

Supporting Information

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**Control of Selectivity in Palladium-Catalyzed Oxidative
Carbocyclization/Borylation of Allenynes****

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Experimental Section

General remarks

Unless otherwise noted, all reagents were used as received from the commercial suppliers. B₂pin₂ was commercially available from Combi-Blocks. Pd(OAc)₂ was obtained from Pressure Chemicals and used without further purification. Palladium-catalyzed cyclizations were performed without any efforts to exclude moisture. Dry solvents (Et₂O, THF) were obtained from a VAC Solvent Purifier. Reactions were monitored using thin-layer chromatography (SiO₂). TLC plates were visualized with UV light (254 nm) or KMnO₄ stain. Flash chromatography was carried out with 60Å (particle size 35-70 μm) normal flash silica gel. NMR spectra were recorded at 400 MHz (¹H) or 500 MHz (¹H) and at 100 MHz (¹³C) or 125 MHz (¹³C), respectively. Chemical shifts (δ) are reported in ppm, using the residual solvent peak in CDCl₃ (δ_H = 7.26 and δ_C = 77.0 ppm) as internal standard, and coupling constants (*J*) are given in Hz. HRMS were recorded using ESI-TOF techniques.

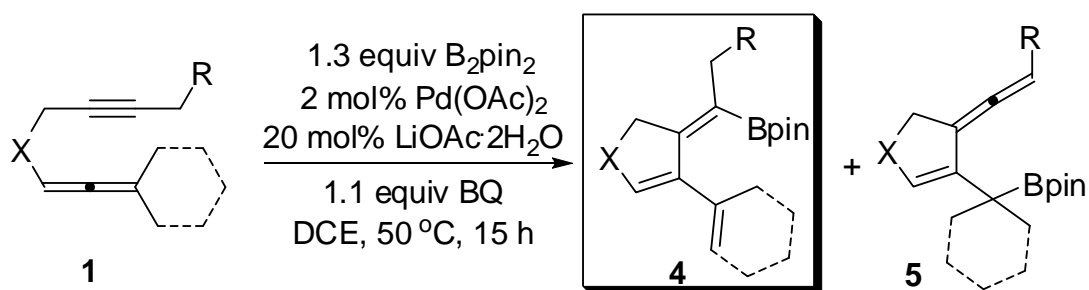
General procedure for preparation of starting materials

All bromoallenes were prepared according to the procedure published by Landor^{1a} with minor modifications.^{1b}

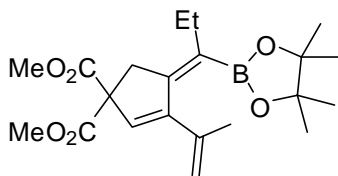
Dimethyl propargylmalonate is commercially available from Aldrich. Other alkyl substituted dimethyl propargylmalonates were prepared as described in the literature.²

Allenynes **1a-g** were prepared as previously described.³

General procedure for oxidative carbocyclization/borylation for the formation of trienes **4**



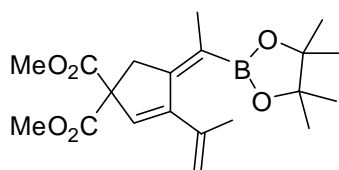
Representative procedure A for the synthesis of **4**. (*E*)-dimethyl 3-(prop-1-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-propylidene)cyclopent-2-ene-1,1-dicarboxylate (**4a**)



To a mixture of B₂pin₂ (33.1 mg, 0.13 mmol), BQ (12.2 mg, 0.11 mmol), Pd(OAc)₂ (0.5 mg, 0.002 mmol), and LiOAc·2H₂O (1.8 mg, 0.02 mmol) were added **1a** (26.0 mg, 0.10 mmol) and 0.5 mL of DCE at rt. The reaction was stirred at 50 °C for 15 h. After the reaction was complete as monitored by TLC, evaporation and column chromatography on silica gel (pentane/ethyl acetate = 10/1) afforded **4a** (27.9 mg,

73%) as a liquid; ^1H NMR (500 MHz, CDCl_3): δ 5.93 (s, 1H), 5.02-5.00 (m, 2H), 3.72 (s, 6H), 3.20 (s, 2H), 2.19 (q, $J = 7.5$ Hz, 2H), 1.95 (s, 3H), 1.26 (s, 12H), 1.02 (t, $J = 7.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 171.1, 151.1, 147.9, 139.8, 129.1, 116.6, 83.3, 63.0, 52.9, 37.5, 27.0, 25.2, 23.8, 13.4; HRMS (ESI): calc. for $\text{C}_{21}\text{H}_{31}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 413.2110; found: 413.2113.

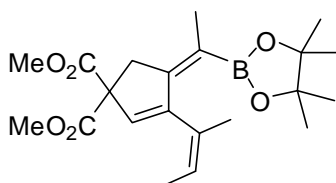
(E)-dimethyl 3-(prop-1-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)cyclopent-2-ene-1,1-dicarboxylate (**4b**).^{3b}



92% isolated yield, solid. ^1H NMR (400 MHz, CDCl_3): δ 5.91 (s, 1H), 5.01-4.96 (m, 2H), 3.72 (s, 6H), 3.16 (s, 2H), 1.95 (s, 3H), 1.78 (s, 3H), 1.24 (s, 12H).

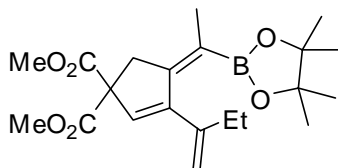
Compound **4c** and **4c'** were obtained as an inseparable mixture.

(E)-dimethyl 3-((*Z*)-but-2-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)cyclopent-2-ene-1,1-dicarboxylate (**Z-4c**)



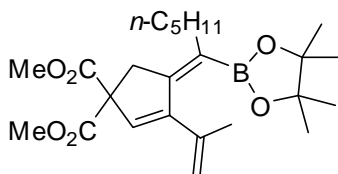
Liquid; ^1H NMR (500 MHz, CDCl_3): δ 5.81 (d, $J = 0.5$ Hz, 1H), 5.37 (qt, $J_1 = 6.5$ Hz, $J_2 = 1.5$ Hz, 1H), 3.71 (s, 6H), 3.14 (s, 2H), 1.84 (d, $J = 1.5$ Hz, 3H), 1.76 (s, 3H), 1.45 (dd, $J_1 = 6.5$ Hz, $J_2 = 1.5$ Hz, 3H), 1.24 (s, 12H); HRMS (ESI): calc. for $\text{C}_{21}\text{H}_{31}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 413.2110; found: 413.2116.

(E)-dimethyl 3-(but-1-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)ethylidene)cyclopent-2-ene-1,1-dicarboxylate (**4c'**)



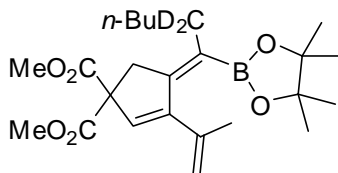
Liquid; ^1H NMR (500 MHz, CDCl_3): δ 5.89 (s, 1H), 4.990 (s, 1H), 4.989 (s, 1H), 3.71 (s, 6H), 3.16 (s, 2H), 2.26 (qt, $J_1 = 7.5$ Hz, $J_2 = 1.5$ Hz, 2H), 1.76 (s, 3H), 1.02 (t, $J = 7.5$ Hz, 3H); HRMS (ESI): calc. for $\text{C}_{21}\text{H}_{31}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 413.2110; found: 413.2116.

(E)-dimethyl 3-(prop-1-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-hexylidene)cyclopent-2-ene-1,1-dicarboxylate (**4d**)



81% isolated yield, liquid. ^1H NMR (400 MHz, CDCl_3): δ 5.92 (s, 1H), 5.01-4.99 (m, 2H), 3.72 (s, 6H), 3.19 (s, 2H), 2.18-2.11 (m, 2H), 1.95 (s, 3H), 1.42-1.35 (m, 2H), 1.34-1.26 (m, 4H), 1.25 (s, 12H), 0.88 (t, $J = 6.8$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 171.1, 151.1, 148.0, 139.8, 129.0, 116.5, 83.3, 63.0, 52.8, 37.7, 34.1, 32.0, 28.6, 25.2, 25.0, 23.8, 22.5, 14.0; HRMS (ESI): calc. for $\text{C}_{24}\text{H}_{37}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 455.2580; found: 455.2577.

$[D_2]$ -*(E)*-dimethyl 3-(prop-1-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl) hexylidene)cyclopent-2-ene-1,1-dicarboxylate ($[D_2]$ -**4d**)

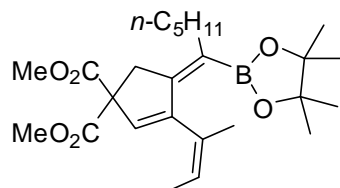


84% isolated yield (Scheme 3b), liquid. ^1H NMR (400 MHz, CDCl_3): δ 5.91 (s, 1H), 5.00-4.98 (m, 2H), 3.70 (s, 6H), 3.18 (s, 2H), 1.93 (t, $J = 1.2$ Hz, 3H), 1.40-1.34 (m, 2H), 1.33-1.25 (m, 4H), 1.23 (s, 12H), 0.87 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 171.0, 151.1, 148.0, 139.8, 129.0, 116.5, 83.2, 62.9, 52.8, 37.7, 31.9, 28.4,

25.2, 24.9, 23.7, 22.5, 13.9; HRMS (ESI): calc. for $C_{24}H_{35}BD_2NaO_6$ $[M+Na]^+$: 457.2705; found: 457.2706.

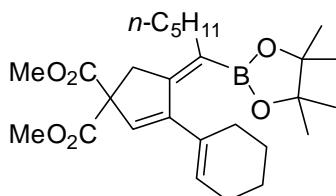
Compound **4e** and **4e'** were obtained as an inseparable mixture.

(*E*)-dimethyl 3-((*Z*)-but-2-en-2-yl)-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-hexylidene)cyclopent-2-ene-1,1-dicarboxylate (**Z-4e**)



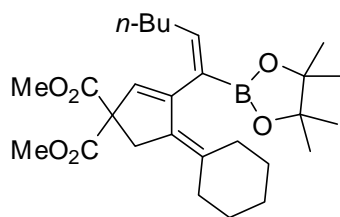
Liquid; ¹H NMR (500 MHz, CDCl₃): δ 5.82 (s, 1H), 5.37 (qq, $J_1 = 6.5$ Hz, $J_2 = 1.5$ Hz, 1H), 3.72 (s, 6H), 3.18 (s, 2H), 2.16-2.10 (m, 2H), 1.86 (t, $J = 6.5$ Hz, 3H), 1.45 (dq, $J_1 = 6.5$ Hz, $J_2 = 1.5$ Hz, 3H), 1.42-1.36 (m, 2H), 1.34-1.26 (m, 4H), 1.25 (s, 12H), 0.88 (t, $J = 6.8$ Hz, 3H); HRMS (ESI): calc. for $C_{25}H_{39}BNaO_6$ $[M+Na]^+$: 469.2736; found: 469.2735.

(*E*)-dimethyl 3-cyclohexenyl-4-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-hexylidene)cyclopent-2-ene-1,1-dicarboxylate (**4f**)



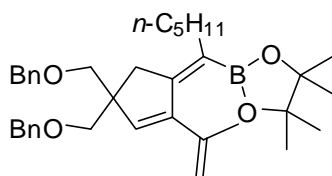
28% isolated yield, liquid. ¹H NMR (500 MHz, CDCl₃): δ 5.91 (s, 1H), 5.77-5.72 (m, 1H), 3.72 (s, 6H), 3.19 (s, 2H), 2.22-2.14 (m, 4H), 2.11-2.04 (m, 2H), 1.70-1.64 (m, 2H), 1.63-1.55 (m, 2H), 1.44-1.36 (m, 2H), 1.34-1.26 (m, 4H), 1.25 (s, 12H), 0.89 (t, $J = 7.0$ Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 171.2, 151.7, 147.6, 133.6, 128.0, 126.3, 82.9, 62.5, 52.8, 38.6, 34.3, 32.1, 28.8, 28.2, 25.3, 25.2, 22.5, 22.4, 21.8, 14.0; HRMS (ESI): calc. for $C_{27}H_{41}BNaO_6$ $[M+Na]^+$: 495.2893; found: 495.2899.

(*Z*)-Dimethyl 4-cyclohexylidene-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-hex-1-enyl)cyclopent-2-ene-1,1-dicarboxylate (**4f'**) (less polar)



20% isolated yield, liquid. ^1H NMR (500 MHz, CDCl_3): δ 5.75 (s, 1H), 5.28-5.22 (m, 1H), 3.73 (s, 6H), 3.10 (s, 2H), 2.21-2.16 (m, 2H), 2.15-2.08 (m, 2H), 2.06-1.97 (m, 2H), 1.68-1.61 (m, 2H), 1.60-1.56 (m, 2H), 1.38-1.32 (m, 2H), 1.31-1.22 (m, 4H), 1.13 (s, 12H), 0.88 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 171.4, 152.5, 143.7, 142.9, 125.9, 121.3, 82.8, 63.8, 52.7, 35.1, 31.8, 31.7, 29.5, 29.0, 27.4, 24.6, 22.6, 22.5, 22.1, 14.0; HRMS (ESI): calc. for $\text{C}_{27}\text{H}_{41}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 495.2893; found: 495.2902.

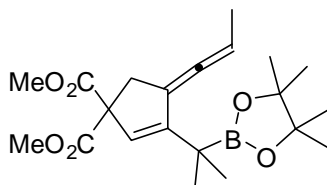
(E)-2-(1-(4,4-Bis(benzyloxymethyl)-2-(prop-1-en-2-yl)cyclopent-2-enylidene)hexyl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (**4g**)



57% isolated yield, liquid. ^1H NMR (500 MHz, CDCl_3): δ 7.35-7.24 (m, 10H), 5.88 (s, 1H), 4.99 (s, 1H), 4.97 (s, 1H), 4.53 (s, 4H), 3.47 (s, 4H), 2.49 (s, 2H), 2.14 (t, $J = 8.0$ Hz, 2H), 1.95 (s, 3H), 1.42-1.35 (m, 2H), 1.34-1.28 (m, 4H), 1.28 (s, 12H), 0.90 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 150.9, 149.4, 141.1, 138.9, 136.1, 128.2, 127.4, 127.3, 115.4, 83.1, 73.4, 73.2, 51.4, 37.6, 33.9, 32.1, 29.0, 25.2, 24.0, 22.6, 14.1; HRMS (ESI): calc. for $\text{C}_{36}\text{H}_{49}\text{BNaO}_4$ $[\text{M}+\text{Na}]^+$: 579.3622; found: 579.3630.

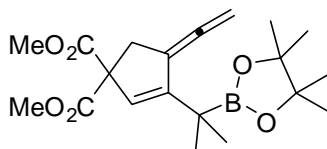
General procedure for oxidative carbocyclization/borylation for the formation of vinylallenes 5

Representative procedure B for the synthesis of **5**. Dimethyl 4-(prop-1-enylidene)-3-(2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)propan-2-yl)cyclopent-2-ene-1,1-dicarboxylate (**5a**)



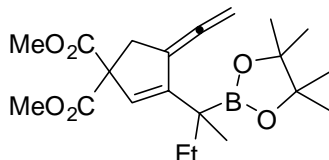
To a mixture of B₂pin₂ (66.2 mg, 0.26 mmol), BQ (24.0 mg, 0.22 mmol), Pd(OAc)₂ (1.0 mg, 0.004 mmol), and BF₃·Et₂O (6 μL, 0.04 mmol) were added **1a** (52.7 mg, 0.20 mmol) and 1.0 mL of THF at rt. The reaction was stirred at 50 °C for 20 h. After the reaction was complete as monitored by TLC, evaporation and column chromatography on silica gel (pentane/ethyl acetate = 10/1) afforded **5a** (59.6 mg, 77%) as a liquid; ¹H NMR (400 MHz, CDCl₃): δ 5.55 (d, *J* = 1.6 Hz, 1H), 5.34-5.23 (m, 1H), 3.716 (s, 3H), 3.715 (s, 3H), 3.19-3.17 (m, 2H), 1.68 (d, *J* = 7.2 Hz, 3H), 1.18 (s, 12H), 1.17 (s, 3H), 1.13 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 199.1, 171.5, 171.3, 153.8, 122.9, 107.1, 91.2, 83.1, 63.5, 52.7, 36.5, 25.0, 24.7, 24.5, 23.9, 23.8, 14.8; HRMS (ESI): calc. for C₂₁H₃₁BNaO₆ [M+Na]⁺: 413.2106; found: 413.2103.

Dimethyl 3-(2'-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)propan-2'-yl)-4-vinylidenecyclopent-2-ene-1,1-dicarboxylate (**5b**)



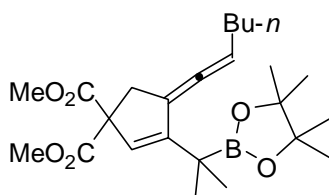
73% isolated yield, liquid. ¹H NMR (500 MHz, CDCl₃): δ 5.57 (t, *J* = 1.5 Hz, 1H), 5.02 (t, *J* = 4.0 Hz, 1H), 5.01 (t, *J* = 4.0 Hz, 1H), 3.74 (s, 6H), 3.25 (t, *J* = 4.0 Hz, 2H), 1.20 (s, 12H), 1.19 (s, 6H); ¹³C NMR (125 MHz, CDCl₃): δ 203.1, 171.2, 153.2, 123.0, 107.4, 83.2, 81.0, 63.7, 52.8, 36.4, 24.6, 23.9; HRMS (ESI): calc. for C₂₀H₂₉BNaO₆ [M+Na]⁺: 399.1949; found: 399.1952.

Dimethyl 3-(2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)butan-2-yl)-4-vinylidene-cyclopent-2-ene-1,1-dicarboxylate (5c)



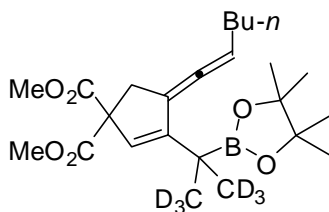
56% isolated yield, liquid. ^1H NMR (400 MHz, CDCl_3): δ 5.58 (s, 1H), 5.04-4.94 (m, 2H), 3.73 (s, 3H), 3.71 (s, 3H), 3.25 (dt, $J_1 = 16.0$ Hz, $J_2 = 4.0$ Hz, 1H), 3.18 (dt, $J_1 = 16.0$ Hz, $J_2 = 4.0$ Hz, 1H), 1.83-1.73 (m, 1H), 1.64-1.54 (m, 1H), 1.192 (s, 6H), 1.188 (s, 6H), 1.14 (s, 3H), 0.74 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 203.0, 171.3, 171.1, 151.1, 124.8, 107.2, 83.1, 80.8, 63.7, 52.7, 36.5, 28.1, 24.7, 24.6, 19.6, 8.6; HRMS (ESI): calc. for $\text{C}_{21}\text{H}_{31}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 413.2106; found: 413.2112.

Dimethyl 4-(hex-1-enylidene)-3-(2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)propan-2-yl)cyclopent-2-ene-1,1-dicarboxylate (5d)



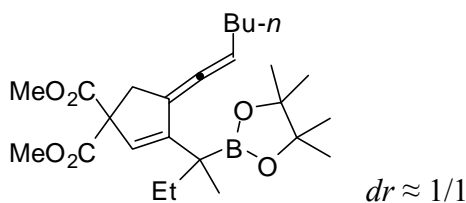
79% isolated yield, liquid. ^1H NMR (400 MHz, CDCl_3): δ 5.55 (d, $J = 1.6$ Hz, 1H), 5.32-5.25 (m, 1H), 3.71 (s, 6H), 3.21 (dd, $J_1 = 16.0$ Hz, $J_2 = 3.6$ Hz, 1H), 3.14 (dd, $J_1 = 16.0$ Hz, $J_2 = 3.6$ Hz, 1H), 2.09-2.01 (m, 2H), 1.42-1.28 (m, 4H), 1.18 (s, 9H), 1.17 (s, 6H), 1.13 (s, 3H), 0.88 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 198.0, 171.5, 171.3, 153.9, 122.9, 107.5, 96.6, 83.1, 63.5, 52.7, 36.8, 31.1, 29.2, 24.7, 24.5, 24.0, 23.8, 22.2, 13.8; HRMS (ESI): calc. for $\text{C}_{24}\text{H}_{37}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 455.2580; found: 455.2582.

[D₆]-Dimethyl 4-(hex-1-enylidene)-3-(2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)propan-2-yl)cyclopent-2-ene-1,1-dicarboxylate ([D₆]-5d)



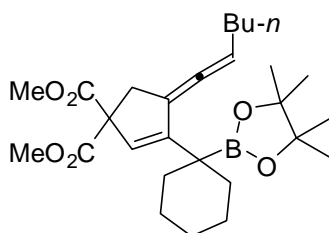
54% isolated yield (Scheme 3c), liquid. ^1H NMR (500 MHz, CDCl_3): δ 5.56 (s, 1H), 5.32-5.26 (m, 1H), 3.73 (s, 6H), 3.23 (dd, $J_1 = 16.0$ Hz, $J_2 = 3.5$ Hz, 1H), 3.15 (dd, $J_1 = 16.0$ Hz, $J_2 = 3.5$ Hz, 1H), 2.09-2.01 (m, 2H), 1.42-1.28 (m, 4H), 1.19 (s, 6H), 1.18 (s, 6H), 0.89 (t, $J = 7.5$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 198.0, 171.6, 171.3, 153.9, 122.8, 107.5, 96.6, 83.1, 63.5, 52.7, 36.9, 31.1, 29.3, 24.8, 24.5, 22.2, 13.9; HRMS (ESI): calc. for $\text{C}_{24}\text{H}_{31}\text{BD}_6\text{NaO}_6$ $[\text{M}+\text{Na}]^+$: 461.2956; found: 461.2951.

Dimethyl 4-(hex-1-enylidene)-3-(2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-butan-2-yl)cyclopent-2-ene-1,1-dicarboxylate (5e)



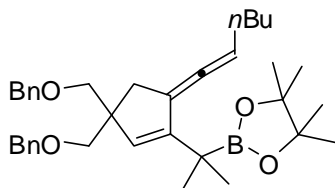
77% isolated yield, liquid. ^1H NMR (400 MHz, CDCl_3): δ 5.61-5.55 (m, 1H), 5.32-5.22 (m, 1H), 3.72-3.68 (m, 6H), 3.25-3.06 (m, 2H), 2.09-1.96 (m, 2H), 1.88-1.76 (m, 0.5H), 1.72-1.62 (m, 1H), 1.51-1.41 (m, 0.5H), 1.40-1.41 (m, 4H), 1.22-1.10 (m, 15H), 0.91-0.82 (m, 3H), 0.81 (t, $J = 7.2$ Hz, 1.5H), 0.66 (t, $J = 7.2$ Hz, 1.5H); ^{13}C NMR (100 MHz, CDCl_3): δ 198.0, 197.9, 171.5, 171.4, 171.3, 171.2, 152.2, 151.4, 125.0, 124.4, 107.5, 107.2, 96.4, 96.1, 83.1, 83.0, 63.5, 63.4, 52.6, 37.0, 36.9, 31.1, 30.8, 29.2, 29.1, 29.0, 27.4, 25.0, 24.9, 24.7, 24.6, 24.4, 22.2, 20.2, 19.4, 13.8, 9.5, 8.0; HRMS (ESI): calc. for $\text{C}_{25}\text{H}_{39}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 469.2732; found: 469.2728.

Dimethyl 4-(hex-1-enylidene)-3-(1-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-cyclohexyl)cyclopent-2-ene-1,1-dicarboxylate (5f)



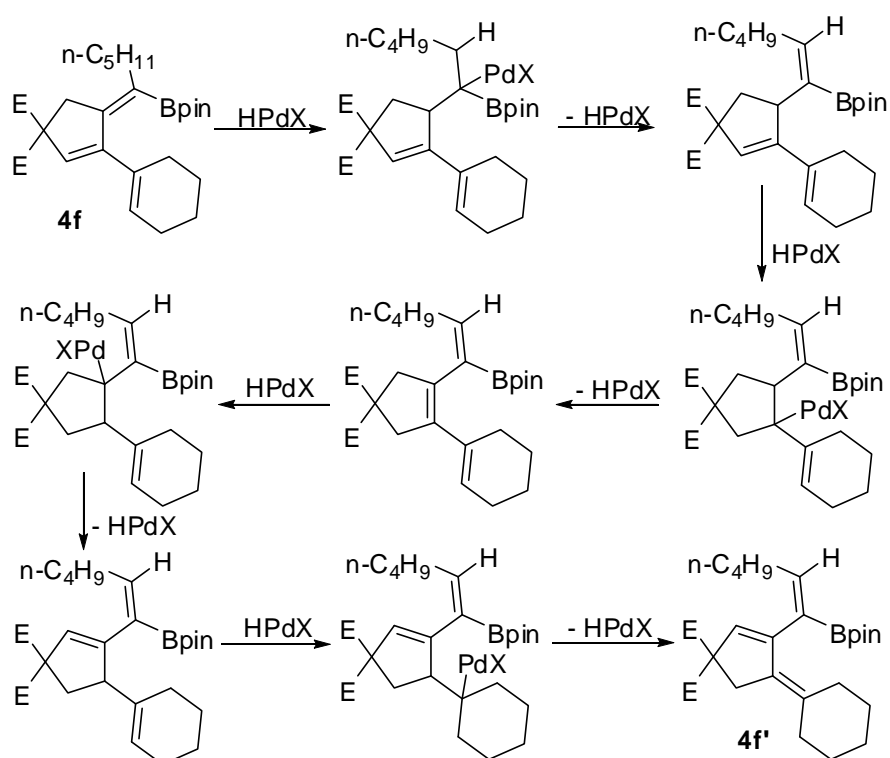
70% isolated yield, liquid. ^1H NMR (400 MHz, CDCl_3): δ 5.59 (d, $J = 1.2$ Hz, 1H), 5.30-5.22 (m, 1H), 3.714 (s, 3H), 3.705 (s, 3H), 3.16 (dd, $J_1 = 16.0$ Hz, $J_2 = 3.6$ Hz, 1H), 3.09 (dd, $J_1 = 16.0$ Hz, $J_2 = 3.6$ Hz, 1H), 2.19-2.01 (m, 4H), 1.70-1.61 (m, 1H), 1.60-1.45 (m, 3H), 1.44-1.25 (m, 8H), 1.19 (s, 6H), 1.18 (s, 6H), 0.89 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 198.5, 171.5, 171.2, 151.8, 124.2, 106.6, 95.8, 83.1, 63.3, 52.6, 37.2, 32.8, 32.6, 31.0, 29.1, 26.4, 25.0, 24.71, 24.66, 24.5, 24.4, 22.1, 13.9; HRMS (ESI): calc. for $\text{C}_{27}\text{H}_{41}\text{BNaO}_6$ $[\text{M}+\text{Na}]^+$: 495.2888; found: 495.2877.

2-(2-(3,3-Bis(benzyloxymethyl)-5-(hex-1-enylidene)cyclopent-1-enyl)propan-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (5g)



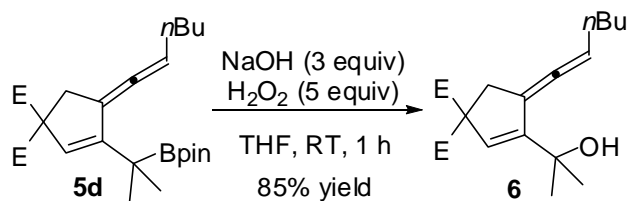
37% isolated yield, liquid. ^1H NMR (500 MHz, CDCl_3): δ 7.35-7.24 (m, 10H), 5.51 (d, $J = 1.5$ Hz, 1H), 5.25-5.20 (m, 1H), 4.55 (s, 4H), 3.53-3.42 (m, 4H), 2.54-2.42 (m, 2H), 2.09-2.01 (m, 2H), 1.44-1.30 (m, 4H), 1.21 (s, 6H), 1.20 (s, 6H), 1.18 (s, 3H), 1.14 (s, 3H), 0.91 (t, $J = 7.0$ Hz, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 198.3, 151.0, 139.1, 139.0, 129.2, 128.2, 127.34, 127.29, 127.21, 127.19, 108.9, 95.2, 82.9, 73.8, 73.7, 73.2, 52.0, 37.0, 31.3, 29.5, 24.8, 24.5, 24.1, 24.0, 22.2, 13.9; HRMS (ESI): calc. for $\text{C}_{36}\text{H}_{49}\text{BNaO}_4$ $[\text{M}+\text{Na}]^+$: 579.3622; found: 579.3623.

Explanation for the formation of 4f' from 4f.

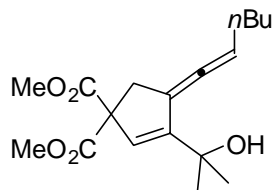


Scheme S1

Application:^{3a}



Dimethyl 4-(hex-1-enylidene)-3-(2-hydroxypropan-2-yl)cyclopent-2-ene-1,1-dicarboxylate (6)^{3c}



To a solution of **5d** (367.7 mg, 0.85 mmol) in THF (17 mL) were added H₂O₂ (0.48 mL, 30% in H₂O, 4.25 mmol), NaOH aqueous solution (0.85 mL, 3M, 2.55 mmol) sequentially at rt for 1 h. When the reaction was complete as monitored by TLC, the mixture was diluted with Et₂O (20 mL) and H₂O (10 mL). The organic layer was separated and the aqueous layer was extracted with diethyl ether (2 x 20 mL). The combined organic layers were dried over Na₂SO₄. Concentration and column chromatography on silica gel (pentane/ethyl acetate = 3:1) afforded **6** (232.1 mg, 85%): liquid; ¹H NMR (500 MHz, CDCl₃): 5.84 (s, 1H), 5.50-5.41 (m, 1H), 3.75 (s, 3H), 3.74 (s, 3H), 3.30-3.20 (m, 2H), 2.14 (s, 1H), 2.13-2.02 (m, 2H), 1.43 (s, 6H), 1.42-1.30 (m, 4H), 0.90 (t, *J* = 7.0 Hz, 3H).

Kinetic Isotope Effect (KIE) Experiments

1. Determination of Intermolecular Competition KIE of the triene formation.

To a mixture of B₂pin₂ (66.0 mg, 0.26 mmol), BQ (24.0 mg, 0.22 mmol), Pd(OAc)₂ (0.9 mg, 0.004 mmol), and LiOAc·2H₂O (4.1 mg, 0.04 mmol) were added **1a** (30.6 mg, 0.10 mmol), 0.5 mL of DCE, [D₂]-**1a** (30.8 mg, 0.10 mmol), and 0.5 mL of DCE sequentially in a vial at rt. Then the reaction was stirred at 50 °C for 1 h, quenched with Et₂O (5 mL), and concentrated under reduced pressure. The yields of **4d** and [D₅]-**4d** were analyzed by ¹H NMR measurement using anisole as the internal standard (11 μL, 0.1 mmol) and the ratio of **4d**/[D₅]-**4d** was analyzed by HRMS and determined by comparing the abundance of the corresponding [M+Na] and [M+5+Na] peaks (assuming **4d** and [D₅]-**4d** have similar ionization pattern).

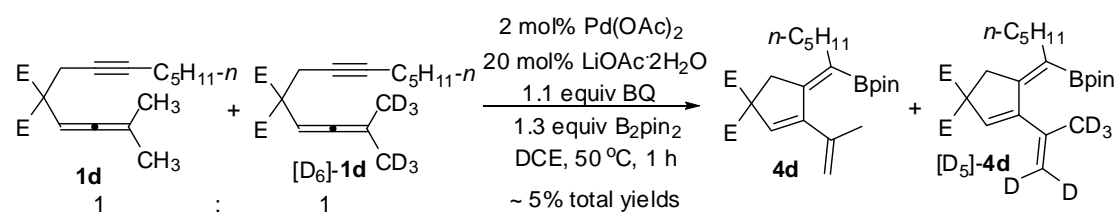


Table S1.

	1st	2nd	average
Abundance of 4d	173744	102314	
Abundance of [D ₅]- 4d	27420	14671	
Abundance of 1d in the end of the reaction	625356	441152	
Abundance of [D ₆]- 1d in the end of the reaction	624172	456367	
The ratio (1d /[D ₆]- 1d) in the end of the reaction	1.00	0.97	
KIE	6.34	6.97	6.66

Display Report

Analysis Info

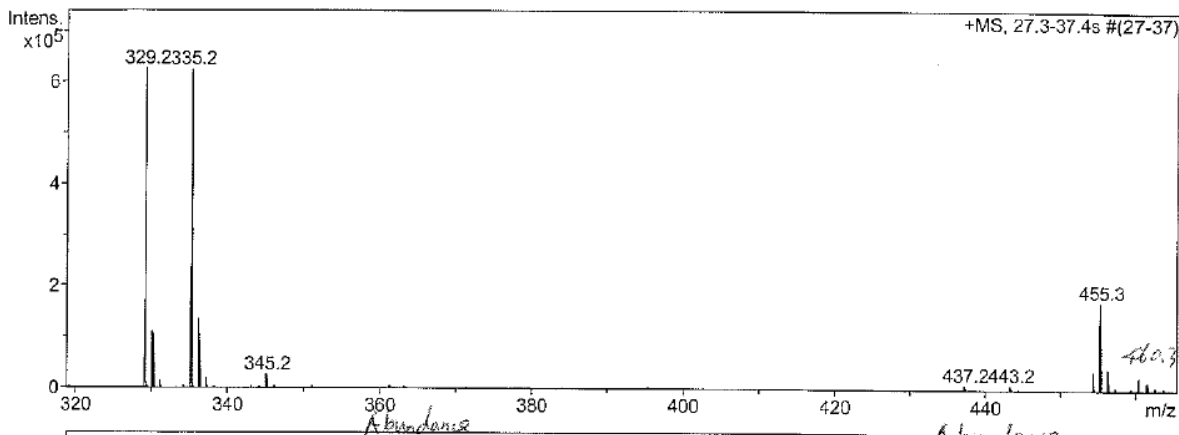
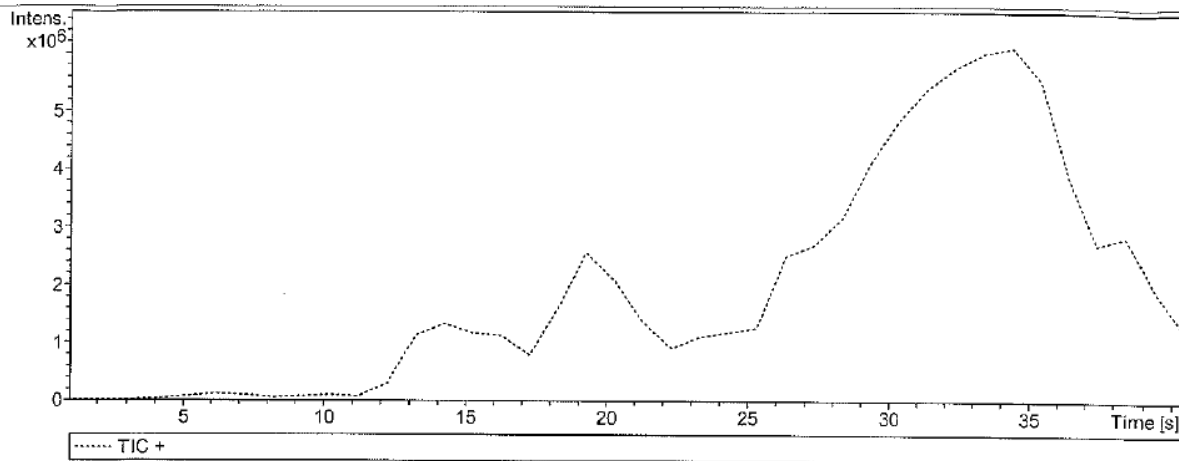
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Operator Carin Larsson
 Instrument / Ser# microTOF 125

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Scan End	2000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



<p>For mw 329 = $\frac{625356}{(1d)} = 1$</p> <p>mw 335 = $\frac{624172}{(Dd)-1d}$</p>	<p>For mw = 455.3 = $\frac{173748}{(4d)} = 6.3$</p> <p>MW = 860 = $\frac{17420}{(Dd)-4d} = 1$</p>
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2. Determination of Intermolecular Competition KIE of the vinylallene formation.

To a mixture of B₂pin₂ (66.0 mg, 0.26 mmol), BQ (24.0 mg, 0.22 mmol), Pd(OAc)₂ (0.9 mg, 0.004 mmol), and BF₃·Et₂O (6 μL, 0.04 mmol) were added **1a** (30.6 mg, 0.10 mmol), 0.5 mL of THF, [D₂]-**1a** (30.8 mg, 0.10 mmol), and 0.5 mL of THF sequentially in a vial at rt. Then the reaction was stirred at 50 °C for 1 h, quenched with Et₂O (5 mL), and concentrated under reduced pressure. The yields and the ratio of **5d** and [D₁]-**5d** were analyzed by ¹H NMR measurement using anisole as the internal standard (11 μL, 0.1 mmol).

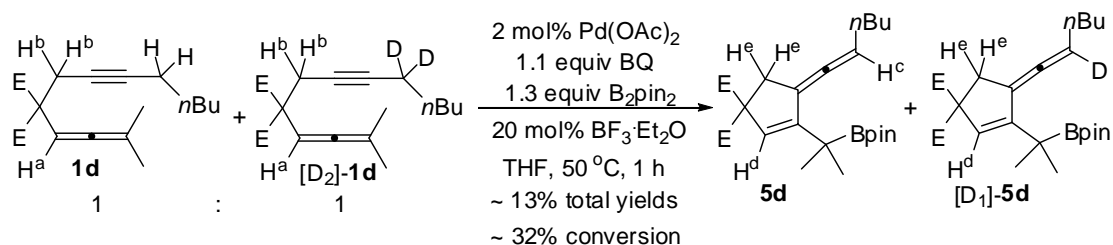
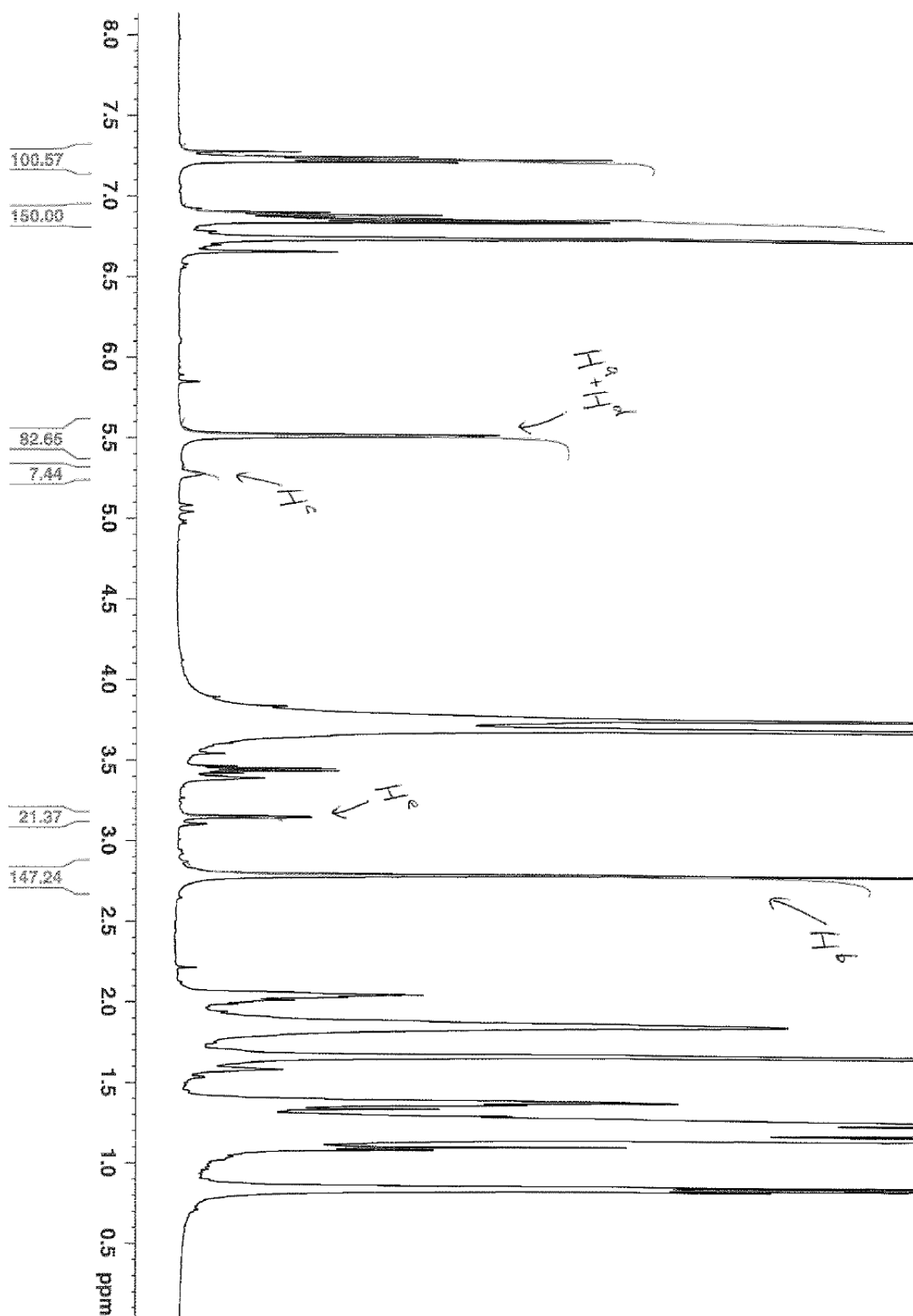


Table S2.

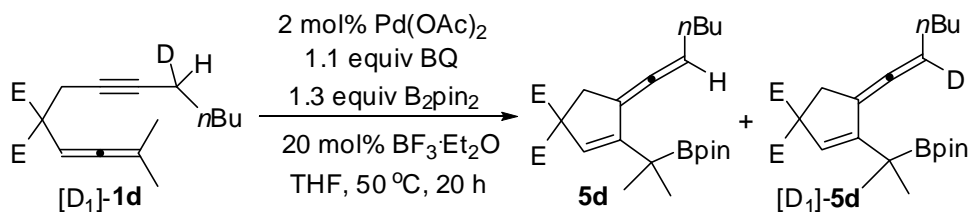
	1st	2nd	3rd	4th	average
Yield of 5d (%)	7.4	9.6	9.9	9.8	
Total yields of 5d + [D₁]-5d (%)	10.5	15.0	12.9	13.6	13.0
Yield of [D₁]-5d (%) (from calculation)	3.1	5.4	3.0	3.8	
Conversion (%)	26	37	33	30	31.5
Ratio between 5d / [D₁]-5d	2.39	1.78	3.30	2.58	2.43
The estimated ratio ([D₂]-1d / 1d) in the end of the reaction	1.12	1.14	1.23	1.09	1.15
KIE	2.53	1.90	3.66	2.81	2.73

Since the ratio of the starting material continuously changes during the reaction, the isotope effect is obtained after a slight correction of this factor. Taking the experiment 1 for example, the ratio (**1d**/**[D₂]-1d**) in the end of the reaction was estimated to be 1:1.12. The yield of (**5d** + **[D₁]-5d**) was 10.5% at a 26% conversion. Here, we assume the 15.5% (26%-10.5%) of the starting materials were converted to by-products without any isotope effect. Then the remaining **1d** should be 50%-(15.5%/2)-7.4% = 34.8% and the remaining **[D₂]-1a** should be 50%-(15.5%/2)-3.1% = 39.1%, respectively. Therefore, the isotope effect calculated from the product ratio and the

change of the starting material ratio is approximately 2.53 (namely, $2.39 \times [(1+1.12)/2]$).



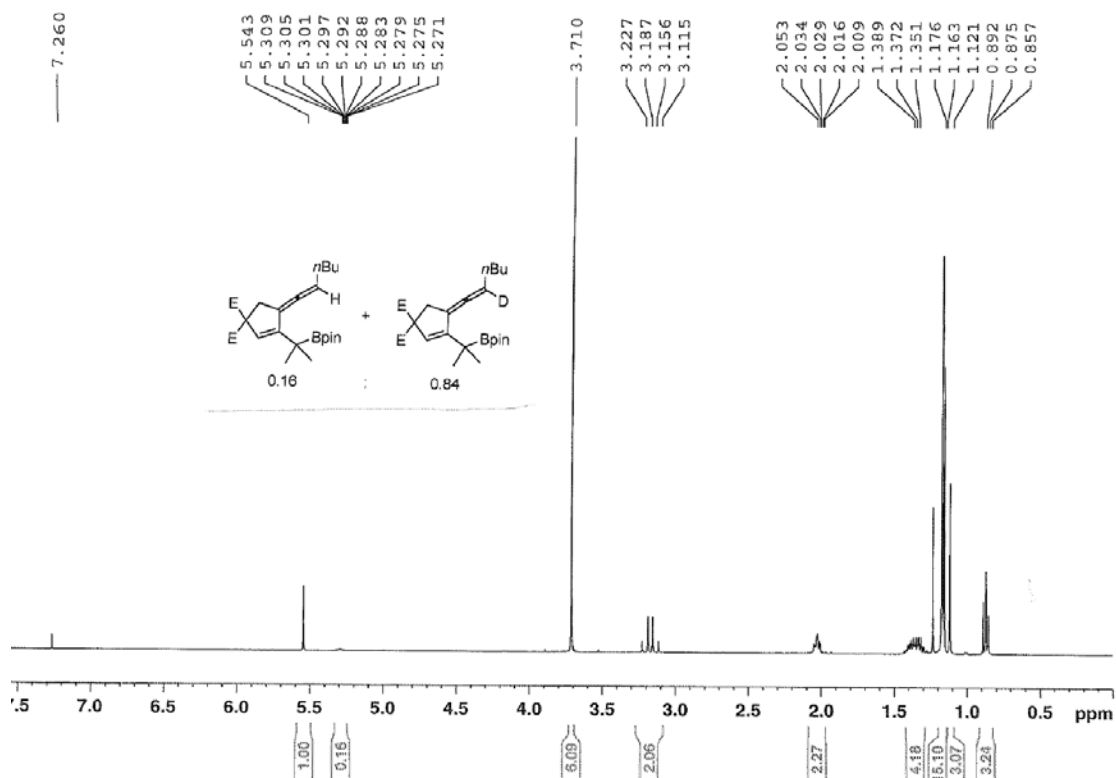
3. Determination of Intramolecular KIE of the vinylallene formation.



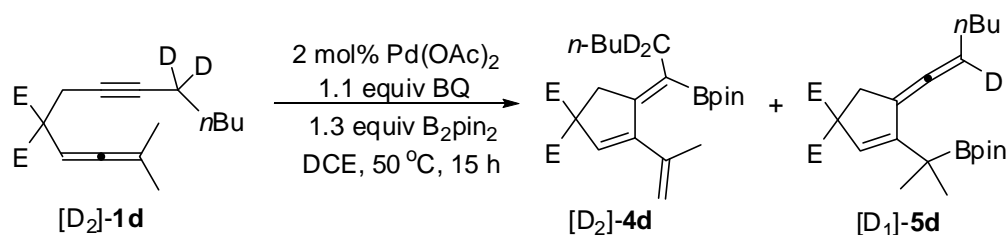
To a mixture of B_2pin_2 (66.0 mg, 0.26 mmol), BQ (24.0 mg, 0.22 mmol), $Pd(OAc)_2$ (0.9 mg, 0.004 mmol), and $BF_3 \cdot Et_2O$ (6 μL , 0.04 mmol) were added $[D_1]-1d$ (61.4 mg, 0.20 mmol) and 1.0 mL of THF sequentially in a vial at rt. Then the reaction was stirred at 50 °C for 20 h, quenched with Et_2O (5 mL), and concentrated under reduced pressure. The yields and the ratio of $5d$ and $[D_1]-5d$ were analyzed by 1H NMR measurement using anisole as the internal standard (22 μL , 0.2 mmol).

Table S3.

	1st	2nd	3rd	average
Yield of $5d$ (%)	12	11	9	
Yield of $[D_1]-5d$ (%)	54	55	57	
Total yields (%)	66	66	66	66
KIE	4.50	5.00	6.33	5.28



4. Effect of isotope substitution at the alkyne part on product distribution.

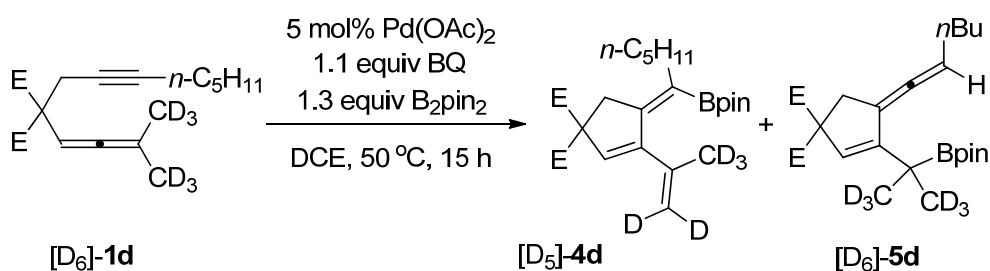


To a mixture of B_2pin_2 (33.0 mg, 0.13 mmol), BQ (12.0 mg, 0.11 mmol), and Pd(OAc)_2 (0.5 mg, 0.002 mmol) were added $[\text{D}_2]\text{-1d}$ (30.8 mg, 0.10 mmol) and 0.5 mL of DCE sequentially in a vial at rt. Then the reaction was stirred at 50 °C for 15 h, quenched with Et_2O (2.5 mL), and concentrated under reduced pressure. The yields and the ratio of $[\text{D}_2]\text{-4d}$ and $[\text{D}_1]\text{-5d}$ were analyzed by ^1H NMR measurement using anisole as the internal standard (11 μL , 0.1 mmol).

Table S4.

	1st	2nd	average
Yield of $[\text{D}_2]\text{-4d}$ (%)	73	75	74
Yield of $[\text{D}_1]\text{-5d}$ (%)	6	6	6
Ratio $[\text{D}_2]\text{-4d}/[\text{D}_1]\text{-5d}$			12.3/1

5. Effect of isotope substitution at allene part on product distribution.



To a mixture of B_2pin_2 (33.0 mg, 0.13 mmol), BQ (12.0 mg, 0.11 mmol), and Pd(OAc)_2 (1.2 mg, 0.005 mmol) were added $[\text{D}_6]\text{-1d}$ (31.2 mg, 0.10 mmol) and 0.5 mL of DCE sequentially in a vial at rt. Then the reaction was stirred at 50 °C for 15 h, quenched with Et_2O (2.5 mL), and concentrated under reduced pressure. The yields and the ratio of $[\text{D}_5]\text{-4d}$ and $[\text{D}_6]\text{-5d}$ were analyzed by ^1H NMR measurement using anisole as the internal standard (11 μL , 0.1 mmol).

The product ratio for this experiment was obtained from the integration of the overlapping CH₂ signals at ca. 3.20 ppm. To enhance accuracy of the integration, the ratio was determined as the average of two methods of integrating the signal, which led to [D₅]-**4d**/[D₆]-**5d** is 1/5.0, and the ¹H NMR yield is 12% and 60%, respectively.

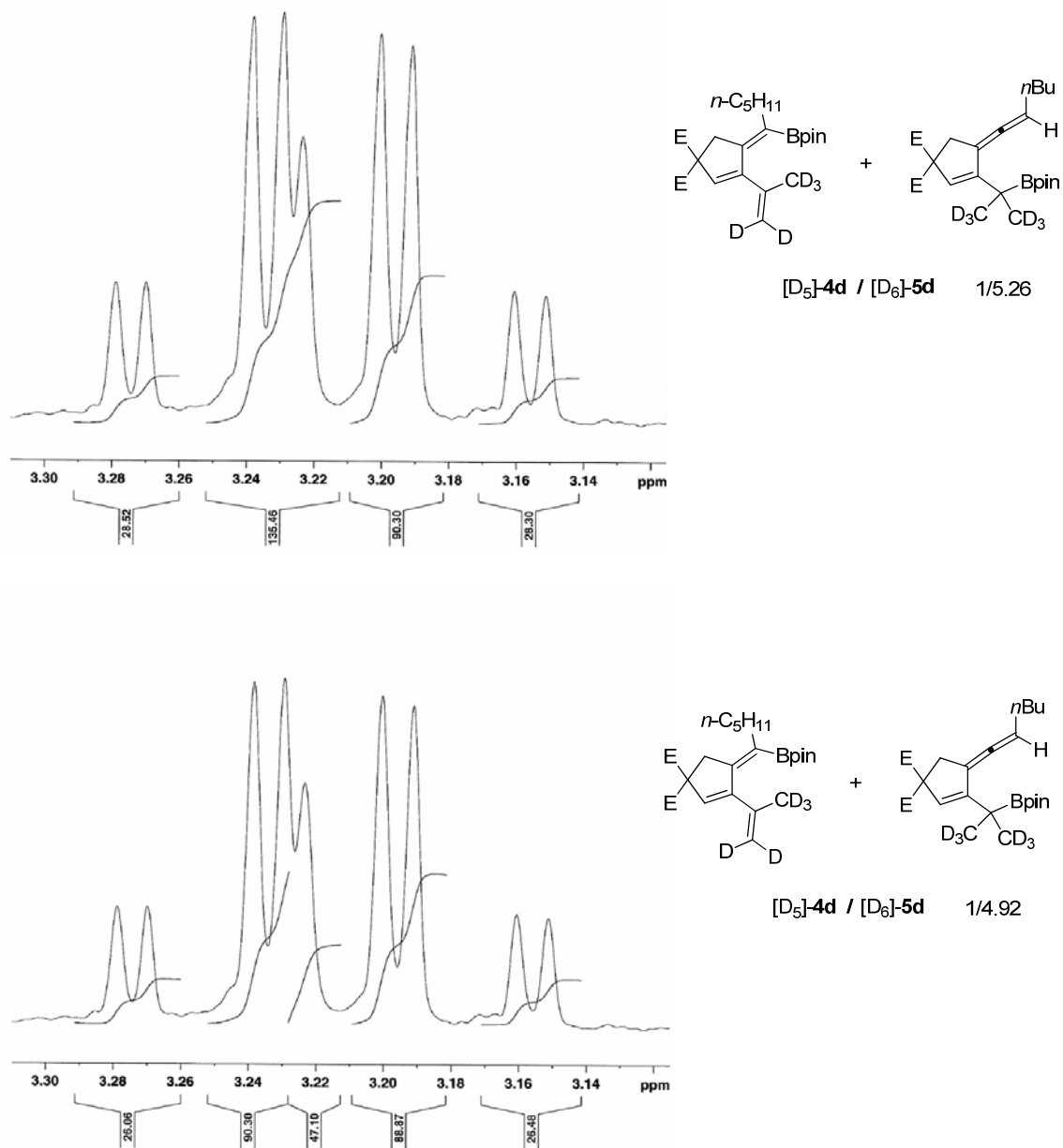
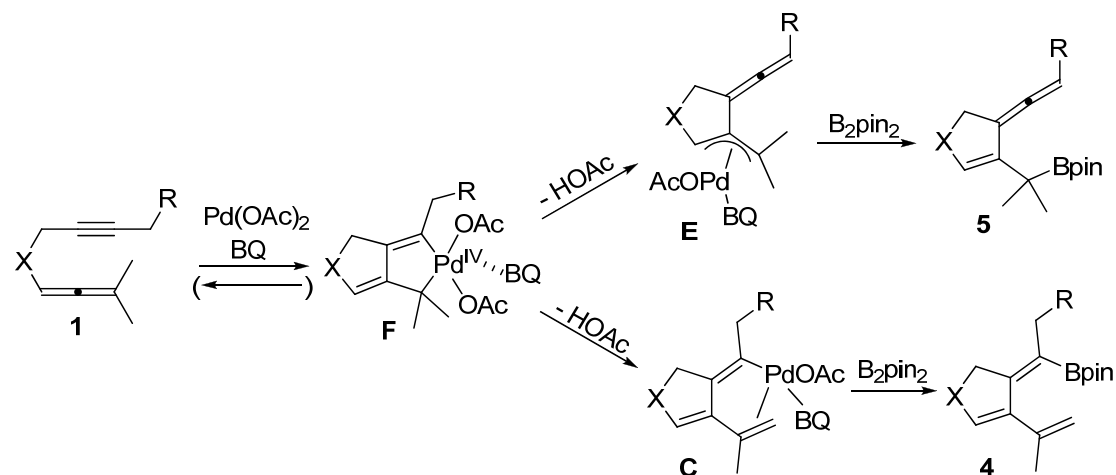


Figure S1.

Alternative mechanism to that proposed in Scheme 5 for oxidative carbocyclization/borylation of allene **1**

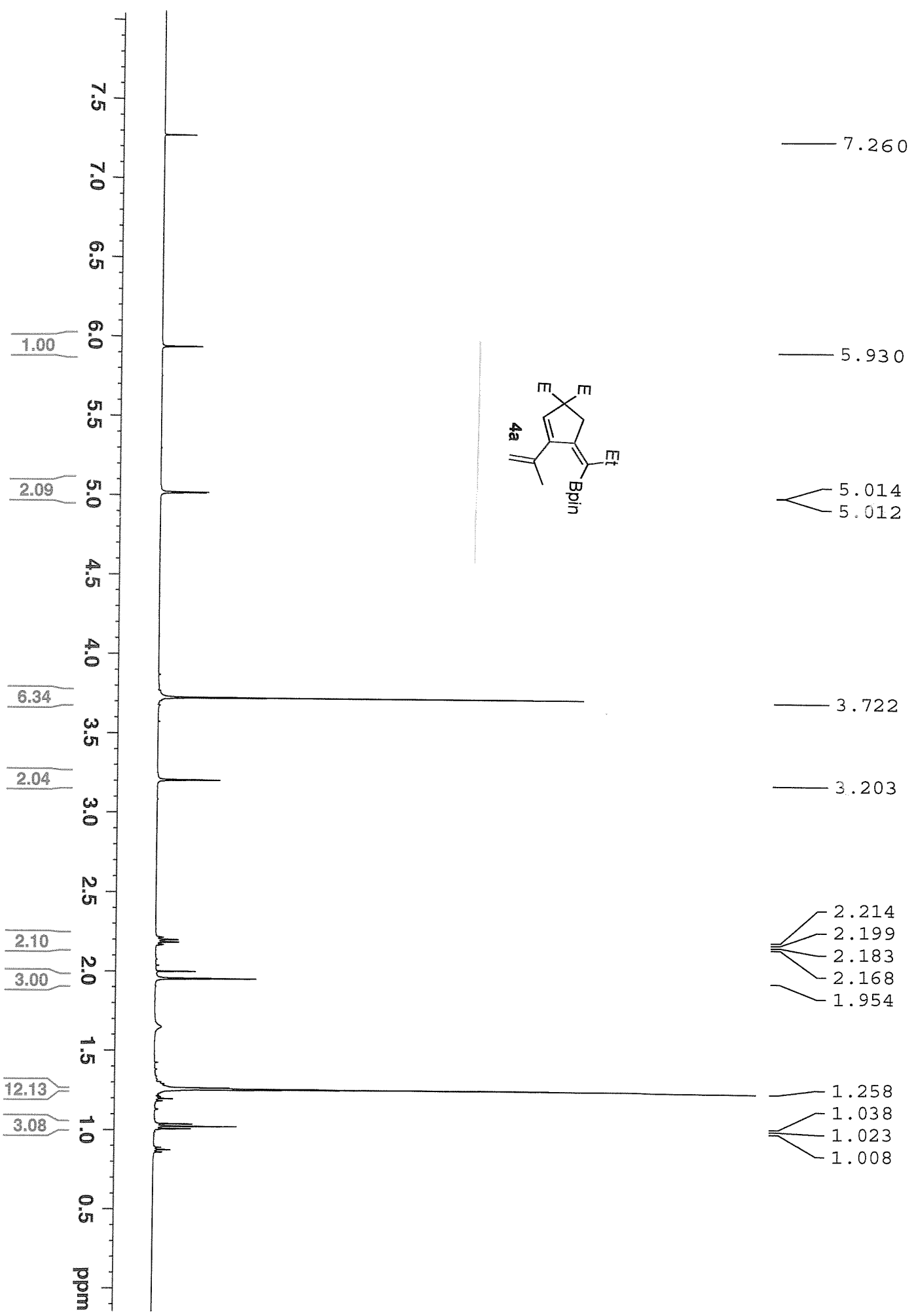
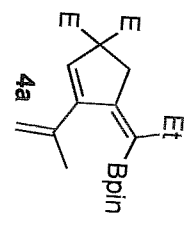
A mechanism involving a pallada(IV)cyclopentene intermediate **F** is also possible, which would generate intermediates **C** and **E** selectively via β -H elimination and subsequent loss of HOAc leading to products **4** and **5**, respectively.

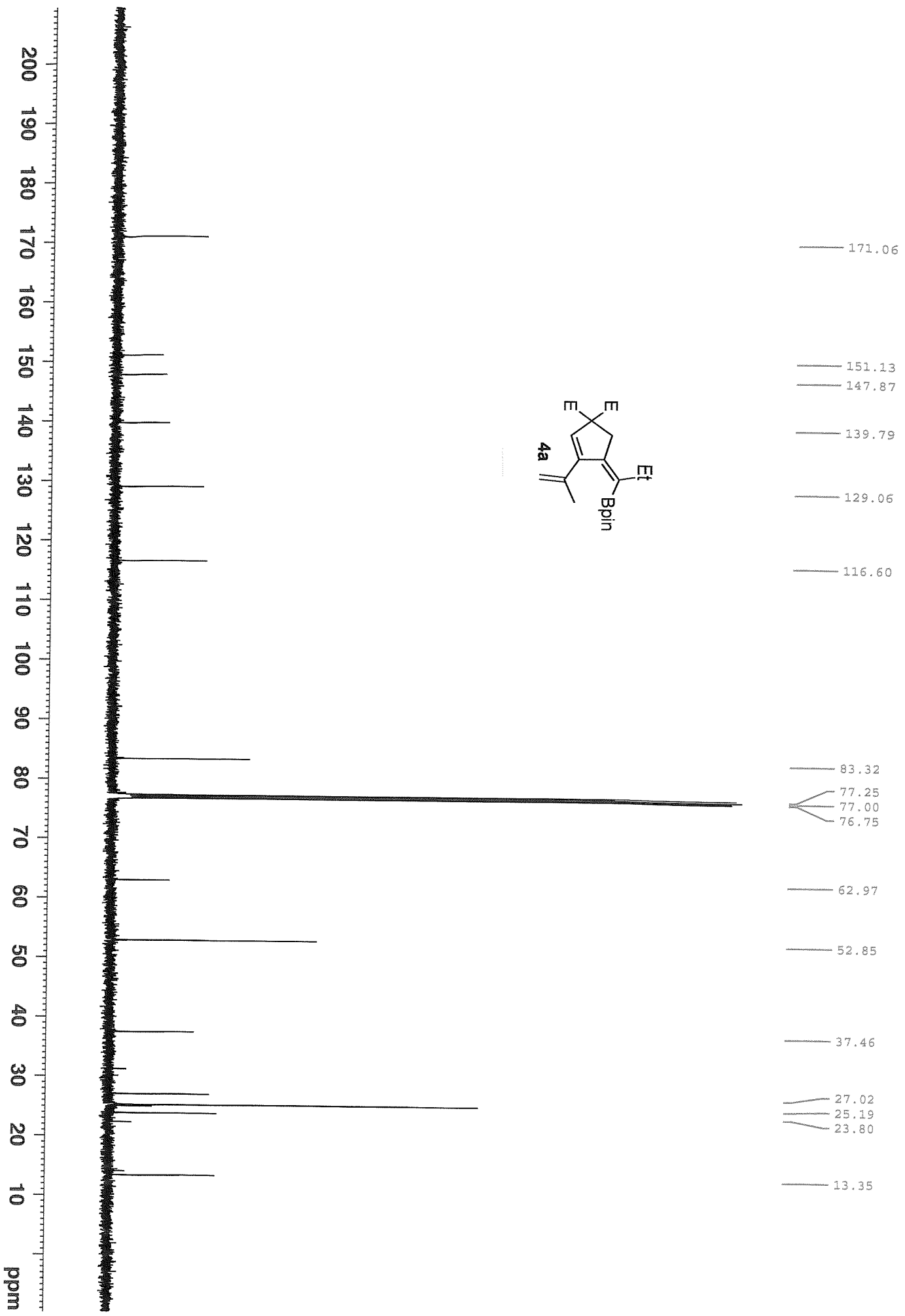


Scheme S2. Proposed alternative mechanism for palladium-catalyzed oxidative selective carbocyclization/borylation of allenes **1**.

References:

1. a) D. K. Black, S. R. Landor, A. N. Patel, P. F. Whiter, *Tetrahedron Lett.* **1963**, *4*, 483; b) A. K. Å. Persson, E. V. Johnston, J.-E. Bäckvall, *Org. Lett.* **2009**, *11*, 3814.
2. C. Sperger, L. H. S. Strand, A. Fiksdahl, *Tetrahedron* **2010**, *66*, 7749.
3. a) V. Pardo-Rodríguez, J. Marco-Martínez, E. Buñuel, D. J. Cárdenas, *Org. Lett.* **2009**, *11*, 4548; b) Y. Deng, T. Bartholomeyzik, A. K. Å. Persson, J. Sun, J.-E. Bäckvall, *Angew. Chem.* **2012**, *124*, 2757; *Angew. Chem. Int. Ed.* **2012**, *51*, 2703; c) Y. Deng, J.-E. Bäckvall, *Angew. Chem. Int. Ed.* **2013**, DOI: 10.1002/anie.201208718.





Mass Spectrum SmartFormula Report

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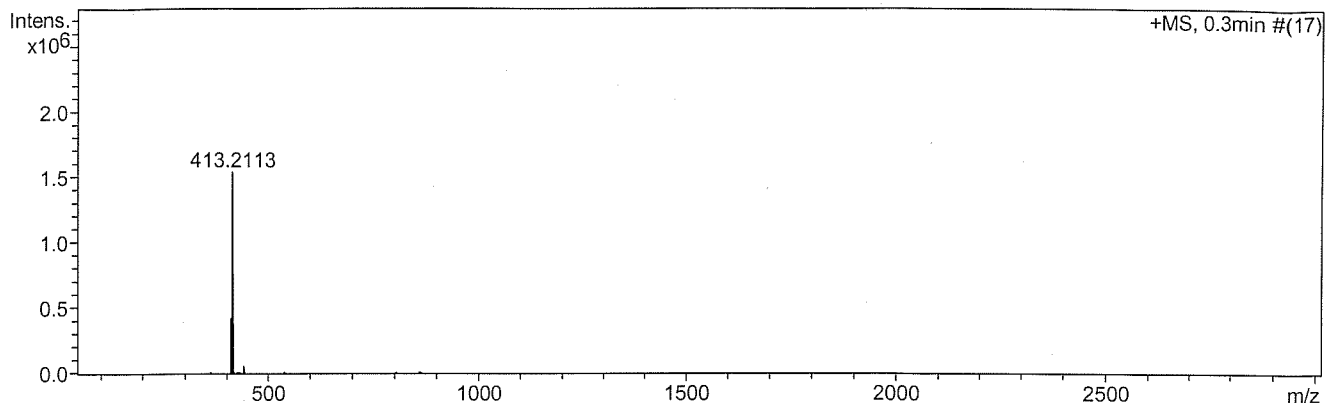
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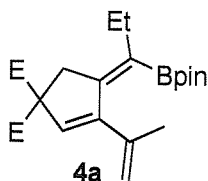
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Instrument / Ser# micrOTOF 125

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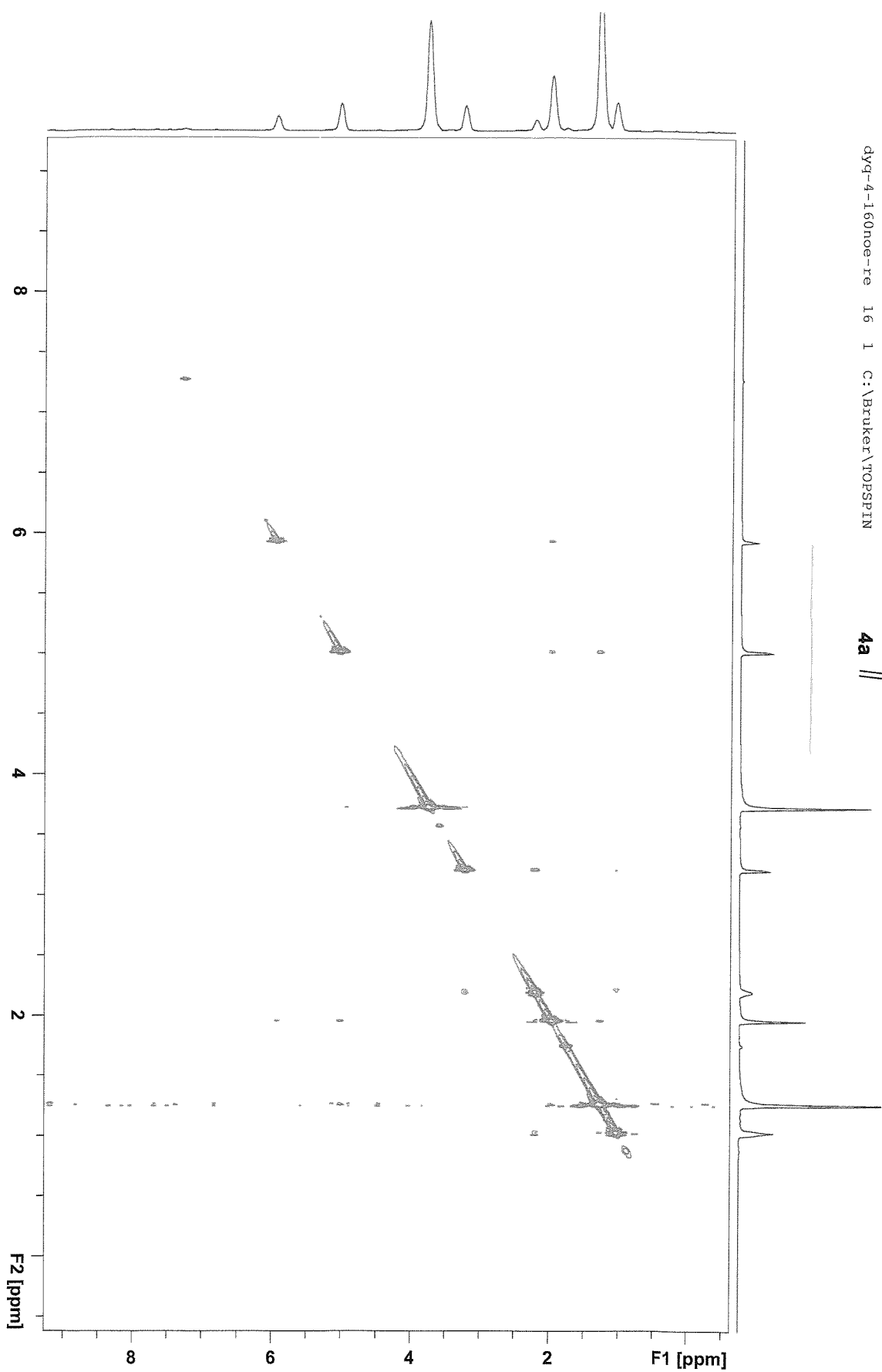
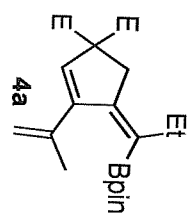
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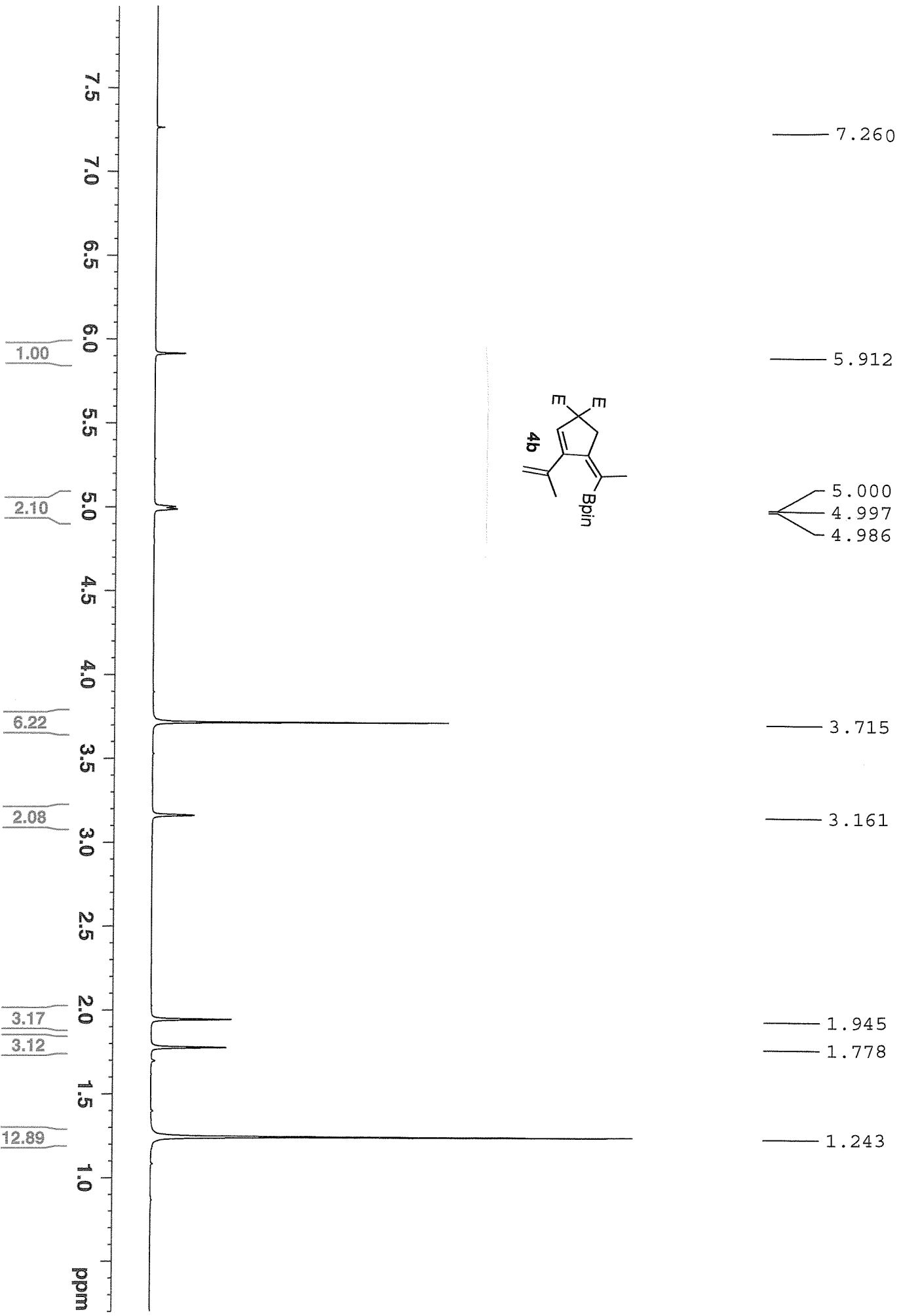


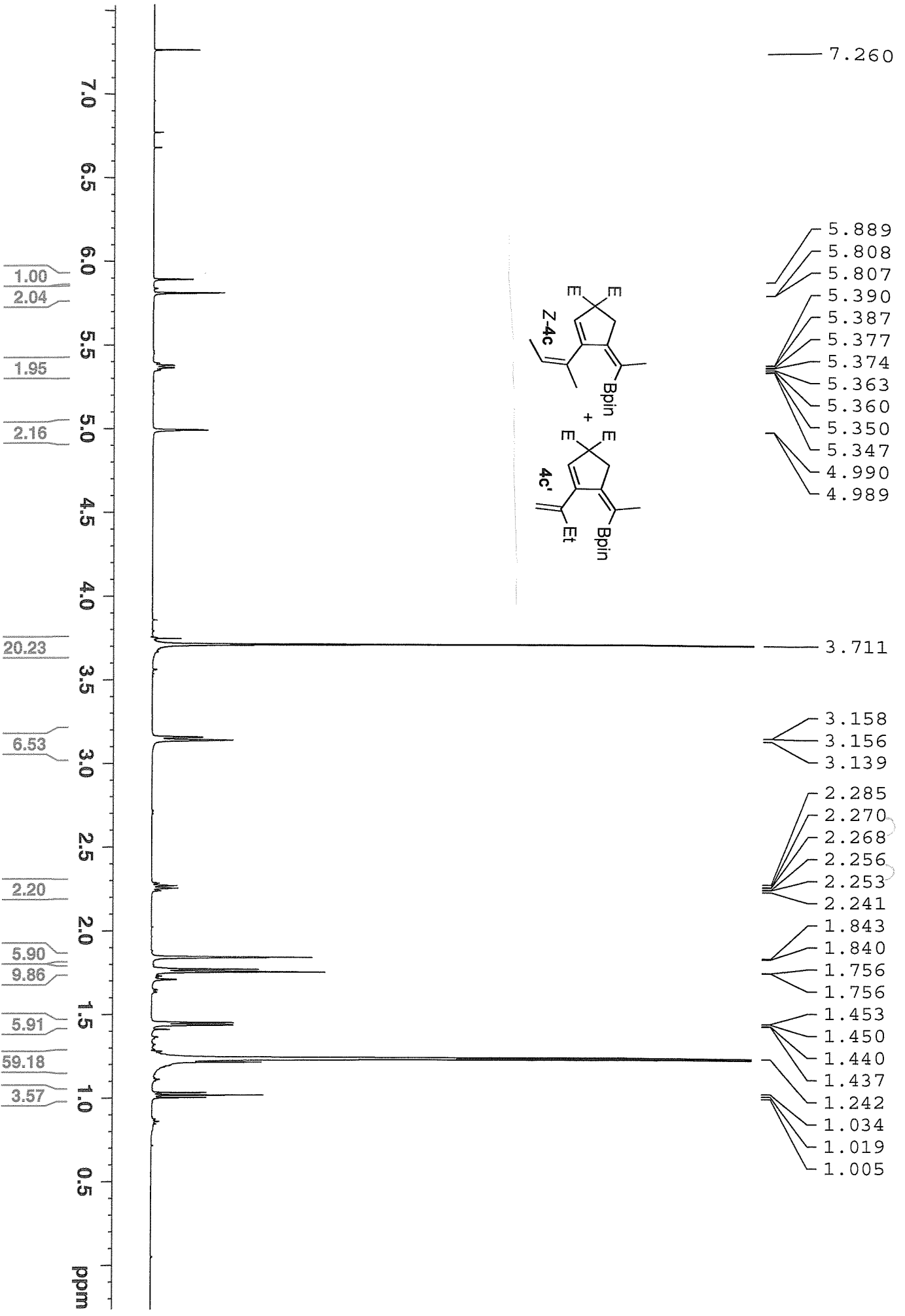
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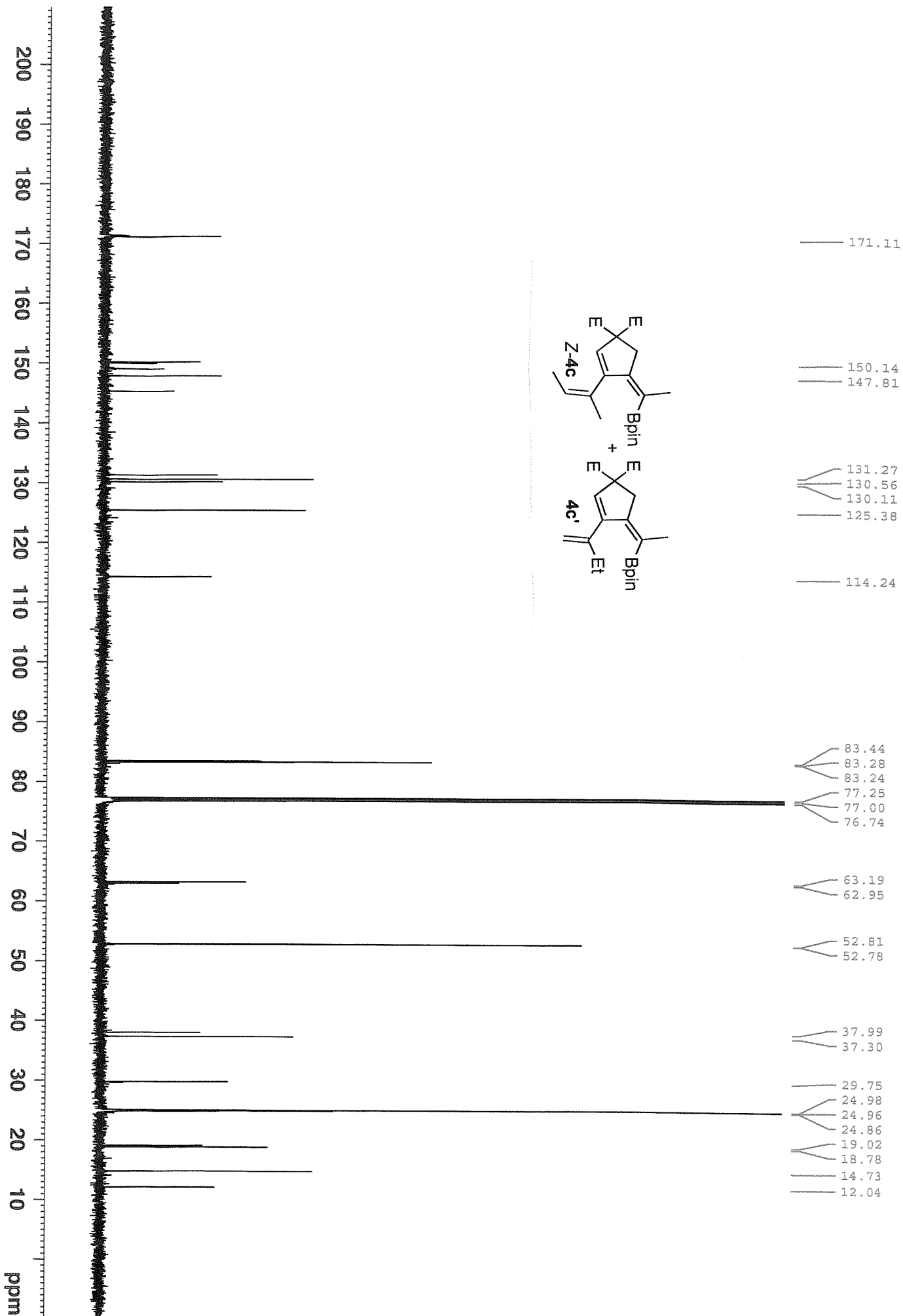


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Mass Spectrum SmartFormula Report

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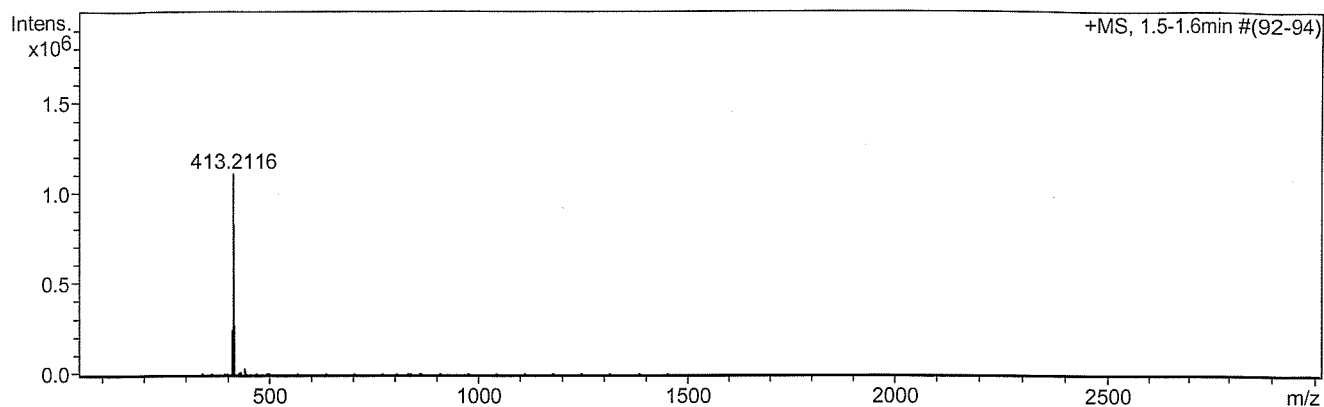
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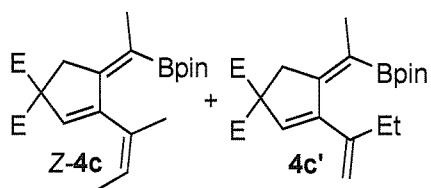
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Instrument / Ser# micrOTOF 125

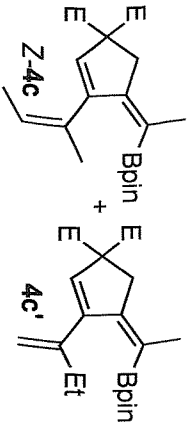
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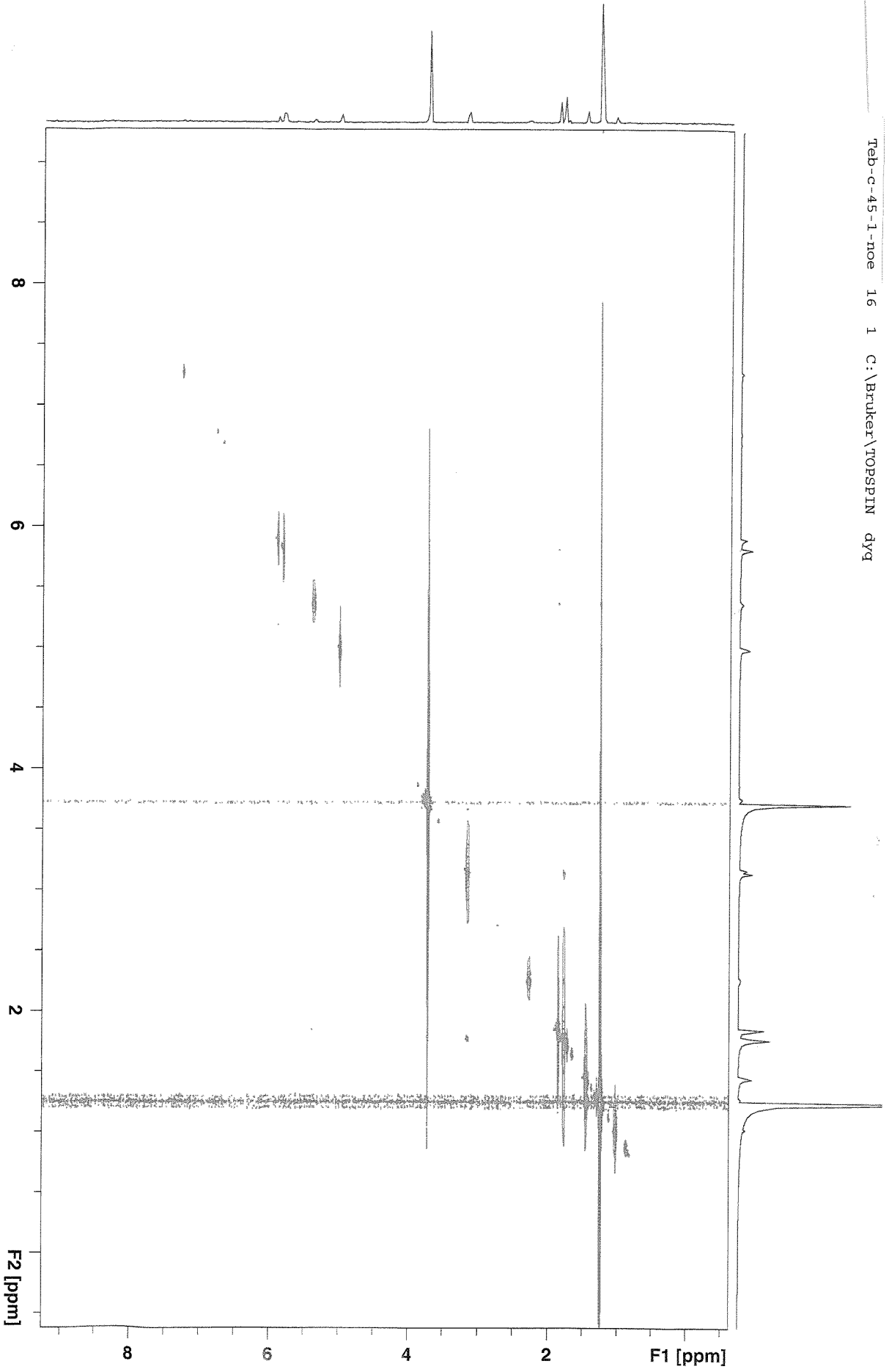


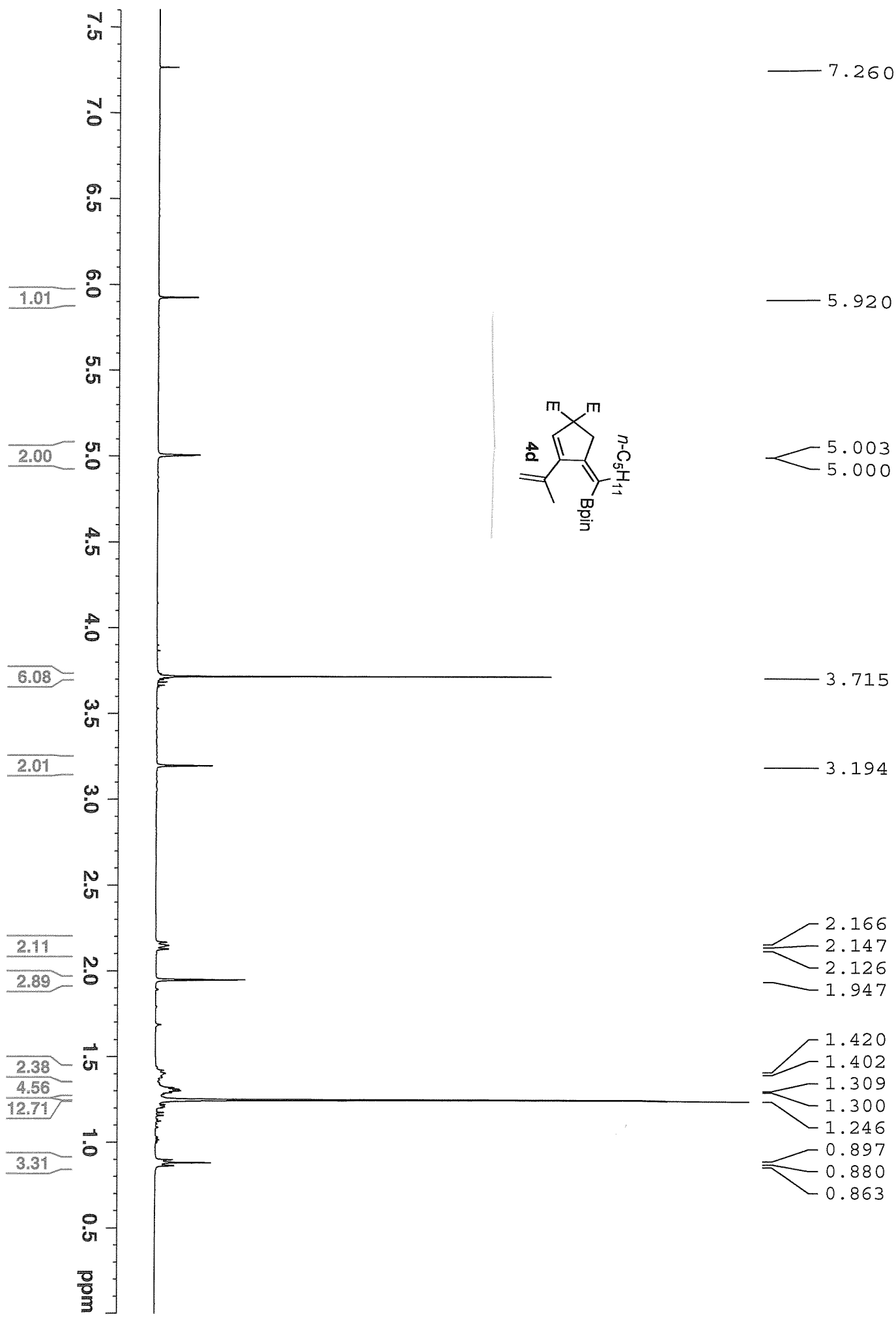
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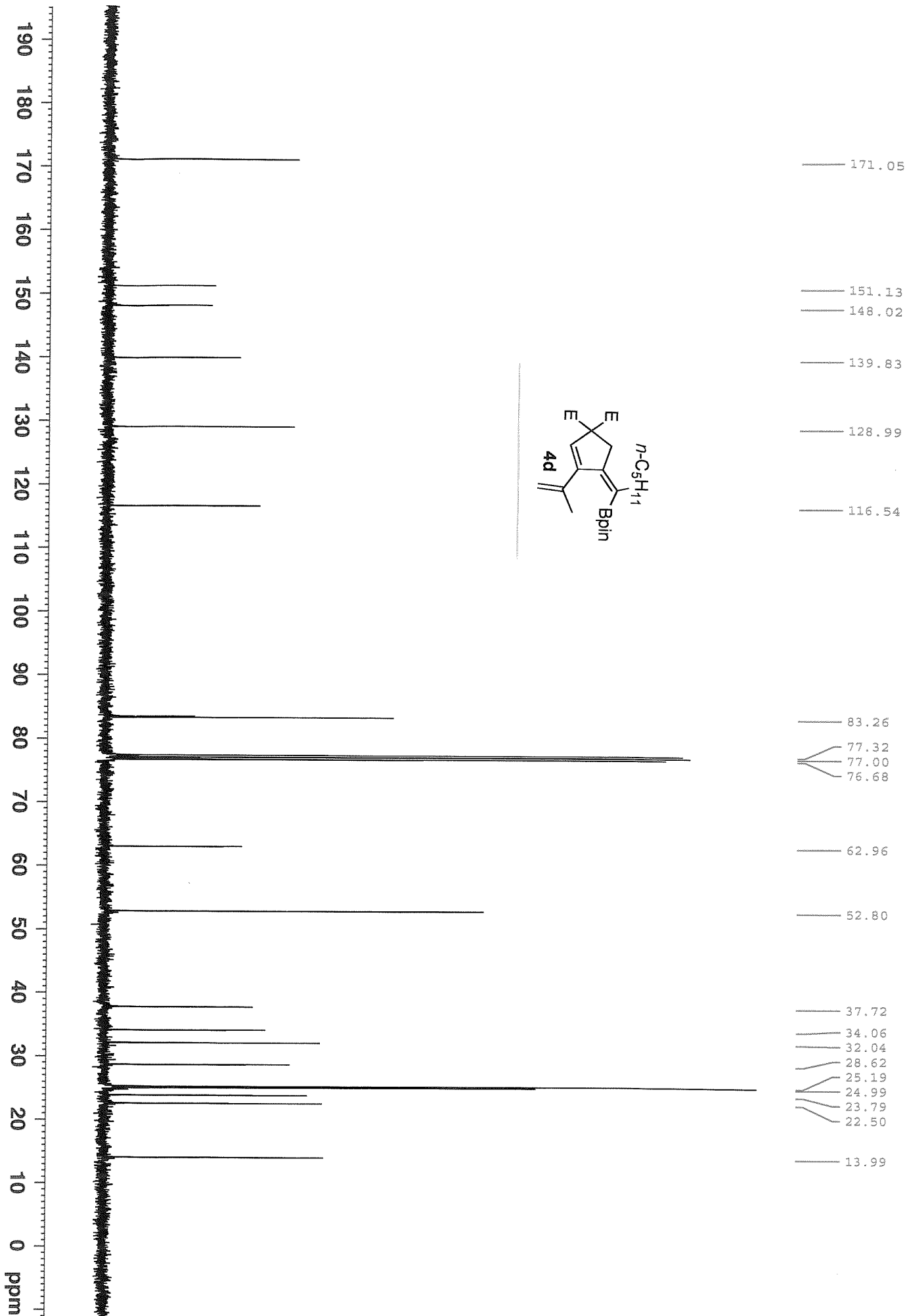
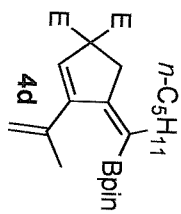




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Mass Spectrum SmartFormula Report

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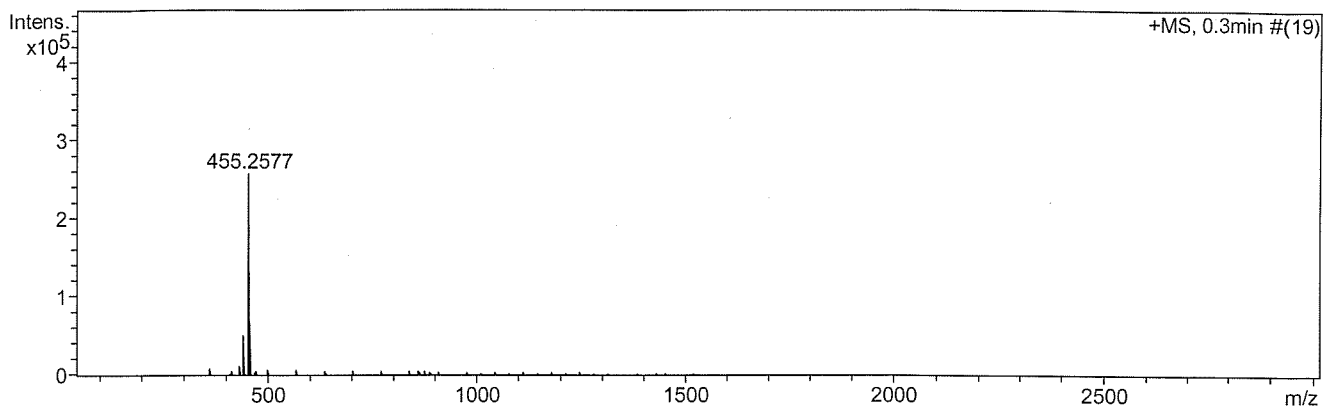
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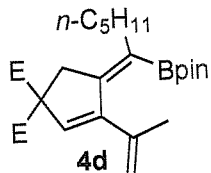
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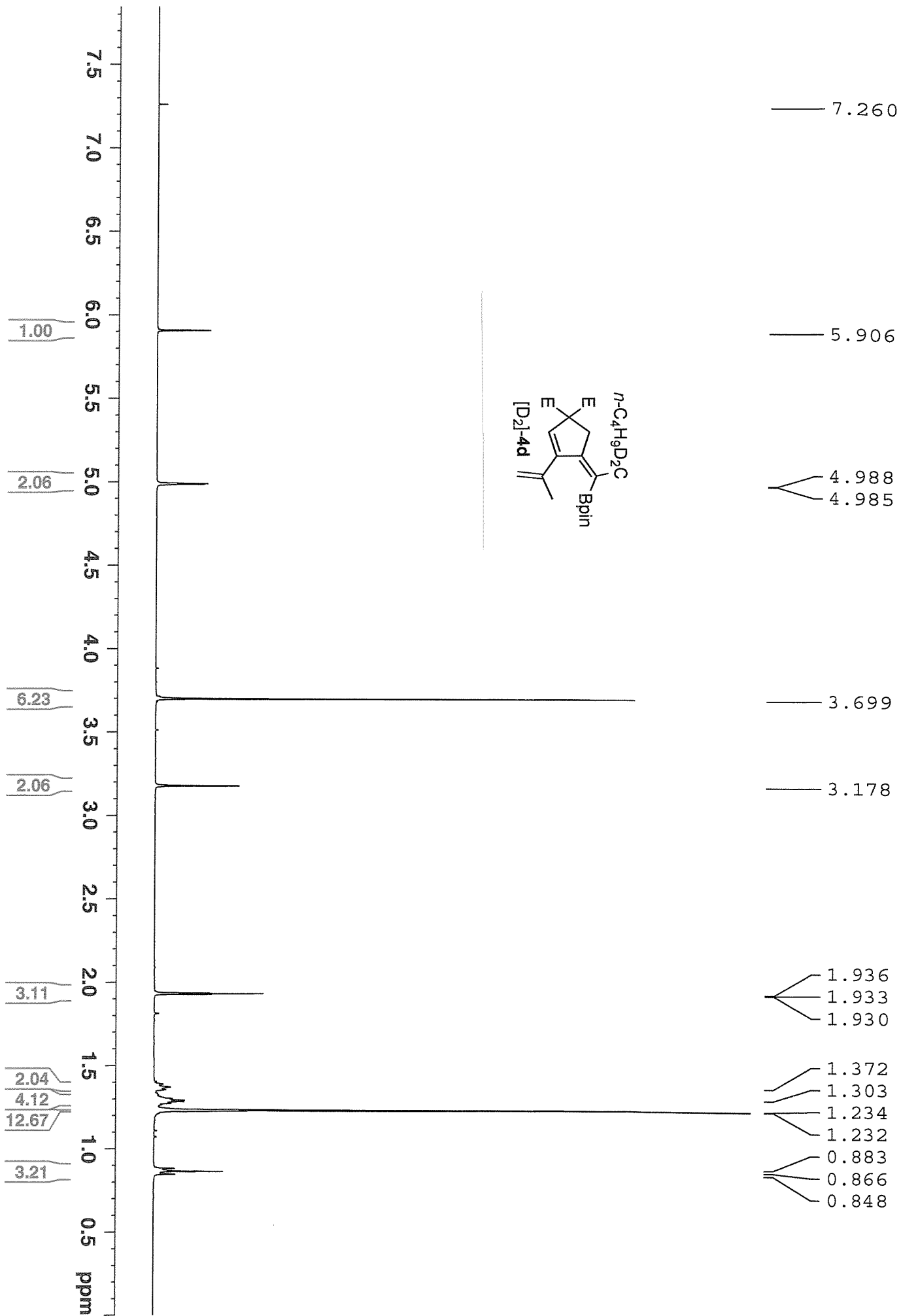
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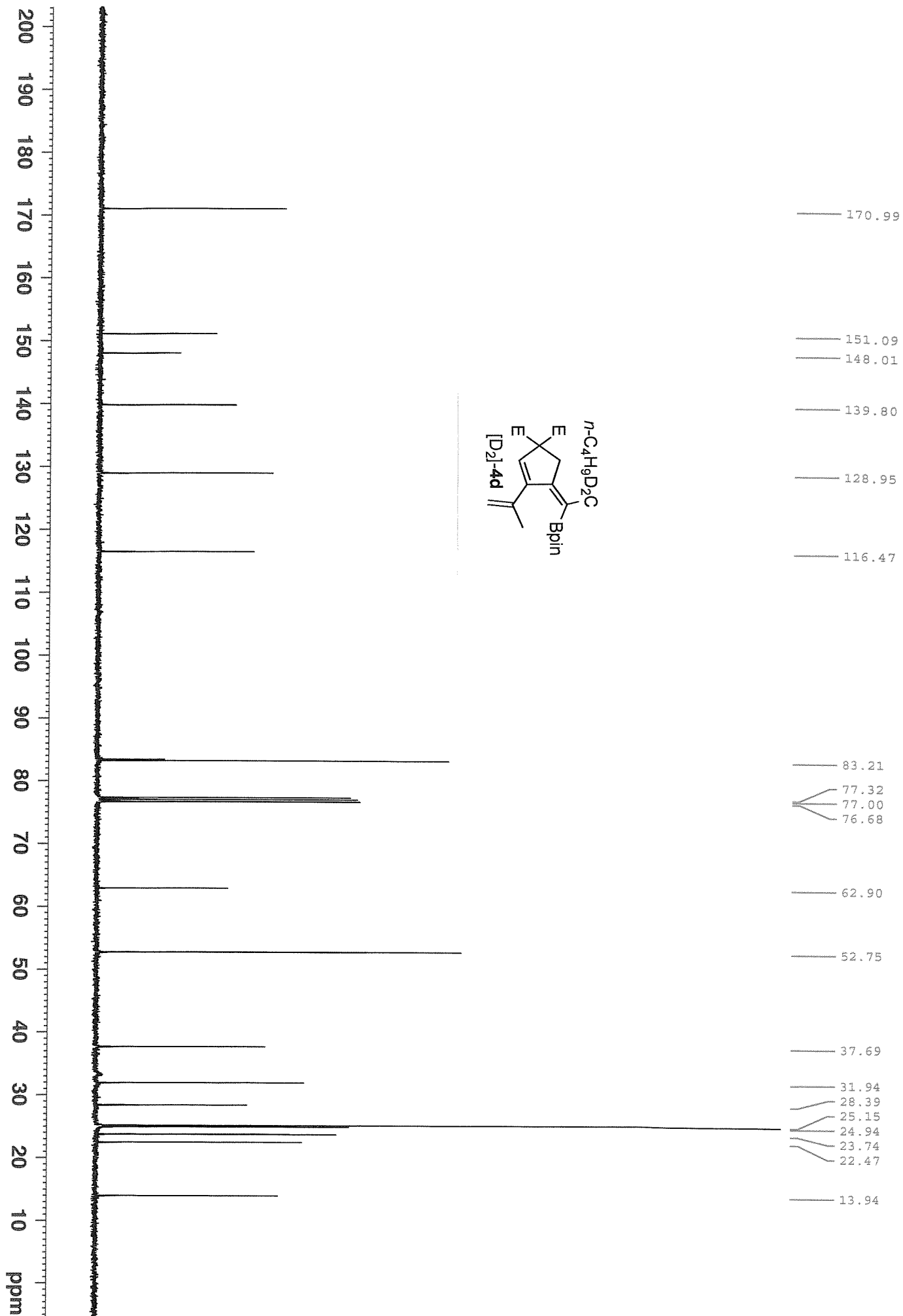
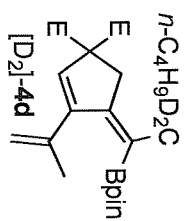
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e ⁻ Conf	N-Rule
455.2577	1	C ₂₄ H ₃₇ BNaO ₆	100.00	455.2580	0.2	0.5	14.2	6.5	even	ok







Mass Spectrum SmartFormula Report

Analysis Info

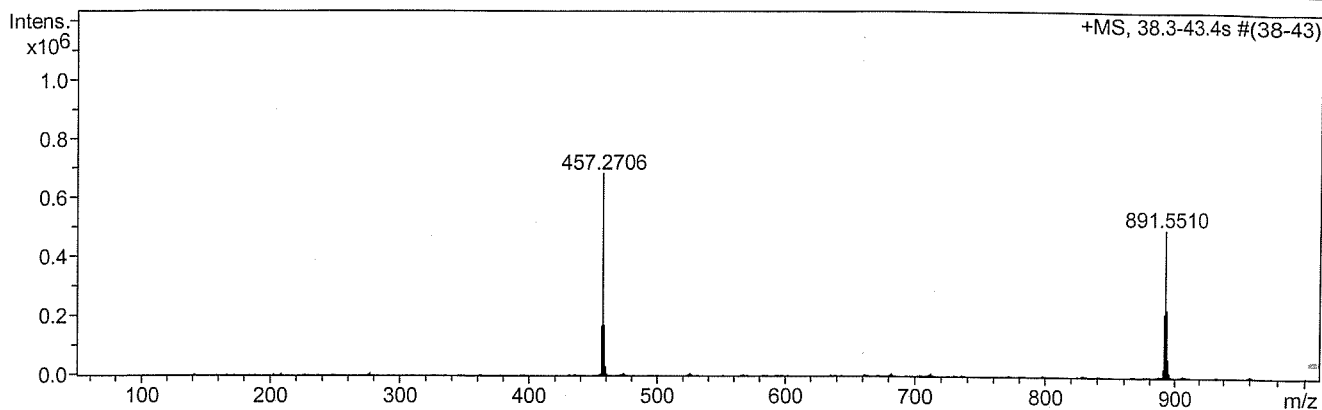
Analysis Name E:\Data2\Youqian\dyq-4-182000001.d
Method Tune_low_pos.m
Sample Name dyq-4-182
Comment

Acquisition Date 2012-12-07 18:28:51

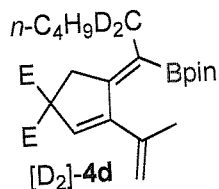
Operator Carin Larsson
Instrument / Ser# micrOTOF 125

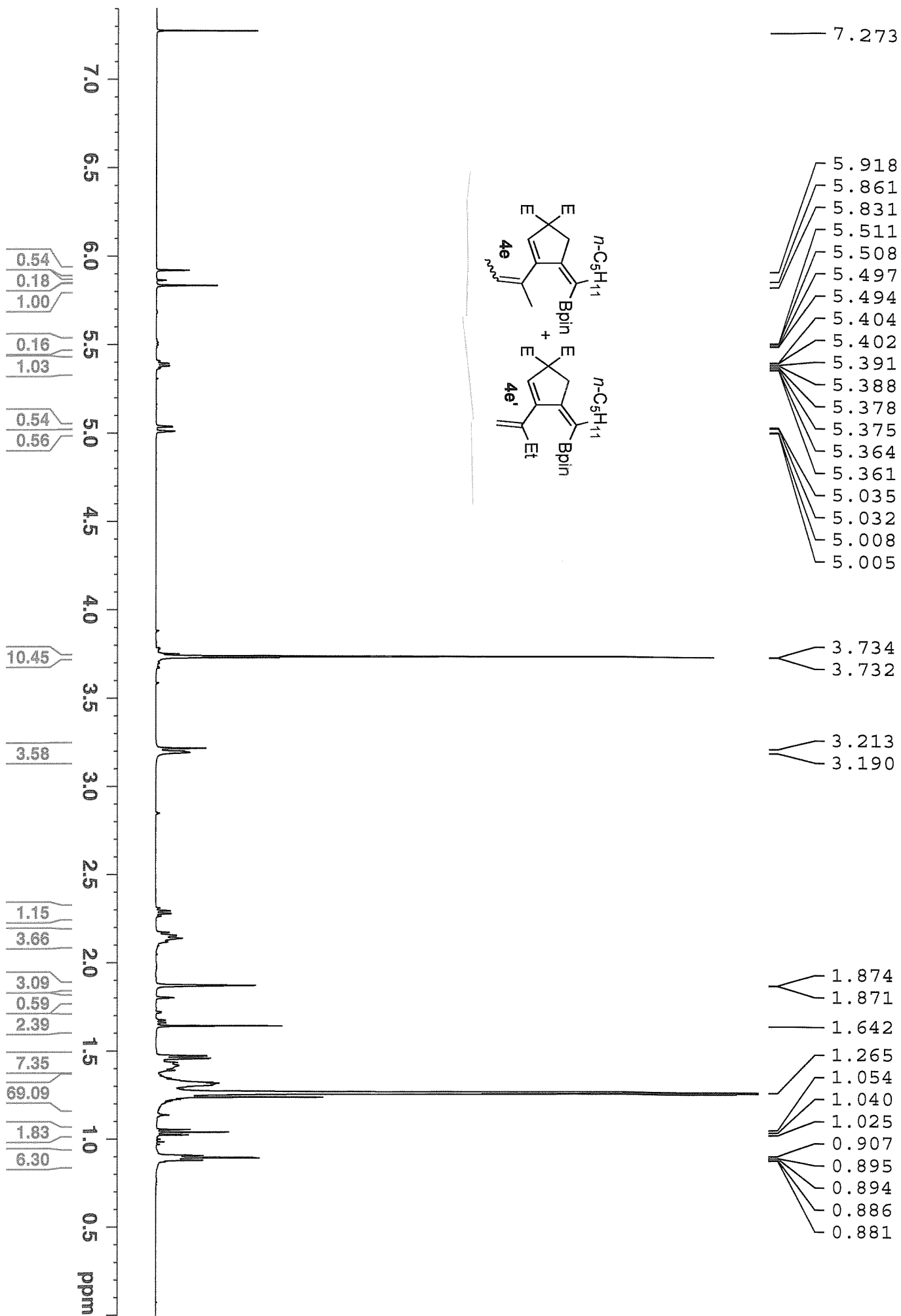
Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4000 V	Set Dry Gas	4.0 l/min
Scan End	1000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 24 H 35 B D 2 Na O 6	457.2706	457.2705	-0.2	0.0





Mass Spectrum SmartFormula Report

Analysis Info

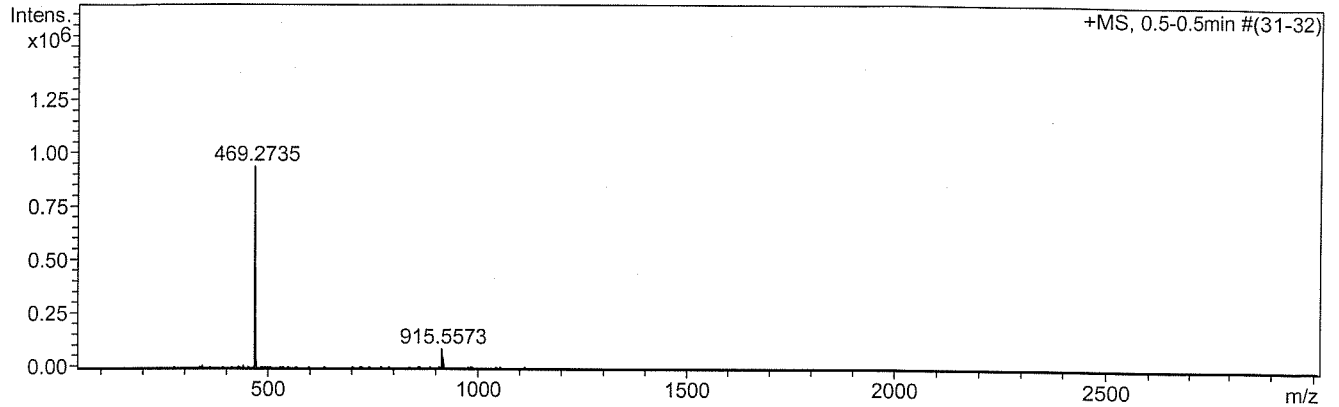
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Method Tune_wide_pos.m
Sample Name dyq-4-163-1
Comment

Acquisition Date 2012-11-17 14:45:16

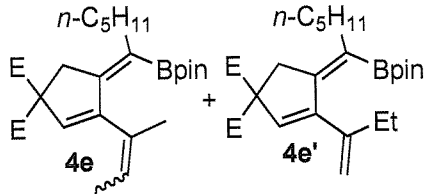
Operator Carin Larsson
Instrument / Ser# micrOTOF 125

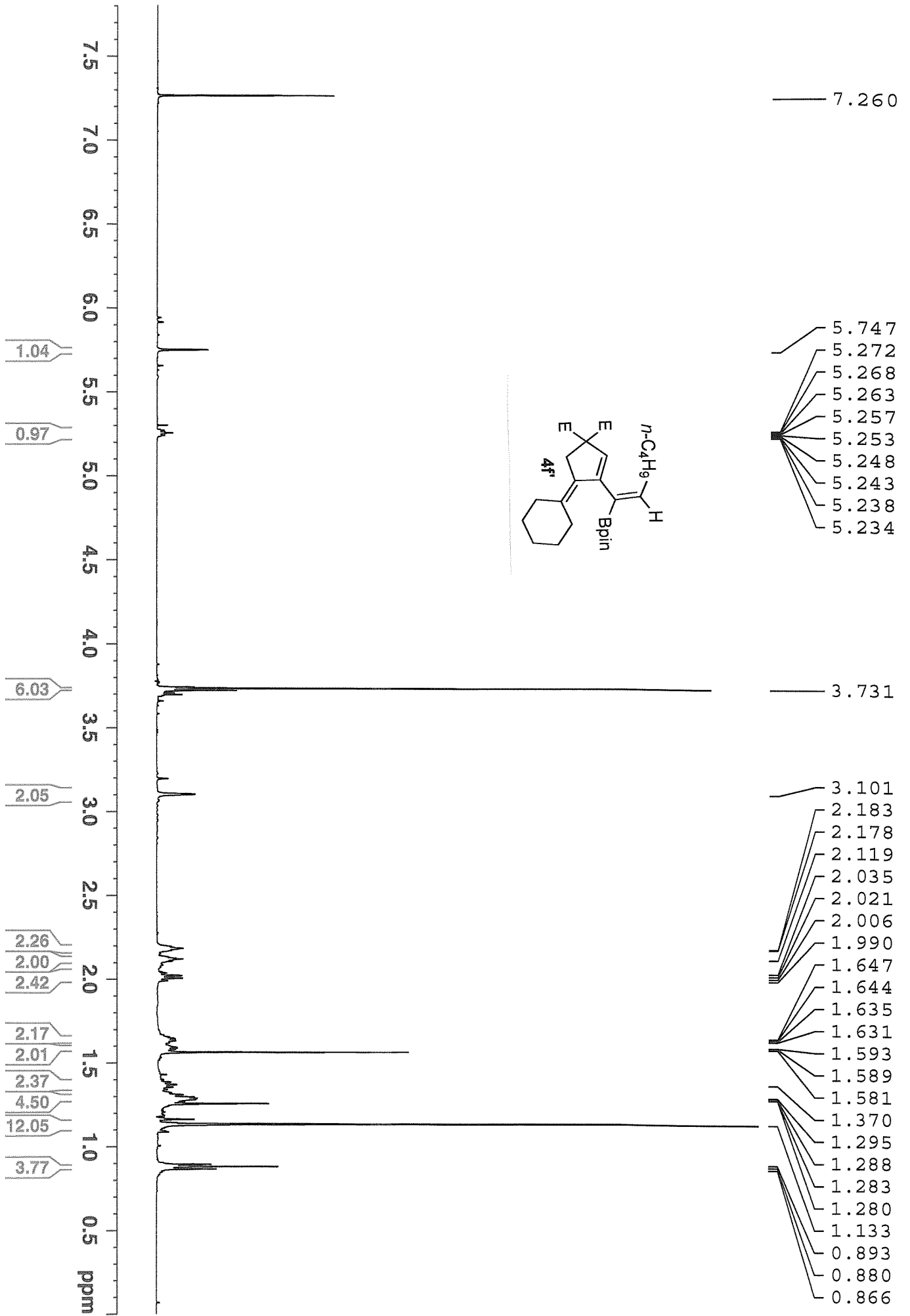
Acquisition Parameter

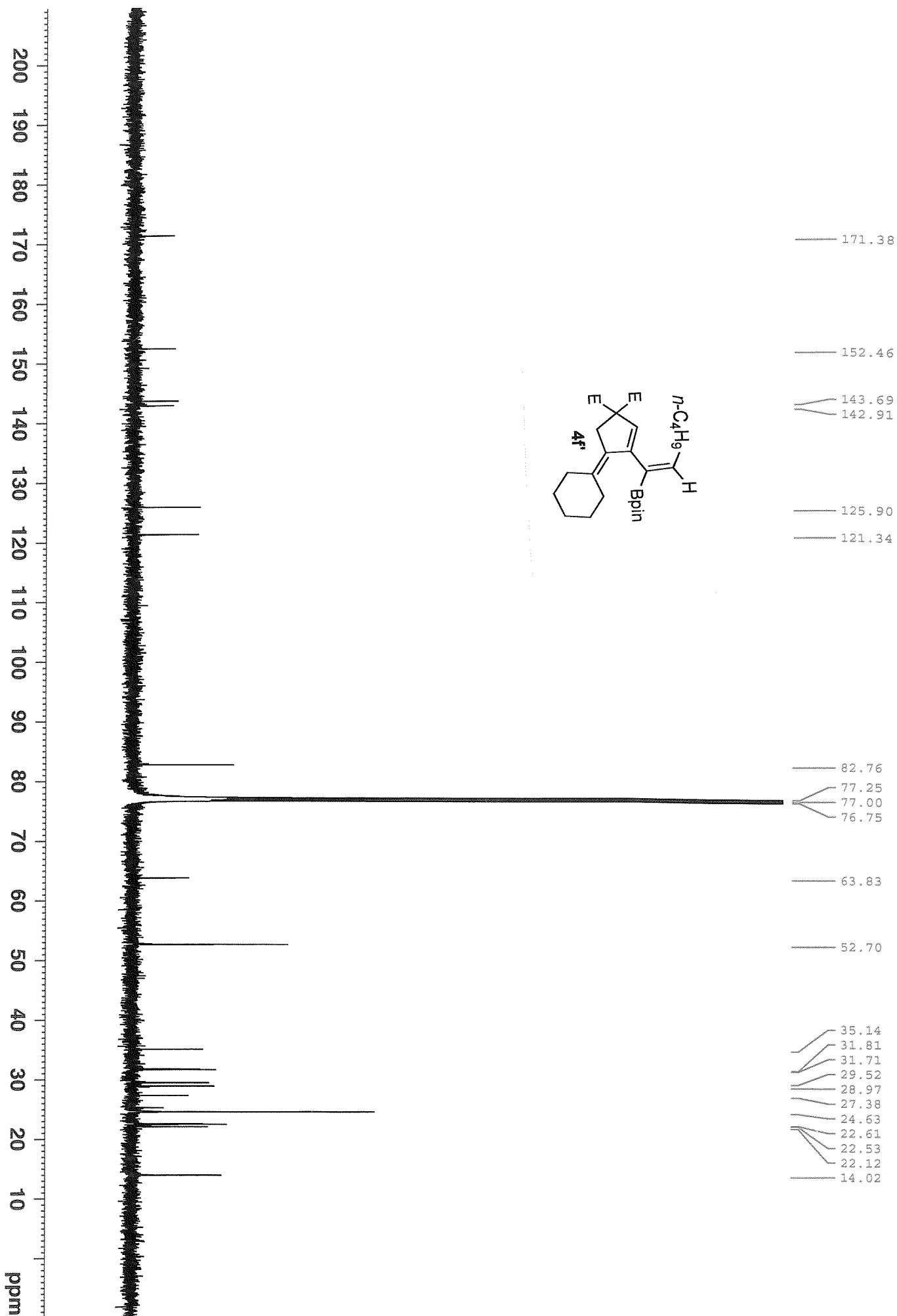
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 25 H 39 B Na O 6	469.2735	469.2736	0.3	0.4







Mass Spectrum SmartFormula Report

Analysis Info

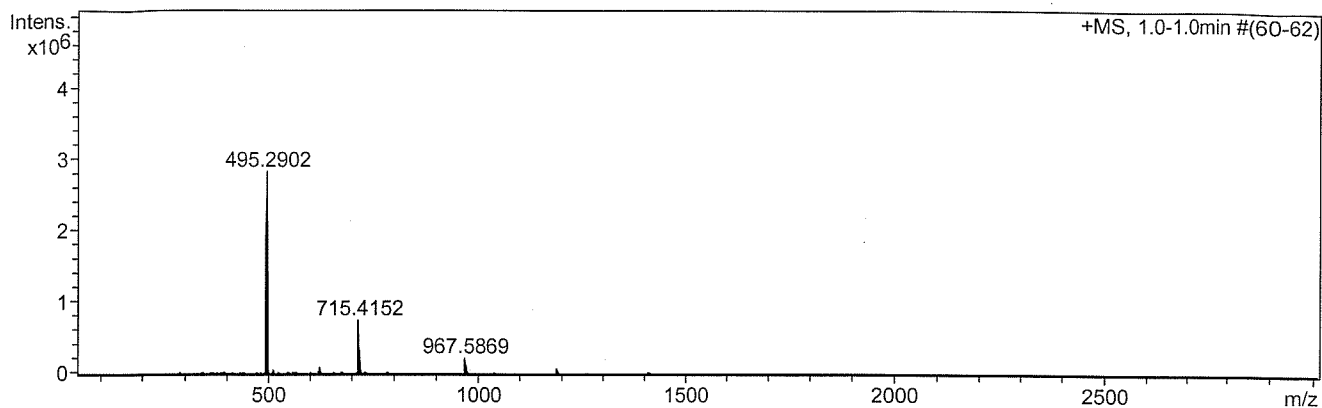
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Method Tune_wide_pos.m
Sample Name dyq-4-162-1
Comment

Acquisition Date 2012-11-16 14:00:49

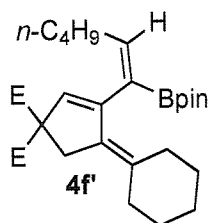
Operator Carin Larsson
Instrument / Ser# micrOTOF 125

Acquisition Parameter

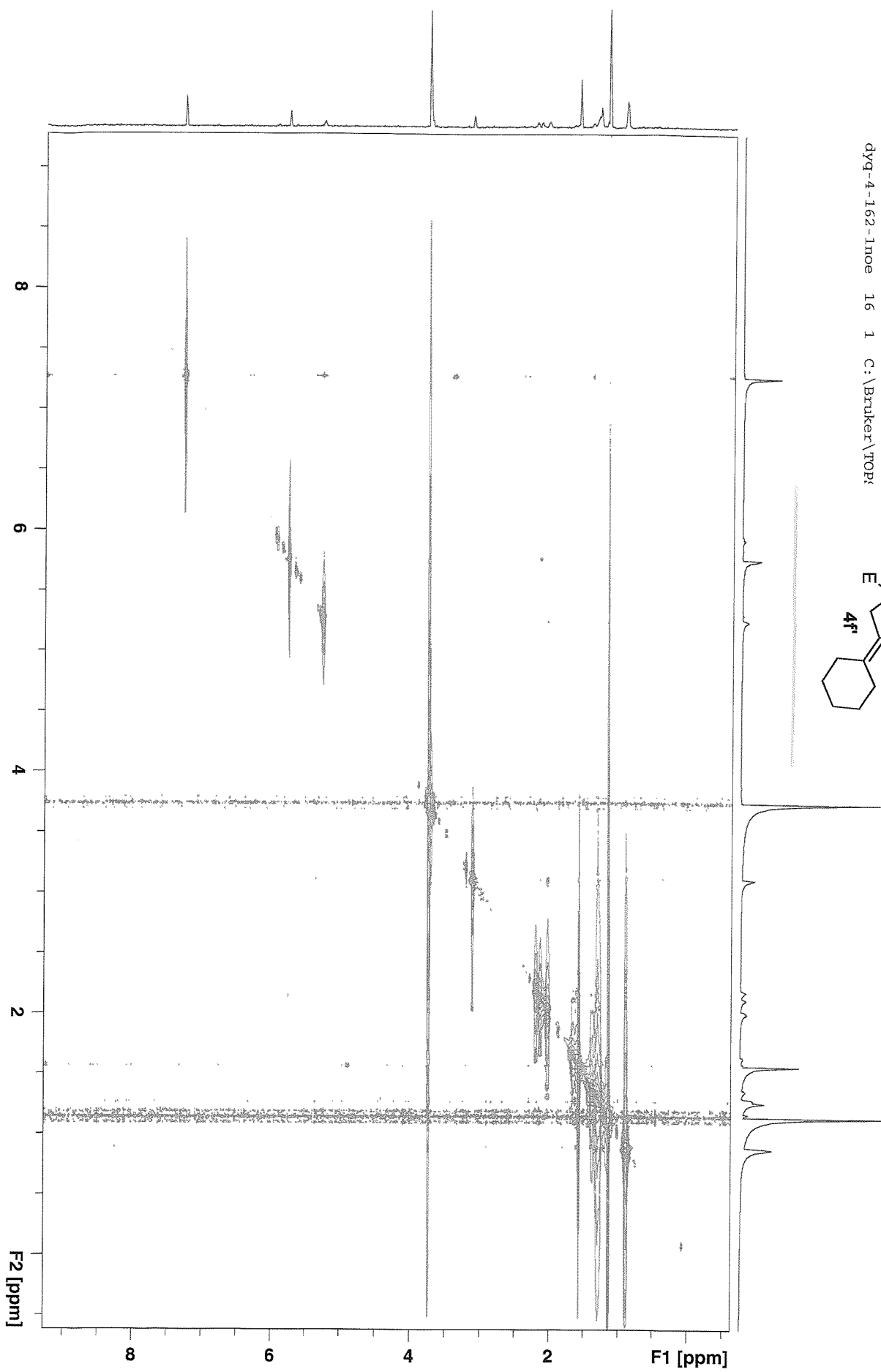
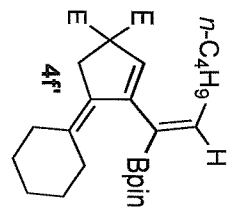
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source

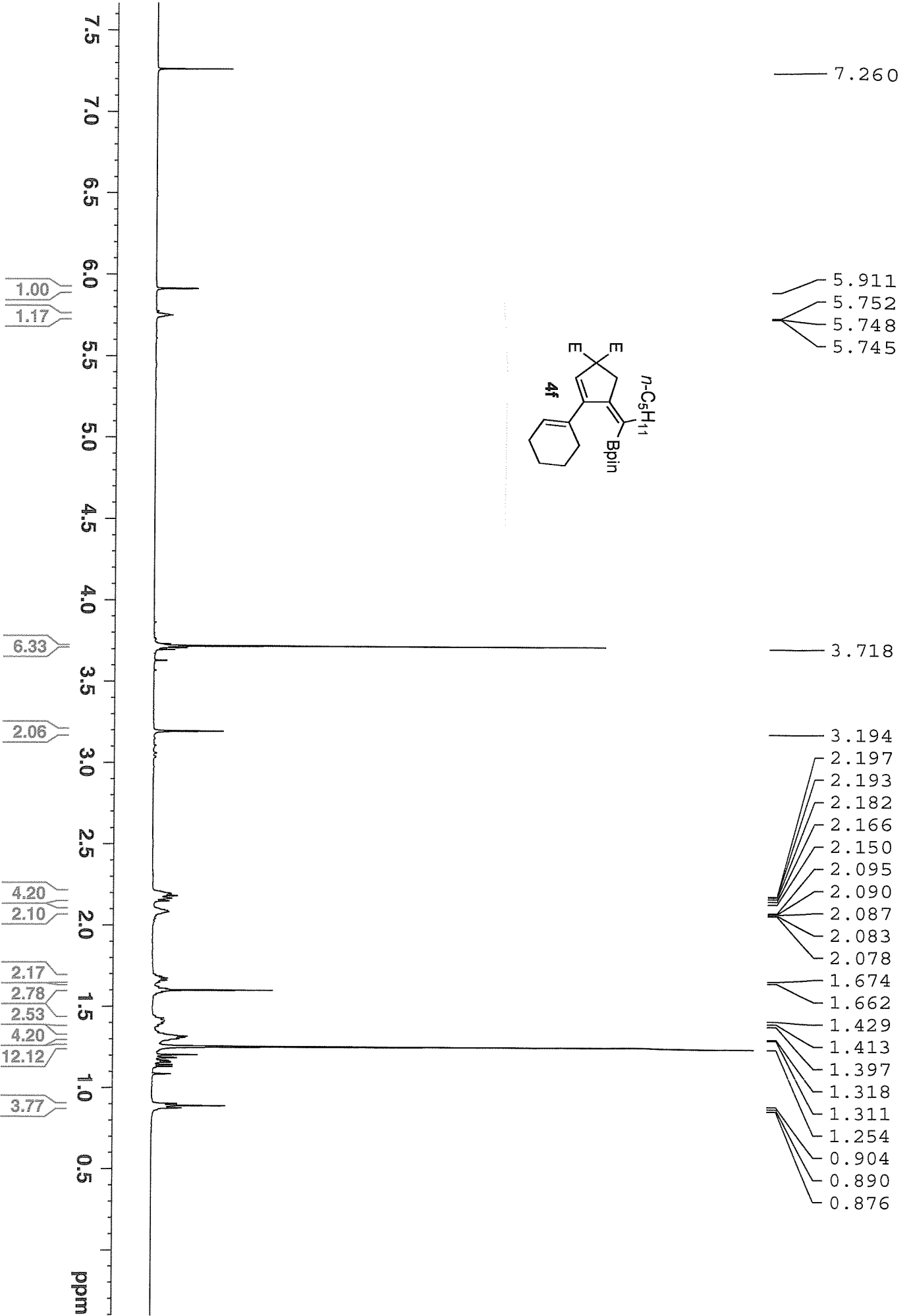


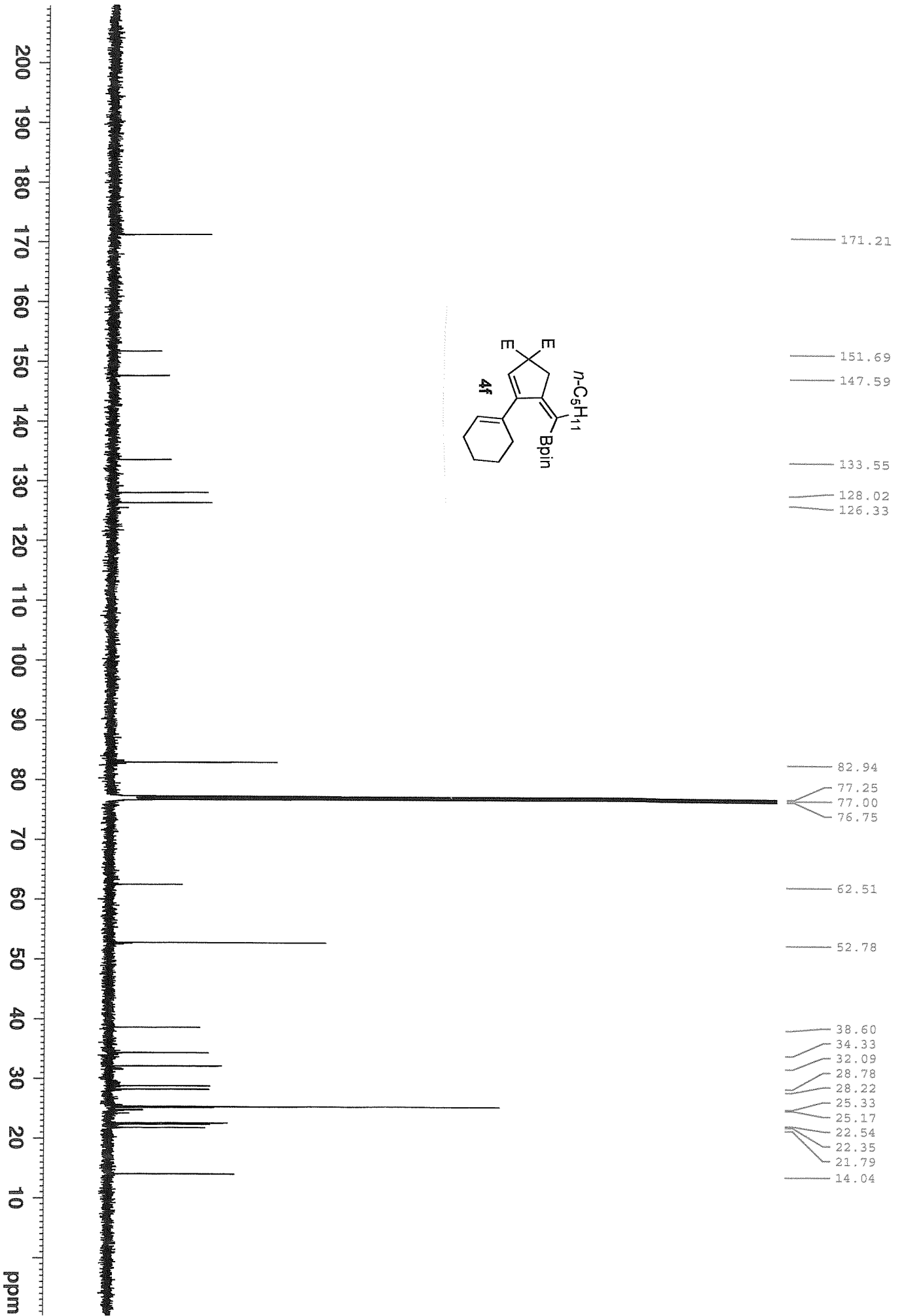
Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 27 H 41 B Na O 6	495.2902	495.2893	-1.9	-2.1



dyg-4-162-Inoe 16 1 C:\Bruker\TOPK







Mass Spectrum SmartFormula Report

Analysis Info

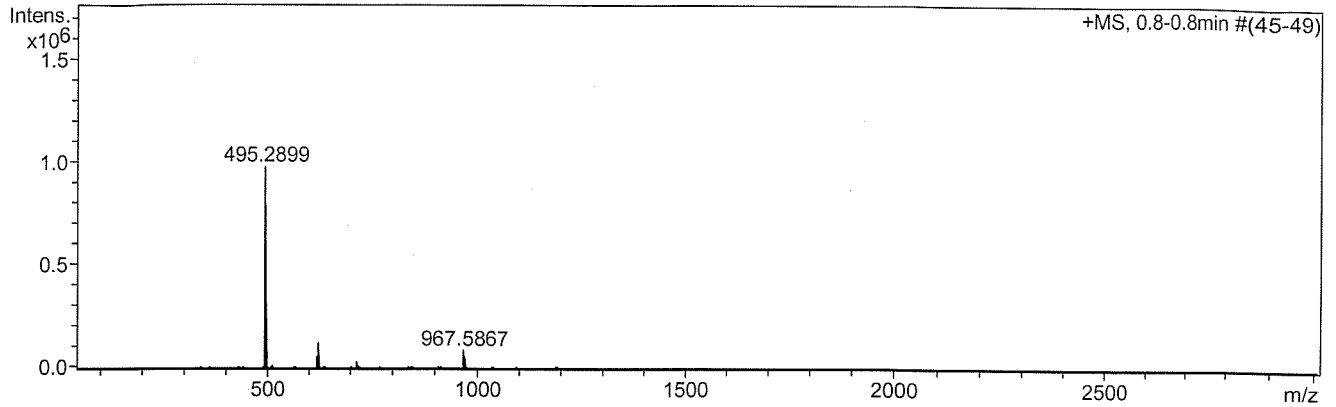
Analysis Name E:\Data2\Youqian\dyq-4-162-2000001.d
Method Tune_wide_pos.m
Sample Name dyq-4-162-2
Comment

Acquisition Date 2012-11-16 14:06:58

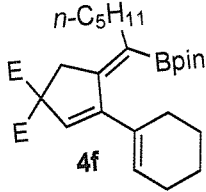
Operator Carin Larsson
Instrument / Ser# micrOTOF 125

Acquisition Parameter

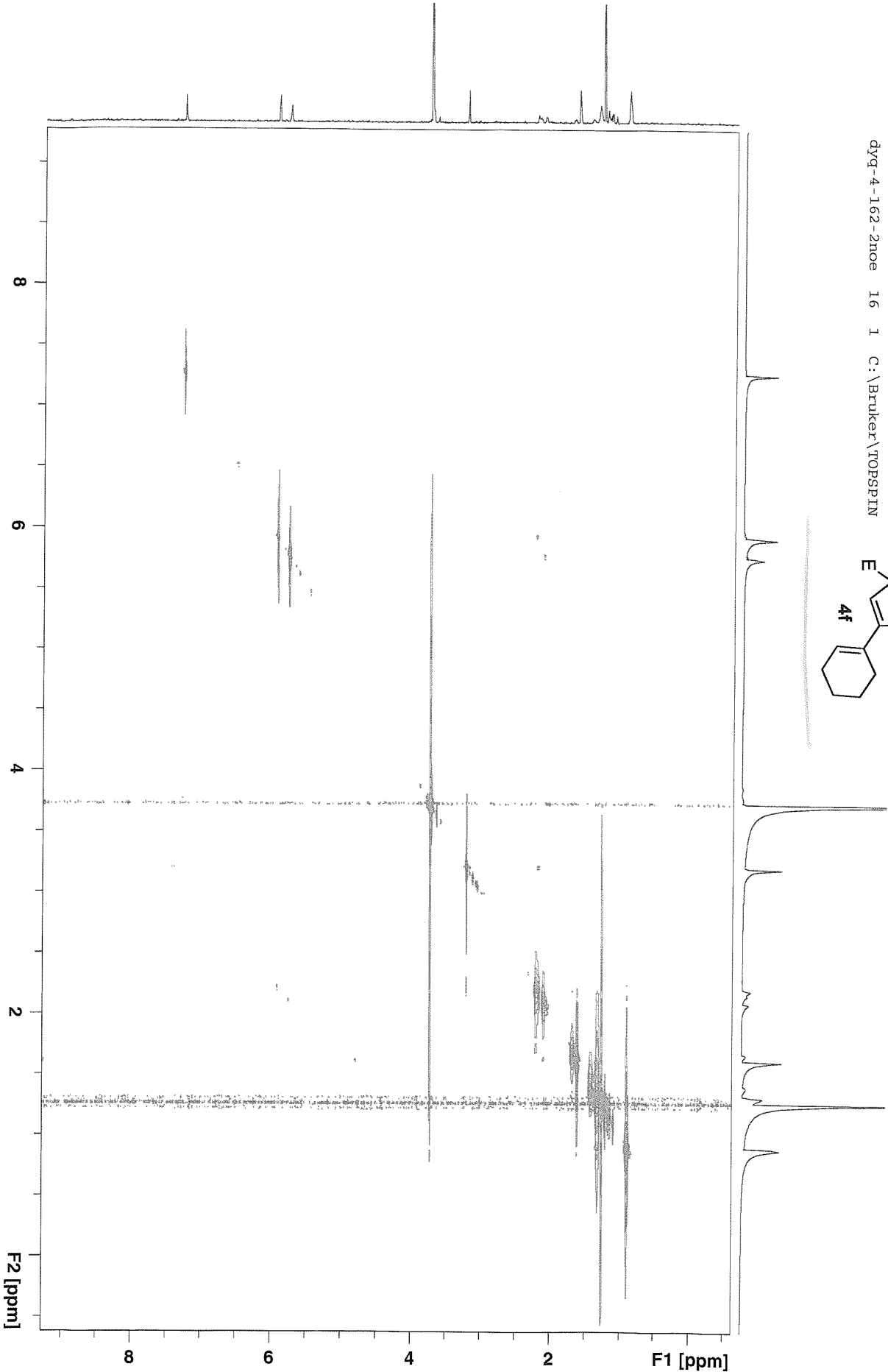
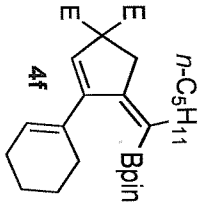
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.3 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source

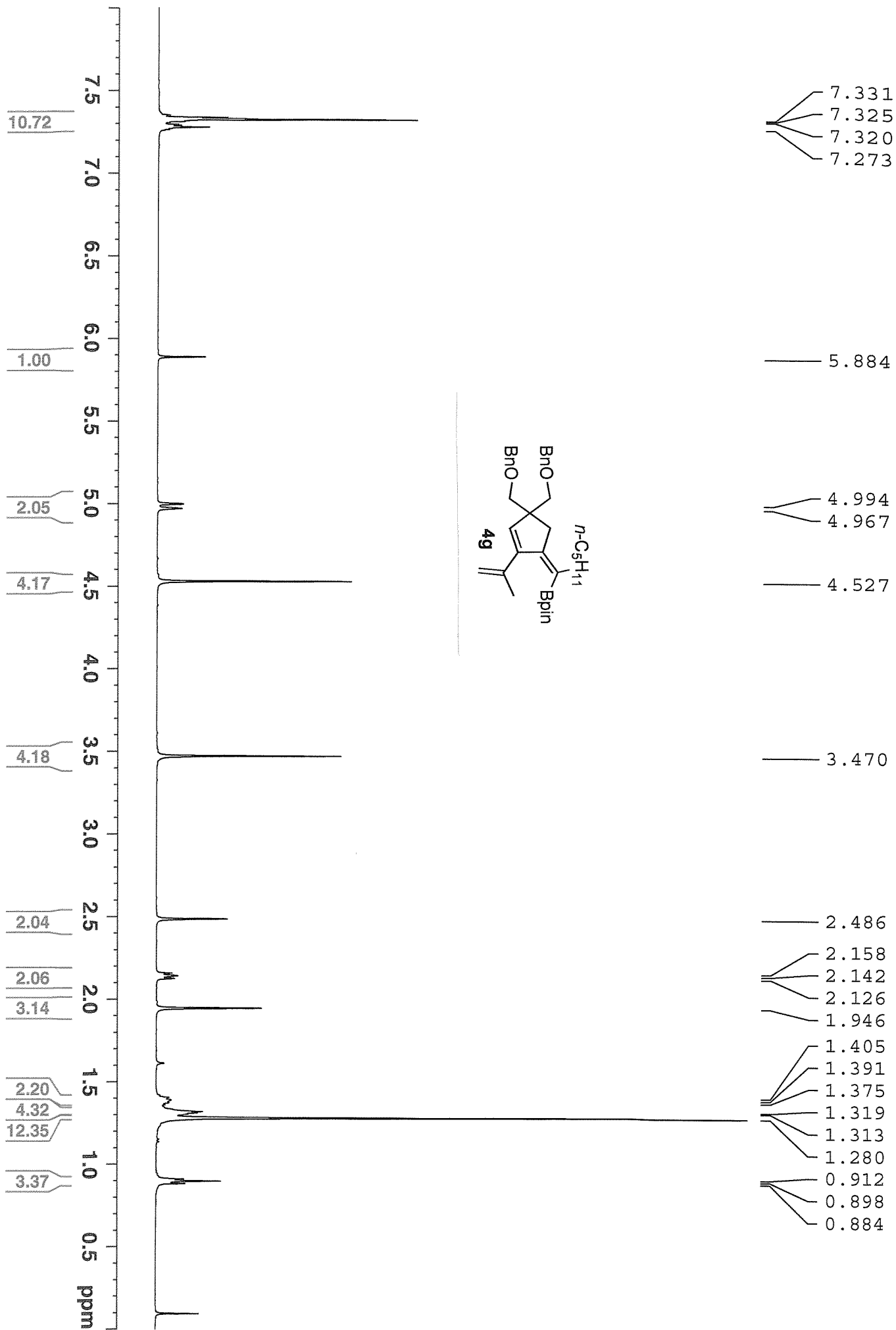


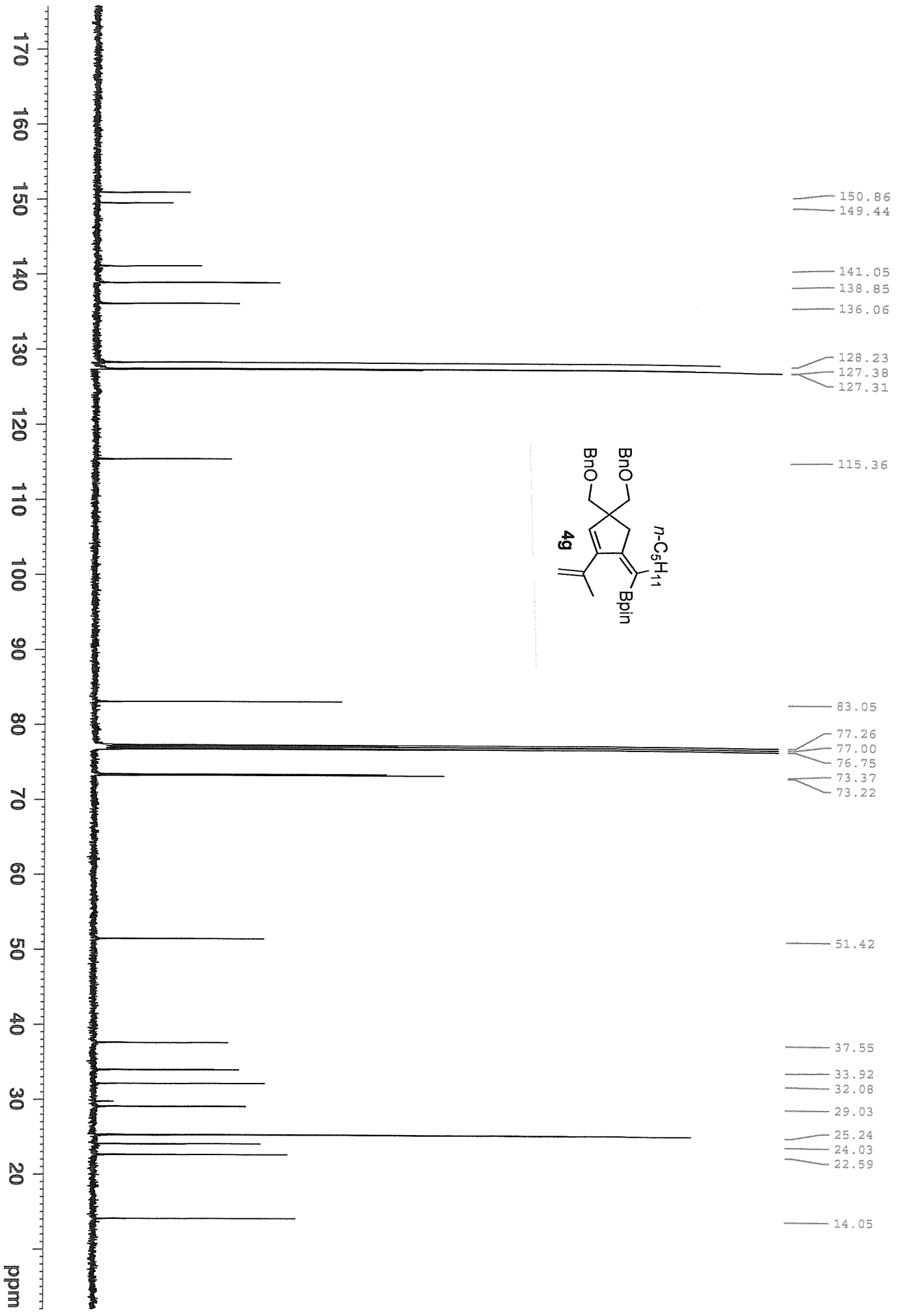
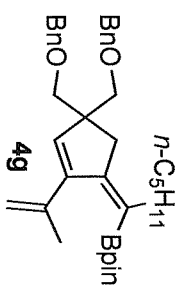
Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 27 H 41 B Na O 6	495.2899	495.2893	-1.2	-3.0



dyg-4-162-2noe 16 1 C:\Bruker\TOPSPIN







Mass Spectrum SmartFormula Report

Analysis Info

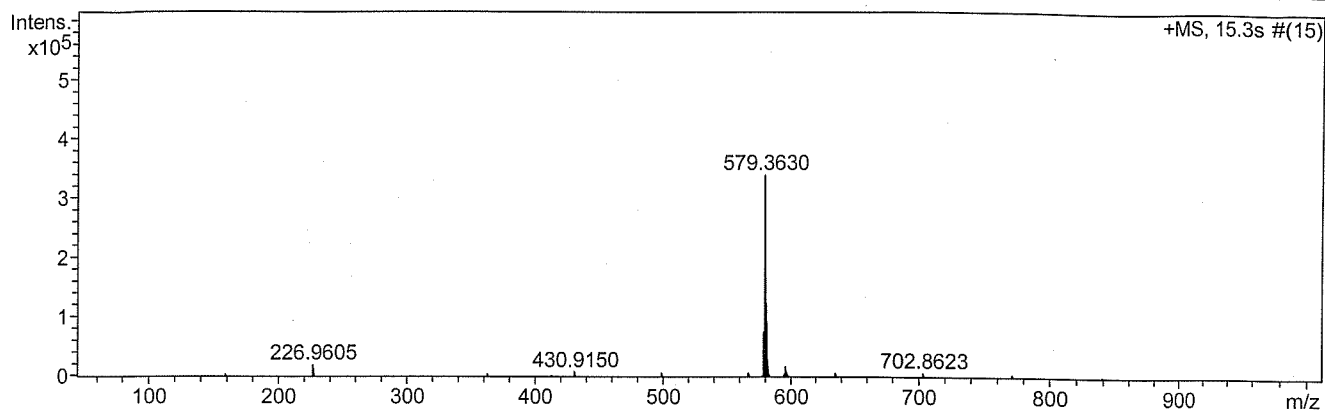
Analysis Name E:\Data2\Youqian\Teb-c-59000001.d
Method Tune_low_pos.m
Sample Name Teb-c-59
Comment

Acquisition Date 2012-12-13 15:42:26

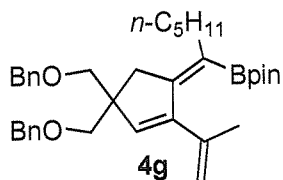
Operator Carin Larsson
Instrument / Ser# micrOTOF 125

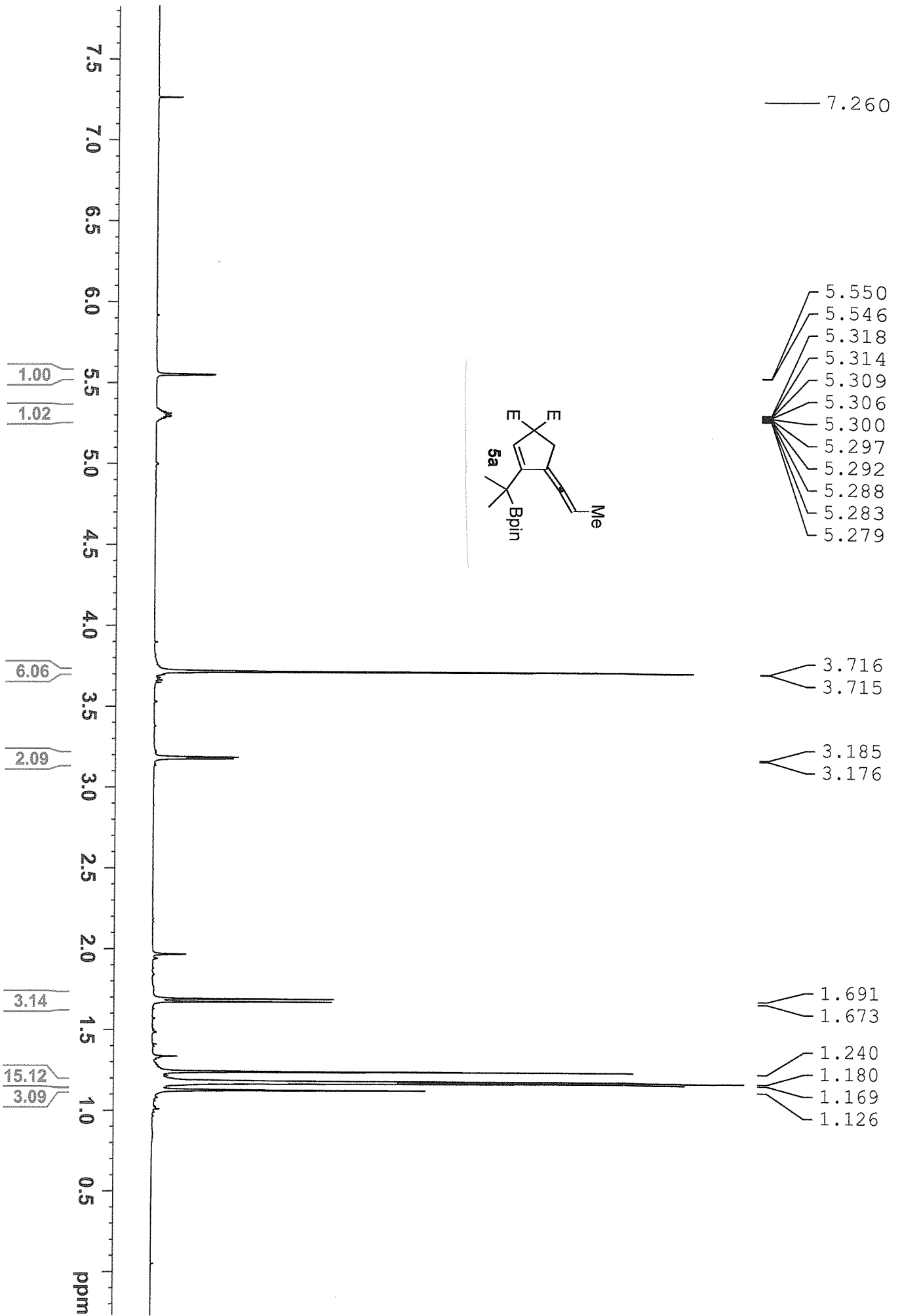
Acquisition Parameter

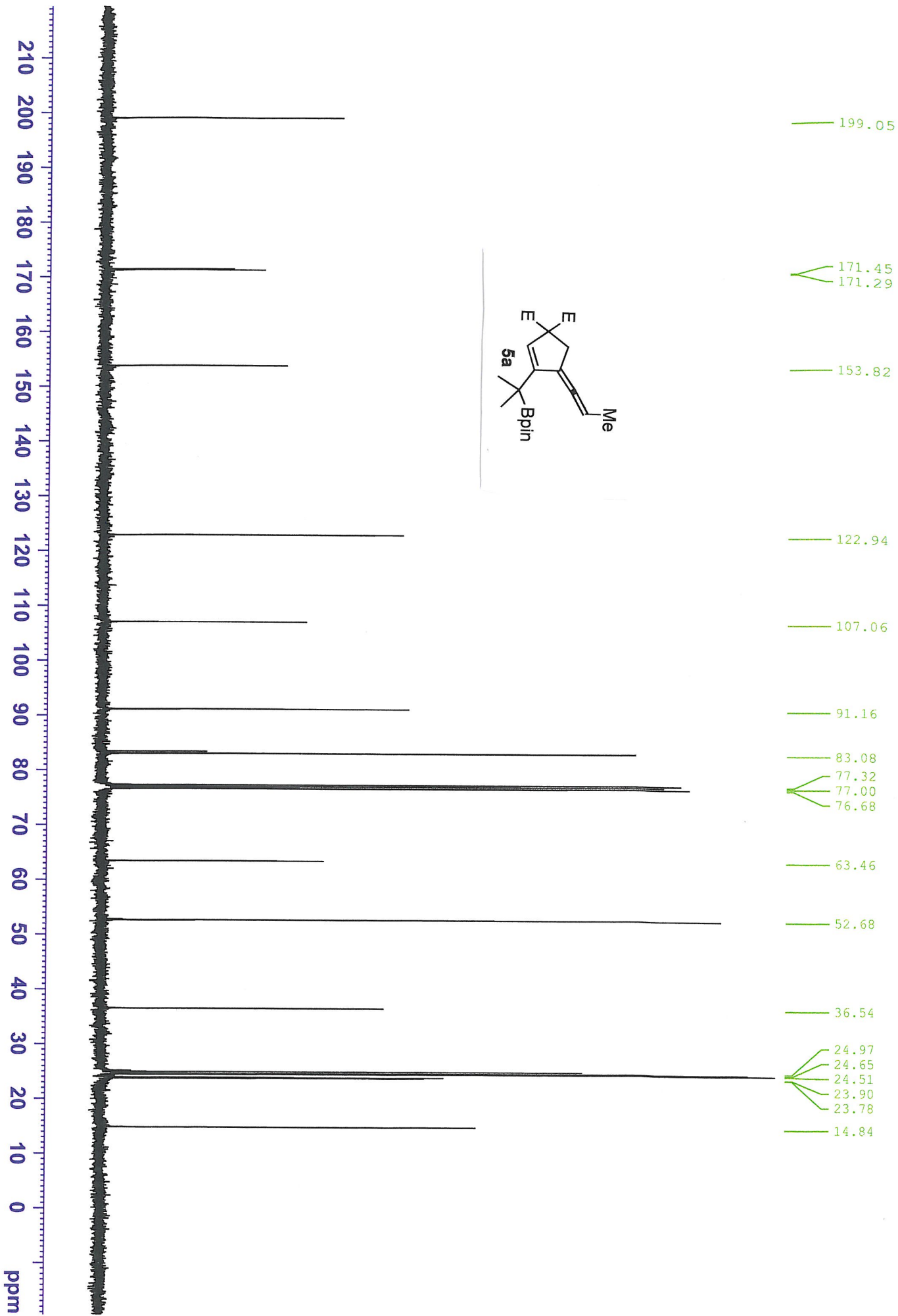
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4000 V	Set Dry Gas	4.0 l/min
Scan End	1000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 36 H 49 B Na O 4	579.3630	579.3622	-1.4	-1.0







Mass Spectrum SmartFormula Report

Analysis Info

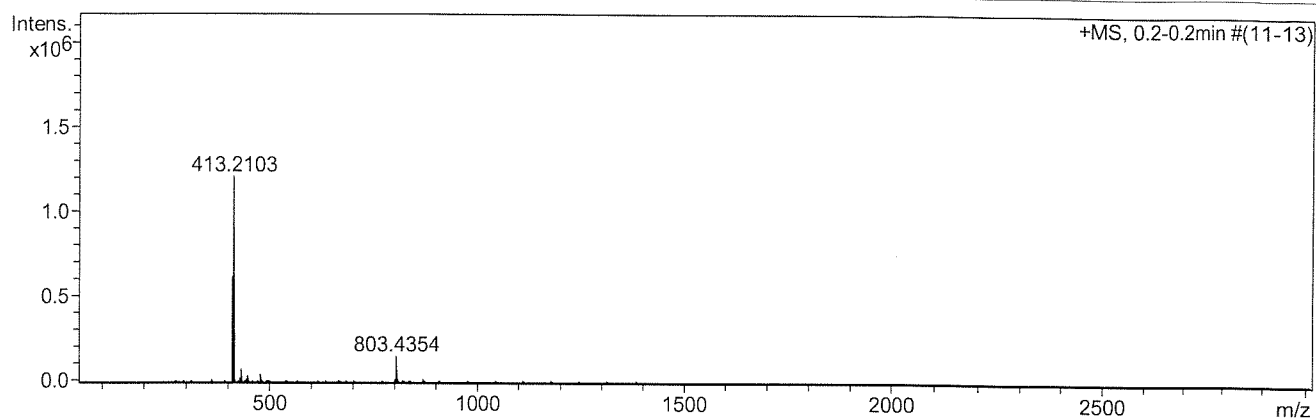
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Method tune_wide_dirk.m
Sample Name
Comment

Acquisition Date 2012-08-15 14:35:56

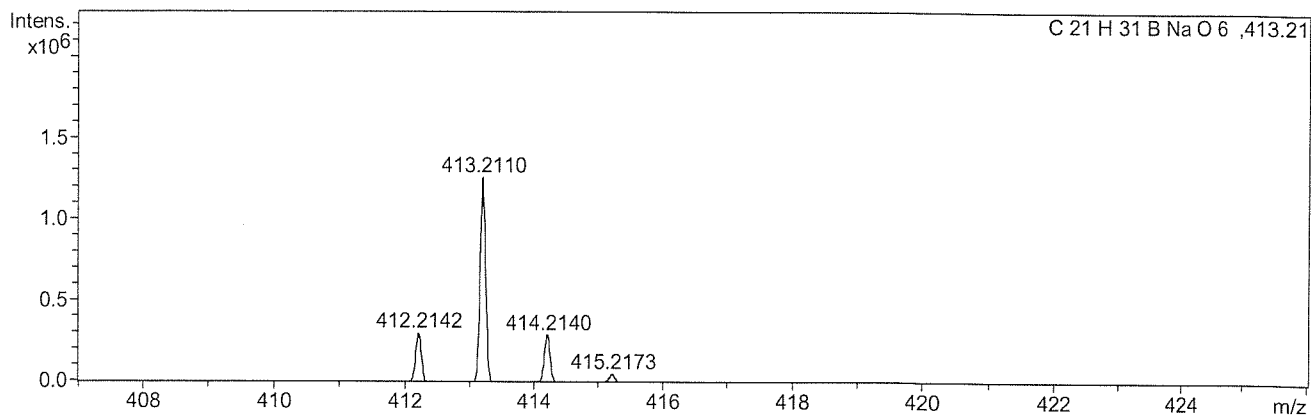
Operator pia
Instrument / Ser# micrOTOF 125

Acquisition Parameter

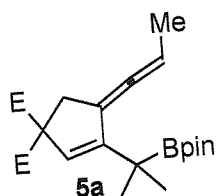
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Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source

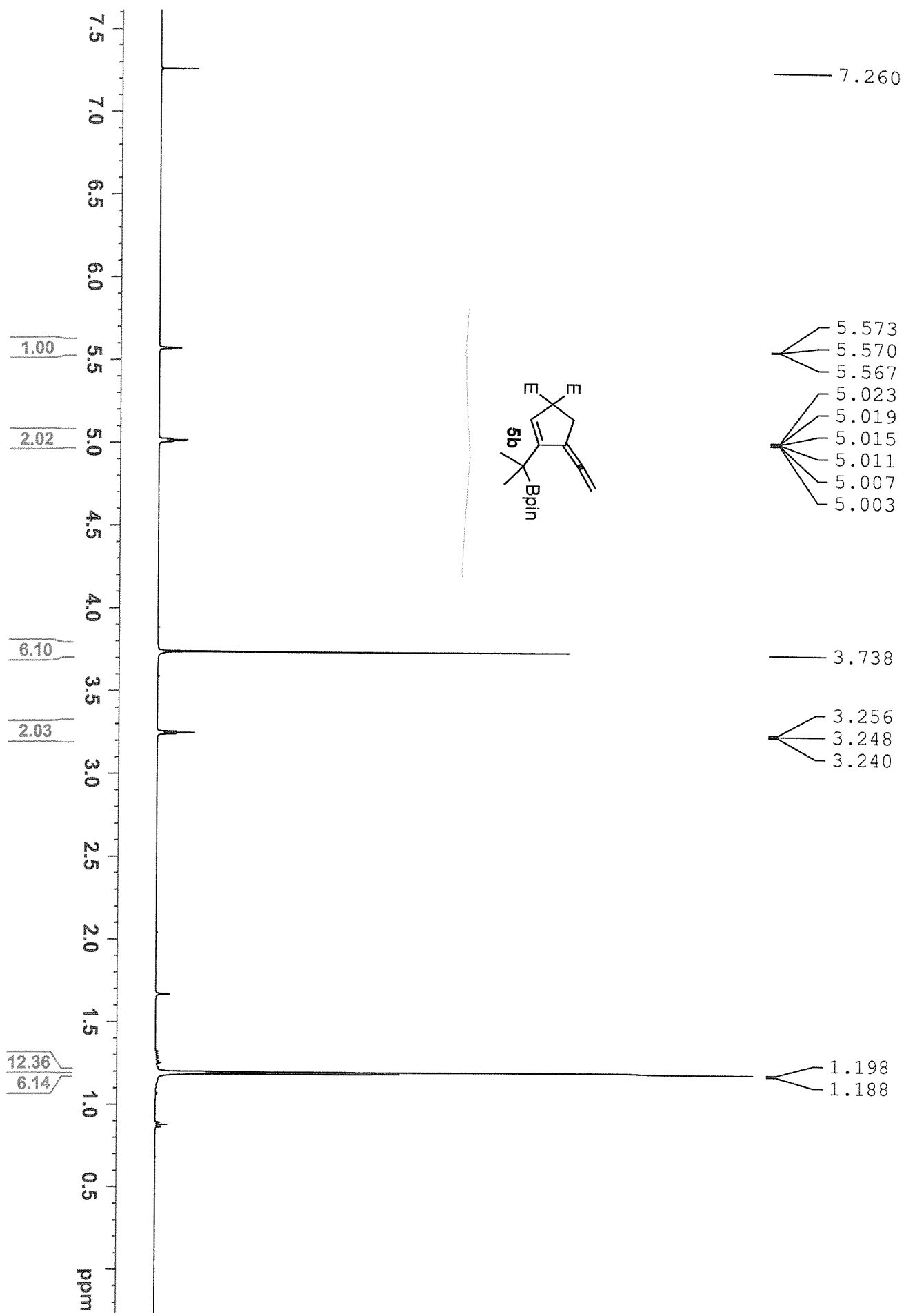


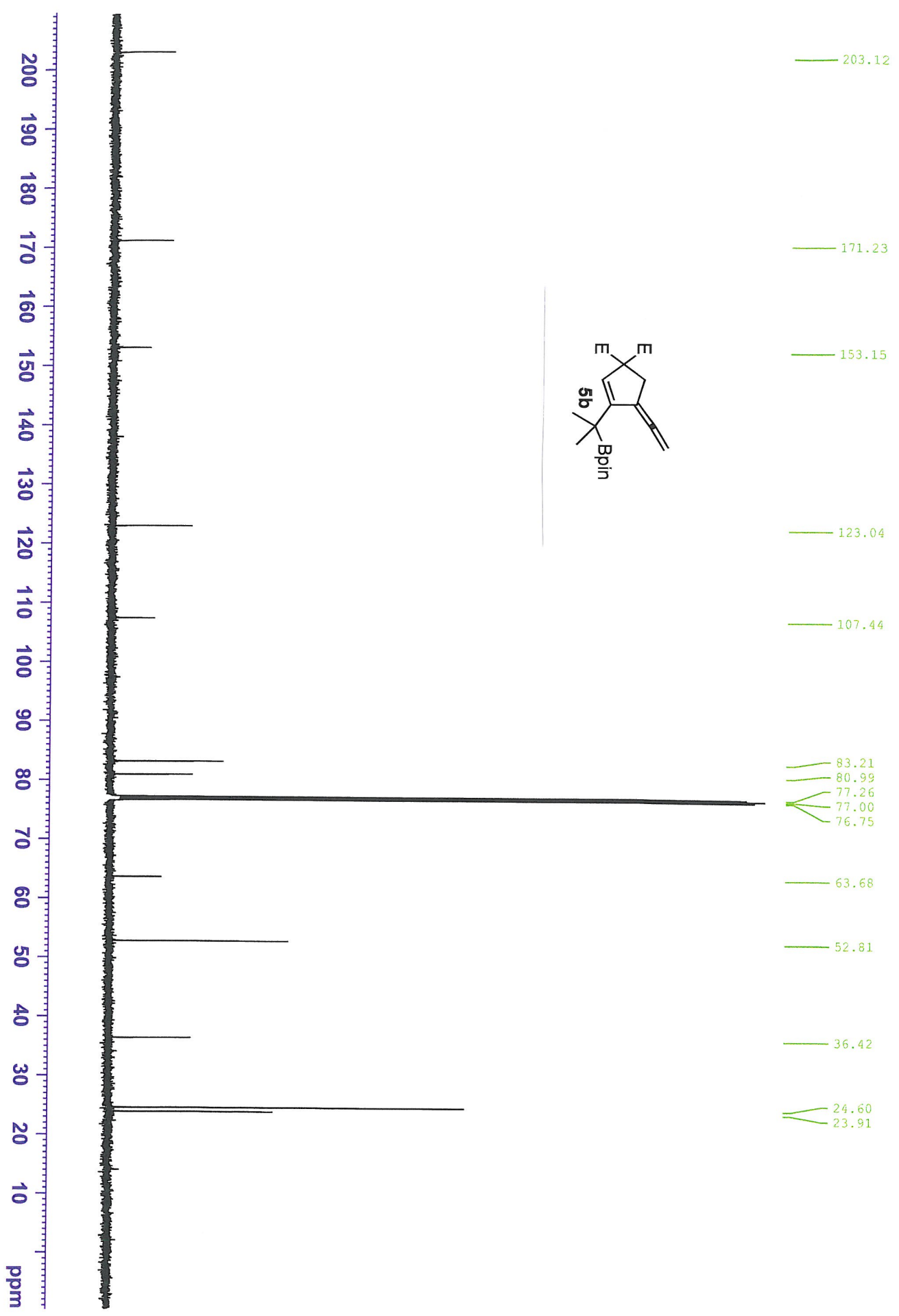
Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C ₂₁ H ₃₁ BNaO ₆	413.2103	413.2106	0.7	4.4



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
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Mass Spectrum SmartFormula Report

Analysis Info

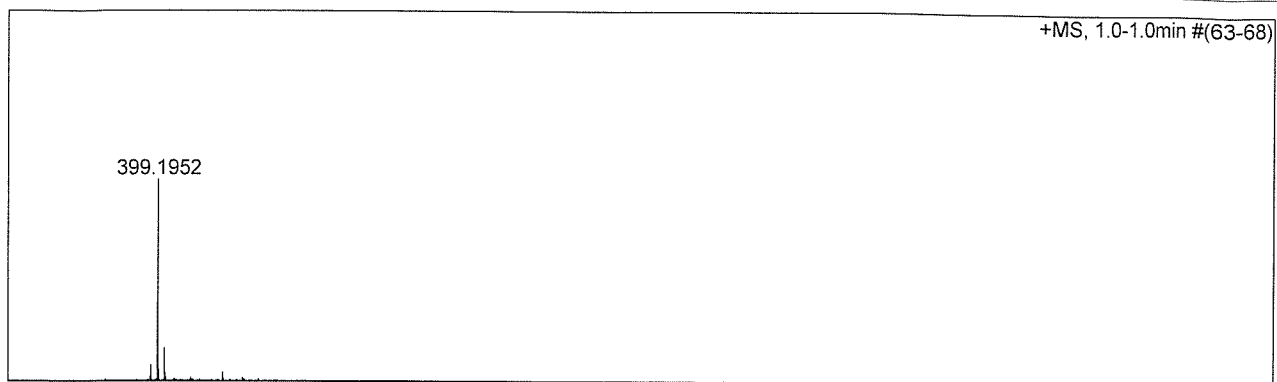
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Method tune_wide_dirk.m
Sample Name dyq-3-33-2
Comment

Acquisition Date 2011-12-07 11:36:40

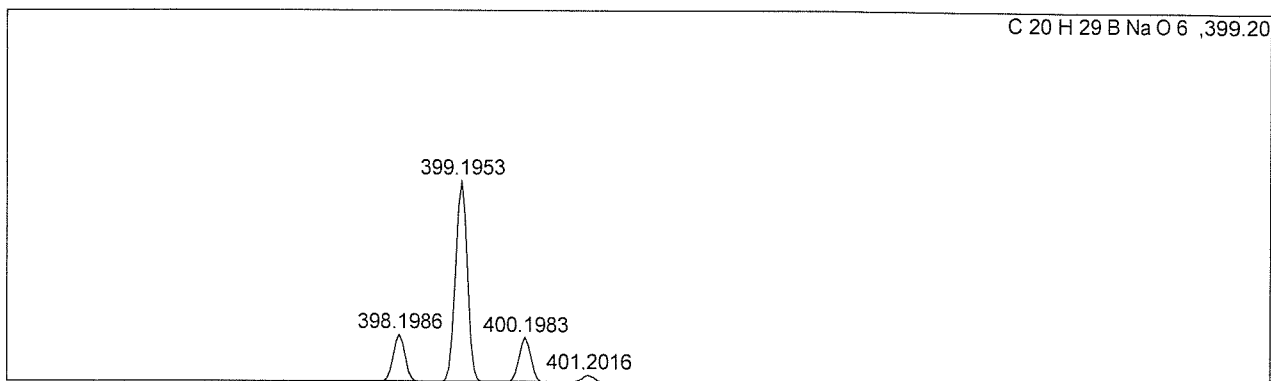
Operator pia
Instrument / Ser# micrOTOF 125

Acquisition Parameter

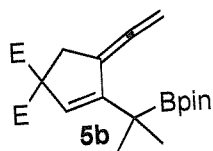
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source

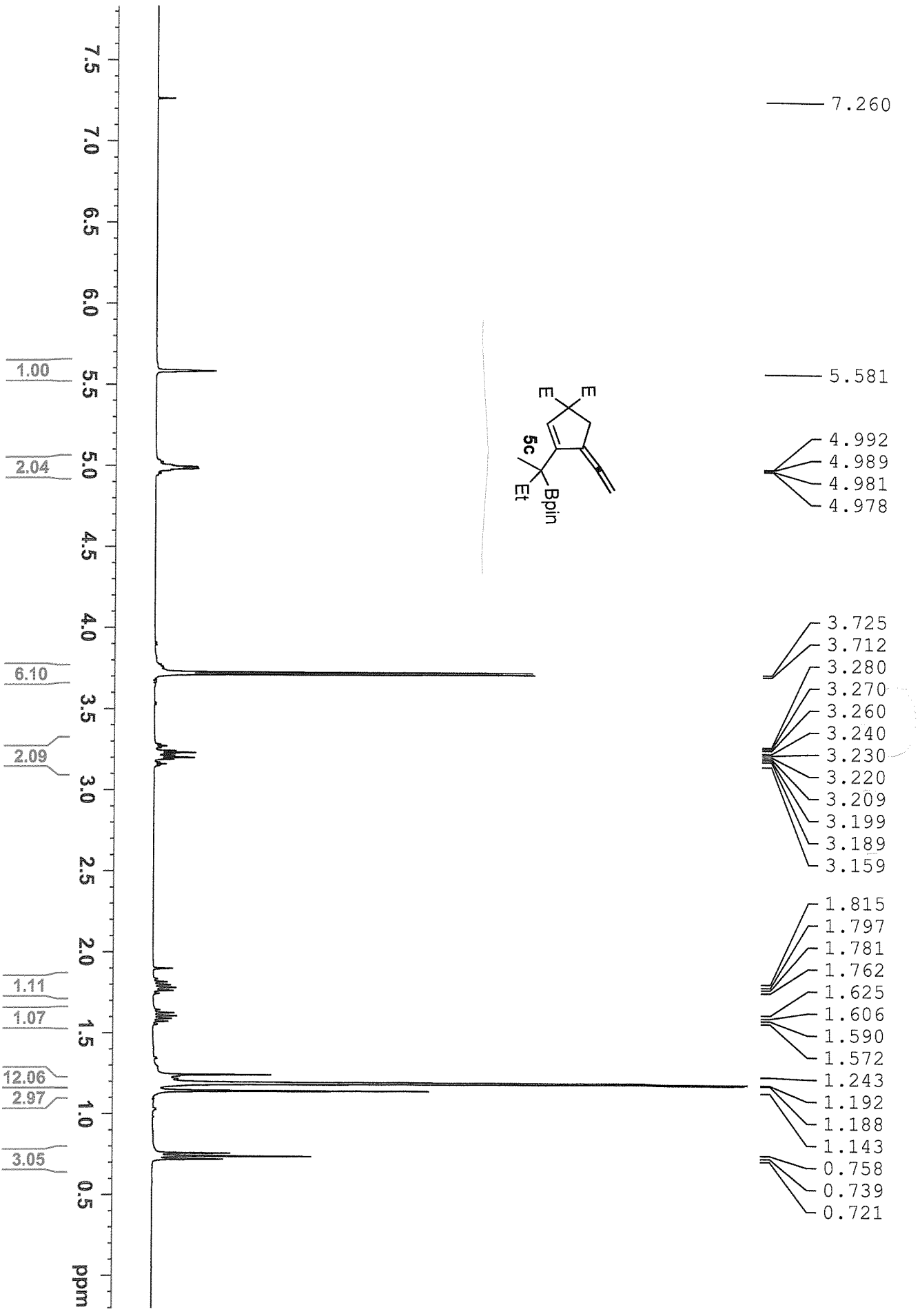


Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C ₂₀ H ₂₉ BNaO ₆	399.1952	399.1949	-0.6	4.7

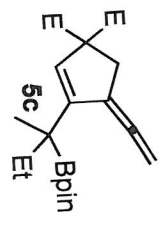
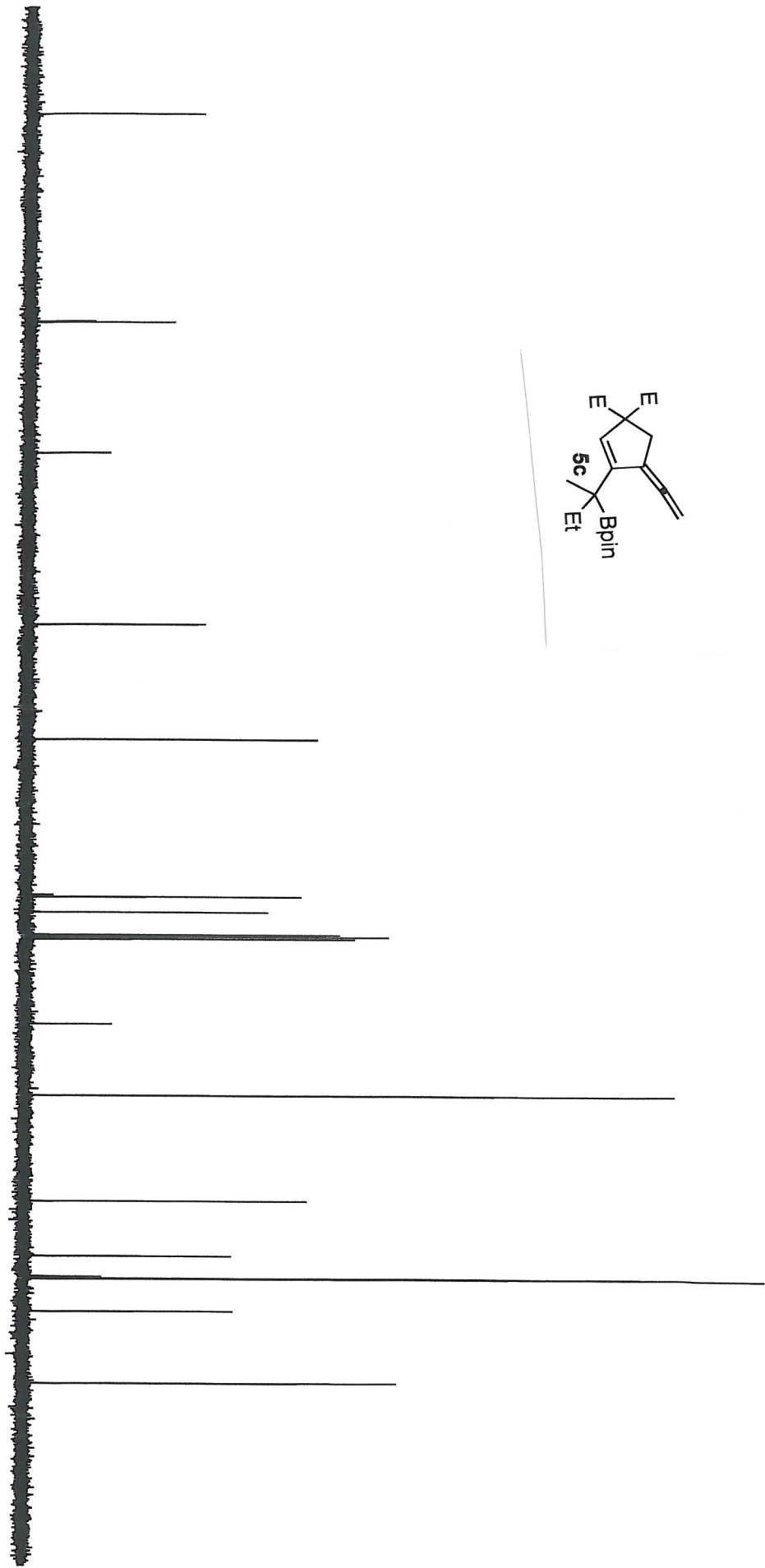


Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
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210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 ppm



- 203.00
- 171.26
- 171.08
- 151.10
- 124.83
- 107.23
- 83.13
- 80.78
- 77.32
- 77.00
- 76.68
- 63.66
- 52.71
- 36.53
- 28.09
- 24.98
- 24.70
- 24.62
- 19.62
- 8.58

Mass Spectrum SmartFormula Report

Analysis Info

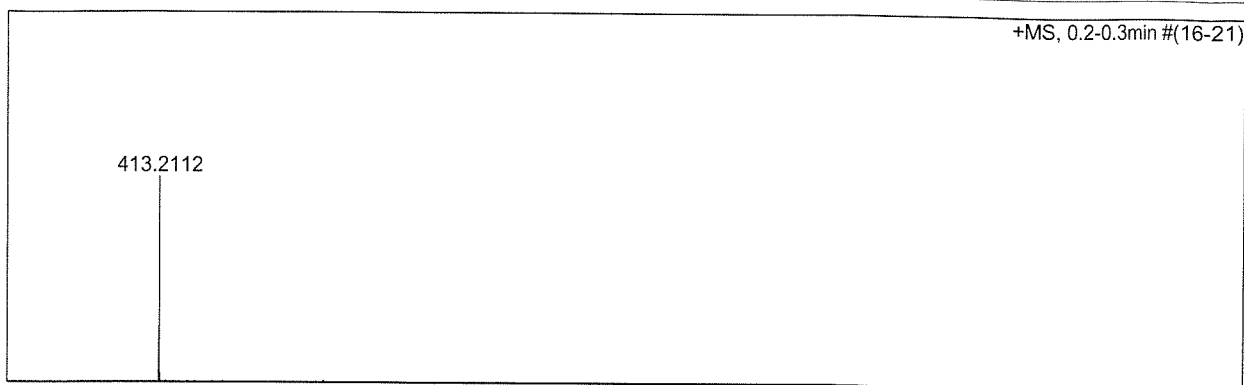
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Method tune_wide_dirk.m
Sample Name dyq-3-179
Comment

Acquisition Date 2012-04-30 10:37:06

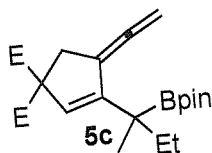
Operator pia
Instrument / Ser# micrOTOF 125

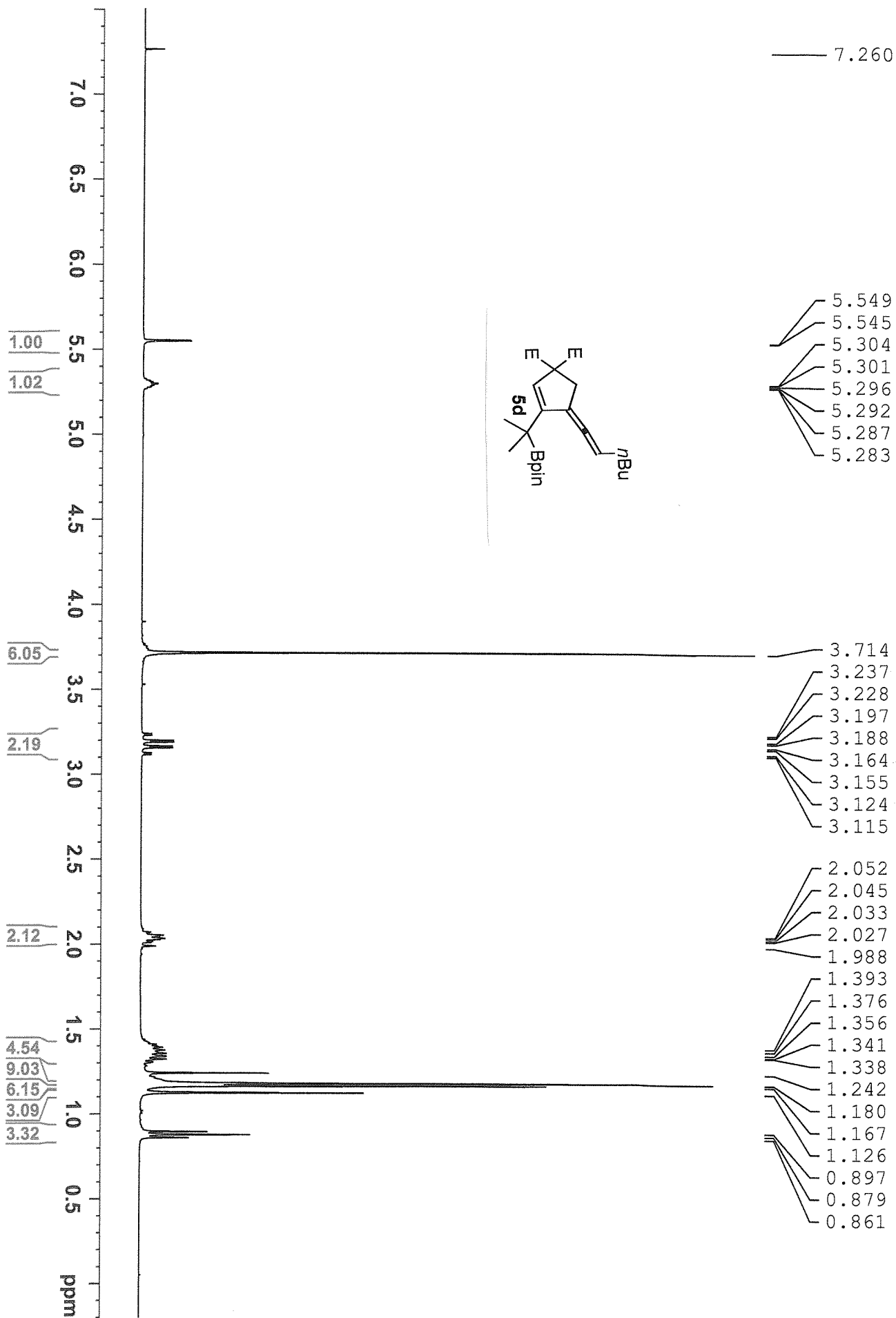
Acquisition Parameter

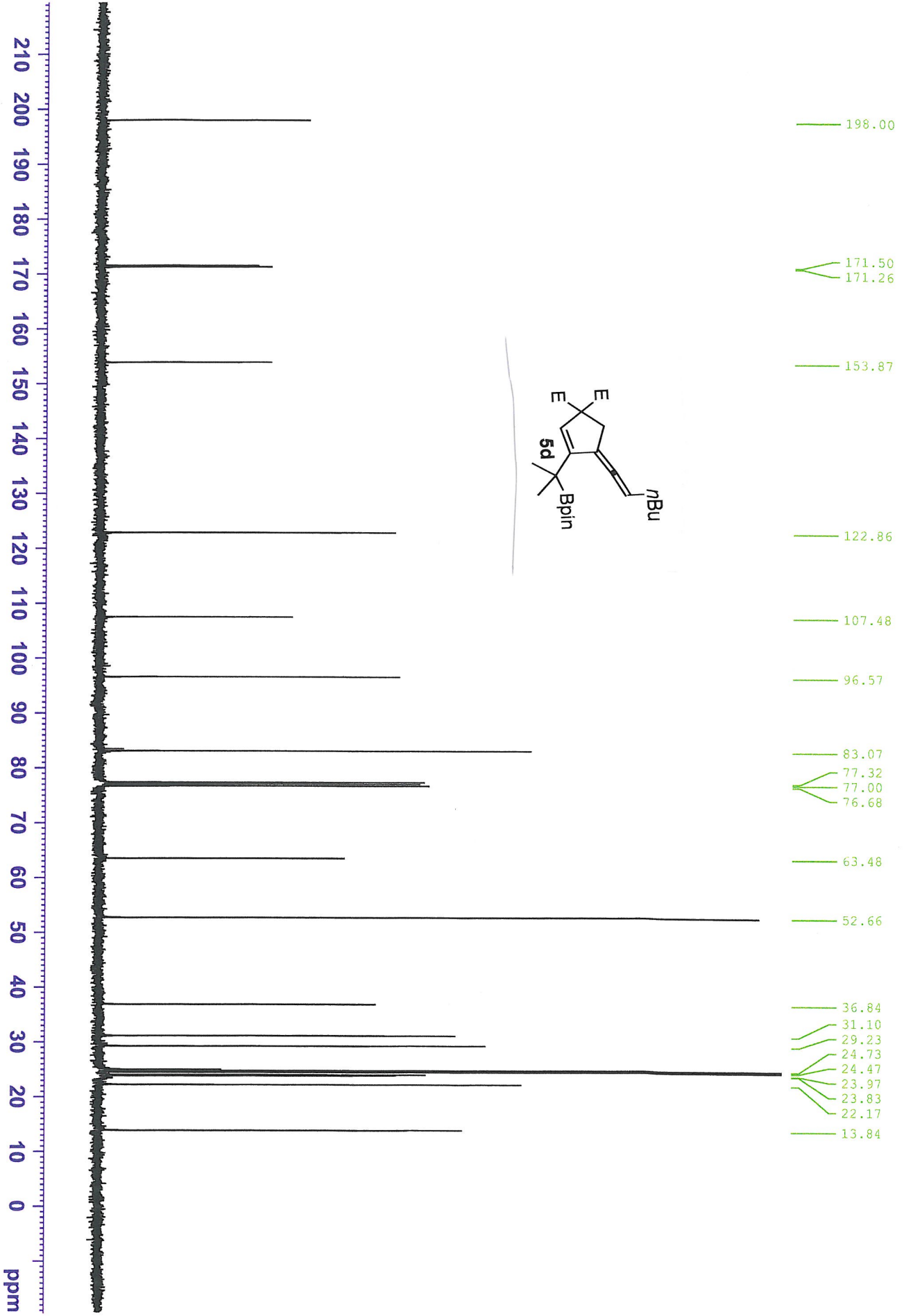
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C ₂₁ H ₃₁ BNaO ₆	413.2112	413.2106	-1.4	1.4







Mass Spectrum SmartFormula Report

Analysis Info

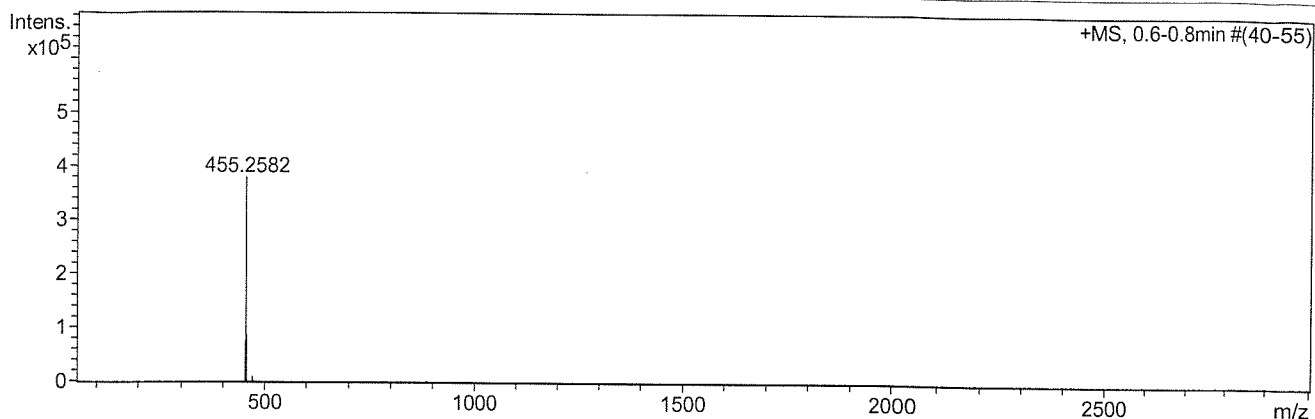
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Method tune_wide_dirk.m
Sample Name dyq-3-167
Comment

Acquisition Date 2012-04-26 18:08:48

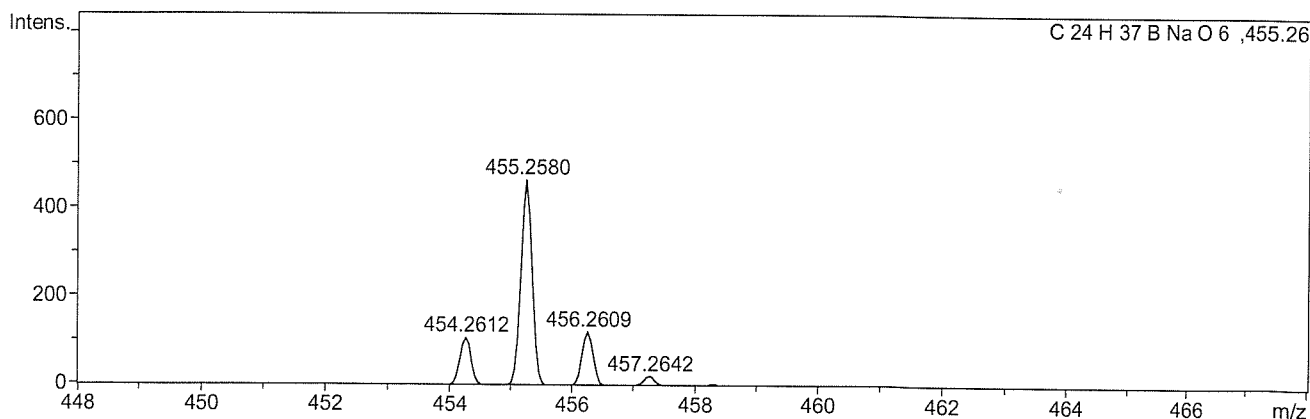
Operator pia
Instrument / Ser# micrOTOF 125

Acquisition Parameter

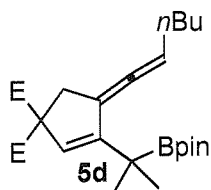
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source

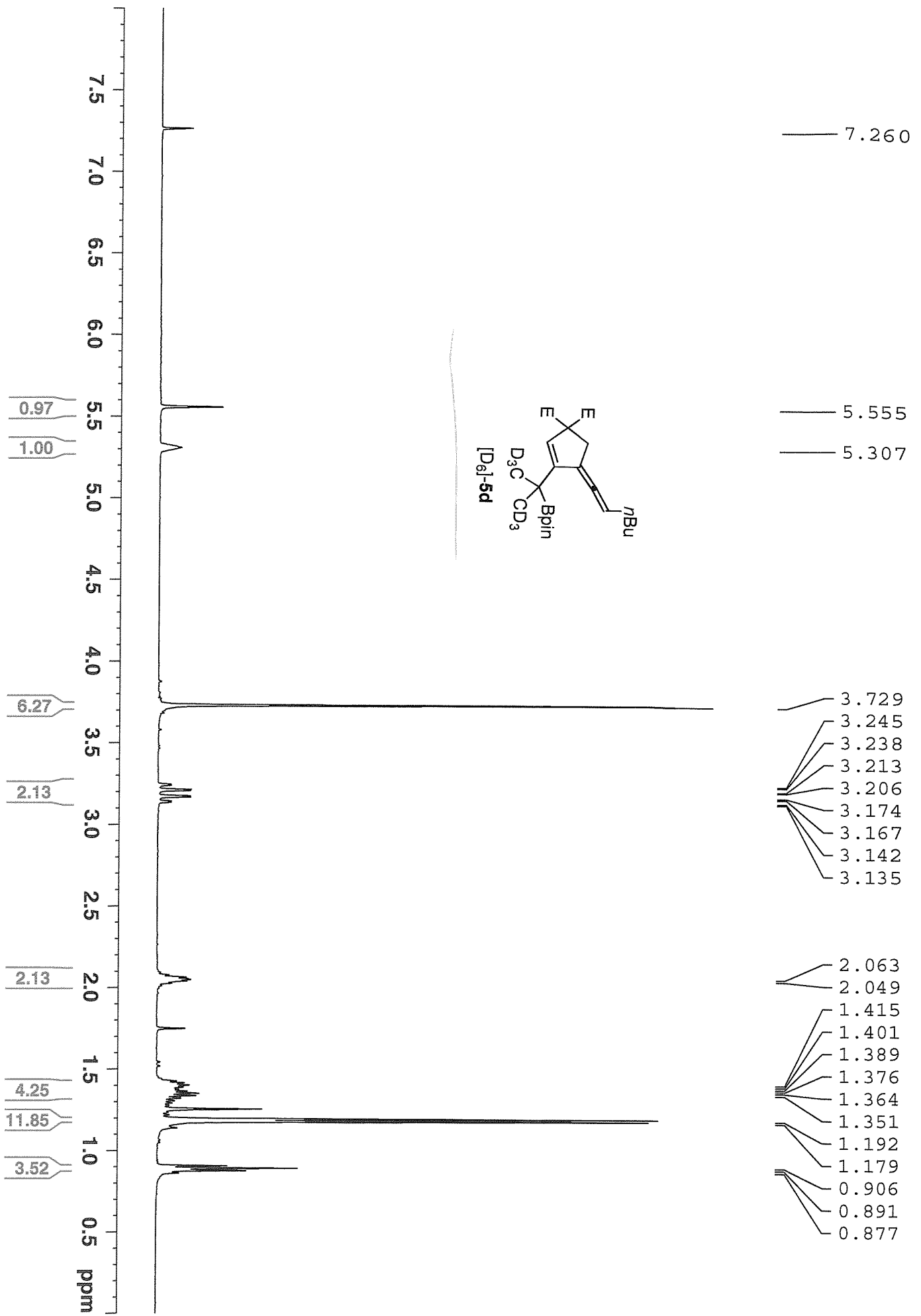
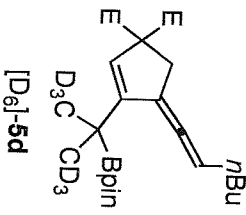


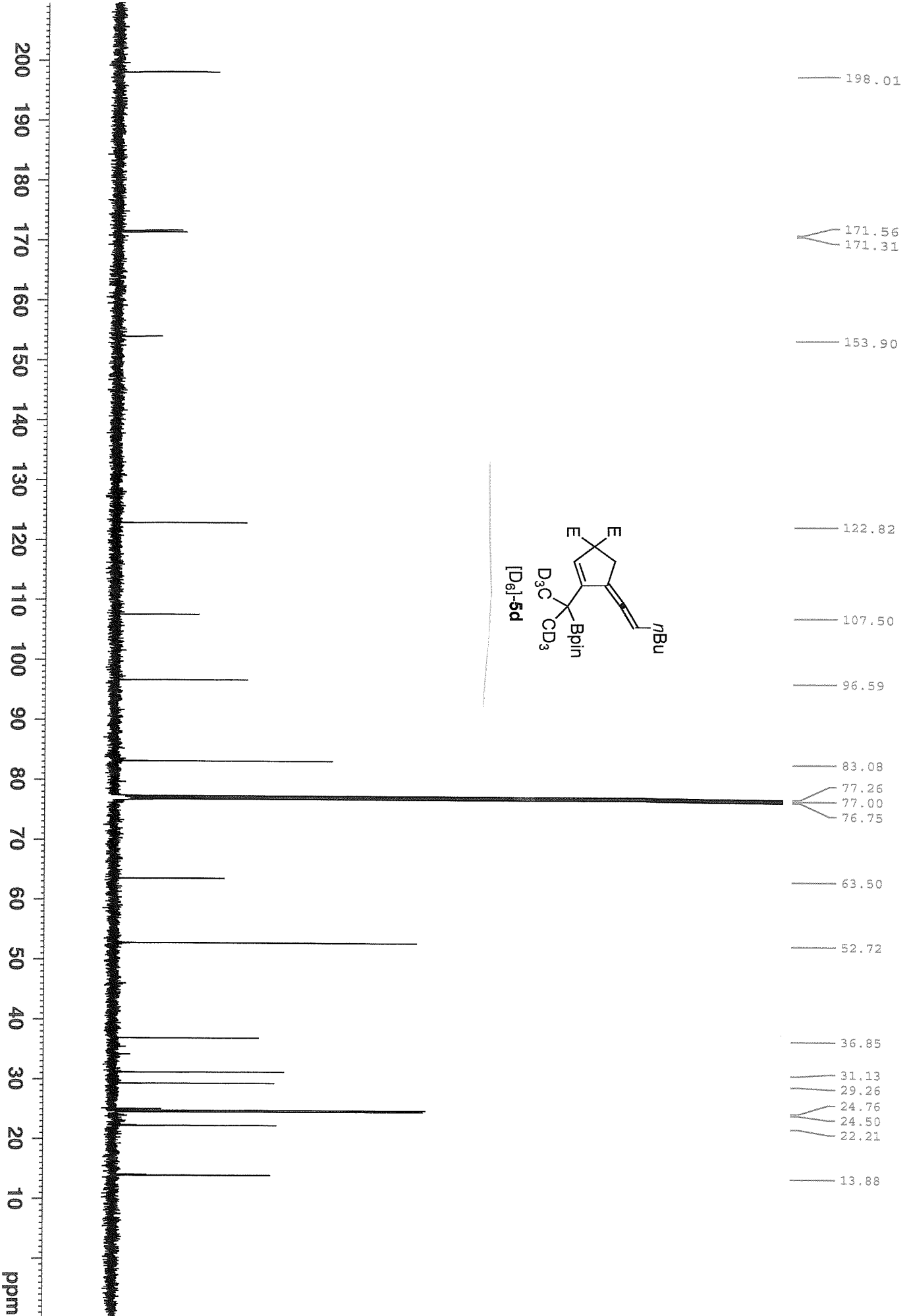
Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 24 H 37 B Na O 6	455.2582	455.2575	-1.5	-0.2
C 24 H 37 B Na O 6	456.2605	455.2575	1.5	-0.2



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
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Mass Spectrum SmartFormula Report

Analysis Info

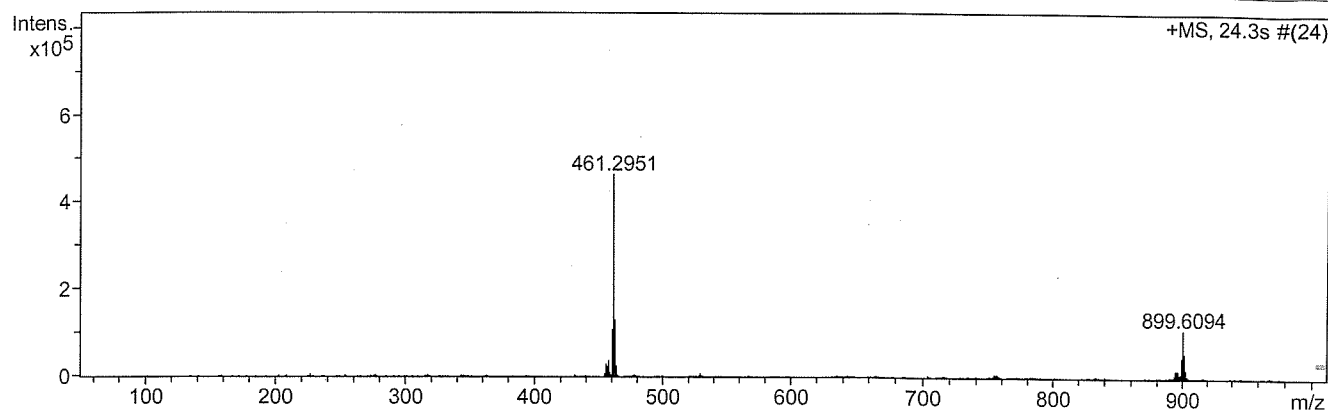
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Method Tune_low_pos.m
Sample Name dyq-4-185
Comment

Acquisition Date 2012-12-07 18:34:02

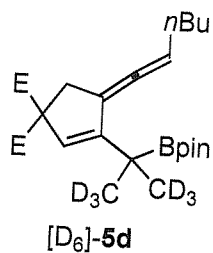
Operator Carin Larsson
Instrument / Ser# micrOTOF 125

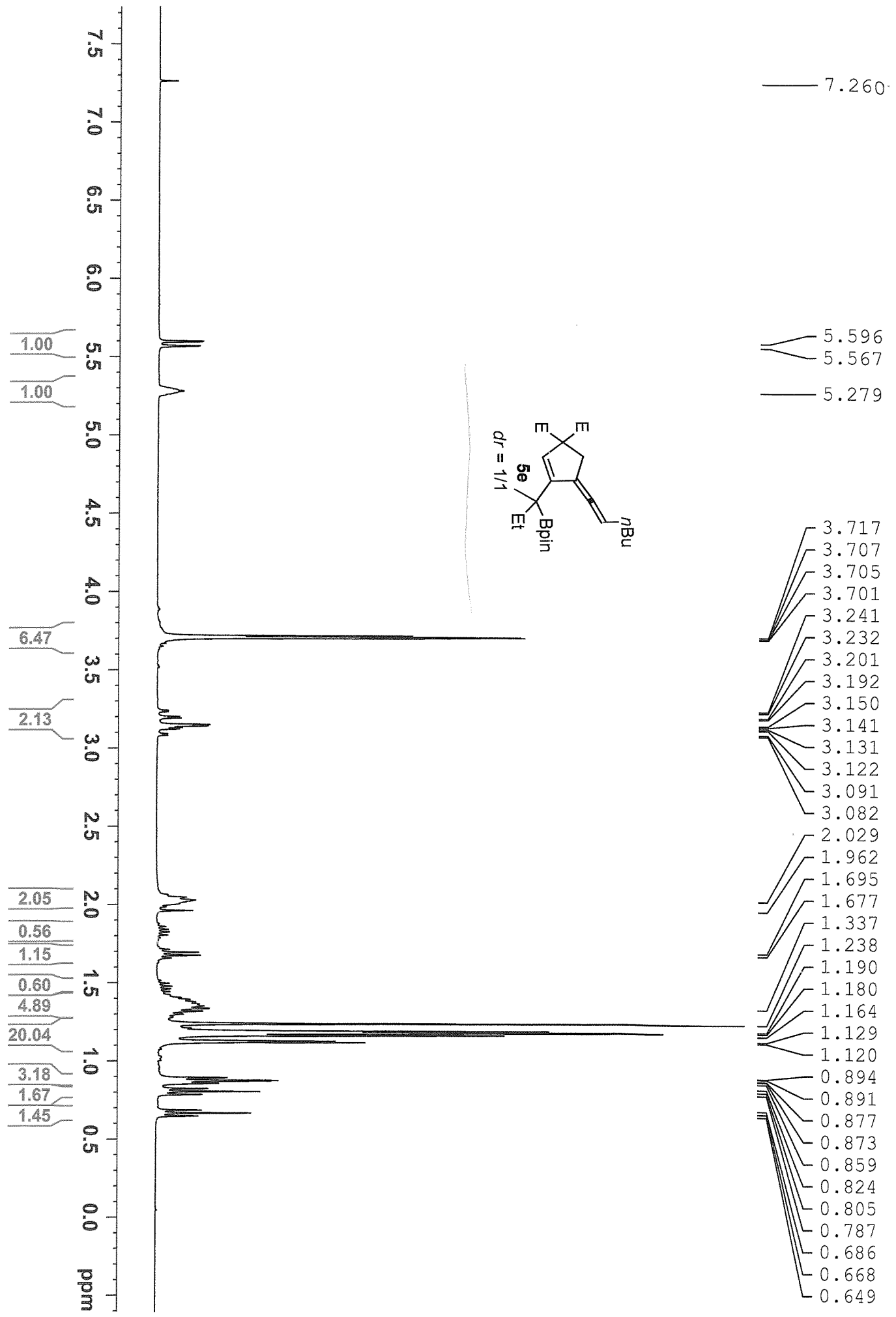
Acquisition Parameter

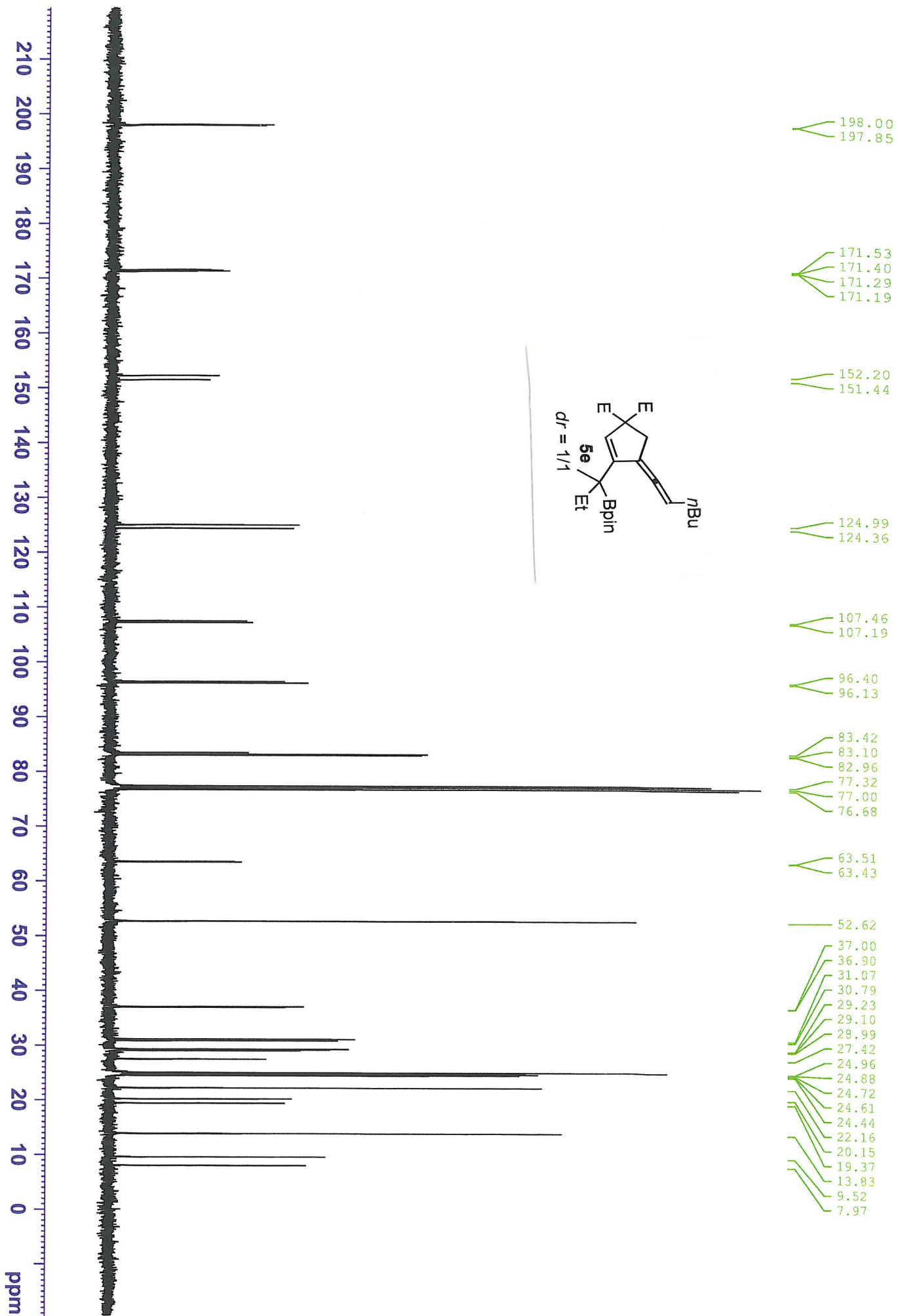
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4000 V	Set Dry Gas	4.0 l/min
Scan End	1000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 24 H 31 B D 6 Na O 6	461.2951	461.2956	0.2	1.7







Mass Spectrum SmartFormula Report

Analysis Info

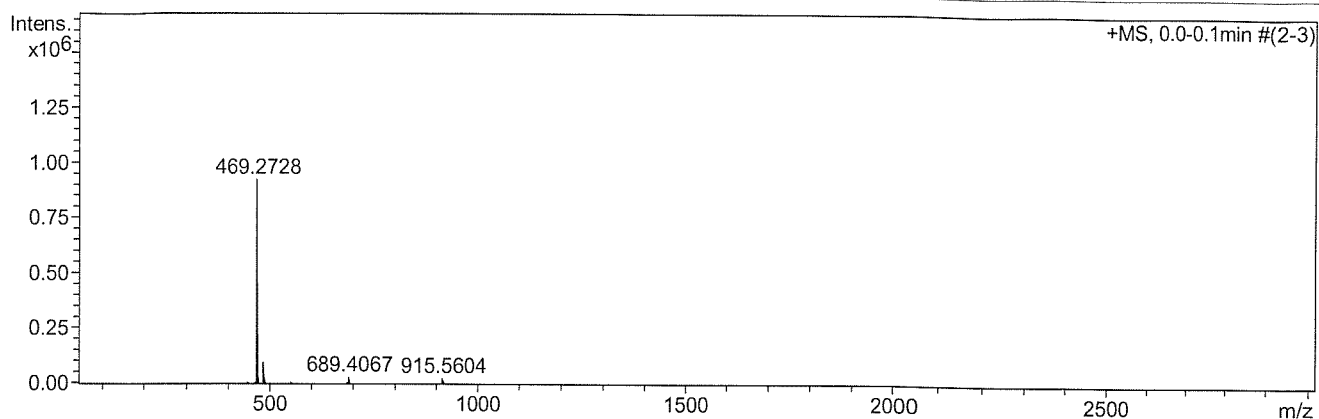
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Method tune_wide_dirk.m
Sample Name dyq-4-10
Comment

Acquisition Date 2012-05-28 17:22:21

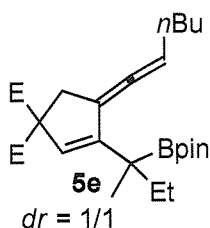
Operator pia
Instrument / Ser# micrOTOF 125

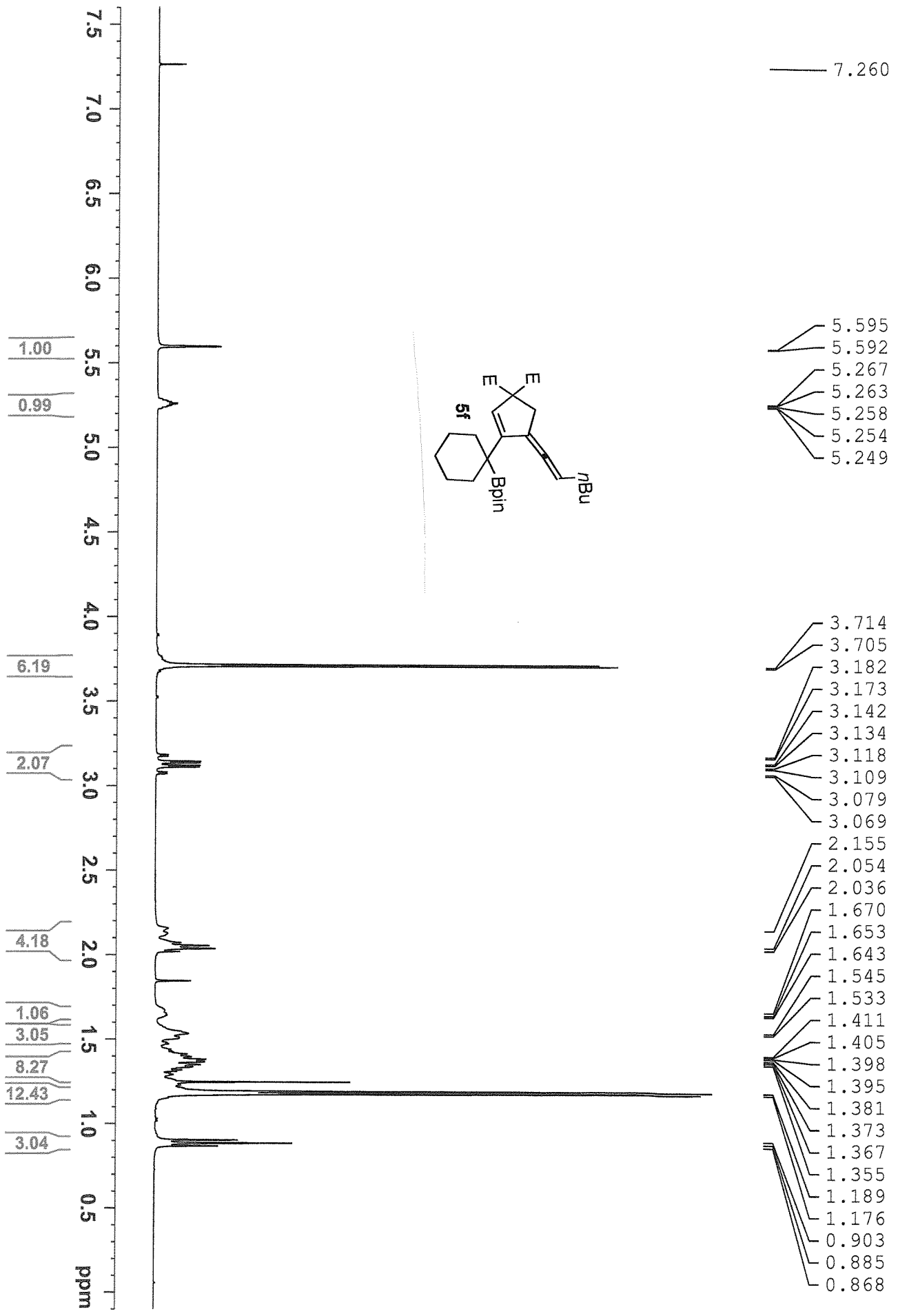
Acquisition Parameter

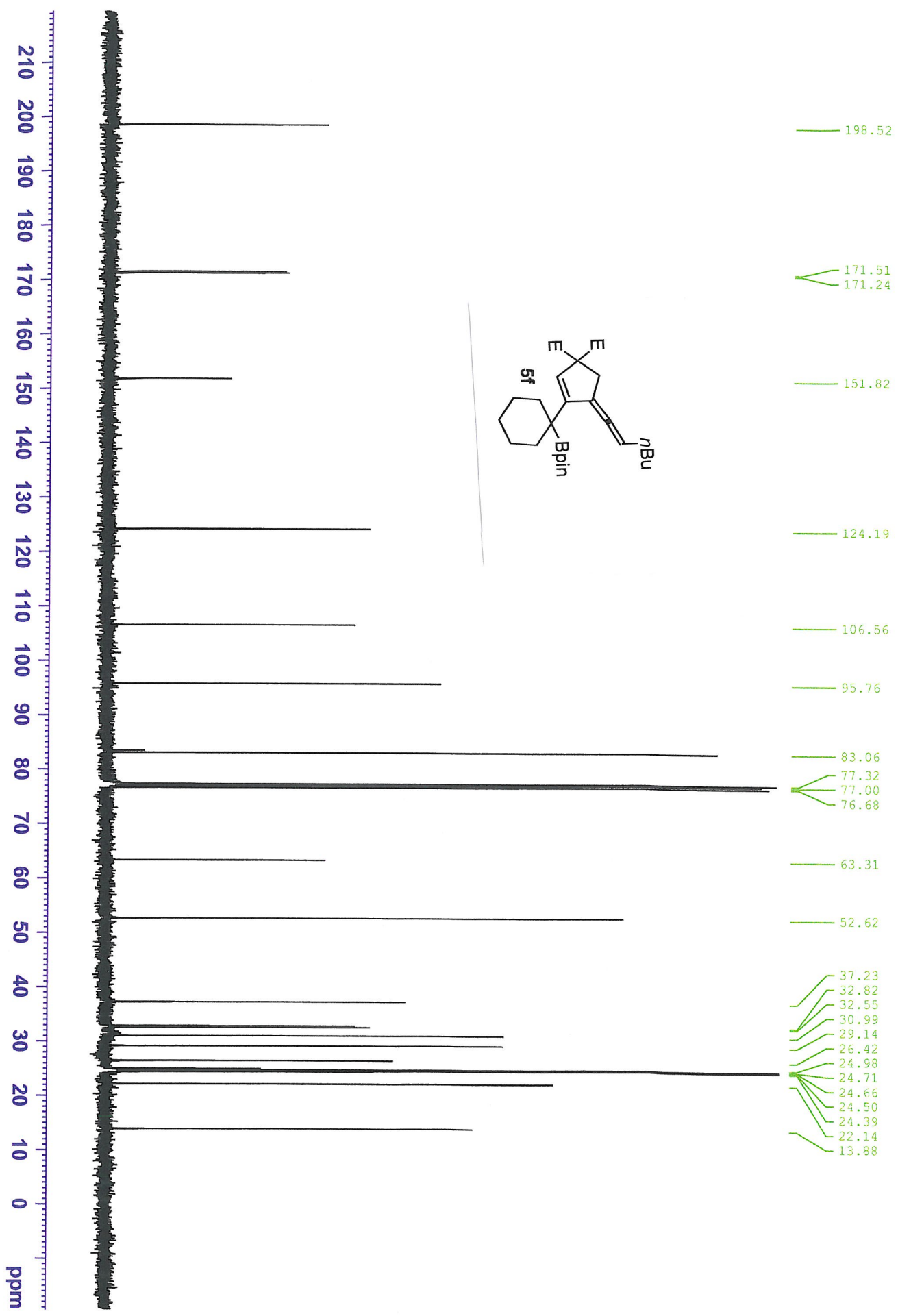
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C ₂₅ H ₃₉ BNaO ₆	469.2728	469.2732	0.9	1.7







Mass Spectrum SmartFormula Report

Analysis Info

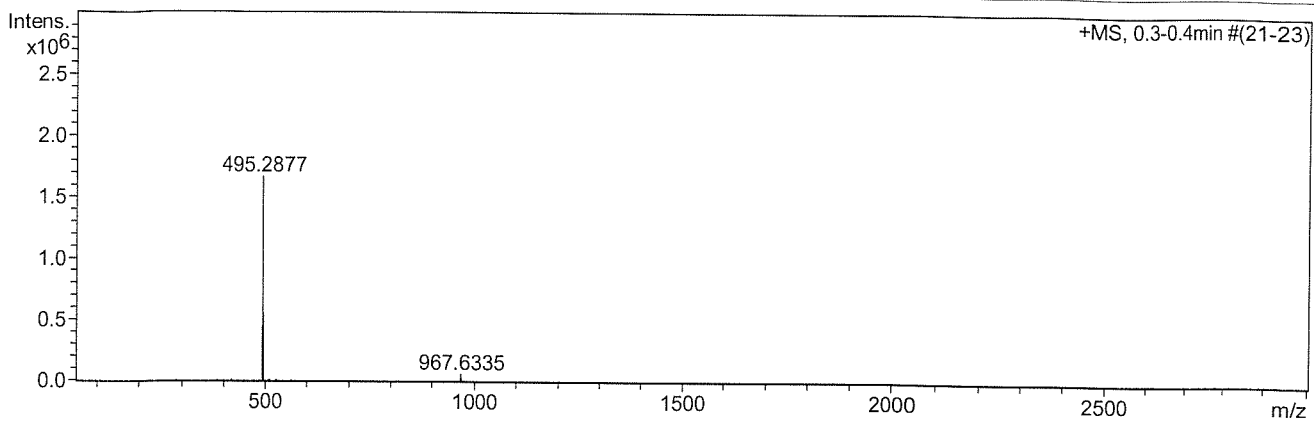
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Method tune_wide_dirk.m
Sample Name dyq-3-171
Comment

Acquisition Date 2012-04-27 18:34:57

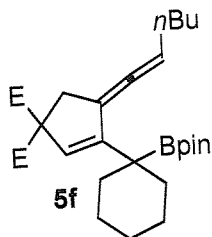
Operator pia
Instrument / Ser# microTOF 125

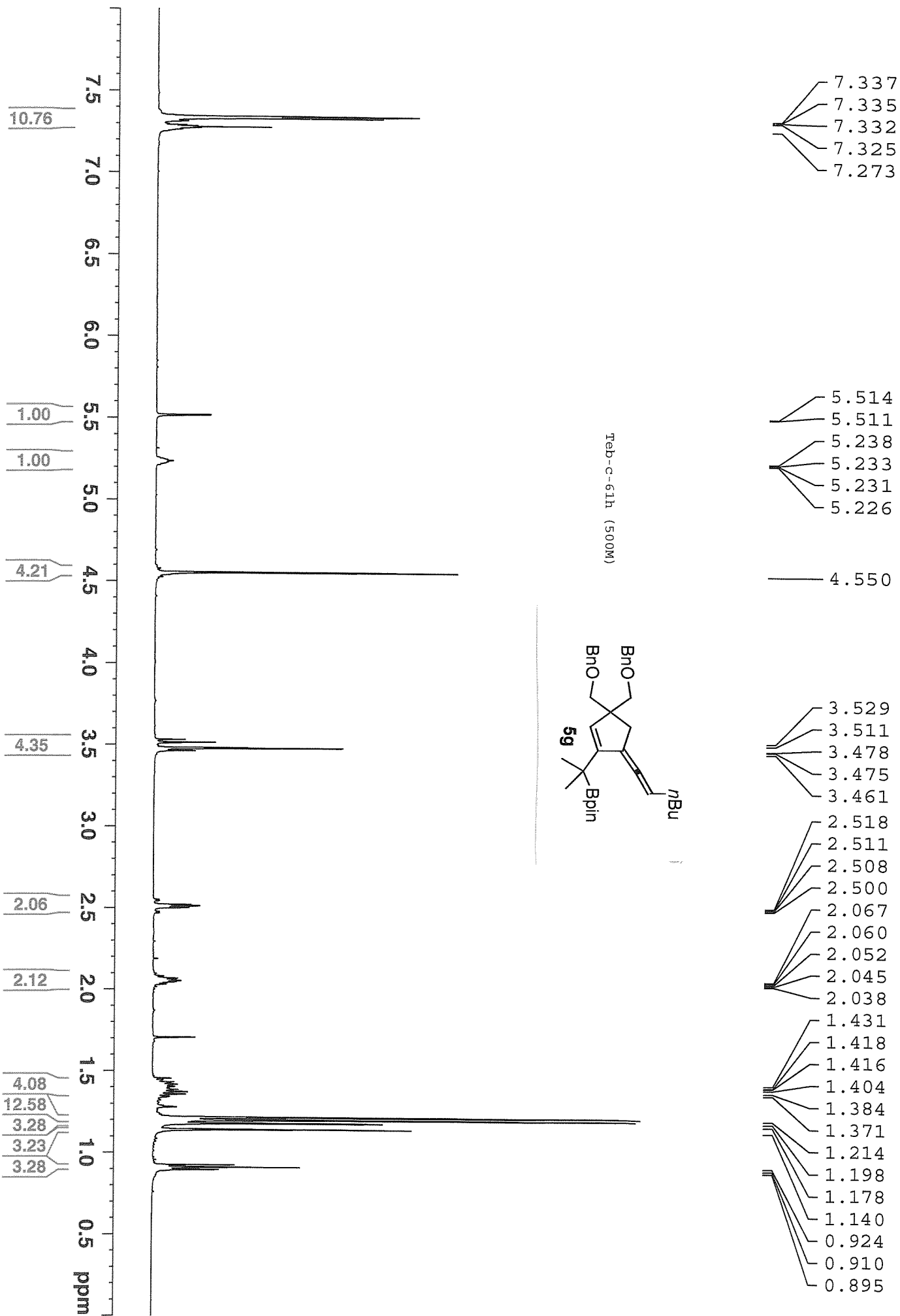
Acquisition Parameter

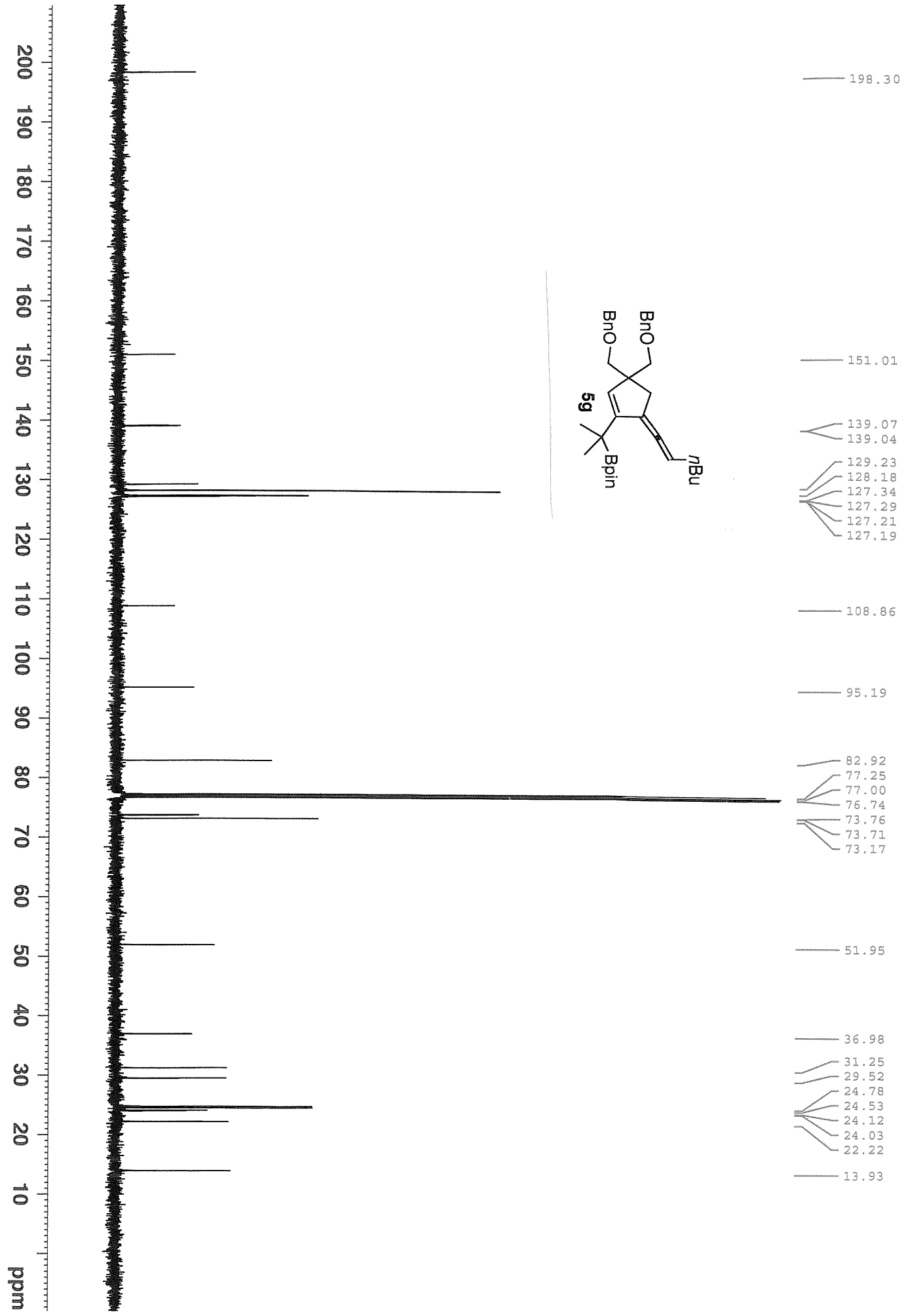
Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C ₂₇ H ₄₁ BNaO ₆	495.2877	495.2888	2.2	4.7







Mass Spectrum SmartFormula Report

Analysis Info

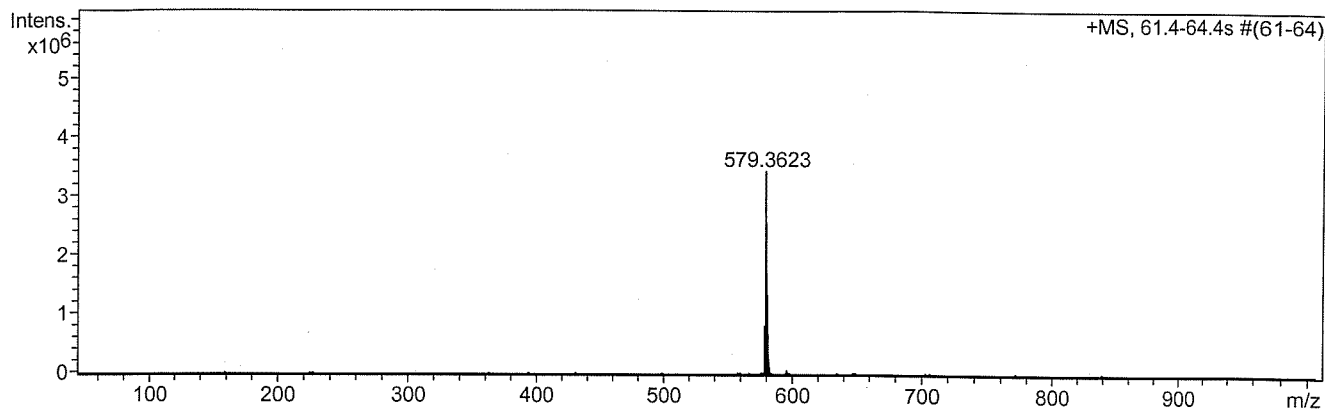
Analysis Name E:\Data2\Youqian\Teb-c-61000001.d
Method Tune_low_pos.m
Sample Name Teb-c-61
Comment

Acquisition Date 2012-12-13 15:47:51

Operator Carin Larsson
Instrument / Ser# micrOTOF 125

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Not active			Set Dry Heater	180 °C
Scan Begin	50 m/z	Set Capillary	4000 V	Set Dry Gas	4.0 l/min
Scan End	1000 m/z	Set End Plate Offset	-500 V	Set Divert Valve	Source



Formula	Meas. m/z	m/z	err [ppm]	Mean err [ppm]
C 36 H 49 B Na O 4	579.3623	579.3622	-0.1	0.3

