

Supplementary Table 1. Characteristics of patients found to harbor U2AF1 mutations.

ID	Dx	Cytogenetics	Sex	U2AF1 mutation	Exome seq	RNA seq
1	sAML	46,XX[20]	F	c.101C>A,p.S34Y,c.104G>T,p.R35L	0	0
2	pAML(M2)	45,XY,-7[17]/45,idem,del(12)(p12)[3]	M	c.101C>A, p.S34Y	0	0
3	pAML(M4)	46,XY[20]	M	c.101C>T, p.S34F	1	0
4	pAML(M4)	46,XY[20]	M	c.101C>T, p.S34F	1	0
5	pAML(M4)	46,XY, add(8)	M	c.101C>T, p.S34F	1	1
6	pAML(M5)	46,XY, add(8)	M	c.101C>A, p.S34Y	1	1
7	pAML(M5)	47,XY,add(3)(p21),del(6)(p21.3),del(11)(q23),del(13)(q14q22),add(14)(p10),del(15)(q22),+19,add(19)(p13.3)[18]/46,XY[2].	M	c.467A>G, p.R156Q	0	0
8	pAML(M5)	45,X,-Y[22]/46,XY[2]	M	c.101C>T, p.S34F	0	0
9	pAML(M6)	N/A	M	c.101C>T, p.S34F	1	1
10	sAML	46,XY[20]	M	c.101C>T, p.S34F	1	0
11	sAML	47,XY,+19[20]	M	mutated	0	0
12	pAML	46,XY,-7,+14[17]/46,XY[3]	M	c.470A>C, p.Q157P	0	0
13	sAML	46,XY,del(20)(q11.2)[12].	M	c.101C>T, p.S34F	1	0
14	sAML	46,X,-X[17]/46,XX[3].	F	c.470A>C, p.Q157P	0	0
15	sAML	46,XY,del(7)(q11.2)[10]/46,XY,add(12)(q24.3)[7]/46,XY[3]	M	c.470A>C, p.Q157P	0	0
16	sAML	46,XY,del(20)(q11.2)[6]/46,idem,del(7)(p12)[2]/46,XY,ider(20)(q10)del(20)(q11.2)[8]/46,XY[4]	M	c.101C>T, p.S34F	0	0
17	sAML	46,XY[20]	M	c.101C>T, p.S34F	0	0
18	sAML	46,XX,del(17)(q24)[6]/46,XX[24]	F	c.101C>T, p.S34F	0	0
19	pAML(M1)	46,XY,del(7q) / 7q-	M	c.101C>T, p.S34F	1	1
20	pAML(M1)	47,XY, add(8)	M	c.470A>C, p.Q157P	1	0
21	pAML(M1)	46,XY,t(8;21)	M	C>G p.G213A	1	0
22	pAML(M1)	47,XY,+11[20] 46,XY,-3,-	M	c.101C>T, p.S34F	0	0
23	pAML(M1)	4,der(4)add(4)(p16)add(4)(q35),del(5)(q13q33),add(5)(p13),-7,del(9)(q31),-13,add(15)(p13),der(17)t(13;17)(q12;p11.2),-21,-22,-22,+mar1,+mar2[cp17]/41,idem,i(13)(q10)[3].	M	whole gene deletion	0	0
24	pAML(M2)	46,XY[20]	M	c.101C>T, p.S34F	1	1
25	pAML(M2)	46,XX[20],del(7q) / 7q-	F	c.101C>A, p.S34Y	1	1
26	sAML	46,XY[18]	M	c.251A>C p.Q84P	1	0
27	CMML-1	46,XY,?inv(20)(q11.2q13)[20]	M	mutated	0	0
28	CMML-1	46,XY[20]	M	c.467A>G, p.R156Q	0	0
29	CMML-1	46,XY[17]	M	mutated	0	0
30	CMML-1	45,XY,-7[18]/46,XY[2]	M	c.470A>C, p.Q157P	0	0
31	CMML-2	46,XY[20]	M	c.470A>C, p.Q157P	0	0
32	CMML-2	46,XY,del(20)(q11.2q13.3)[19]/46,XY[1].	M	c.101C>T, p.S34F	0	0
33	CMML-2	46,XY[20]	M	c.101C>T, p.S34F	0	0
34	RAEB	44,XY,-3,-5,-7,+9,del(11)(p12)[2]/43,idem,-9,-18,+mar[4]/46,XY[8]	M	c.470A>C, p.Q157P	0	0
35	RAEB	47, XX, del(3)(q21),del(5)(q13),del(12)(q21q24.3),+mar1[7]/45,sl,+3,-del(3),-7,-18,-mar1,+mar2[cp12 one is 4n]/46XX[1]	F	c.470A>C, p.Q157P	0	0
36	RAEB	46,XY,del(20)(q11.2)[2]/46,idem,-4,+8[1]/46,XY,der(20)del(20)(p11.2)del(20)(q11.2)[2]/46,XY[6].	M	c.101C>T, p.S34F	0	0
37	RAEB	46,XX,?add(7)(q22) or ?del(7)(q22q22)[5]/46,XX[15]	F	c.101C>T, p.S34F	0	0
38	RARS	Del(5), -7, -20, +19, +22	M	c.101C>T, p.S34F	0	0
39	RCMD	46,XY,del(20)(q11.2)[3]/46,idem,del(5)(q12q33)[15]/46,XY[2]	M	c.101C>T, p.S34F	0	0
40	RCUD MDS	46,XX,del(20)(q11.2)[20]	M	c.470A>G p.Q157R	1	0
41	tMDS	46,XX,add(12)(p13)[6]/46,XX[14]	F	c.470A>C, p.Q157P	0	0

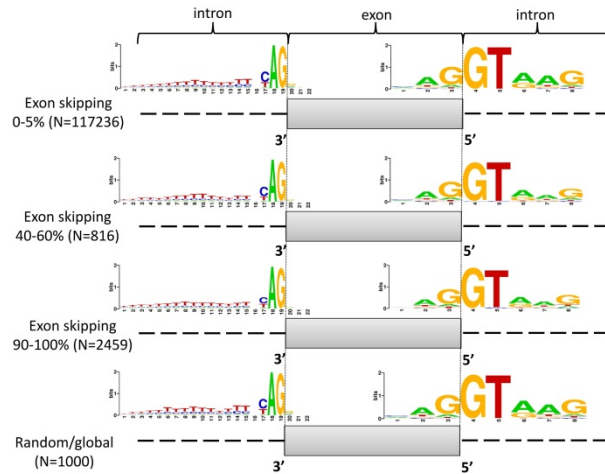
Supplementary Table 3. U2AF1 mutational status of patients with myeloid malignancies

WHO classification	U2AF1 mut	tested
MDS	12	116
Low grade	7	79
RA/RCMD	5	48
RARS/RCMD-RS	2	31
Advanced grade	5	37
RAEB1/2	5	37
MDS/MPN	12	76
CMML1/2	12	76
AML	22	332
Primary AML	15	259
Secondary AML	7	73

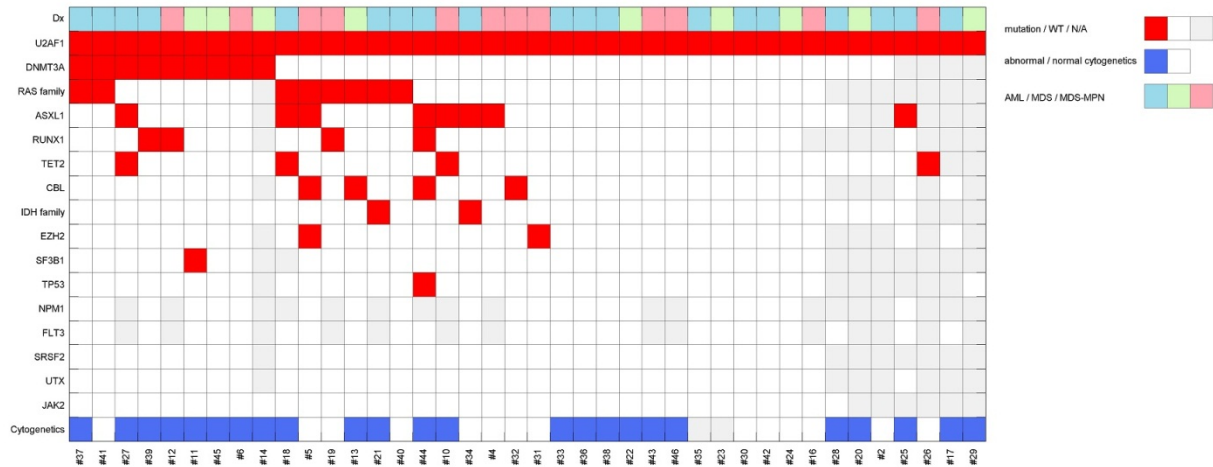
Supplementary Table 1. Characteristics and data types available for patients that were found to carry U2AF1 mutations.

Supplementary Table 2. Exon usage levels in genes that were found to be differentially spliced in U2AF1 mutants. Provided is additional information about the number of mutations and copy number abnormalities found in these genes in public databases (COSMIC) as well as from an in-house database of patients.

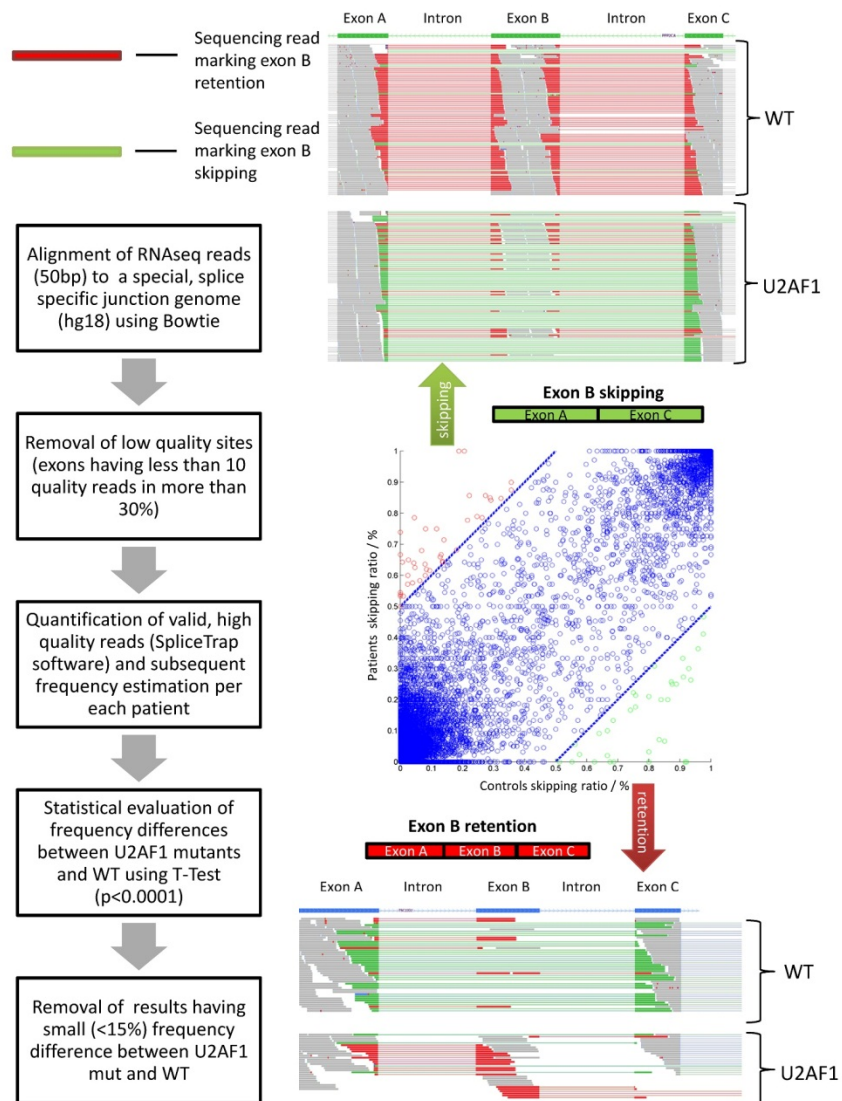
Supplementary Table 3. Characteristics of patients used in this study. The total number of patients studied is 524, with 116 patients diagnosed with MDS, 76 with MDS/MPN and 332 with AML. The total number of patients harboring U2AF1 mutations is 46 (12 MDS, 12 MDS/MPN and 22 AML).



Supplementary Figure 1. Frequencies of nucleotides at 3' and 5' splice sites flanking exons in control RNA samples. The top three lines show all exons grouped by their frequency of skipping. The bottom line shows a randomly selected set of 1,000 exons. Nucleotide frequencies are represented using WebLogo software. The height of each stack represents the information content of that position in bits. The height of each letter represents the frequency of occurrence of each nucleotide at each position.

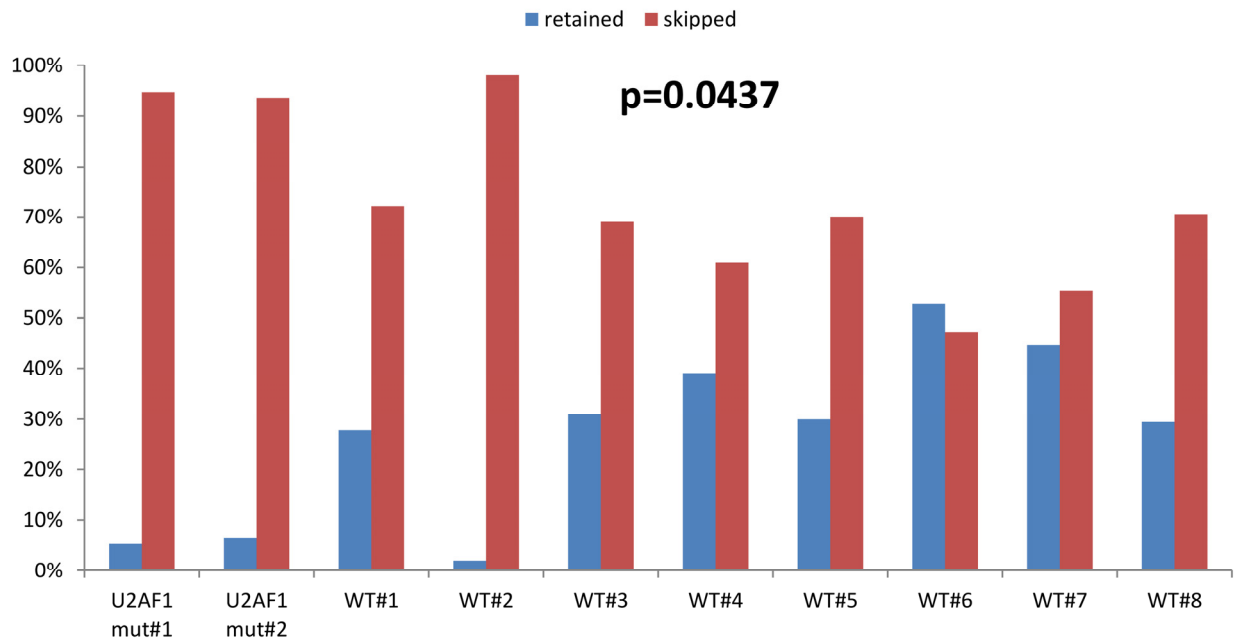


Supplementary Figure 2. Additional mutations found in patients with U2AF1 mutations. Each column represents one patient. The top row represents the diagnosis of the patient (AML – blue, MDS/MPN – pink, MDS – green). The bottom row represents the individual karyotype of each patient (abnormal – blue, normal – white). The remaining rows represent genes that were screened for mutations in the U2AF1 mutated patients. Each row is a separate gene as listed. Red color indicates the presence of a mutation, white color indicates a WT sequence and light grey indicates an unknown status (sequencing not done).



Supplementary Figure 3. Work flow for the identification of candidate exons harboring differential exon usage between U2AF1 mutants and WT. Two major patterns of changes in alternative splicing were identified: 1) increased exon skipping in patients with U2AF1 mutations as shown by an excess of green reads supporting exclusion of the middle exon as depicted in the upper panel and 2) increased exon retention in patients with U2AF1 mutation as shown by an excess of red reads supporting inclusion of the middle exon as depicted in the lower panel. The center panel shows a scatter plot of exon skipping in

control cell RNA versus U2AF1 mutant cell RNA. The lines show the 15% difference cutoff limit used in selecting the most affected exons.



Supplementary Figure 4. Patients exon usage ratio of CEP164 gene (exon7) using intensity of bands on a gel.