45327
1-	100
# Library Score Dot Product Prob. (%) Rev-Dot Name	2020 2030 2040 2050 2050 2050 2070 2080 ▲Kratos MALDI-TOF AXIMA-CFR: (  Head to Tail MF=13 RMF=117  ▼LipidA-PP 64:36:0: LipidA-PP 114/
1 lipidblast-neg 13 117 0.85 198 LipidA-PP 64:36:0; LipidA-PP (14/14/18/18/30-	Difference Head to Tail Side by Side Subtraction / 13 117R 0.85P
2 lipidblast-neg 13 117 0.85 198 Lipidb-PP 64:36:0; Lipidb-PP (16/16/14/18/30-)	Name: LipidA-PP 64:36:0; LipidA-PP [14/14/18/18
3 lipidblast-neg 13 117 0.85 198 LipidA-PP 64:36:0; LipidA-PP [16/16/16/16/16/30-] 4 lipidblast-neg 12 117 0.95 199 LipidA-PP 64:36:0; LipidA-PP [16/16/16/16/16/30-]	100- 100- MW: 2048 ID#: 88254 DB: lipidblast-neg
5 lipidblast-neg 13 117 0.85 198 LipidA-PP 68:32:0: LipidA-PP [16/16/18/18/30-1	2048.49327 7 largest peaks:
6 lipidblast-neg 13 117 0.85 198 LipidA-PP 64:36:0; LipidA-PP [18/18/10/18/30-]	504 1804.28955 999.00   1950.51637 600.00   204 1764 22191 250 00   1966 51129 50 00
7 lipidblast-neg 13 117 0.85 198 LipidA-PP 64:36:0; LipidA-PP [18/18/12/16/30-]	7.m/z Values and Intensities:
8 lipidblast-neg 13 117 U.85 198 LipidA-PP 64:36:0; LipidA-PP [18/18/14/14/30-]	2020 2030 2040 2050 2060 2070 2080 1666.24501 300.00 [M·H]·P04H3·R2·O·FA    [M·▼
Names Structures Hit List	[lipidblast-neg] LipidA-PP 64:36:0; LipidA-PP [14/14/18
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide

### Name: SHIMADZU KRATOS MALDI TRAP TOF; [M-H]-; SQDG 36:5 <u>MW:</u> 839 <u>ID#:</u> 65 <u>DB:</u> Spec. List <u>Comment:</u> SQDG(20:5/16:0); [M-H]-; Prec. m/z: 839.8; Sulfoquinovosyldiacylglyceride - antiviral aktive Substanzen; Diss Universitt Erlangen-Nürnberg; 2009; Ivonne Naumann <u>3 largest peaks:</u>



Name: SHIMADZU LCMS-IT-TOF; [M-H]-; PS 32:0 <u>MW:</u> 734 <u>ID#:</u> 66 <u>DB:</u> Spec. List <u>Comment:</u> PS(16:0/16:0); [M-H]-; Prec. m/z: 734.4672; PS(16:0/16:0); Identification of Phospholipid Molecular Species Using Neutral Loss Survey and MS3 Analysis; Shimadzu Technical Report 4 largest peaks:



Name: SHIMADZU LCMS-IT-TOF; [M-H]-; SQDG 34:3 <u>MW:</u> 815 <u>ID#:</u> 67 <u>DB</u>: Spec. List <u>Comment:</u> SQDG(34:3); [M-H]-; Prec. m/z: 815.499; SQDG(34:3);A Chloroplastic UDP-Glucose Pyrophosphorylase from Arabidopsis Is the Committed Enzyme for the First Step of Sulfolipid Biosynthesis;Plant Cell Okazaki et al. 21: 892 ; http://www.plantcell.org/cgi/data/tpc.108.063925/DC1/1; 3 largest peaks:



Name: Thermo Finnigan DecaXP ion trap; [M+Na]+; DGDG 34:1 <u>MW:</u> 941 <u>ID#:</u> 68 <u>DB:</u> Spec. List <u>Comment:</u> DGDG(18:1/16:0); DGDG from K. brevis; [M+Na]+; Prec. m/z: 941.41; <u>6 largest peaks:</u>



<u>Name:</u> Thermo Fisher Exactive Orbitrap; [M-H]-; PS MIX <u>MW:</u> 834 <u>ID#:</u> 69 <u>DB:</u> Spec. List <u>Comment:</u> PS MIX; M-H]-; Prec. m/z: 834.5299; C46H77O10NP; Analysis of whole lipid extracts using on-line high resolution LC-MS; Catharina Crone, Eric Genin and Helmut Muenster <u>10 largest peaks:</u>

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· X h f A A M Y A A A A A A A A A A A A A A A A	
🔞 🖕 🚘 🔲 1. Thermo Fisher Exactive Orbitrap; [M-I 🔍 🛞 🖳 🔎 🥘 🚳	
	Name: Thermo Eicher Exactive Orbitran: (M.H.): PS MIX
	100- 747.4981 MW: 834 ID#: 69 DB: Spec. List
62 L JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]; MGDG 36:4	Comment: PS MIX; M-H]; Prec. m/z: 834.5299; C46H77010NP; Analysis of 10 largest peaks:
63 L JEULJMS-HX11UA/11UA/1Andem MS; FAB; [M-H]-; NA 64 L Kratos MALDI-TOF AXIMA-CFR: [M-H]-; Lipid A	
65 L SHIMADZU KRATOS MALDI TRAP TOF; (M-H)-; SQDG 36:5	(Spec. List) Themo Fisher E>
67 L SHIMADZU LCMS+T+T0F; [M-H]; PS 32:0	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
68 L Thermo Finnigan DecaXP ion trap; [M+Na]+; DGDG 34:1	
E I hermo Fisher Exactive Urbitrap; [M-H]-; PS MIX     Thermo Finnigan LCQ ion trap; ESI: [M+NH4]+; DGDG 36;4	100-
71 L Thermo Finnigan LCQ DECA ion trap; [M·H]-; Lipid A	419.25/6
	50-
Names Structures Spec List	480.3100 621.4589 701.5142
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370 total spectra	
1000-1	283.26354 419.25639
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10-	
	100-
# Library Score Dot Product Prob. (%) Rev-Dot Name	▲ Thermo Fisher Exactive Orbitrap:   Head to Tail MF=385 RMF=648 ▼PS 40:6; (M-H]-: GPS er(18:0/22:6
1 lipidblast-neg 385 648 42.1 845 PS 40:6; [M-H]-; GPSer(18:0/22:6(4Z,7Z,10Z,13	<u>∫ Difference</u> <u>A Head to Tail</u> <u>Side by Side</u> <u>A</u> <u>Subtraction</u> <u>385 648R 42.1P</u>
2 lipidblastneg 385 648 42.1 845 PS 40:6; [M-H]-; GPSer(22:6(4Z,7Z,10Z,13Z,16Z	747 49647 Name: PS 40:6; [M-H]; GPSer(18:0/22:6(4Z,7Z,1)
4 lipidblastneg 193 414 0.61 773 PS 40:6; [M-H]-; GPSet(20:1(112)/20:5(52,82,11	100- <u>Comment:</u> Parent=834.52850 Mz_exact=834.5285
5 lipidblast-neg 193 414 0.61 773 PS 40:6; [M-H]-; GPSer(20:1(13E)/20:5(5Z,8Z,11	7 largest peaks:
6 lipidblast-neg 193 414 0.61 773 PS 40:6; [M-H]-; GPSer(20:1(13Z)/20:5(5Z,8Z,11	419.25639 283.26354 100.00   327.23226 100.00
7 iipiapiaseneg 133 414 0.61 773 PS 40:6; [M-H]-; GPSet(20:5(52,82,112,142,172 8 lipidblast-neg 193 414 0.61 773 PS 40:6: [M-H]-; GPSet(20:5(52,82,112,142,172 -	0 7 m/z Values and Intensities:
	(lipidblast-neg) PS 40:6; [M-H]-; GPSer(18:0/22:6/4Z,7Z
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PS-MIX

3<sup>rd</sup> probability group

<u>Name:</u> Thermo Finnigan LCQ ion trap; ESI; [M+NH4]+; DGDG 36:4 <u>MW:</u> 958 <u>ID#:</u> 70 <u>DB:</u> Spec. List <u>Comment:</u> DGDG 36:4; DGDG(18:1/18:3); [M+NH4]+; Prec. m/z: 958.4; Simultaneous analysis of glycolipids and phospholids molecular species in avocado (Persea americana Mill) fruit; http://dx.doi.org/10.1016/j.chroma.2006.10.022 <u>10 largest peaks:</u>

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🚳 Þ 🗃 📮 1. Thermo Finnigan LCQ ion trap; ESI; [I 🔄 🛞 🖺 😥 🍭 🚳	
# Src. Name	Name: Thermo Finnigan LCQ ion trap; ESI; [M+NH4]+; DGDG 36:4
61 L JEOL JMS-HX110A/110A Tandem MS; FAB; [M-H]; GM3	100- 001-3 MW: 958 ID#: 70 DB; Spec. List Comment: DGDG 36:4: DGDG(18:1/18:3): [M+NH4]+: Prec. m/z: 958 4: Sir
62 L JEOLJMS-HX110A/110A Tandem MS; FAB; [M-H]; MGDG 36:4	339.2   933.0 <u>10 largest peaks:</u>
64 L Kratos MALDI-TOF AXIMA-CFR; [M-H]-; Lipid A	
65 L SHIMADZU KRATOS MALDI TRAP TOF; [M-H]; SQDG 36:5	Spec. List) Thermo Finnigan
67 L SHIMADZU LUMSHT-TDF; [M-H]; PS 32:0	Plot/Text of Search Spectrum Plot of Search Spectrum A Plot/Text of Spec List
68 L Thermo Finnigan DecaXP ion trap; [M+Na]+; DGDG 34:1	691 3
69 L Thermo Fisher Exactive Orbitrap; [M-H]; PS MIX 70 L Thermo Finningen LCD ion trans ESI: [M-NH4]+: DGDG 36:4	100-
71 L Thermo Finnigan LCQ DECA ion trap; [M-H]-; Lipid A	
	50- 617.3
Names Structures / Spec List	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	- 0 <u>339.2 425.4 506.6 591.9</u> 797.5 933.0
total spectra	
10-	504
	- 100- 455.26233 581.40309 743.45591
	360 420 480 540 600 660 720 780 840 900 960
# Library Score Dot Product Prob. (%) Rev-Dot Name	▲ Thermo Finnigan LCQ ion trap; ESI Head to Tail MF=12 RMF=240 I▼DGDG 35:0; [M+Na]+; DGDG(13:
1 lipidblast-pos 19 340 5.67 707 TG 58:6; [M+Na]+; TG(18:3/18:3/22:0)	Difference A Head to Tail Side by Side A Subtraction / 12 240R 4.22P
2 lipidblast-pos 14 277 4.57 577 TG 59:5; [M+Li]+; TG(17:0/20:5/22:0)	A55 26233 7/3 /5591 Name: DGDG 35:0; [M+Na]+; DGDG(13:0/22:0)
4 linidblast-pos 14 277 4.57 577 TG 59:5; [M+Li]+; TG(17:1/20:4722:0)	100- 100-
5 lipidblast-pos 14 277 4.57 577 TG 58:6; [M+Na]+; TG(16:1/20:5/22:0)	
6 lipidblast-pos 12 240 4.22 500 DGDG 35:0; [M+Na]+; DGDG(13:0/22:0)	455.26233 999.00   581.40309 999.00   517.3
7 lipidblast-pos 12 240 4.22 500 DGDG 35:0; [M+Na]+; DGDG(22:0/13:0)	455.26233 999.00 [M+Na]-sn2-C6H1005 (-162)
8 lipidbias(meg 11 223 4.05 452 Subla 44:2; [M-H]; Subla(22:0/22:2(132,162,)) -	420 560 700 840 581.4U309 999.00 [M+Na]-sn1-C6H1005(-162)
Names Structures Hit List	Inplablasr-pos) DaDia 35:0; [M+Naj+; DiaDia[13:0722:1]
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide

(exclude M+Li)

Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; Lipid A <u>MW:</u> 1796 <u>ID#:</u> 71 <u>DB:</u> Spec. List <u>Comment:</u> Lipid A from E coli; [M-H]-; Prec. m/z: 1796.3; C94H178N2O25P2; LipidA-PP-[R2(14:0)(3-OH)/R3(14:0)(3-OH)/R2'(14:0)/R3'14:0)/R2'-3-O-(12:0)/R3'-3O-(14:0)]; DOI: 10.1002/jms.614; Structural analysis of lipid A from Escherichia coli O157:H7:K- using thin-layer chromatography and ion-trap mass spectrometry <u>10 largest peaks:</u>



Name: Thermo Finnigan LCQ ion trap; ESI; [M+NH4]+; MGDGD MIX <u>MW:</u> 800 <u>ID#:</u> 72 <u>DB</u>: Spec. List <u>Comment:</u> MGDGD MIX; MGDG(18:2/18:0)+MGDG(18:1/18:1); [M+NH4]+; Prec. m/z: 800.4; Simultaneous analysis of glycolipids and phospholids molecular species in avocado (Persea americana Mill) fruit; http://dx.doi.org/10.1016/j.chroma.2006.10.022 <u>6 largest peaks:</u>

MGDG MIX



Name: Thermo LCQ iontrap ESI; [M+NH4]+; TG 52:2 <u>MW:</u> 876 <u>ID#:</u> 73 <u>DB:</u> Spec. List <u>Comment:</u> TAG 52:2; TAG(18:1/18:1/16:0); [M+NH4]+; Prec. m/z: 876.9; 1,3-dioleoyl-2-palmitoylglycerol (OPO); Structural Characterization of Triacylglycerols Using Electrospray Ionization-MSn Ion-Trap MS; 10.1007/s11746-003-0676-2 <u>4 largest peaks:</u>



### Name: Thermo LCQ iontrap ESI; [M+NH4]+; TG 48:1 <u>MW:</u> 822 <u>ID#:</u> 74 <u>DB:</u> Spec. List <u>Comment:</u> TAG(14:0/18:1/16:0); [M+NH4]+; Prec. m/z: 822.6; 1-myristoyl-2-oleoyl-3-palmitoyl glycerol (MOP); Structural Characterization of Triacylglycerols Using Electrospray Ionization-MSn Ion-Trap MS; 10.1007/s11746-003-0676-2 <u>5 largest peaks:</u>



<u>Name:</u> Thermo LCQ iontrap ESI; [M+NH4]+; TG 54:1 <u>MW:</u> 906 <u>ID#:</u> 75 <u>DB:</u> Spec. List <u>Comment:</u> TAG; [M-H]-; Prec. m/z: 906.7; 1,3-distearoyl-2-oleoyl glycerol (SOS); Structural Characterization of Triacylglycerols Using Electrospray Ionization-MSn Ion-Trap MS; 10.1007/s11746-003-0676-2 <u>4 largest peaks:</u>



<u>Name:</u> Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM1 35:0 <u>MW:</u> 1013 <u>ID#:</u> 76 <u>DB:</u> Spec. List <u>Comment:</u> PIM1(19:0/16:0); [M-H]-; Prec. m/z: 1013.7; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. I. PIMs and lyso-PIMs. <u>10 largest peaks:</u>

757 999.00 |



Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 35:0 <u>MW:</u> 1175 <u>ID#:</u> 77 <u>DB:</u> Spec. List <u>Comment:</u> PIM2(19:0/16:0); [M-H]-; Prec. m/z: 1175.6; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. I. PIMs and Iyso-PIMs.

10 largest peaks:



Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 51:0 <u>MW</u>: 1413 <u>ID#</u>: 78 <u>DB</u>: Spec. List <u>Comment</u>: PIM2(16:0/19:0/16:0); [M-H]-; Prec. m/z: 1413.9; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. II. Monoacyl- and Diacyl-PIMs <u>10 largest peaks</u>:



Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 69:2 <u>MW:</u> 1676 <u>ID#:</u> 79 <u>DB:</u> Spec. List <u>Comment:</u> PIM2(16:0/18:1/19:0/16:1); [M-H]-; Prec. m/z: 1676.1; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. II. Monoacyl- and Diacyl-PIMs 10 largest peaks:



Peptide

Peptide

Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 69:1 <u>MW</u>: 1678 <u>ID#</u>: 80 <u>DB</u>: Spec. List <u>Comment</u>: PIM2(16:0/18:1/19:0/16:0); [M-H]-; Prec. m/z: 1678.1; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. II. Monoacyl- and Diacyl-PIMs <u>10 largest peaks</u>:



Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 70:0 <u>MW</u>: 1694 <u>ID#</u>: 81 <u>DB</u>: Spec. List <u>Comment</u>: PIM2(16:0/19:0/19:0/16:0); [M-H]-; Prec. m/z: 1694.2; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. II. Monoacyl- and Diacyl-PIMs <u>10 largest peaks</u>:



Name: Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM3(19:0/16:0) MW: 1337 ID#: 82 DB: Spec. List

<u>Comment:</u> PIM3(19:0/16:0); [M-H]-; Prec. m/z: 1337.9; Hsu and Turk, Structural characterization of phosphatidyl-myo-inositol mannosides from Mycobacterium bovis Bacillus Calmette Gurin by multiple-stage quadrupole ion-trap mass spectrometry with electrospray ionization. I. PIMs and Iyso-PIMs.

7 largest peaks:

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#       Src.       Name         73       L       Thermo LCQ iontrap ESI; [M+NH4]+; TG 52:2         74       L       Thermo LCQ iontrap ESI; [M+NH4]+; TG 54:1         75       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN1 35:0         76       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN1 35:0         77       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 35:0         78       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 51:0         78       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 51:0         79       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         79       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         79       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         79       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         80       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         81       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         82       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         83       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         84       L       Thermo Finnigan LCQ DECA ion trap; [M+]; PIN2 50:0         81       L       Th	0/16:0)
# Library       Score       Dot Product       Prob. (%)       Rev-Dot       Name         Difference       Head to Tail       Side by Side       Subtraction	=
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Name: Thermo LCQ Deca XP MAX ion trap; [M+Na]+; DGDG 34:1 <u>MW:</u> 941 <u>ID#:</u> 83 <u>DB:</u> Spec. List <u>Comment:</u> DGDG(18:1/16:0) from K. brevis; [M+Na]+; Prec. m/z: 941.41; Mono- and digalactosyldiacylglycerol composition of dinoflagellates. II. Lepidodinium chlorophorum, Karenia brevis, and Kryptoperidinium foliaceum, three dinoflagellates with aberrant plastids Jeffrey D. Leblond ; Andrew D. Lasiter;European Journal of Phycology, 1469-4433, Volume 44, Issue 2, First published 2009, Pages 199 - 205 <u>6 largest peaks:</u>



Name: Thermo LCQ Deca XP MAX ion trap ESI; [M+H]+; NA <u>MW:</u> 787 <u>ID#:</u> 84 <u>DB:</u> Spec. List <u>Comment:</u> wrong prec (786.6); PC 36:2; PC(18:2/18:0); [M+H]+; Prec. m/z: 787.3; Profiling of phospholipids in lipoproteins by multiplexed hollow fiber flow field-flow fractionation and nanoflow liquid chromatographytandem mass spectrometry ;http://dx.doi.org/10.1016/j.chroma.2010.01.006 5 Jargest peaks:



Name: Thermo LTQ; [M+H]+;PC 34:1; <u>MW:</u> N/A <u>ID#:</u> 85 <u>DB:</u> Spec. List <u>Comment:</u> PEPMASS=760.61; PC 34:1; [M+H]+ ; Prec. m/z: 760.5856; GPCho(16:0/18:1(11E)); C42H82NO8P 10 largest peaks:



Name: Thermo Finnigan LTQ linear ion trap; [M+Na]+; TG 52:1 <u>MW:</u> 883 <u>ID#:</u> 196 <u>DB:</u> Text File <u>Comment:</u> TAG(16:0/18:1/18:0); [M+Na]+; Prec. m/z: 883.6; Electrospray Ionization Multiple-Stage Linear Ion-trap Mass Spectrometry for Structural Elucidation of Triacylglycerols: Assignment of Fatty Acyl Groups on the Glycerol Backbone and Location of Double Bonds; Fong-Fu Hsu and John Turk 7 largest peaks:



<u>Name:</u> Thermo Finnigan LTQ linear ion trap; [M+NH4]+; TG 52:1 <u>MW:</u> 878 <u>ID#:</u> 87 <u>DB:</u> Spec. List <u>Comment:</u> TAG(16:0/18:0/18:1); [M+NH4]+; Prec. m/z: 878.7; Electrospray Ionization Multiple-Stage Linear Ion-trap Mass Spectrometry for Structural Elucidation of Triacylglycerols: Assignment of Fatty Acyl Groups on the Glycerol Backbone and Location of Double Bonds; Fong-Fu Hsu and John Turk <u>6 largest peaks:</u>



<u>Name:</u> Thermo Finnigan LTQ linear ion trap; [M+Li]+; TG 52:1 <u>MW:</u> 867 <u>ID#:</u> 88 <u>DB:</u> Spec. List <u>Comment:</u> TAG(16:0/18:1/18:0); [M+Li]+; Prec. m/z: 867.6; Electrospray Ionization Multiple-Stage Linear Ion-trap Mass Spectrometry for Structural Elucidation of Triacylglycerols: Assignment of Fatty Acyl Groups on the Glycerol Backbone and Location of Double Bonds; Fong-Fu Hsu and John Turk <u>7 largest peaks:</u>



<u>Name:</u> Thermo Finnigan LTQ linear ion trap; [M+Li]+; TG 50:1 <u>MW:</u> 839 <u>ID#:</u> 89 <u>DB:</u> Spec. List <u>Comment:</u> TAG(16:0/18:1/16:0); [M+Li]+; Prec. m/z: 839.0; Electrospray Ionization Multiple-Stage Linear Ion-trap Mass Spectrometry for Structural Elucidation of Triacylglycerols: Assignment of Fatty Acyl Groups on the Glycerol Backbone and Location of Double Bonds; Fong-Fu Hsu and John Turk <u>6 largest peaks:</u>

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t Sto Name	Name: Thermo Finnigan LTQ linear ion trap; [M+Li]+; TG 50:1
79 L Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 69:2	100- 583 MW: 839 ID#: 899 DB: Spec. List Comment: TAC(15:0/19:1/16:0): Mul ils: Prec. m/z: 829 0: Electrospray lon
80 L Thermo Finnigan LCQ DECA ion trap; [M-H]; PIM2 69:1	839 <u>6 largest peaks:</u>
81 L Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM2 70:0 82 L Thermo Finnigan LCQ DECA ion trap: [M-H]-; PIM3(19:0/16:0)	
83 L Thermo LCQ Deca XP MAX ion trap; [M+Na]+; DGDG 34:1	Spec. List) Thermo Finnigan
84 L Thermo LCQ Deca XP MAX ion trap ESI; [M+H]+; NA	Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /
85 L Thermo El Q; [M+H]+,PC 34:1; 86 L Thermo Finnigan LTQ linear ion trap: [M+Na]+; TG 52:1	
87 L Thermo Finnigan LTQ linear ion trap; [M+NH4]+; TG 52:1	100- 583
88 L Thermo Finnigan LTQ linear ion trap; (M+Li)+; TG 52:1	
09 L Thems Finders LTO faces in here 041 3 - TO 2010	50
Names / Structures / Specifiet	557 000
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custompc+hpos.msp; custompc+hapos.msp; hilic-urine; lipidblast-heg; pc-ac-heg.msp; pc-form-heg.msp; lipidblast-pos; 230370 total spectra	
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	<sup>100</sup> 557.51181
	560 580 600 620 640 660 680 700 720 740 760 780 800 820 840
📕 📕 Library Score Dot Product Prob. (%) Rev-Dot Name 🛕	▲ Thermo Finnigan LTQ linear ion tra Head to Tail MF=169 RMF=895 I▼TG 50:1; [M+Li]+; TG(16:0/16:0/1
1 lipidblast-pos 169 895 16.1 949 TG 50:1; [M+Li]+; TG(16:0/16:0/18:1)	LITTERENCE A Head to Tail A Side by Side A Subtraction / 169 895R 16.1P
2 lipidblast-pos 169 895 16.1 949 16.50(1) [M+Li]+; 16(16:0/16:0/18:1) 2 lipidblast-pos 24 596 0.61 707 TG 49(2) [MuNaturi TG(16:0/16:0/17:2)	Name: TG 50:1; [M+Li]+; TG(16:0/16:0/18:1)
4 inidialscepts 34 350 0.01 for 10 452, (MHXay, 10(10.010017.2) 1	100- Comment: Parent=839.76753 Mz_exact=839.7675
5 lipidblast-pos 22 487 0.40 577 TG 50:1; [M+Li]+; TG[16:0/17:0/17:1]	2 largest peaks:
6 lipidblast-pos 22 487 0.40 577 TG 49:2; [M+Na]+; TG(16:0/16:1/17:1)	557.51181 999.00   583.52745 999.00   2 m/z Values and Intensities:
7 lipidblast-neg 12 344 0.28 408 SQDG 36:5; [M-H]-; SQDG(16:0/20:5(5Z,8Z,11Z	01 557.51181 999.00 [M+Li]·sn3+Li
8 lipidblast-neg 12 344 0.28 408 SQDG 36:5; [M-H]-; SQDG(20:5(52,82,11Z,14Z,	600 660 720 780 840 583.52745 999.00 [M+Li]·sn1+Li    [M+Li]·sn2+Li
Names & Structures / UB 1:-4	lipidblast-pos) TG 50.1; (M+Li)+; TG(16:0/16:0/18:1)
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	Peptide Peptide //

<u>Name:</u> Thermo Finnigan LTQ linear ion trap; [M+Li]+; TG 60:12 <u>MW:</u> 957 <u>ID#:</u> 90 <u>DB:</u> Spec. List <u>Comment:</u> TAG(20:4/20:4/20:4); [M+Li]+; Prec. m/z: 957.6; Electrospray Ionization Multiple-Stage Linear Ion-trap Mass Spectrometry for Structural Elucidation of Triacylglycerols: Assignment of Fatty Acyl Groups on the Glycerol Backbone and Location of Double Bonds; Fong-Fu Hsu and John Turk <u>7 largest peaks:</u>

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	1. 70.0010
	.ij+; 1G 60:12
oz         Comment: TAG(20:4/20:4); [M+Li]+; Prec. m           83         L         Thermo LCQ Deca XP MAX ion trap; [M+Na]+; DGDG 34:1         957.6         Zlargest peaks;	1/z: 957.6; Electrospray Ion
84 L Thermo LCQ Deca XP MAX ion trap ESI; [M+H]+; NA	, 361 10.00  483 1
85 L Thermo Li U; [M+H]+; PC 34; 1; 86 L Thermo Finnigan LTQ linear ion trap; [M+Na]+; TG 52;1 (Gene Lin) Thermo Finnigan LTQ linear ion trap; [M+Na]+; TG 52;1	
87 L Thermo Finnigan LTQ linear ion trap; [M+NH4]+; TG 52:1 Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List	ist /
88 L Thermo Finnigan LTQ linear ion trap; [M+Li]+; TG 50:1	
90 L Thermo Finnigan LTQ linear ion trap; [M+Li]+; TG 60:12 100-	
91 L Thermo Fisher LTQ-FT; [M-H]-; GM2-alpha 92 L Thermo Fisher LTQ-FT: (M-H)-PG-20-2	
Names Structures Spec List	957.6
custompo+bnos msp: custompo+papos msp: bilic-urine: linidblast-pen; pc-ac-pen msp: pc-form-pen msp: linidblast-pos; 235370 0 311 361 483	
total spectra	
10-	
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🗰 Library Score Dot Product Prob. (%) Rev-Dot Name 🔺 🔺 Thermo Finnigan LTQ linear ion tra Head to Tail MF=3 RMF=978	60:12; [M+Li]+; TG(20:4/20:4/
1 lipidblast-pos 3 978 2.17 1e+3 TG 60:12; [M+Li]+; TG(20:4/20:4/20:4) 🗌 🚺 Difference 🔪 Head to Tail 🖉 Side by Side 🔪 Subtraction 🖊	3 978R 2.17P
2 lipidblast-pos 0 621 1.91 632 TG 59:5; [M+Li]+; TG(19:0/20:1/20:4)	i]+; TG(20:4/20:4/20:4)
3 lipidblast-pos 0 621 1.91 632 1G 58:6; [M+Na]+; 1G [18:1/20:1/20:4] 100- 4 lipidblast-pos 0 564 1.91 577 TG 59:5: [M+I]+; 1G [17:1/20:4/22:1] 100-	<u>/B:</u> lipidblast-pos 75189 Mz_exact=957 7518
5 lipidblast-pos 0 564 1.91 577 TG 59:5; [M+Li]+; TG[17:1/20:4/22:0]	
6 lipidblast-pos 0 564 1.91 577 TG 59:5; [M+Li]+; TG(18:1/20:4/21:0) 504 1.91 577 505 504 1.91 507 505 505 504 505 505 505 505 505 505 505	isities:
7 lipidblast-pos 0 564 1.91 577 TG 60:12; [M+Li]+; TG(18:2/20:4/22:6)	+Li]-sn1+Li    [M+Li]-sn2+Li
8 lipidblast-pos U 564 1.91 577 TG 60:12; [M+Li]+; TG(18:3/20:4/22:5) 🔽 450 600 750 900 🔂 <u>Synonyms:</u>	
Image: Names (Structures / Structures / Structures / Structures / Structures / Hit List         Image: Plot/Text of Hit / Plot of Hit / Structures / Struct	
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Peptide Peptide	

Name: Thermo Fisher LTQ-FT; [M-H]-; GM2-alpha

<u>MW:</u> 1354 <u>ID#:</u> 91 <u>DB:</u> Spec. List <u>Comment:</u> GM2alpha ganglioside (putative); [M-H]-; ; Prec. m/z: 1354.783;Method for Lipidomic Analysis: p53 Expression Modulation of Sulfatide, Ganglioside, and Phospholipid Composition of U87 MG Glioblastoma Cells; Anal. Chem., 2007, 79 (22), pp 84238430; 10.1021/ac071413m 9 largest peaks:

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<u> </u>	
# Src. Name	1063.5 Name: Thermo Fisher LTQ-FT; [M-H]-; GM2-alpha
82 L Thermo Finnigan LCQ DECA ion trap; [M-H]-; PIM3(19:0/16:0)	TUUH <u>MW:</u> 1394 <u>IDH:</u> SFec. List Comment: GM2alpha ganglioside (putative): [M-H]-: : Prec. m/z: 1354.783:M
83 L Inerro LCU Deca XP MAX (on trap; [M+Na+; DibDG 34:1	536.4 <u>9 largest peaks:</u>
85 L Thermol TD (M+H)+PC 341.	
86 L Thermo Finnigan LTQ linear ion trap; [M+Na]+; TG 52:1	300 680.4 100.00 1336.5 100.00 1138.6 50.00 1021.3 30.00
87 L Thermo Finnigan LTQ linear ion trap; [M+NH4]+; TG 52:1	Plot/Text of Search Spectrum $\lambda$ Plot of Search Spectrum $\lambda$ Plot/Text of Spec List
88 L Thermo Finnigan LTQ linear ion trap; [M+L]+176 52:1	
90 L Thermo Finnigan LTQ integring that its 16 50.1	860.5 1063.5
91 L Thermo Fisher LTG #104 hor Mark 5 Mark	100-
92 L Thermo Fisher LTQ-FT; [M-H]; PG 36:2	
	50-
Names Structures / Spec List	536.4 698.4 1156.5
custompo+hpos.msp: custompo+hapos.msp: hilic-urine: lipidblast-heg: pc-ac-heg.msp: pc-form-heg.msp: lipidblast-pos: 235370	1021.3 1198.6 1336.5
total spectra	
10-	698.55677
	50-
	1063.68930
1-	
	1354.7847
	560 630 700 770 840 910 980 1050 1120 1190 1260 1330
# Library Score Dot Product Prob. (%) Rev-Dot Name	▲ Thermo Fisher LTQ-FT; [M-H]-; GN Head to Tail MF=168 RMF=436 ▼[glycan]-Cer 34:1; GM2(d18:1/16:)
1 lipidblast-neg 168 436 50.0 617 [glycan]-Cer 34:1; GM2(d18:1/16:0)); [M-H]-; GalNAc	∫ Difference À Head to Tail ∠ Side by Side À Subtraction / 168 436R 50.0P
2 lipidblast-neg 168 436 50.0 617 [glycan]-Cer 34:1; Ganglioside; [M-H]-; NeuAcalpha2	Name: [glycan]-Cer 34:1; GM2(d18:1/16:0)); [M-H]
	100-1354.7847 <u>MW:</u> 1354 <u>ID#</u> : 79298 <u>DB:</u> lipidblast-neg
	Lormment: Parent=1354.78471 Mz_exact=1354.76
	50-
	5 m/z Values and Intensities;
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	(inidblast-neg) [ducan).Cer 34:1: 6M2(d18:1/16:00): [M]
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Name: Thermo Fisher LTQ-FT; [M-H]; PG 36:2 <u>MW</u>: 773 <u>ID#</u>: 92 <u>DB</u>: Spec. List <u>Comment:</u> PG 36:2; [M-H]; Prec. m/z: 773.5325; Remodeling of phosphatidylglycerol in Synechocystis PCC6803 ;http://dx.doi.org/10.1016/j.bbalip.2009.10.009 <u>7 largest peaks</u>:



<u>Name:</u> Thermo Fisher LTQ-FT; [M+NH4]+; TG 48:2 MW: 820 ID#: 93 DB: Spec. List

Comment: TAG (14:0,16:0,18:2); [M+NH4]+; Prec. m/z: 820.7394;High mass measurement accuracy MS/MS utilizing LTQ-FT; Application of the accurate mass and time tag approach in studies of the human blood lipidome;Jie Ding, Christina M. Sorensen, Navdeep Jaitly, Hongliang Jiang, Daniel J. Orton, Matthew E. Monroe, Ronald J. Moore, Richard D. Smith, Thomas O. Metz;

#### 4 largest peaks:


<u>Name:</u> Thermo Fisher LTQ-FT; [M+NH4]+; TG 50:2 MW: 848 ID#: 94 DB: Spec. List

<u>Comment:</u> TAG (16:0,16:1,18:1) TG 50:2 (putative); [M+NH4]+; Prec. m/z: 848.7707; High mass measurement accuracy MS/MS utilizing LTQ-FT; Application of the accurate mass and time tag approach in studies of the human blood lipidome; Jie Ding, Christina M. Sorensen, Navdeep Jaitly, Hongliang Jiang, Daniel J. Orton, Matthew E. Monroe, Ronald J. Moore, Richard D. Smith, Thomas O. Metz; <u>3 largest peaks:</u>

📕 NIST MS Search 2.0 - [Peptide, Presearch Default - 7 spectra] - 🗆 × \_ 8 × 👗 🖻 🖻 🎒 🔠 👎 🌆 👎 🖬 #/z 🔶 🌹 1. Thermo Fisher LTQ-FT; [M+NH4]+; T( 🔽 🛞 🗄 🚇 🔍 ىلىستا 60 🍉 😂 Name: Thermo Fisher LTQ-FT; [M+NH4]+; TG 50:2 # Src. Name 100-575.5017 ٠ MW: 848 ID#: 94 DB: Spec. List 91 Thermo Fisher LTQ-FT; [M-H]-; GM2-alpha L Comment: TAG (16:0.16:1.18:1) TG 50:2 (putative); [M+NH4]+; Prec. m/z; 8] 92 Thermo Fisher LTQ-FT; [M-H]; PG 36:2 L 3 largest peak: 93 Thermo Fisher LTQ-FT; [M+NH4]+; TG 48:2 1 575.5017 999.00 | 549.4861 420.00 | 577.5176 350.00 | 94 Thermo Fisher LTQ-FT; [M+NH4]+: TG 50:2 540 720 • 3 m/z Values and Intensities: 95 Thermo MALDI LTQ ion trap; [M-H]-; Lipid A L F (Spec. List) Thermo Fisher L1 96 Thermo LTQ Orbitrap; ???; MGDG 34:6; L 97 1 Thermo LTQ Orbitrap; [M+NH4]+; TG 52:2 ; 98 L Thermo LTQ Orbitrap; [M-H]- ;PC 32:0 575,5017 99 Thermo LTQ Orbitrap : TG 52:2 L 100-100 Thermo Orbitrap Velos ESI ; [M-H]-; GM1(d18:1/18:0); 1 101 Thermo Fisher LTQ with DESI; [M-H]-; PS 36:1 Т Thanks 1 VO (2000) - 04 10 - CL 70-10 1.01 50 549,4861 Names 🖌 Structures 🖊 Spec List custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370 total spectra 10-831.76751 50-100-549.48798 1000 900 800 700 600 500 400 300 200 ά 100 840 540 570 6Ó0 630 660 690 720 750 780 810 # Library Score Dot Product Prob. (%) Rev-Dot Name Thermo Fisher LTQ-FT; [M+NH4]+ Head to Tail MF=568 RMF=941 [TG 50:2; [M+NH4]+; TG[16:0/16: Difference  $\lambda$  Head to Tail  $\bigwedge$  Side by Side  $\lambda$  Subtraction , 568 941R 97.3P 568 941 97.3 941 lipidblast-pos TG 50:2; [M+NH4]+; TG(16:0/16:1/18:1) 2 lipidblast-pos 78 506 0.97 674 TG 50:2; [M+NH4]+; TG(16:0/16:0/18:2) Name: TG 50:2; [M+NH4]+; TG(16:0/16:1/18:1) 506 674 3 lipidblast-pos 78 0.97 TG 50:2; [M+NH4]+; TG(16:0/17:1/17:1) MW: 848 ID#: 75736 DB: lipidblast-pos 100H 559 4 lipidblast-pos 56 420 0.38 TG 50:2; [M+NH4]+; TG(16:0/17:0/17:2) Comment: Parent=848.77025 Mz exact=848.7702 34 299 674 4 largest peaks: lipidblast-pos 0.15 TG 50:2: [M+NH4]+: TG(16:1/16:1/18:0) 50-549.48798 999.00 | 575.50362 999.00 | 577.5 26 559 6 lipidblast-pos 248 0.11 TG 50:2; [M+NH4]+; TG(14:1/16:1/20:0) 831.76751 4 m/z Values and Intensities: lipidblast-pos 26 248 0.11 559 TG 50:2; [M+NH4]+; TG(16:1/17:0/17:1) 7 549.48798 999.00 [M+NH4]-sn3-18 575.50362 999.00 [M+NH4]-sn1-18 560 630 700 770 840 (lipidblast-pos) TG 50:2; [M+NH4]+; TG(16:0/16:1/18:1 Names / Structures / Hit List Plot/Text of Hit / Plot of Hit / Other Search MSMS Lib. Search Names Compare Librarian Peptide Peptide

Name: Thermo MALDI LTQ ion trap; [M-H]-; Lipid A <u>MW:</u> 1823 <u>ID#:</u> 95 <u>DB:</u> Spec. List <u>Comment:</u> Lipid A from MKM10 F. tularensis; [M-H]-; Prec. m/z: 1823.9; C94H178N2O25P2; LipidA-PP-[14/14/14/14/14/14]; DOI: 10.1002/jms.614; Identification of LpxL, a Late Acyltransferase of Francisella tularensis; 10.1128/IAI.01288-06 10 largest peaks:



Name: Thermo LTQ Orbitrap; ???; MGDG 34:6;

MW: 769 ID#: 96 DB: Spec. List

<u>Comment:</u> MGDG 34:6; wrong adduct [M+H]+; Prec. m/z: 769.48;RT 10.45 min ; Advanced Mass Spectrometry Methods for Analysis of Lipids from Photosynthetic Organisms; Bettina Seiwert, Patrick Giavalisco, and Lothar Willmitzer; dx.doi.org/10.1007/978-90-481-2863-1\_20 <u>3 largest peaks:</u>



<u>Name:</u> Thermo LTQ Orbitrap; [M+NH4]+; TG 52:2 ; <u>MW:</u> 876 <u>ID#:</u> 97 <u>DB:</u> Spec. List <u>Comment:</u> TAG(18:1/18:1/16:0); TAG OOP; APCI ; Prec. m/z: 876.7995; Computer assisted Interpretation of Triacylglycerols Mass Spectra;Josef Cvacka and Edita Kofronová <u>4 largest peaks:</u>

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#     Src.     Name       91     L     Thermo Fisher LTQ-FT; [M-H]; GM2-alpha       92     L     Thermo Fisher LTQ-FT; [M-H]; PG 36:2       93     L     Thermo Fisher LTQ-FT; [M+NH4]+; TG 48:2       94     L     Thermo Fisher LTQ-FT; [M+NH4]+; TG 50:2       95     L     Thermo ALDI LTQ ion trap; [M-H]; Lipid A       96     L     Thermo LTQ Orbitrap; ???; MGDG 34:6;       97     L     Thermo ITQ Orbitrap; ???; MGDG 34:6;	100-     876.7995       100-     876.7995       100-     100- <td< th=""></td<>	
37   L   Thermo LTQ Orbitrap; [M-H]; PC 32:0     98   L   Thermo LTQ Orbitrap; TG 52:2     100   L   Thermo Orbitrap (TG 52:2)     100   L   Thermo Orbitrap (TG 52:2)     101   L   Thermo Fisher LTQ with DESI; [M-H]; PS 36:1     102   L   Thermo Fisher LTQ with DESI; [M-H]; PS 36:1     103   L   Thermo Fisher LTQ with DESI; [M-H]; PS 36:1     104   Names & Structures /   Spec List	1 100- 500- 500- 500- 577.5180 603.5335	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370 total spectra	859.774	
1000-1 100- 10-	50- 50-	
	- 100- 577.51926 570 500 520 550 500 720 750 750 910 940 970	
# Library Score Dot Product Prob. (%) Rev-Dot Name	A Thermo LTQ Orbitrap; [M+NH4]+; T Head to Tail MF=766 RMF=980  ▼TG 52:2; [M+NH4]+; TG(16:0/18:	
1   lipidblast-pos   765   980   78.2   992   TG 52:2; [M+NH4]+; TG[16:0718:1718:1]     2   lipidblast-pos   53   460   0.78   585   TG 52:2; [M+NH4]+; TG[16:0718:1718:1]     3   lipidblast-pos   53   460   0.78   585   TG 52:2; [M+NH4]+; TG[16:1718:0718:1]     4   lipidblast-pos   53   460   0.78   585   TG 52:2; [M+NH4]+; TG[16:1718:0718:1]     5   lipidblast-pos   53   460   0.78   585   TG 52:2; [M+NH4]+; TG[16:0718:0720:1]     4   lipidblast-pos   53   460   0.78   585   TG 52:2; [M+NH4]+; TG[16:0716:0720:2]     5   lipidblast-pos   49   440   0.66   713   TG 52:2; [M+NH4]+; TG[16:0716:0720:2]     6   custompc+n   42   258   0.50   310   PC 41:3; [M+Na]+; GPCho[15:1[32]/26:2[52,92] ♥     7   custompc+n   42   258   0.50   310   PC 41:3; [M+Na]+; GPCho[15:1[32]/26:2[52,92] ♥     8   custompc+n   42   258   0.50   310   PC 41:3; [M+Na]+; GPCho[15:1[32]/26:2[52,92] ♥   Imames     Names	Name:     TG 52:2;     [M+NH4]+;     TG [16:0/18:1/18:1]       100-	
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<u>Name:</u> Thermo LTQ Orbitrap; [M-H]- ;PC 32:0 <u>MW:</u> 792 <u>ID#:</u> 98 <u>DB:</u> Spec. List <u>Comment:</u> PC 32:0; PC(14:0/18:0); [M-H]- ; Prec. m/z: 792.57; Essential Lipidomics Experiments Using the LTQ Orbitrap Hybrid Mass Spectrometer; Thermo Application Note 367 <u>1 largest peaks:</u>

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🚳 🍃 🖻 📫 1. Thermo LTQ Orbitrap; [M-H]- ;PC 32:( 🔤 🛞 🖳 🔍 🍭 🔍	
# Src. Name	Name: Thermo LTQ Orbitrap; [M-H]- ;PC 32:0
91 L Thermo Fisher LTQ-FT; [M-H]; GM2-alpha 92 L Thermo Fisher LTD-FT; [M-H]: PG 36:2	100- <u>Comment:</u> PC 32:0; PC(14:0/18:0); [M-H]- ; Prec. m/z: 792.57; Essential Lip
93 L Thermo Fisher LTQ-FT; [M+NH4]+; TG 48:2	0 1 1 argest peaks: 718.53778 999.00 1
94 L Thermo Fisher LTQ-FT; [M+NH4]+; TG 50:2 95 L Thermo MALDILTQ ion tran: [M-H]-: Lipid A	750 800 1 m/z Values and Intensities:
96 L Thermo LTQ Orbitrap; ???; MGDG 34:6;	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
97 L Thermo LTQ Urbitrap; [M+NH4]+; TG 52:2 ; 98 L Thermo LTQ Orbitrap; [M-H]+; PC 32:0	
99 L Thermo LTQ Orbitrap ; TG 52:2	100-
100 L Thermo Fisher LTQ with DESI; [M-H]; PS 36:1	
100 I Theme 1900 (colored BALID - OL 70:10	50-
Names Structures Spec List	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370 total spectra	
	792.57544
	50-
	100- 718 53866
# Library Score Dot Product Prob. (%) Rev-Dot Name	[▲Thermo LTQ Orbitrap; [M-H]- ;PC1 Head to Tail MF=0 RMF=912 [▼PC 32:0; [M-Ac-H]-; GPCho(16:0/1
1 pc-ac-neg.m 0 912 4.76 913 PC 32:0; [M-Ac-H]-; GPCho(16:0/16:0)	Difference A Head to Tail A Side by Side A Subtraction 0 912R 4.76P
2 pc-ac-neg.m 0 876 4.76 877 PC 32:0; [M-Ac-H]-; GPCho(5:0/25:0) 3 pc-ac-neg.m 0 876 4.76 877 PC 32:0; [M-Ac-H]-; GPCho(7:0/25:0)	Name: PC 32:0; [M-Ac-H]; GPCho(16:0/16:0)
4 pc-ac-neg.m 0 876 4.76 877 PC 32:0; [M-Ac-H]-; GPCho(8:0/24:0)	Comment: Parent=792.57544 Mz_exact=792.5754
5 pc-ac-neg.m 0 876 4.76 877 PC 32:0; [M-Ac-H]-; GPCho(9:0/23:0)	50- 50- 50- 50-
7 pc-ac-neg.m 0 876 4.76 877 PC 32:0; [M-Ac-H]-; GPCho[10:0/22:0]	792.57544 3m/z Values and Intensities:
8 pc-ac-neg.m 0 876 4.76 877 PC 32:0; [M-Ac-H]-; GPCho(12:0/20:0)	718.53866 999.00 [M-CH3]-(-15)
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or Help, press F1	Peptide Peptide //

<u>Name:</u> Thermo LTQ Orbitrap ; TG 52:2 <u>MW:</u> 876 <u>ID#:</u> 99 <u>DB:</u> Spec. List <u>Comment:</u> TAG(18:1/18:1/16:0); APCI ; Prec. m/z: 876.7995; Computer assisted Interpretation of Triacylglycerols Mass Spectra;Josef Cvacka and Edita Kofronová <u>4 largest peaks:</u>



Name: Thermo Orbitrap Velos ESI ; [M-H]-; GM1(d18:1/18:0); MW: 1544 ID#: 100 DB: Spec. List Comment: GM1 ganglioside; GM1(d18:1/18:0); [M-H]-; Prec. m/z: 1544.9; ; Localization, Imaging and Structural Analysis of Sialylated Glycosphingolipids in Brain Tissue Sections by Mass Spectrometry.; Benoit Colsch1, Shelley N. Jackson1, Sucharita M. Dutta2, Alice M. Delvolv1, Amina S, Woods1,: ASMS2010: http://www.asms.org/asms10pdf/ASMS201020798.0375VER.4.pdf

10 largest peaks:



<u>Name:</u> Thermo Fisher LTQ with DESI; [M-H]-; PS 36:1 <u>MW:</u> 788 <u>ID#:</u> 101 <u>DB:</u> Spec. List <u>Comment:</u> PS(18:0/18:1); [M-H]-; Prec. m/z: 788.2; Desorption Electrospray Ionization (DESI) Mass Spectrometry and Tandem Mass Spectrometry (MS/MS) of Phospholipids and Sphingolipids: Ionization, Adduct Formation, and Fragmentation; Nicholas E. Manicke, Justin M. Wiseman, Demian R. Ifa and R. Graham Cooks; <u>6 largest peaks:</u>

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Image:
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# Src Name Name: Thermo Fisher LTO with DESI: [M-H]: PS 36:1
95 L Thermo MALDI LTQ ion trap; [M-H]-; Lipid A 100-701.2 MW: 788 ID#: 101 DB: Spec. List
96 L Thermo LTQ Orbitrap; ???; MGDG 34:6; 97 L Thermo LTQ Orbitrap; ???; MGDG 34:6; 97 419.2 6 largest peaks:
97 L Thermo Li Q Urbitrap; (M+NH4)+; TG 52/2 ; 98 L Thermo LTQ Orbitrap; (M+NH4)+; TG 52/2 ; 98 J Thermo LTQ Orbitrap; (M+NH4)+; TG 52/2 ; 99 J Thermo LTQ Orbitrap; (M+NH4)+; TG 52/2 ; 98 J Thermo LTQ Orbitrap; (M+NH4)+; TG 52/2 ; 99 J Thermo LTQ Orbitrap; (M+NH4)+; TG 52/2 ; 90 J Thermo LTQ Orbitrap; (M+NH4)+; TG 52/2 ; 91 J THERMO LTQ Orbitrap; (M+NH4)+; 91 J THERMO LTQ Orbit
99 L Thermo LTQ Orbitrap ; TG 52:2 (Spec. List) Thermo Fisher L1
100 L Thermo Orbitrap Velos ESI ; [M-H]; GM1(d18:1/18:0); 101 L Thermo Eicher LTD with DECI- (M H1: PS 30:1
102 L Themo LXQ iontrap: [M-H]; CL 76:10
103 L Thermo LXQ iontrap; ESI; [M-H]-; PS 38:4 100-
104 L Thermo LXQ iontrap; ESI; (M-H); PS 422 MX
Names / Structures / Spec List / 10.0
total spectra
1000 <sub>1</sub> 417.24079
100-
10-
300 350 400 450 500 550 600 650 700 750 800
H Library Score Dot Product Product Product Name A Interno Fisher LTQ with DESt; [M] Head to Tail M F=75 RMF=853 (▼P5.35; 1; [M-H]; GPSer[18:0/18:1] Interno Fisher LTQ with DESt; [M] Head to Tail M F=75 RMF=853 (▼P5.35; 1; [M-H]; GPSer[18:0/18:1] Difference A Head to Tail A Side by Side A Subtraction / 75.863R 1.70P
2 birthasher 75 863 1.70 866 PS 300; (min; di Sec(100/101(112))
3 lipidblast-neg 75 863 1.70 866 PS 36:1; [M-H]; GPSer[18:0/18:1[132]] 100 701.51215 Name: PS 36:1; [M-H]; GPSer[18:0/18:1[14]]
4 lipidblast-neg 75 863 1.70 866 PS 36:1; [M-H]; GPSer(18:0/18:1(17Z))
5 lipidblast-neg 75 863 1.70 866 PS 36.1; [M-H]; GPSer(18:0/18:1(4E)) 50-
6 lipidblast-neg /5 863 1./U 866 PS 36.1; [M-H]; GPSer(18:0/18:1(62)) 417.24079 281.24790 100.00   283.26354 100.00   283.26354 100.00   283.26354 100.00
7 ipipulastrieg 75 003 1.70 000 F5 30.1; [M-H]; GFSet[16.0/10.1(72,1)]
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Peptide Peptide

Name: Thermo LXQ iontrap; [M-H]-; CL 76:10 <u>MW</u>: 1499 <u>ID#</u>: 102 <u>DB</u>: Spec. List <u>Comment</u>: CL 76:10; MIX; [M-H]-; Prec. m/z: 1448.0; Oxidative lipidomics of hyperoxic acute lung injury: mass spectrometric characterization of cardiolipin and phosphatidylserine peroxidation; doi:10.1152/ajplung.00035.2010 10 largest peaks:



Name: Thermo LXQ iontrap; ESI; [M-H]-; PS 38:4 <u>MW:</u> 810 <u>ID#:</u> 103 <u>DB:</u> Spec. List <u>Comment:</u> PS 38:4; PS(18:0/20:4); [M-H]-; Prec. m/z: 810.5; Oxidative lipidomics of hyperoxic acute lung injury: mass spectrometric characterization of cardiolipin and phosphatidylserine peroxidation; doi:10.1152/ajplung.00035.2010 7 largest peaks:



Name: Thermo LXQ iontrap; ESI; [M-H]-; PS 42:2 MIX <u>MW:</u> 870 <u>ID#:</u> 104 <u>DB:</u> Spec. List <u>Comment:</u> PS 42:2 mix (ox); PS(18:1/24:1); [M-H]-; Prec. m/z: 870.5; Oxidative lipidomics of hyperoxic acute lung injury: mass spectrometric characterization of cardiolipin and phosphatidylserine peroxidation; doi:10.1152/ajplung.00035.2010 <u>10 largest peaks:</u>



Name: Thermo LTQ Orbitrap; [M-H]-; NA <u>MW:</u> 772 <u>ID#:</u> 105 <u>DB:</u> Spec. List <u>Comment:</u> PE mix (unresolved); [M-H]-; Prec. m/z: 772.5; Shotgun Lipidomics Identifies a Paired Rule for the Presence of Isomeric Ether Phospholipid Molecular Species; Kui Yang, Zhongdan Zhao, Richard W. Gross, Xianlin Han <u>10 largest peaks:</u>

PE MIX (unresolved)



Name: Thermo LTQ Orbitrap; [M-H]-; plasmenyl-PE 38:4 <u>MW:</u> 750 <u>ID#:</u> 176 <u>DB:</u> Text File <u>Comment:</u> plasmenyl-PE 18:0-20:4 (plasmenylethanolamine); [M-H]-; Prec. m/z: 750.54; Shotgun Lipidomics Identifies a Paired Rule for the Presence of Isomeric Ether Phospholipid Molecular Species; Kui Yang, Zhongdan Zhao, Richard W. Gross, Xianlin Han <u>8 largest peaks:</u>



<u>Name:</u> Thermo Finnigan TSQ-7000 triple quadrupole; [M+Li]+; DG 34:1 <u>MW:</u> 601 <u>ID#:</u> 107 <u>DB:</u> Spec. List <u>Comment:</u> DAG(16:0/18:1); [M+Li]+; Prec. m/z: 601.7; Electrospray Ionization/Mass Spectrometric Analyses of Human Promonocytic U937 Cell Glycerolipids and Evidence That Differentiation Is Associated with Membrane Lipid Composition Changes That Facilitate Phospholipase A2 Activation; DOI 10.1074/jbc.M908342199 <u>10 largest peaks:</u>



Name: Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]-; CL 68:3 <u>MW:</u> 1401 <u>ID#:</u> 174 <u>DB:</u> Text File

<u>Comment:</u> Cardiolipin CL (18:1/16:0/18:1/16:1); CL 68:3; [M-H]-; Prec. m/z: 1401.9; Structural Characterization of Cardiolipin by Tandem Quadrupole and Multiple-Stage Quadrupole Ion-Trap Mass Spectrometry with Electrospray Ionization;John Turk, Elizabeth R. Rhoades, David G. Russell, Yixin Shi and Eduardo A. Groisman;http://dx.doi.org/10.1016/j.jasms.2004.12.015 10 largest peaks:

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#   Src.   Name     107   A   Thermo Finnigan TSQ-7000 triple quadrupole; [M+L]+; DG 34:1     108   A   Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M+H]+; CL 68:3     109   A   Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M+H]+; CL 68:3     100   A   Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M+H]+; CL 68:2     110   A   Thermo Finnigan TSQ Quantum Triple Quadrupole; [M+H]+; PC 32:0     112   A   Thermo Finnigan TSQ Quantum Triple Quadrupole; [M+H]+; PC 32:0     113   A   Thermo Finnigan TSQ Quantum Triple Quadrupole; [M+H]+; PC 32:0     114   A   Waters AutoSpec magnetic sector MS; [M+H]+; PC 36:2     115   A   Waters micro QTOF ; [M+Na]+; MGDG 38:4     116   A   Waters Micromass Q-TOF Micro ; [M-H]+; PG 44:12     117   A   Waters MicroMass QqQ triple quadrupole; [M-H]+; SQDG 30:0     100   A   Waters MicroMass QqQ triple quadrupole; [M-H]+; CL 74:0     115   A   Waters MicroMass QqQ triple quadrupole; [M-H]+; SQDG 30:0     116   A   Waters MicroMass QqQ triple quadrupole; [M-H]+; CL 74:0     117   A   Waters MicroMass QqQ triple quadrupole; [M-H]+; CL 74:0     118   A	100   281     100   281     100   101     100   101     100   101     100   101     100   101     100   101     100   101     100   101     101   101     101   101     101   101     101   101     101   101     101   101     101   101     101   101     101   101     101   101     101   111     101   111     102   111     103   111     104   111     105   111     106   1111     107   1111     100   1111     1100   1111     1100   1111     1100   1111     1111   1111     1111   1111     1111   1111     1111<	
# Library Score Dot Product Prob. (%) Rev-Dot Name	240 360 480 600 720 840 360 1080 1200 1320 ▲Themo Finnigan MAT TSQ 7000 Head to Tail MF=366 RMF=582 ▼CL 68:3: (M-H1: CL(16:0/18:1/16)	
1 lipidblast-neg 368 584 35.3 604 CL 68:3; [M-H]-; CL(16:1/18:1/18:1/16:0)	Difference A Head to Tail Side by Side A Subtraction / 366 582R 32.6P	
2   lipidblastneg   366   592   32.6   607   CL 68:3; [M-H]; CL(16:0/18:1/16:1/18:1)     3   lipidblastneg   336   549   9.06   584   CL 68:3; [M-H]; CL(16:0/18:1/18:1/16:0)     4   lipidblastneg   333   545   8.00   569   CL 68:3; [M-H]; CL(16:0/18:1/18:1/18:1/16:1)     5   lipidblastneg   306   515   2.35   549   CL 68:3; [M-H]; CL(16:0/18:1/18:1/18:1)     6   lipidblastneg   306   515   2.35   549   CL 68:3; [M-H]; CL(16:0/18:1/16:1/18:1)     7   lipidblastneg   306   515   2.35   549   CL 68:3; [M-H]; CL(16:0/18:1/16:1/18:1)     7   lipidblastneg   302   510   1.99   548   CL 68:3; [M-H]; CL(16:0/18:1/16:0/18:2)     8   lipidblastneg   301   509   1.91   542   CL 68:3; [M-H]; CL(16:0/18:1/16:1/18:1)   ▼     1   Mames   Structures   Hit List   Hit List   Lib. Search   Other Search   Names   Compare   Librarian   MSMS	100   671.46489     50   417.24045     0   1145.73998     280   560     840   1120     1100   1120     1112   1100     1112   1100     11145.73998   671.46489     11145.73998   10.13724625     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11145.73998   11.145.73998     11120   1120     11120   1120     11120   1120     11120   1120     11120   1120     11120   1120     11120   1120     11120   1120     111120   1120     11120   1120 </th	
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Name: Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]-; CL 72:8 <u>MW:</u> 1448 <u>ID#:</u> 109 <u>DB:</u> Spec. List <u>Comment:</u> CL 72:8; Cardiolipin CL (18:2/18:2/18:2); CL 72:8; [M-H]-; Prec. m/z: 1448.0; CARDIOLIPIN LOSS DURING MYOCARDIAL ISCHEMIA; Am J Physiol Heart Circ Physiol 280: H2770-H2778, 2001 3 largest peaks:



<u>Name:</u> Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]-; CL 68:2 <u>MW:</u> 1403 <u>ID#:</u> 110 <u>DB:</u> Spec. List <u>Comment:</u> Cardiolipin CL (18:1/16:0/18:1/16:0); CL 68:2; [M-H]-; Prec. m/z: 1403.9; Structural Characterization of Cardiolipin by Tandem Quadrupole and Multiple-Stage Quadrupole Ion-Trap Mass Spectrometry with Electrospray Ionization <u>10 largest peaks:</u>



Name: Thermo Finnigan TSQ Quantum Triple Quadrupole; [M+H]+; PC 32:0 <u>MW</u>: 734 <u>ID#</u>: 111 <u>DB</u>: Spec. List <u>Comment</u>: PC 32:0; PC(16:0/16:0); [M-H]-; Prec. m/z: 734.8; Lipidomics: An analysis of cellular lipids by ESI-MS; Stephen Milne, Pavlina Ivanova, JeVrey Forrester, H. Alex Brown; Methods 39 (2006) 92-103 <u>6 largest peaks</u>:



<u>Name:</u> Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]-; PI 32:0 <u>MW:</u> 809 <u>ID#:</u> 112 <u>DB:</u> Spec. List <u>Comment:</u> PI 32:0; PI(16:0/16:0); [M-H]-; Prec. m/z: 809.7; Lipidomics: An analysis of cellular lipids by ESI-MS; Stephen Milne, Pavlina Ivanova, JeVrey Forrester, H. Alex Brown; Methods 39 (2006) 92-103 <u>10 largest peaks</u>



Name: Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]-; PS 36:2 <u>MW:</u> 786 <u>ID#:</u> 113 <u>DB:</u> Spec. List <u>Comment:</u> PS 36:2; PC(18:1/18:1); [M-H]-; Prec. m/z: 786.8; Lipidomics: An analysis of cellular lipids by ESI-MS; Stephen Milne, Pavlina Ivanova, JeVrey Forrester, H. Alex Brown; Methods 39 (2006) 92-103 7 largest peaks:



Name: Waters AutoSpec magnetic sector MS; [M+H]+; PC 24:0 MW: 622 ID#: 114 DB: Spec. List

Comment: PC(12:0/12:0); Matrix-assisted laser desorption/ionization mass spectrometry of phospholipids; D. J. Harvey; Journal of Mass Spectrometry, Volume 30, Issue 9 (p 1333-1346)

5 largest peaks:

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t Src. Name	Name: Waters AutoSpec magnetic sector MS; [M+H]+; PC 24:0	
109 L Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]-; CL 72:8	100 422.6 MW: 622 ID#: 114 DB: Spec. List	
110 L Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]; CL 68:2	5 largest peaks:	
112 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]; PI 32:0	622.7 999.00 422.6 620.00 440.3 600.00 644.6 600.00 585.5 10 520 650 5 m/z Values and Intensities:	
113 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]-; PS 36:2	(Spec. List) Waters AutoSper	
115 L Waters micro QTOF ; [M+Na]+; MGDG 38:4	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List	
116 L Waters Micromass Q-TOF Micro ; [M-H]; PG 44:12	622.7	
117 L Waters MicroMass Ugu triple quadrupole; [M-H]+; 5U/D 30:0	1 100-	
119 L Waters QTOF Premier; [M-H]-; Ac2PIM2	422.6 440.3 644.6	
100 L MALLO OTOF Deserve AUDI, DA 04.1	50-	
Names Structures Spec List	595.5	
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370		
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1 lipidplast-pos 236 514 72.4 533 PC 24:0; [M+H]+; GPC ho(12:0/12:0) = 2 custompo+b 196 407 14.7 496 PC 24:0; [M+H]+; GPC ho(12:0/12:0)		
3 lipidblast-pos 138 355 3.05 612 MGDG 23:0; [M+NH4]+; MGDG(11:0/12:0)	100 563.37127 Name: PC 24:0; [M+H]+; GPCho[12:0/12:0]	
4 lipidblast-pos 138 355 3.05 612 MGDG 23:0; [M+NH4]+; MGDG(12:0/11:0)	Comment: Parent=622.44477 Mz_exact=622.4447	
5 lipidblast-pos 79 229 0.55 395 MGDG 25:0; [M+NH4-C0]+; MGDG(12:0/13:0)	50- 604.43421 563 37127 999 00 1 422 26725 600 00 1 440 2	
6 lipidblast-pos /9 229 0.55 395 MGDG 25:0; [M+NH4-C0]+; MGDG (13:0/12:0) 7 lipidblast-pos 61 192 0.29 216 DS 24:0; [M+NH4-C0]+; MGDG (13:0/12:0)	5 m/z Values and Intensities:	
7 iipiuulastineg 61 163 0.26 316 F3 24.0; [MiH]; GPSet(7.0/17.0) 8 iipidblastineg 61 183 0.28 316 PS 24.0; [MiH]; GPSer(17:0/7:0)	0 422.26725 600.00 [M+H]·sn1·H20    [M+H]·sn2·H	
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Name: Waters micro QTOF ; [M+Na]+; MGDG 38:4 <u>MW:</u> 829 <u>ID#:</u> 115 <u>DB:</u> Spec. List <u>Comment:</u> MGDG(20:1;18:3); NAPOLITANO et al ;J. Agric. Food Chem., Vol. 55, No. 25, 2007; Novel Galactolipids from the Leaves of Ipomoea batatas L.: Characterization by Liquid Chromatography Coupled with Electrospray IonizationQuadrupole Time-of-Flight Tandem Mass Spectrometry <u>10 largest peaks:</u>

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#     Src.     Name       109     I     Thermo Finningan MAT TSD 7000 Triple Quadrupole: [MiHL: CL 72:8]	100-333.4 783.8 <u>Name</u> , waters micro Q10F; (M+Na)+; MGDG 36.4 <u>MW:</u> 829 [D#: 115 <u>DB:</u> Spec. List	
110 L Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]; CL 68:2	519.5 Comment: MGDG(20:1;18:3); NAPOLITANO et al ;J. Agric. Food Chem., Vol.—	
111 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M+H]+; PC 32:0		
113 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]-; PS 36:2	350 /UU 335.4 400.00 341.4 400.00 829.9 400.00 519.5 350.00 830.9 30 ▲	
114 L Waters AutoSpec magnetic sector MS; [M+H]+; PC 24:0	Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /	
115 L Waters micro UTUF; (M+Na)+; MGDG 38:4 116 L Waters Micromass Q-TOF Micro : (M-H)-; PG 44:12		
117 L Waters MicroMass QqQ triple quadrupole; [M-H]-; SQDG 30:0		
118 L Waters QqQ triple quadropole VG Quattro II; [M-H]-; GM1	243.2	
120 L Michael OTOF Denvice (MUL) DA 24.1	1 50 8299 8299	
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total spectra		
1 <sup>100</sup> 1		
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# Library Score Dot Product Prob. (%) Rev-Dot Name	▲Waters micro QTOF; [M+Na]+; M Head to Tail MF=170 RMF=298 ▼MGDG 38:4; [M+Na]+; MGDG(18)	
1 lipidblast-pos 170 298 5.67 957 MGDG 38:4; [M+Na]+; MGDG(18:3(6Z,9Z,12Z)/	Difference A Head to Tail Side by Side A Subtraction / 170 298R 5.67P	
2 lipidblast-pos 170 298 5.67 957 MGDG 38:4; [M+Na]+; MGDG(18:3(62,92,122)/	Mame: MGDG 38:4; [M+Na]+; MGDG(18:3(6Z,9Z,	
3 lipidbiast-pos 170 298 5.67 957 Mid.Did 38:4; [M+Na]+; Mid.Did 18:3(62,32,122)/ 4 lipidblast-pos 170 298 5.67 957 MiGDG 38:4; [M+Na]+; MiGDG(18:3(62,92,122)/	100- 100- 100- <u>MW: 829 ID#: 12497 DB: lipidblast-pos</u> Comment: Parent=829 58057 Mz_exact=829 5805	
5 lipidblast-pos 170 298 5.67 957 MGDG 38:4; [M+Na]+; MGDG[18:3(9Z,12Z,15Z]	2 largest peaks:	
6 lipidblast-pos 170 298 5.67 957 MGDG 38:4; [M+Na]+; MGDG(18:3(92,122,152)	519.29357 999.00   551.35613 999.00   2 m/z Values and Intensities	
7 lipidblast-pos 170 298 5.67 957 MGDG 38:4; [M+Na]+; MGDG(18:3(92,122,152)	519.29357 999.00 [M+Na]-sn2	
8 IIpidpiast-pos 170 238 5.67 357 MGDG 38:4; [M+Na]+; MGDG [18:3(92,122,152]	260 390 520 650 780 551.35613 999.00 [M+Na]-sn1	
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Name: Waters Micromass Q-TOF Micro ; [M-H]-; PG 44:12 <u>MW:</u> 865 <u>ID#:</u> 116 <u>DB:</u> Spec. List <u>Comment:</u> 22:6/22:6-GPG; [M-H]- ; mz=865.336; Song et al ;Algorithms for automatic processing of data from mass spectrometric analyses of lipids; Journal of chromatography. B, Analytical technologies in the biomedical and life sciences [1570-0232] Song (2009) volume: 877 issue: 26 page: 2847-2854 10 largest peaks:



Name: Waters MicroMass QqQ triple quadrupole; [M-H]-; SQDG 30:0 <u>MW:</u> 765 <u>ID#:</u> 117 <u>DB:</u> Spec. List <u>Comment:</u> SQDG 30:0; [M-H]-; Prec. m/z: 765.77; CID spectrum; Glyco- and sphingophosphonolipids from the medusa Phyllorhiza punctata: NMR and ESI-MS/MS fingerprints; http://dx.doi.org/10.1016/j.chemphyslip.2006.11.001 <u>10 largest peaks:</u>



Name: Waters QqQ triple quadropole VG Quattro II; [M-H]-; GM1 <u>MW:</u> 1572 <u>ID#:</u> 118 <u>DB:</u> Spec. List <u>Comment:</u> GM1 (d20:1/C18:0)(?); [M-H]-; Prec. m/z: 1572.5; GM1-gangliosidosis in a cross-bred dog confirmed by detection of GM1-ganglioside using electrospray ionisation-tandem mass spectrometry;DOI 10.1007/s004010000187 <u>10 largest peaks:</u>

Comment from Publication:

Purified bovine brain monosialoganglioside-GM1 consisting of GM1(d18:1–C18:0) and GM1 (d20:1–C18:0) was obtained from Sigma (Poole, UK).

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# Src. Name	Lang 1 1572 E Name: Waters QqQ triple quadropole VG Quattro II; [M-H]; GM1	
109 L Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M-H]; CL 72:8	100-250.1 1572.5 MW: 1572 ID#: 118 DB: Spec. List 916.5 Comment: GM1 (d20:1/C18:0)(2): [M-H]: Prec. m/z: 1572.5: GM1-gangligsid	
110 L Thermo Finnigan MAT TSQ 7000 Triple Quadrupole; [M+H]+; CL 68:2 111 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M+H]+; PC 32:0		
112 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]; PI 32:0		
113 L Thermo Finnigan TSQ Quantum Triple Quadrupole; [M-H]; PS 36:2 114 J Waters AutoSpec magnetic sector MS ( [M+H]+; PC 24:0	(Spec. List) Waters QqQ triple	
115 L Waters micro QTOF ; [M+Na]+; MGDG 38:4	Plot/Text of Search Spectrum A Plot of Search Spectrum A Plot/Text of Spec List A	
116 L Waters Micromass Q-TOF Micro ; [M-H]-; PG 44:12 117 L Waters MicroMass QoD triple guadrupple: [M-H]-: SDDG 30:0	100 290.1 1572.5	
118 L Waters QqQ triple quadropole VG Quattro II; [M-H]-; GM1	100-	
119 L Waters QTOF Premier; (M-H); Ac2PIM2	916.5	
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# Library Score Dot Product Prob. (%) Rev-Dot Name	▲Waters QqQ triple quadropole VG Head to Tail MF=353 RMF=659  ▼[glycan]-Cer 38:1; GM1(d18:1720:	
1 lipidblast-neg 353 659 10.9 895 [glycan]-Cer 38:1; Ganglioside; [M-H]-; NeuAcalp	Difference A Head to Tail A Side by Side A Subtraction / 353 659R 10.9P	
2 ipidolast-neg 353 659 10.9 855 [gij/can]-Cer 3617 (MT(d18:17200)); [M-H]-; Gal—	1572.9001 Name: [glycan]-Cer 38:1; GM1(d18:1/20:0)); [M-H]	
4 lipidblast-neg 353 659 10.9 895 [glycan]-Cer 38:1; GM1alpha(d18:1/20:0)); [M-H]	100- Comment: Parent=1572.90013 Mz_exact=1572.90	
5 lipidblast-neg 353 659 10.9 895 [glycan]-Cer 38:1; sialyl-lactotetraosylceramide(d1	1281.80472 50 1281.80472 500 00 1 22 1572 90013 999 00 1 1281 80472 500 00 1 22	
b lipidplast-neg 353 659 10.9 895 [glycan]-Cer 38:1; sialyl[2-6]lactotetraosylceramid; 7 lipidplast-neg 353 659 10.9 895 [glycan]-Cer 38:1; Ganglioside; [M-H]: Galbeta1.	592.56651 5 m/2 Values and Intensities:	
8 lipidblast-neg 353 659 10.9 895 [glycan]-Cer 38:1; Ganglioside; [M-H]-; NeuAcalp 🖵	290.08799 400.00 fon CTTHT6N08-[290.0873: 290 580 870 1160 1450 592.56651 200.00 ion ceramide	
	(lipidblast-neg) [glycan]-Cer 38:1; GM1(d18:1/20:0)); [M	
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Name: Waters QTOF Premier; [M-H]-; Ac2PIM2 <u>MW:</u> 1175 <u>ID#:</u> 119 <u>DB:</u> Spec. List <u>Comment:</u> Ac2PIM2(methyl-18:0/16:0); [M-H]-; Prec. m/z: 1175.6753; Synthesis and Structure of Phosphatidylinositol Dimannoside;J. Org. Chem., 2007, 72 (9), pp 3282-3288; 10.1021/jo0625599 10 largest peaks:



Name: Waters QTOF Premier; [M-H]-; Ganglioside; [glycan]-Cer(d18:1, C24:1) <u>MW:</u> 1627 <u>ID#:</u> 162 <u>DB:</u> Text File <u>Comment:</u> IV3Neu5Ac-nLc4Cer (d18:1, C24:1); [M-H]-; Prec. m/z: 1627.10; Mass Spectrometry ReviewsVolume 29, Issue 3, DOI:10.1002/mas.20253 <u>10 largest peaks:</u>

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🚳 ⊳ 😅 📫   1. Waters QTOF Premier; [M-H]-; Gangli 🗾 🛞 🖺 🔛 🧶 🔍		
tt Src Name	Name: Waters 0T0F Premier: [M-H1: Ganglioside: [glycan]-Cer(d18:1, C24:1	
118 A Waters QqQ triple quadropole VG Quattro II; [M-H]-; GM1	100-250.16 MW: 1627 ID#: 162 DB: Text File	
119 A Waters QTOF Premier; [M-H]; Ac2PIM2	1627.1 10 largest peaks:	
12U A Waters U I UF Premier; [M-H]-; Ganglioside; [glycan]-Cer[d18:1, U24:1] 121 A Waters OTOE Premier; [M-H]-; Ganglioside; [glycan]-Cer[d18:1, U24:1]		
121 A Waters QTOF Fremier, [M-H]-, Cangiloside, [giydan]-cel(010.1, C10.0)	790 1580 1173.89 31.00 1155.90 18.00 646.68 12.00 470.23 1.00	
123 A Waters QTOF Premier; [M-H]-; NA	Itext File) Waters QTUF Prei	
124 A Waters QTOF Premier; [M-H]-; PE 34:0		
125 A Waters UTUF Premier; [M-H]-; PE 34:1	290.16	
127 A Waters OTOF Premier: [M-H]-: NA	1004	
128 A Waters QTOF Premier; [M-H]-; PS 34:1		
	50- 1627.1	
Names Structures Spec List	1335.95	
[ipidblast-neg; lipidblast-pos; custompc+hpos.msp; custompc+napos.msp; pc-ac-neg.msp; pc-form-neg.msp; 234420 total spectra	470,23 646,68 808,73 970,81 1173,89	
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	646.61343 808.66625	
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1	100-	
t Library Score Dot Product Prob (%) Rev-Dot Name	350 480 500 720 840 950 1080 1200 1320 1440 1550	
1 lipidplast-neg 65 806 11.1 834 [glycani-Cer 42:2: Ganglioside: [M-H]-: NeuAcalo	Difference A Head to Tail Side by Side A Subtraction 65 806R 11.1P	
2 lipidblast-neg 65 806 11.1 834 [glycan]-Cer 42:2; GM1(d18:1/24:1)); [M-H]-; Gall	Name: [dwaw].Cer 42:2: GM1(d19:1/24:1)): [M,H]	
3 lipidblast-neg 65 806 11.1 834 [glycan]-Cer 42:2; cis GM1, GM1b(d18:1/24:1)); [	1626.9470 Nw: 1626 ID#; 79312 DB; lipidblast-neg	
4 lipidblast-neg 65 806 11.1 834 [glycan]-Cer 42:2; GM1alpha(d18:1/24:1)); [M-H]	Comment: Parent=1626.94708 Mz_exact=1626.94	
5 lipidblast-neg 65 806 11.1 834 [glycan]-Cer 42:2; sialyl-lactotetraosylceramide(d1	1335.85167 <u>5 largest peaks:</u> 1626.94708.999.001.1335.85167.500.001.20	
6 lipidblast-neg 65 806 11.1 834 [glycan]-Cer 42:2; sialyl(2-6)lactotetraosylceramid	646.61343 5 m/z Values and Intensities:	
7 lipidplast-neg 65 806 11.1 834 [glycan]-Cer 42:2; Ganglioside; [M-H]-; Galbeta1- 0 [widthast-neg 65 000 11.1 004 [etwach] Cer 40:0; Cer 40:0; Cer 40:0; M-H)-; Calbeta1-	0 290.08759 400.00 ion C11H16N08- (290.08759	
o iipiuulaseney oo ooo iii.i oo4 (giycan)-teri4∠:2; taangiloside; (M-H)-; NeuAcab	300 600 900 1200 1500 646.61.343 200.00 ion ceramide	
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Name: Waters QTOF Premier; [M-H]-; Ganglioside; [glycan]-Cer(d18:1, C16:0) <u>MW:</u> 1516 <u>ID#:</u> 161 <u>DB:</u> Text File <u>Comment:</u> IV3Neu5Ac-nLc4Cer(d18:1, C16:0); [M-H]-; Prec. m/z: 1516.95; Mass Spectrometry ReviewsVolume 29, Issue 3, DOI:10.1002/mas.20253 10 largest peaks:



Name: Waters QTOF Premier; [M-H]-; PA 34:1 <u>MW:</u> 673 <u>ID#:</u> 120 <u>DB:</u> Spec. List <u>Comment:</u> PA 34:1 ; PS(16:0/18:1); [M-H]-; Prec. m/z: 673.5; Profiles of photosynthetic glycerolipids in three strains of Skeletonema determined by UPLC-Q-TOF-MS; Journal of Applied Phycology; DOI: 10.1007/s10811-010-9553-3 <u>8 largest peaks:</u>



Name: Waters QTOF Premier; [M-H]-; NA <u>MW:</u> 716 <u>ID#:</u> 121 <u>DB:</u> Spec. List <u>Comment:</u> PC 32:1 (wrong prec m/z (730, 776.790) ; [M-H]-; Prec. m/z: 716.5; Profiles of photosynthetic glycerolipids in three strains of Skeletonema determined by UPLC-Q-TOF-MS; Journal of Applied Phycology; DOI: 10.1007/s10811-010-9553-3 <u>4 largest peaks:</u>

Wrong precursor, wrong ID, probably mixed spectrum

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🚳 🍃 🚔 📮 1. Waters QTOF Premier; [M-H]-; NA 💽 🛞 🖭	
#   Src.   Name     115   L   Waters micro QTOF; [M+Na]+; MGDG 38:4     116   L   Waters Micromass Q-TOF Micro; [M-H]; PG 44:12     117   L   Waters MicroMass QqU triple quadrupole; [M-H]; SQDG 30:0     118   L   Waters QqU triple quadrupole VG Quattro II; [M-H]; SQDG 30:0     119   L   Waters QqD F Premier; [M-H]; Ac2PIM2     120   L   Waters QTOF Premier; [M-H]; PA 34:1     121   L   Waters QTOF Premier; [M-H]; PA 34:1     122   L   Waters QTOF Premier; [M-H]; PE 34:0     123   L   Waters QTOF Premier; [M-H]; PE 34:1     124   L   Waters QTOF Premier; [M-H]; PE 34:1     125   L   Waters QTOF Premier; [M-H]; PE 34:1     125   L   Waters QTOF Premier; [M-H]; PG 34:1     125   L   Waters QTOF Premier; [M-H]; PG 34:1     126   Waters QTOF Premier; [M-H]; PG 34:1     127   L   Waters QTOF Premier; [M-H]; PG 34:1     125   L   Waters QTOF Premier; [M-H]; PG 34:1     126   Waters QTOF Premier; [M-H]; PG 34:1     127   L   Waters QTOF Premier; [M-H]; PG 34:1     126 <td>Name:     Waters QTOF Premier; [M-H]: NA.       100-     506.3202       230     460       690     590       (Spec. List)     227.1755       100-     100-       9     200       9     0.127.1755       9     0.1281.2328       9     0.1281.2328       100-     100-       9     100-       9     100-       9     100-       9     100-       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     100-</td>	Name:     Waters QTOF Premier; [M-H]: NA.       100-     506.3202       230     460       690     590       (Spec. List)     227.1755       100-     100-       9     200       9     0.127.1755       9     0.1281.2328       9     0.1281.2328       100-     100-       9     100-       9     100-       9     100-       9     100-       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     281.2328       100-     100-
# Library Score Dot Product Prob. (%) Rev-Dot Name	▲ Waters QTOF Premier; [M-H]; NA Head to Tail MF=29 RMF=556 ▼PE 34:1; [M-H]; GPEtn(14:0/20:1
1 lipidblast-neg 29 556 0.99 657 PE 34:1; [M	M-H]-; GPEtn(14:0/20:1(11E))
2 lipidblast-neg 29 556 0.99 657 PE 34:1; [N	M-H]; GPEth(14:0/20:1(11Z))
3 lipidblast-neg 29 556 0.99 657 PE 34:1; [N	M-H]; GPEth(14:0/20:(1(3E)) 100-
4 lipidblast-neg 29 556 0.99 657 PE 34:1; [A	M-H]; GPEth[14//2011[132]]
D ipidblast-neg 29 556 0.99 657 PE 34:1; (M	M-H; CarEm(20:1(11E)/14:0) 50- 227.20088 999.001 309.27918 999.001 424.2
o iipiubiaseneg 23 336 0.33 637 PE 34:1; (n 7 lipidblasteneg 29 559 0.99 657 DE 24:1; (n	MHD; cpcut(20.1(12)/14.0) 424.24662 488.31426 50.001
r iipidulastineg 20 556 0.35 657 PE 34:1; (f 8 linidblastineg 29 556 0.99 657 PE 34:1; (f	M-H): GPEm(20:1(132)/14:0) 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0
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<u>Name:</u> Waters QTOF Premier; [M-H]-; PE 34:0 <u>MW:</u> 718 <u>ID#:</u> 122 <u>DB:</u> Spec. List <u>Comment:</u> PE(17:0/17:0); [M-H]-; Prec. m/z: 718.4834; A Systems Biology Strategy Reveals Biological Pathways and Plasma Biomarker Candidates for Potentially Toxic Statin-Induced Changes in Muscle; 10.1371/journal.pone.0000097 <u>5 largest peaks:</u>

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Here Name: Waters OTOE Premier: MHL: PE 24:0	
#     Src.     Name     Maile:     Waters gifter Heiner, [M+1]; 1: 04:0       115     L     Waters micro QTOF ; [M+Nal+; MGDG 38:4     100-     718.4832     MW: 718 [D#; 122 DB; Spec. List	
116 L Waters Micromass Q-TOF Micro ; [M-H]; PG 44:12	
117 L Waters MicroMass QQQ triple quadrupole; [M-H]; SQDG 30:0	⊆,
119 L Waters QTOF Premier; [M-H]; Ac2PIM2 (Spec_List) Waters QTOF Pr	<u> </u>
120 L Waters QTOF Premier; [M-H]; PA 34:1 121 L Vision OTOF Duration [M-H]; PA 34:1 Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /	
121 L Waters QTOF Premier; [M-H]; NA	-1
123 L Waters QTOF Premier; [M-H]; PE 34:1	
124 L Waters QTOF Premier; [M-H]-; PG 34:1	534
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Custompc+hpos msp: custompc+napos msp: hilic-urine: lipidblast-neg: pc-ac-neg msp: pc-form-neg msp: lipidblast-pos: 235370	
total spectra	
1000-1 466.29353	
# Library Score Dot Product Prob. (%) Rev-Dot Name	17:0
1 lipidblast-neg 70 864 6.16 960 PE 34:0; [M-H]-; GPEtn(17:0/17:0) — 🖓 🗋 Difference 🔪 Head to Tail 🖉 Side by Side \lambda Subtraction / 70 864R 6	6P
2 lipidblast-pos 4 268 0.97 306 MGDG 32:1; [M+NH4-CO]+; MGDG(15:1(92)/17 [Name: PE 34:0; [M-H]; GPEth(17:0/17:0)	
3 ipidolast-pos 4 268 0.97 306 Mid/u 3211 (M+NH4-CU)+/ Mid/u (17:0/15:1(3, 100- 4 linidolast-pos 3 206 0.93 500 MGDG 3211 (M+NH4-CU)+/ MGDG(15:0/17:1(9, 100-	F III
5 lipidblast-pos 3 206 0.93 500 MGDG 32:1; [M+NH4-CO]+; MGDG (17:1(92)/15 3]	
6 custompc+h 3 100 0.93 262 PC 31:1; [M+H]+; GPCho(14:1(9Z)/17:0) 466,29353 200.00 446,29353 200.00 446,29353 200.00 446	
7 custompc+h 3 100 0.93 262 PC 31:1; [M+H]+; GPCho(17:0/14:1(9Z))	
8 lipidblast-neg 2 196 0.89 224 PS 31:1; [M-H]; GPSer(14:1(92)/17:0) 300 400 500 600 700 448.28297 50.00 [M-H]-sn1-H20 [[M-H]-sn2	
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Name: Waters QTOF Premier; [M-H]-; PE 34:1 <u>MW:</u> 716 <u>ID#:</u> 123 <u>DB:</u> Spec. List <u>Comment:</u> PE 34:1; PE(16:0/18:1); [M-H]-; Prec. m/z: 716.5; Profiles of photosynthetic glycerolipids in three strains of Skeletonema determined by UPLC-Q-TOF-MS; Journal of Applied Phycology; DOI: 10.1007/s10811-010-9553-3 <u>5 largest peaks:</u>

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tt Src Name	Name: Waters QTOF Premier: IM-H1: PE 34:1
115 L Waters micro QTOF ; [M+Na]+; MGDG 38:4	100- 281.2205 MW: 716 ID#, 123 DB; Spec. List
116 L Waters Micromass Q-TOF Micro ; [M-H] ; PG 44:12	716 5341 5 largest peaks:
118 L Waters QqQ triple quadropole VG Quattro II; [M-H]-; GM1	281.2205 999.00   255.2099 600.00   140.0014 100.00   452.2726 5C
119 L Waters QTOF Premier; [M-H]; Ac2PIM2	(Spec. List) Waters QTOF Pr
120 L Waters QTOF Premier; [M-H]-; PA 34:1 121 L Waters QTOF Premier; [M-H]-; NA	Plot/Text of Search Spectrum Plot of Search Spectrum Plot/Text of Spec List
122 L Waters QTOF Premier; [M-H]; PE 34:0	281 2205
123 L Waters QTOF Premier; [M-H]-; PE 34:1	100-
125 L Waters QTOF Premier; [M-H]; PS 40:1	
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total spectra	452 27790
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t Library Score Dot Product Prob 1%1 Bey-Dot Name	150 200 250 300 350 400 450 500 550 600 650 700
1 lipidblast-neg 404 895 5.50 923 PE 34:1; [M	M-H]; GPEtn(16:0/18:1(11E))
2 lipidblast-neg 404 895 5.50 923 PE 34:1; [M	4-H]; GPEtn(16:0/18:1(11Z))
3 lipidblast-neg 404 895 5.50 923 PE 34:1; [M	A-H]; GPEtn(16:0/18:1(13Z)) 255.23226 MW/: 716 ID#: 107994 DB: lipidblast-neg
4 lipidblast-neg 404 895 5.50 923 PE 34:1; [M 5 lipidblast-neg 404 995 5.50 923 PE 34:1; [M	A-H]; GPE tr[16:0/18:1[172]] G [argest peaks:
6 lipidblast-neg 404 895 5.50 923 PE 34:1; [M	MHP; dF Edi(16:0/16:1(42)) 50- MHP; GPEtr(16:0/18:1(62)) 50- 452.27200 255.23226 999.00   281.24790 999.00   452.2
7 lipidblast-neg 404 895 5.50 923 PE 34:1; [M	4-H]; GPEtn(16:0/18:1(7Z)) 450.28298 50.001 6 m/z Values and Intensities;
8 lipidblast-neg 404 895 5.50 923 PE 34:1;[M	4-H]; GPEtn(16:0/18:1(9E))
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Name: Waters QTOF Premier; [M-H]-; PG 34:1 <u>MW:</u> 747 <u>ID#:</u> 124 <u>DB:</u> Spec. List <u>Comment:</u> PG 34:1; PG(16:0/18:1:0); [M-H]-; Prec. m/z: 747.5; Profiles of photosynthetic glycerolipids in three strains of Skeletonema determined by UPLC-Q-TOF-MS; Journal of Applied Phycology; DOI: 10.1007/s10811-010-9553-3 <u>8 largest peaks</u>:



Name: Waters QTOF Premier; [M-H]-; PS 40:1 <u>MW:</u> 844 <u>ID#:</u> 125 <u>DB:</u> Spec. List <u>Comment:</u> Unknown wrongly assigned as PS(18:1/22:0) but no FA 22:0 m/z 339.3; [M-H]-; Prec. m/z: 844.6063; RT: 9.71 min; Lipidomics: Study of Total Phospholipids in Immortalized Liver Cells Exposed to Different Fatty Acid Substrates: Pagliasotti,Prenni,Ryan,Rainville,Baker; 720001756EN; <u>10 largest peaks:</u>

Unknown wrongly assigned FA missing

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	Name: Waters OTOF Premier: (M-H): PS 40:1	
+ 510. Name	100- <u>MW:</u> 844 <u>ID#:</u> 125 <u>DB:</u> Spec. List	
116 L Waters Micromass Q-TOF Micro ; [M-H]-; PG 44:12	Comment: Unknown wrongly assigned as PS(16:1722:0) but no FA 22:0 m/2  770.5710 10 largest peaks:	
117 L Waters MicroMass UqU triple quadrupole; [M-H]-; SUDG 30:0		
119 L Waters QTOF Premier; [M-H]-; Ac2PIM2	(Spec. List) Waters QTOF Pr	
120 L Waters QTOF Premier; [M-H]-; PA 34:1	Plot/Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /	
121 L Waters QTOF Premier; [M-H]; PE 34:0		
123 L Waters QTOF Premier; [M-H] ; PE 34:1	100-	
124 L Waters UT UF Premier; [M-H]-; PG 34:1		
120 L Middle Grof Frender, (M.11), DC 24.1	504	
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custompe+bnos msp: custompe+papos msp: bilie-urine: linidblast-pen; pe-ae-pen msp; pe-form-pen msp; linidblast-pos; 235370	506.3250 776.3710 844.6067	
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1 pc-ac-neg.m 113 506 1.07 655 PC 36:2; [M-Ac-H]; GPCho(18:1(11E)/18:1(11E)	Difference A Head to Tail Side by Side A Subtraction / 113 506R 1.07P	
2 pc-ac-neg.m 113 506 1.07 655 PC 36:2; [M-Ac-H] ; GPCho(18:1(11E)/18:1(11Z)	Name: PC 36/2: [M-Ac-H]-: GPCho(18:1(11E)/18:1	
3 pc-ac-neg.m 113 506 1.07 655 PC 36:2; [M-Ac-H]-; GPCho(18:1(11E)/18:1(13Z)	100- 770.56997 <u>MW:</u> 844 ID#: 2569 <u>DB</u> : pc-ac-neg.msp	
4 pc-ac-neg.m 113 506 1.07 655 PC 36:2; [M-Ac-H]; GPCho(18:1(11E)/18:1(17Z)	Comment: Parent=844.60675 Mz_exact=844.6067	
5 pc-ac-neg.m 113 506 1.07 655 PC 36.2; [M-Ac-H]; (Ar-Life)/16.1(46.)] 6 pc-ac-neg.m. 113 506 1.07 655 PC 36.2; [M-Ac-H]; (BPCho[18:1(11F)/18:1(67)])	50- 770.56997 999.00   281.24790 100.00   844.6	
7 pc-ac-neg.m 113 506 1.07 655 PC 36:2; [M-Ac-H]-; GPCho[18:1(11E)/18:1(7Z)]	- <u>3 m/z Values and Intensities:</u> 281 24790, 100 00, EA sn1 II EA sn2	
8 pc-ac-neg.m 113 506 1.07 655 PC 36:2; [M-Ac-H]-; GPCho(18:1(11E)/18:1(9E)) 🗸	360 480 600 720 840 770.56997 999.00 [M-CH3]- (-15) ▼	
	(pc-ac-neg.msp) PC 36:2; [M-Ac-H]; GPCho(18:1(11E)	
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	Peptide Peptide //	

Name: Waters QTOF Premier; [M-H]-; PS 34:1 <u>MW:</u> 760 <u>ID#:</u> 126 <u>DB:</u> Spec. List <u>Comment:</u> PS 34:1; PS(16:0/18:1); [M-H]-; Prec. m/z: 760.5; Profiles of photosynthetic glycerolipids in three strains of Skeletonema determined by UPLC-Q-TOF-MS; Journal of Applied Phycology; DOI: 10.1007/s10811-010-9553-3 <u>9 largest peaks:</u>



Name: Waters QTOF Premier; [M-H]-; PS 36:1 <u>MW:</u> 788 <u>ID#:</u> 127 <u>DB:</u> Spec. List <u>Comment:</u> PS 36:1; PS(16:0/18:1); [M-H]-; Prec. m/z: 788.5; Profiles of photosynthetic glycerolipids in three strains of Skeletonema determined by UPLC-Q-TOF-MS; Journal of Applied Phycology; DOI: 10.1007/s10811-010-9553-3 9 largest peaks:


<u>Name:</u> Waters QTOF Premier; [M+Na]+; MGDG 34:6 <u>MW:</u> 769 <u>ID#:</u> 128 <u>DB:</u> Spec. List <u>Comment:</u> MGDG(18:3/16:3); [M+Na]+; Prec. m/z: 769.4885; Global characterization of the photosynthetic glycerolipids from a marine diatom Stephanodiscus sp. by ultra performance liquid chromatography coupled with electrospray ionization-quadrupole-time of flight mass spectrometry ;http://dx.doi.org/10.1016/j.aca.2010.01.026

7 largest peaks:



<u>Name:</u> Waters QTOF Premier; [M-H]-; PG 34:1 <u>MW:</u> 747 <u>ID#:</u> 129 <u>DB:</u> Spec. List <u>Comment:</u> PG(16:0/18:1); [M-H]-; Prec. m/z: 747.5078; Global characterization of the photosynthetic glycerolipids from a marine diatom Stephanodiscus sp. by ultra performance liquid chromatography coupled with electrospray ionization-quadrupole-time of flight mass spectrometry ;http://dx.doi.org/10.1016/j.aca.2010.01.026 <u>6 largest peaks:</u>

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# Src. Name	100 281.2436 Name: Waters QTOF Premier; [M-H]; PG 34:1
12U L Waters QTOF Premier; [M-H]-; PA 34:1 121 L Waters QTOF Premier; [M-H]-; NA	747.5076 Comment: PG(16:0/18:1); [M-H]; Prec. m/z: 747.5078; Global characterizati
122 L Waters QTOF Premier; [M-H]; PE 34:0	
123 L Waters QTOF Premier; [M-H]; PE 34:1	360 720 227.0278 20.00
125 L Waters QTOF Premier; [M-H]; PS 40:1	Spec. List) Waters QTOF Pr
126 L Waters QTOF Premier; [M-H]; PS 34:1	
127 L Waters QTOF Premier; [M+N]+; PS 36:1 128 L Waters QTOF Premier; [M+Na]+; MGDG 34:6	281.2436
129 L Waters QTOF Premier; [M-H]-; PG 34:1	
130 L Waters QTOF Premier; [M-H]-; SQDG 34:2	
A Structures I Structures I	747.5078
custompc+hpos.msp; custompc+hapos.msp; hilic-urine; lipidblast-heg; pc-ac-heg.msp; pc-form-heg.msp; lipidblast-pos; 235370 total spectra	
1000-1	391.22513 465.26191
100-	50-
10-	
	100- 255 2222
# Library Score Dot Product Prob. (%) Rev-Dot Name	A Waters QTOF Premier; [M-H]-; PGI Head to Tail MF=190 RMF=696  ▼PG 34:1; [M-H]-; GPGro(16:0/18:1
1 lipidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPGro(16:0/18:1(11E))	Difference Head to Tail Side by Side Subtraction / 190 696R 4.63P
2 lipidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPGro(16:0/18:1(11Z))	Name: PG 34:1; [M-H]; GPGro(16:0/18:1(11E))
3 lipidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPG-ra(16:0718:11[13∠]) 4 lipidblast-neg 190 696 4.62 779 PC 34:1; [M-H]-; GPG-ra(16:0719:11[72])	100- 203.23226 <u>MW:</u> 747 ID#: 113470 DB: lipidblast-neg
5 linidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPGro(16:0/16:1(172)]	Blargest peaks:
6 lipidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPGro(16:0/18:1(6Z))	50- 255.23226 999.00   281.24790 999.00   391.2 391.22513 492.27247 200.00   491.27255 200.00   509.21
7 lipidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPGro(16:0/18:1(7Z))	43.27247 200.001 431.27733 200.001 303.27 8 m/z Values and Intensities:
8 lipidblast-neg 190 696 4.63 779 PG 34:1; [M-H]-; GPGro(16:0/18:1(9E))	260 390 520 650 255.23226 999.00 sn1 FA
Names Structures / Hit List	I (lipidblast-neg) PG 34:1; [M-H]; GPGro(16:0/18:1(11E)
Lib Search Names Compare Librarian MSMS	
	Peptide Peptide //.

Name: Waters QTOF Premier; [M-H]-; SQDG 34:2 <u>MW:</u> 817 <u>ID#:</u> 130 <u>DB:</u> Spec. List <u>Comment:</u> SQDG(16:0/18:2); [M-H]-; Prec. m/z: 817.5167; Global characterization of the photosynthetic glycerolipids from a marine diatom Stephanodiscus sp. by ultra performance liquid chromatography coupled with electrospray ionization-quadrupole-time of flight mass spectrometry ;http://dx.doi.org/10.1016/j.aca.2010.01.026 8 largest peaks



Name: Waters Synapt HDMS; [M+Na]+; PC 32:0 <u>MW:</u> 756 <u>ID#:</u> 131 <u>DB:</u> Spec. List <u>Comment:</u> PC 32:0; [M+Na]+; Prec. m/z: 756.5524; PC(16:0/16:0) 756.5524; Direct Tissue Imaging and Characterization of Phospholipids Using MALDI SYNAPT HDMS System; Emmanuelle Claude, Marten Snel, Therese McKenna, James Langridge; APNT10011751 <u>9 largest peaks:</u>

🗲 NIST MS Search 2.0 - [Peptide, Presearch Default - 208 spectra]	
Eile Search View Tools Options Window Help	
🚳 🍉 🗃 🚎 1. Waters Synapt HDMS; [M+Na]+; PC 🖅 🛞 🖭 😥 🍭 🚳	
# Src. Name	756 FEO/ Name: Waters Synapt HDMS; [M+Na]+; PC 32:0
121 L Waters QTOF Premier; [M-H] ; NA	146.9827 MW: 756 ID#: 131 DB: Spec. List Comment: PC 32:0; [M+Na]+: Prec. m/z: 756.5524; PC(16:0/16:0) 756.5524
122 L Waters QT UF Premier; [M-H]-; PE 34:0 123 L Waters QT OF Premier; [M-H]-; PE 34:1	9 largest peaks:
124 L Waters QTOF Premier; [M-H]; PG 34:1	400 756.5524 359.00   146.5827 660.00   637.4801 510.00   573.4851 500
125 L Waters QTOF Premier; [M-H]; PS 40:1 126 L Waters QTOF Premier; [M-H]; PS 34:1	. List) Waters Synapt H
127 L Waters QTOF Premier; [M-H]; PS 36:1	Text of Search Spectrum / Plot of Search Spectrum / Plot/Text of Spec List /
128 L Waters QTOF Premier; [M+Na]+; MGDG 34:6	756.5524
125 L Waters QTOF Premier; [M-H]-; PG 34:1	
131 L Waters Synapt HDMS; [M+Na]+; PC 32:0	146.9827
50	575,4651 057,4001
Names Structures Spec List	86.0943 184.0753
custompc+hpos.msp; custompc+napos.msp; hilic-urine; lipidblast-neg; pc-ac-neg.msp; pc-form-neg.msp; lipidblast-pos; 235370	4/8.3405
	441.23832
10-	573 48586
	697.47840
	120 180 240 300 360 420 480 540 600 660 720
📕 📕 Library Score Dot Product Prob. (%) Rev-Dot Name	ters Synapt HDMS; [M+Na]+; I Head to Tail MF=333 RMF=635 ▼PC 32:0; [M+Na]+; GPCho(16:0/1
1 lipidblast-pos 393 635 12.1 974 PC 32:0; (M+Na)+; GPCho(16:0/16:0)	rence A Head to Tail A side by side A subtraction 393 635R 12.1P
2 ipidplast-pos 363 611 4.04 946 PC 32.0; [M+Na]+; GPCho[6.0726.0]	697.47840 Name: PC 32:0; [M+Na]+; GPCho(16:0/16:0)
4 lipidblast-pos 369 611 4.04 946 PC 32:0; [M+Na]+; GPCho(8:0/24:0)	Comment: Parent=756.55190 Mz_exact=756.5519
5 lipidblast-pos 369 611 4.04 946 PC 32:0; [M+Na]+; GPCho(9:0/23:0)	5/3.48586 5 largest peaks:
6 lipidblast-pos 369 611 4.04 946 PC 32:0; [M+Na]+; GPCho(10:0/22:0)	57.47640 555.00 573.46566 600.00 ( 441.2
7 ipidplast-pos 369 611 4.04 946 PC 32:0; [M+Na]+; GPCho[1]:0/21:0] 8 ipidplast-pos 369 611 4.04 946 PC 32:0; [M+Na]+; GPCho(12:0/20:0)	441.23832 40.00 [M+Na]-59-sn1    [M+Na]-59-st
	150 300 450 500 750 300.3102 20.00 [m+na]stit=20 it [m+na]stit.▼
Names Structures / Hit List Plot/	Text of Hit / Plot of Hit /
Lib. Search Other Search Names Compare Librarian MSMS	
	Peptide Peptide

Name: Waters Synapt HDMS w MALDI; [M-H]-; PE 34:1 <u>MW:</u> 716 <u>ID#:</u> 148 <u>DB:</u> Text File <u>Comment:</u> PE 34:1; [M-H]-; Prec. m/z: 716.65; PE(16:0/18:1) ; Structural mass spectrometry analysis of lipid changes in a Drosophila epilepsy model brain ; Michal Kliman, Niranjana Vijayakrishnan, Lily Wang, John T. Tapp, Kendal Broadie and John A. McLean; DOI:10.1039/B927494D 10 largest peaks:



# LipidBlast evaluation using Agilent 6530 QTOF-MS/MS spectra

Curator: Tobias Kind Spectra: John K. Meissen and Brian DeFelice PI: Oliver Fiehn Date: Sept 2011

Spectra: LipidBlast Molecule Images: Avanti Polar Lipids or Sigma

RAW MGF or MSP MS/MS spectra can be found under <a href="http://fiehnlab.ucdavis.edu/projects/LipidBlast">http://fiehnlab.ucdavis.edu/projects/LipidBlast</a>

# **NIST MSPepSearch**

Software for batch interpretation of tandem mass spectra Independent from LipidBlast. Can be freely obtained from <u>http://peptide.nist.gov/</u>

Library search settings for accurate mass spectrometry: Precursor accuracy +/-0.008 Da; product ion accuracy +/-0.008 Da;

Library Search Options	MSPepSearch
Library Search Options       Image: Search MS/MS         Search MS/MS       Libraries       Automation       Limits       Constraints         MS/MS       Search m/z       Tolerance         Precursor ±       0.008         Product ions ±       0.008         Ignore       Precursor       Peptide       MS/MS       Options         Use alt.       peak matching	Input   Input     Input
	Min. match factor (MF) to output (0-999)       Image: Include precursor m/z in the output         Min. peak intensity (1-999)       Image: Include Hit-Unknown precursor m/z difference         Min. peak intensity (1-999)       Image: Im
OK Cancel Help	Done Run Close

MSPepSearch	
Input (.MGF Mascot generic or .MSP NIST MS-type)	File O Folder Open Remove selected
Z:\Lipid MGF\Pos\A852758_PlasmenyIPE_18_18d9_0005.m	ngf
Output directory path:	Select
Z:\Lipid MGF\Pos	
MS/MS libraries to search	Select Remove selected
Z:\lipidblast\LipidBlast-pos	
Z:\lipidblast\CustomPC+Napos.msp Z:\lipidblast\CustomPC+Napos.msp	
	Reset to defaults
- Ontions	
Presearch mode O Fast O Standard	m/z limits Min. 0 Max. 2000 -1 = infinity
Search tolerance settings	
Precursor ion tolerance, m/z units .008	re peaks around precursor Fragment peak m/z tolerance .008
Min. match factor (MF) to output (0-999)	<ul> <li>Include precursor m/z in the output</li> <li>Include Hit-Unknown precursor m/z difference</li> </ul>
Min. peak intensity (1-999)	Output the input spectrum number     Calculate rev-dot
Max number of output hits	Use number of replicates
Show spectra without matches	I Q-TDF ▼ Set program priority above normal
	Load libraries in memory
Done	Run Close

1) Standard Cardiolipin - ESI(-) CL(1'-[14:0/14:0],3'-[14:0/14:0]); C65H132N2O17P2



ID in LipidBlast – single hit OK <u>Name:</u> CL 56:0; [M-H]-; CL(14:0/14:0/14:0/14:0) <u>MW:</u> 1239 <u>ID#:</u> 22921 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=1239.83921 Mz\_exact=1239.83921 ; CL 56:0; [M-H]-; CL(14:0/14:0/14:0/14:0); C65H126O17P2 <u>Source:</u> A710332\_CL\_14\_14\_14\_14\_0003.d, MS/MS of 1239.839965820 1+ at 1.4901 Automatic LipidBlast explanation of fragments: Cardiolipin CL(1'-[14:0/14:0],3'-[14:0/14:0]); C65H132N2O17P2



227.20098	100.00	sn1 FA    sn2 FA    sn3 FA    sn4 FA
363.19353	300.00	sn2+C3H6PO4 (+137.00)    sn4+C3H6PO4 (+137.00)
381.20409	200.00	sn2+C3H6PO4+H2O    sn4+C3H6PO4+H2O
591.40233	999.00	sn1+sn2+C3H6PO4 (+137.00)    sn3+sn4+C3H6PO4 (+137.00)
647.42854	100.00	sn1+sn2+C6H10O5P (+193.026)    sn3+sn4+C6H10O5P (+193.026)
727.39488	300.00	sn1+sn2+C6H11P2O8 (+272.9929)    sn3+sn4+C6H11P2O8 (+272.9929)
1011.63041	50.00	[M-H]-sn1

2) Standard Cardiolipin ESI(-) CL(1'-[18:1/18:1(9Z)],3'-[18:1/18:1(9Z)])



#### ID in LipidBlast – First HIT

<u>Name:</u> CL 72:4; [M-H]-; CL(18:1/18:1/18:1/18:1) <u>MW:</u> 1456 <u>ID#:</u> 33228 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=1456.02701 Mz\_exact=1456.02701 ; CL 72:4; [M-H]-; CL(18:1/18:1/18:1/18:1); C81H150O17P2 <u>7 largest peaks:</u>



<u>Name:</u> plasmenyl-PE 36:1; [M-H]-; PE(P-18:0/18:1(11E)) <u>MW:</u> 728 <u>ID#:</u> 123312 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=728.55944 Mz\_exact=728.55944 ; plasmenyl-PE 36:1; [M-H]-; PE(P-18:0/18:1(11E)); C41H80NO7P 4) Standard Phosphatidic Acid: PA(16:0/18:1(9Z)) - ESI(-)



### ID in LipidBlast – First HIT

<u>Name:</u> PA 34:1; [M-H]-; GPA(16:0/18:1(11E)) – equivalent with GPA(16:0/18:1(9Z)) <u>MW:</u> 673 <u>ID#:</u> 102518 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=673.48083 Mz\_exact=673.48083 ; PA 34:1; [M-H]-; GPA(16:0/18:1(11E)); C37H71O8P



<u>Name:</u> PE 37:4; [M-H]-; GPEtn(17:0/20:4(5E,8E,11E,14E)) - equivalent with GPEtn(17:0/20:4(5Z,8Z,11Z,14Z)) <u>MW:</u> 752 <u>ID#:</u> 108245 <u>DB:</u> lipidblast-neg

Comment: Parent=752.52306 Mz\_exact=752.52306; PE 37:4; [M-H]-; GPEtn(17:0/20:4(5E,8E,11E,14E)); C42H76NO8P



<u>Name:</u> PE 31:1; [M-H]-; GPEtn(14:1(9Z)/17:0) <u>MW:</u> 674 <u>ID#:</u> 107768 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=674.47607 Mz\_exact=674.47607 ; PE 31:1; [M-H]-; GPEtn(14:1(9Z)/17:0); C36H70NO8P



<u>Name:</u> PE 36:1; [M-H]-; GPEtn(18:0/18:1(11E)) <u>MW:</u> 744 <u>ID#:</u> 108438 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=744.55431 Mz\_exact=744.55431 ; PE 36:1; [M-H]-; GPEtn(18:0/18:1(11E)); C41H80NO8P



ID in LipidBlast – First HIT <u>Name:</u> lysoPE 18:1; [M-H]-; PE(18:1(11E)/0:0) <u>MW:</u> 478 <u>ID#:</u> 95782 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=478.29336 Mz\_exact=478.29336 ; lysoPE 18:1; [M-H]-; PE(18:1(11E)/0:0); C23H46NO7P

# 9) Standard phosphatidylglycerol – ESI(-) PG(17:0/14:1(9Z))







Name: PG 31:1; [M-H]-; GPGro(14:1(9Z)/17:0) MW: 705 ID#: 113244 DB: lipidblast-neg Comment: Parent=705.47063 Mz\_exact=705.47063 ; PG 31:1; [M-H]-; GPGro(14:1(9Z)/17:0); C37H71O10P 11) Standard phosphatidylinositol – ESI(-) PI(17:0/14:1(9Z))



ID in LipidBlast – First HIT <u>Name:</u> PI 31:1; [M-H]-; GPIns(14:1(9Z)/17:0) <u>MW:</u> 793 <u>ID#:</u> 118720 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=793.48669 Mz\_exact=793.48669 ; PI 31:1; [M-H]-; GPIns(14:1(9Z)/17:0); C40H75O13P



<u>Name:</u> PI 37:4; [M-H]-; GPIns(17:0/20:4(5E,8E,11E,14E)) <u>MW:</u> 871 <u>ID#:</u> 119197 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=871.53368 Mz\_exact=871.53368 ; PI 37:4; [M-H]-; GPIns(17:0/20:4(5E,8E,11E,14E)); C46H81O13P



<u>Name:</u> PS 28:0; [M-H]-; GPSer(14:0/14:0) <u>MW:</u> 678 <u>ID#:</u> 124337 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=678.43462 Mz\_exact=678.43462 ; PS 28:0; [M-H]-; GPSer(14:0/14:0); C34H66NO10P



<u>Name:</u> PS 34:1; [M-H]-; GPSer(16:0/18:1(11E)) equivalent with GPSer(16:0/18:1(9Z))

MW: 760 ID#: 124644 DB: lipidblast-neg

Comment: Parent=760.51287 Mz\_exact=760.51287 ; PS 34:1; [M-H]-; GPSer(16:0/18:1(11E)); C40H76NO10P



# ID in LipidBlast – First HIT <u>Name:</u> PS 31:1; [M-H]-; GPSer(17:0/14:1(9Z)) <u>MW:</u> 718 <u>ID#:</u> 124856 <u>DB:</u> lipidblast-neg <u>Comment:</u> Parent=718.46593 Mz\_exact=718.46593 ; PS 31:1; [M-H]-; GPSer(17:0/14:1(9Z)); C37H70NO10P



Name: PS 37:4; [M-H]-; GPSer(17:0/20:4(5E,8E,11E,14E)) equivalent to GPSer(17:0/20:4(5Z,8Z,11Z,14Z)) <u>MW:</u> 796 <u>ID#</u>: 124895 <u>DB</u>: lipidblast-neg <u>Comment:</u> Parent=796.51287 Mz\_exact=796.51287 ; PS 37:4; [M-H]-; GPSer(17:0/20:4(5E,8E,11E,14E)); C43H76NO10P

# 17) Standards – ESI(+)



Cardiolipin CL(1'-[14:0/14:0],3'-[14:0/14:0]); C65H132N2O17P2

double negative charge, usually measured in negative mode.



Cardiolipin ESI(-) CL(1'-[18:1/18:1(9Z)],3'-[18:1/18:1(9Z)])

double negative charge, usually measured in negative mode.

18) Standards ESI(+) plasmenylphosphatidylethanolamine; GPEtn(18:0p/18:1(9Z))



# ID in LipidBlast – First HIT – issue with batch version

<u>Name:</u> plasmenyl-PE 36:1; [M+H]+; PE(P-18:0/18:1(11E)) equivalent to PE(P-18:0/18:1(11Z))

# MW: 730 ID#: 59466 DB: lipidblast-pos

Comment: Parent=730.57508 Mz\_exact=730.57508 ; plasmenyl-PE 36:1; [M+H]+; PE(P-18:0/18:1(11E)); C41H80NO7P



ID in LipidBlast – First HIT <u>Name:</u> lysoPC 14:0; [M+H]+; PC(14:0/0:0) <u>MW:</u> 468 <u>ID#:</u> 9185 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=468.30900 Mz\_exact=468.30900 ; lysoPC 14:0; [M+H]+; PC(14:0/0:0); C22H46NO7P <u>4 largest peaks:</u> 20) Standards ESI(+); Phosphatidylcholine; PC(17:0/14:1(9Z))





ID in LipidBlast – First HIT <u>Name:</u> PC 31:1; [M+H]+; GPCho(14:1(9Z)/17:0) <u>MW:</u> 718 <u>ID#:</u> 1382 <u>DB:</u> custompc+hpos.msp <u>Comment:</u> Parent=718.53870 Mz\_exact=718.53870; PC 31:1; [M+H]+; GPCho(14:1(9Z)/17:0); C39H76NO8P

21) Standards ESI(+); phosphatidylethanolamine; PE(17:0/20:4(5Z,8Z,11Z,14Z))



ID in LipidBlast – First HIT <u>Name:</u> PE 37:4; [M+H]+; GPEtn(17:0/20:4(5E,8E,11E,14E)) <u>MW:</u> 754 <u>ID#:</u> 49431 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=754.53870 Mz\_exact=754.53870; PE 37:4; [M+H]+; GPEtn(17:0/20:4(5E,8E,11E,14E)); C42H76NO8P





#### Correct ID in LipidBlast - not first HIT - multiple probabilities (few fragments in pos mode)

<u>Name:</u> PE 31:1; [M+H]+; GPEtn(14:1(9Z)/17:0) <u>MW:</u> 676 <u>ID#:</u> 48954 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=676.49171 Mz\_exact=676.49171 ; PE 31:1; [M+H]+; GPEtn(14:1(9Z)/17:0); C36H70NO8P <u>8 largest peaks:</u>



<u>Name:</u> PE 36:1; [M+H]+; GPEtn(18:0/18:1(11E)) <u>MW:</u> 746 <u>ID#</u>: 49624 <u>DB</u>: lipidblast-pos <u>Comment:</u> Parent=746.56995 Mz\_exact=746.56995 ; PE 36:1; [M+H]+; GPEtn(18:0/18:1(11E)); C41H80NO8P 24) Standard Lyso-phosphatidylethanolamine – ESI(+) PE(18:1(9Z)/0:0



#### ID in LipidBlast – First HIT

<u>Name:</u> lysoPE 18:1; [M+H]+; PE(18:1(11E)/0:0) <u>MW:</u> 480 <u>ID#:</u> 9360 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=480.30900 Mz\_exact=480.30900 ; lysoPE 18:1; [M+H]+; PE(18:1(11E)/0:0); C23H46NO7P <u>5 largest peaks:</u>



<u>Name:</u> PS 28:0; [M+H]+; GPSer(14:0/14:0) <u>MW:</u> 680 <u>ID#:</u> 60713 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=680.45026 Mz\_exact=680.45026 ; PS 28:0; [M+H]+; GPSer(14:0/14:0); C34H66NO10P <u>4 largest peaks:</u>



<u>Name:</u> PS 34:1; [M+H]+; GPSer(16:0/18:1(11E)) <u>MW:</u> 762 <u>ID#:</u> 61020 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=762.52851 Mz\_exact=762.52851 ; PS 34:1; [M+H]+; GPSer(16:0/18:1(11E)); C40H76NO10P



<u>Name:</u> PS 31:1; [M+H]+; GPSer(14:1(9Z)/17:0) <u>MW:</u> 720 <u>ID#:</u> 60794 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=720.48157 Mz\_exact=720.48157 ; PS 31:1; [M+H]+; GPSer(14:1(9Z)/17:0); C37H70NO10P


## ID in LipidBlast – First HIT

<u>Name:</u> PS 37:4; [M+H]+; GPSer(17:0/20:4(5E,8E,11E,14E)) <u>MW:</u> 798 <u>ID#:</u> 61271 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=798.52851 Mz\_exact=798.52851 ; PS 37:4; [M+H]+; GPSer(17:0/20:4(5E,8E,11E,14E)); C43H76NO10P <u>6 largest peaks:</u>



<u>Name:</u> SM 36:2; [M]+; SM(d18:1(4E)/18:1(9Z)) <u>MW:</u> 729 <u>ID#:</u> 70192 <u>DB:</u> lipidblast-pos <u>Comment:</u> Parent=729.59106 Mz\_exact=729.59106 ; SM 36:2; [M]+; SM(d18:1(4E)/18:1(9Z)); C41H82N2O6P <u>4 largest peaks:</u>

## Supplement Literature Collection for LipidBlast MS/MS library;

http://fiehnlab.ucdavis.edu/projects/LipidBlast

For questions please contact the curator: Dr. Tobias Kind (Date: May 2010) <u>http://fiehnlab.ucdavis.edu/staff/kind/</u>

The development of the MS/MS LipidBlast libraries is based on own experimental data and more than ~300 external literature references. Not all references can be included into the publication due to publishing limits and therefore are given here. References from the paper itself may not be referenced here.

Phospholipids in general [1-45]

Phosphatidylcholine (PC) [46-49]

Phosphatidylethanolamine (PE) [50-52]

Phosphatidyldiacylglycerol (PG) [53]

<u>Phosphatidylinositol (PI)</u> [54-57]

Phosphatidic Acid (PA) [58]

Plasmalogens and lysophospholipids [59, 60]

**Fatty Acids and Acyls** [61-65]

Ceramides (Cer) [66-74]

Cardiolipins (CL) [75-81]

MAG-DAG-TAG Mono-Di-Triacylglycerides
[82-96]

<u>Gangliosides + Sulfatides</u> [97-103]

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<u>Historic</u> [153-155]

Biocrates + ABI Kit [156]

Chlorinated Lipids
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Odd Number of carbons in fatty acids [158-160]

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Mass Spectrometer Types [261, 262]

MALDI, FAB and ion mobility and others like DESI, FI, FD, SIMS [58, 263-269]

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End of supplement