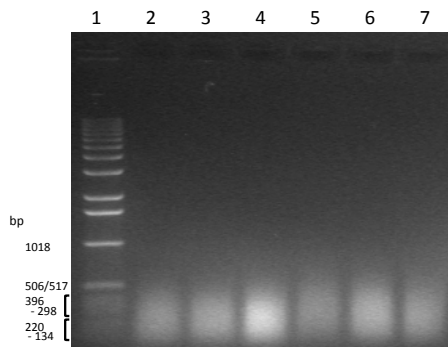


Supplementary Methods

Splinkerette with acoustic shearing for preparation of transposon insertion site libraries for Illumina sequencing

Genomic DNA was isolated from tumor samples by overnight digestion in lysis buffer (0.1 M Tris pH 8.5, 0.2 M NaCl, 5 mM EDTA, 0.2% SDS, 0.1 mg/ml Proteinase K) at 55°C, followed by 2-propanol precipitation, 70% ethanol wash, and resuspension overnight in TE buffer (10mM Tris, 1 mM EDTA, pH 7.4) at 55°C.

To shear DNA into fragments of 200 – 400 bp size, 3 µg of DNA in a volume of 120 µl TE were sonicated in a Covaris E210 acoustic focusing instrument (parameter settings: Time: 600 sec, Duty Cycle: 5%, Intensity: 2, Cycles per burst: 50). DNA was purified with a Qiaquick spin column (QIAGEN) and eluted in a volume of 30 µl EB buffer. For quality control, 5 µl of the eluted DNA was analyzed on a 0.8% agarose gel.



Example of sheared DNA separated on 0.8% agarose gel

Lane 1: 1 kb ladder, Invitrogen

Lane 2-7: DNA sheared with Covaris E210 sonicator

DNA Fragment End repair

14.5 µl Qiaquick purified genomic DNA, sheared by Covaris sonication

2 µl 10x T4 DNA ligase buffer w/ 10 mM ATP (NEB)

1 µl dNTP (10 mM)

1 µl T4 DNA Polymerase

1 µl Klenow DNA Polymerase

0.5 µl T4 Polynucleotid kinase

20 µl

30 min incubation at RT

DNA was purified with a Qiaquick spin column and eluted in 42 µl EB.

A-tailing of DNA fragments

42 µl Qiaquick purified end-repaired DNA
1 µl dATP (10 mM)
5 µl 10x PCR buffer ThermoPol (NEB)
2 µl Taq

50 µl

15 min incubation at 72°C

The entire reaction was EtOH precipitated by using 2 µg Glycogen as carrier and resuspended in 7.5 µl H₂O.

Preparing Splinkerette adaptor

(final adaptor conc. 25 µM)

Mix:

25 µl Adaptor oligo "plus" strand (100 µM)
25 µl Adaptor oligo "minus" strand (100 µM)
20 µl NEB restriction buffer 2
30 µl H₂O

Boil for 2 min, let cool down at RT. Store at -20°C.

Ligation of Splinkerette adaptor

7.5 µl A-tailed DNA fragments
0.5 µl Adaptor SPLK-T (25 µM)
1 µl 10x T4 Ligase buffer
1 µl T4 Ligase
--
10 µl

Ligation was performed o/n at 16°C

The ligation was purified with a Qiaquick spin column and eluted in 40 µl EB.

First Splinkerette PCR

2.5 µl Qiaquick purified ligation product

5 µl 5x Phusion HF buffer

0.5 µl dNTP (10 mM)

0.5 µl Sp-1 (10µM)

0.5 µl PB5-1 (10µM)

0.25 µl Phusion enzyme

15.75 µl H₂O

25µl

3 min 95°C

35 cycles: 15 sec 95°C, 30 sec 65°C, 30 sec 72°C

Second Splinkerette PCR

A 1:200 dilution of the first PCR was prepared with H₂O

2.5µl 1:200 dilution of first PCR

5 µl 5x Phusion HF buffer

0.5 µl dNTP (10 mM)

0.5 µl P7-Sp-2 (10µM)

0.5 µl P5-XX-PB5-2 (10µM) (XX = barcode number)

0.25 µl Phusion enzyme

15.75 µl H₂O

25µl

3 min 95°C

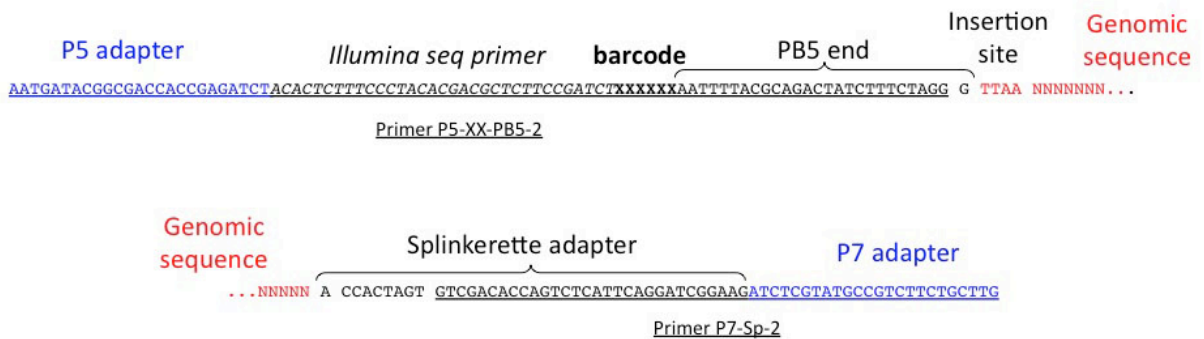
35 cycles: 15 sec 95°C, 30 sec 60°C, 30 sec 72°C

12.5 µl of the PCR product was separated on a 0.8% agarose gel

PCR product was cut out in a size range of 200 to 400 bp size, purified with a Qiaquick spin column and eluted in 30 µl EB.

DNA concentration was determined with a Nanodrop device. To reach a final concentration of the pooled sample of >30 ng/μl in 20 μl H₂O, appropriate amounts of all samples were pooled together, EtOH precipitated, and resuspended in 20 μl H₂O, and submitted to the sequencing facility at Mount Sinai School of Medicine. Samples were mixed with 50% PhiX Genomic DNA library (Illumina) to increase cluster diversity on the Illumina chip. Sample pools were sequenced on a single lane of an Illumina HiSeq 2000 Instrument as single unpaired reads with a length of 100 bases.

Structure of final Splinkerette PCR product



Structure of PB5 insertion site library is shown, PB3 library has same structure, except PB3 end part (see Oligonucleotide Sequences for details).

Oligonucleotide Sequences

Oligonucleotide sequences			
Purpose	Name	Sequence 5' - 3'	Description
Splinkerette linker	SpkTp	CGAAGAGTAACCGTTGCTAGGAGAGACCGTGGCTGAATGAGACTGGTGTGACACTAGTGG* ^T	Splinkerette T-overhang linker, plus strand (* = phosphorothioate bond)
	SpkTm	(5' phospho) CCACATAGTGTGCACACCAGTCTCAATTTTTTTTTTCAAAAAA	Splinkerette T-overhang linker, minus strand
First round Splinkerette PCR	Sp-1	GAGTAACCGTTGCTAGGAGAG	Primer on Splinkerette linker
	PB3-1	CGCATGATTATCTTTAACGTACGTAC	Primer on PB3 end
	PB5-1	CAAGAATGCATGCGTCAATTTACGC	Primer on PB5 end
Second round Splinkerette PCR	P7-Sp-2	CAAGCAGAAGACGGCATAACGAGATCTCCGATCTCTGAATGAGACTGGTGTGAC	Illumina P7 adapter + primer on Splinkerette linker
	P5-01-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTATCAGC AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 01 + primer on PB5 end
	P5-02-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCGATGT AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 02 + primer on PB5 end
	P5-03-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTTAGGC AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 03 + primer on PB5 end
	P5-04-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTGACCA AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 04 + primer on PB5 end
	P5-05-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGACAT AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 05 + primer on PB5 end
	P5-06-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCAGATC AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 06 + primer on PB5 end
	P5-07-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTACTTGA AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 07 + primer on PB5 end
	P5-08-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGATCAG AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 08 + primer on PB5 end
	P5-09-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTAGCTT AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 09 + primer on PB5 end
	P5-10-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGGCTAG AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 10 + primer on PB5 end
	P5-11-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCTTGTA AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 11 + primer on PB5 end
	P5-12-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCGTGT AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 12 + primer on PB5 end
	P5-13-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCGATG AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 13 + primer on PB5 end
	P5-14-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTACATCG AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 14 + primer on PB5 end
	P5-15-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGCCTAA AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 15 + primer on PB5 end
	P5-16-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTGGTCA AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 16 + primer on PB5 end
	P5-17-PB5-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCACTGT AATTTTACGCAGACTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 17 + primer on PB5 end
	P5-01-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTATCAGC CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 01 + primer on PB3 end
	P5-02-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCGATGT CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 02 + primer on PB3 end
	P5-03-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTTAGGC CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 03 + primer on PB3 end
	P5-04-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTGACCA CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 04 + primer on PB3 end
	P5-05-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTACATGT CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 05 + primer on PB3 end
	P5-06-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGCCAAT CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 06 + primer on PB3 end
	P5-07-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCAGATC CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 07 + primer on PB3 end
	P5-08-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTACTTGA CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 08 + primer on PB3 end
	P5-09-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGATCAG CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 09 + primer on PB3 end
	P5-10-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGGCTAG CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 10 + primer on PB3 end
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	P5-12-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCGTGT CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 12 + primer on PB3 end
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	P5-14-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTGCCTAA CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 14 + primer on PB3 end
	P5-15-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTGGTCA CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 15 + primer on PB3 end
	P5-16-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTTAGCTT CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 16 + primer on PB3 end
P5-17-PB3-2	AATGATACGGCGACCACCGAGATCTACACTCTTCCCTACACGACGCTCTCCGATCTCACTGT CGTCACAATATGATTATCTTTCTAGG	Illumina P5 adapter + sequencing primer site + barcode 17 + primer on PB3 end	