Supplementary Material

April 17, 2013

Reconciled Databases Recently there has been an emergence of aggregated databases that reconcile entries from multiple *child* databases into a single *parent* database. Databases such as BKM-React (Lang *et al.*, 2011), MetRxn (Kumar *et al.*, 2012) and MNXref (Bernard *et al.*, 2012) offer an important resource to access metabolic and enzymatic information. However, these aggregated databases are limited by the substantial effort required to keep up to date with their respective *child* databases and license restrictions on redistribution. Generally chemical structure line notations, SMILES and InChI (Warr, 2011), are used to reconcile metabolites in these resources. How metabolites and reactions are merged can vary, one resource may merge entities at different protonation state whilst others may kept them separate. A molecule with unspecified stereo-chemistry may be merged with one which is fully specified, or different stereo isomers may all be collapsed to a generic entry.

Chemical File Formats Metingear can load from several file formats including, Chemical Markup Language (Kuhn *et al.*, 2007), Mol (Warr, 2011), The IUPAC International Chemical Identifier (InChI) and SMILES (Warr, 2011). Models can be exported in SBML with annotations of cross-references and InChI. Mol and CML files provide the exact depiction of what the original author of the structure drew whilst InChI and SMILES requires a new structure diagram is generated. Currently direct drawing of structures is not available. The JChemPaint (http://jchempaint.github.com/) (Krause *et al.*, 2000) application uses a incompatible version of the CDK library.

Services Metingear currently provides services to access; ChEBI (Matos *et al.*, 2010), MetaCyc (Caspi *et al.*, 2012), KEGG Compound (Kanehisa *et al.*, 2012), LIPID Maps (Sud *et al.*, 2007), Human Metabolome Database (HMDB) (Wishart *et al.*, 2009), PubChem-Compound (Bolton *et al.*, 2008) and UniProt (The UniProt Consortium, 2012). Each service, except PubChem, has a loader which allows the user to update the resource with the latest available version. When the download is small enough and available the resource can be updated automatically, in the cases of KEGG and MetaCyc where a fee or registration is required the file can be specified as a location on the local file system (see. https://github.com/johnmay/metingear/wiki/Resources). The loaders create a searchable index in the specified folder. The services then check for this index when a new tool is opened, if the index has been created then the service is available. As with the cross-references each service is linked with MIRIAM registry (Juty *et al.*, 2012) information. This allows Metingear to recognise which resource a metabolite is annotated with and try and locate a required service (e.g. name search, structure download). If there is no local index service loaded then it will default to a web-service query. The services load dynamically at runtime and thus it is possible to add custom services which may connect to in-house databases or web-services and provided specialised compounds. This feature also makes it very easy to integrate new resources (such as the reconciled databases) and keep up to date with the existing resources. Recently HMDB changed their download format, to accommodate this change, only a new loader for the format was required. The existing loader has been kept for legacy and still usable but the existing HMDB services access did not need to be changed.

Inconsistencies Annotating previously published models revealed inconsistencies which could not be easily identified in the original spreadsheet. A model of *Lactobacillus plantarum* WCFS1 (Teusink *et al.*, 2006) was found to be missing a reaction equation for reaction UGMDDS2 (Fig. S1). Also in a model of *Bacillus subtilis* (*i*Bsu1103) (Henry *et al.*, 2009), three reactions were found to reference a metabolite not found in the metabolites table (Fig. S2a and S2b). These inconsistencies demonstrate the use of specialised software in curation of larger reconstructions. These inconsistencies were identified automatically when a model is loaded from Excel. Other inconsistencies checks are carried out in the background of the model but do not declare an error. The mass and

charge balance of a reaction is indicated by *scales* icon which tips to which ever side is heavier or is balanced when the reaction is balanced. Structures attached to metabolites are checked as to whether they match encoded formulas and charges (which can be imported and extracted). This indication serves only as a hint that something might be wrong as the charge and formula annotations may be absent.

Figure 1: Subset of *Lactobacillus plantarum* WCFS1 reactions The reactions as listed in the the 'reaction info' sheet of 'Supplementary material IV.xls' of Teusink *et al.* (2006). The reaction in row 746 with the identifier UGMDDS2 is missing a reaction equation and is instead replaced with the name of another reaction.

Additional Features In addition to handling metabolites and reactions their is also support for genes and gene products. These can be imported from the European Nucleotide Archive (ENA) XML (Amid *et al.*, 2012) and fasta formats. When importing models from a spreadsheet the locus of the reaction is often annotated. This locus annotation can be paired with the gene/gene product information to provide a model which is enriched with sequence as well as chemical information. Although not the primary purpose Metingear can also run a homology search using a locally installed BLAST (Altschul *et al.*, 1990) instance and transfer the annotations from homologous sequences. We are currently focused on metabolite annotation but in future we will improve the gene and gene product linking to be more automated.

A real-time search, undo/redo edit support, star rating and sub-collections help in general navigation. All entities can be easily renamed merged and split allowing the flexibility when editing a reconstruction. Each entity can have it's name and abbreviation changed the primary identifier assigned automatically. Each model is encoded with taxonomy information and when available compartments are annotated with Gene Ontology Terms (Camon, 2003) in the SBML output.

Each metabolite with a structure, molecular formula and charge indicates via *structural validity* whether the structure matches the given formula and charge. The formula and charge can often be imported from the spreadsheets or SBML notes and provides a check as to whether the attached structure is correct. Reactions indicate whether their participants are balanced (mass only) and whether they are transport reactions.

Internally a binary format is used for the reconstructions, this format provides very rapid loading and saving of reconstructions. Draft reconstructions from the model-SEED (Henry *et al.*, 2010) can be directly imported via the spreadsheet format without having to select which fields are present. Metingear can also create and export a stoichiometric matrix to a tabular file or to a '.sif' which can be loaded in Cytoscape (http://www.cytoscape.org/). The chemical structure of metabolites in the models can be exported to a single structured-data file (SDF) (Warr, 2011).

A primitive but functional dialog plugin framework allows one to extend Metingear with their own tools (https://github.com/johnmay/metingear/wiki/Plugable-Dialogs).

Table 1: Available annotations - each annotation can be added one or more times to a model component. Some annotations may have restrictions on where they can be added (e.g. flux annotations can only be added to reactions)

622 rxn03164 623 rxn03167 624 rxn03175 625 rxn03194 626 rxn03239 627 rxn03240 628 rxn03241	UDP-N-acetylmuramoyl-L-alanyl-C cpd00002 + cpd02964 + cpd00731 => cpd00008 + cpd(2-Amino-4-hydroxy-6-(erythro-1, $\frac{1}{2}$ 3 cpd00001 + cpd02978 => 3 cpd00009 + 3 cpd00067 - $N-(5'-Phospho-D-ribosylformiminc cpd02979 < = > cpd02991$ (S)-2-Aceto-2-hydroxybutanoate p cpd03049 + cpd00094 <= > cpd00056 + cpd00498 $(S)-3-Hydroxyhexadecanoyl-CoA: cpd00003 + cpd03113 <=> cpd00067 + cpd00004 + cpd$ $(S)-3$ -Hydroxyhexadecanoyl-CoA cpd03113 <=> cpd00001 + cpd03126 $(S)-3$ -Hydroxytetradecanoyl-CoA ł cpd03115 <=> cpd00001 + cpd03127	
642 rxn03409 643 rxn03424 644 rxn03435 645 rxn03436 646 rxn03437 647 rxn03445 648 rxn03481	Undecaprenyl-diphospho-N-acetyli cpd00002 + cpd00013 + cpd03495 => cpd00008 + cpd(L-erythro-4-Hydroxyglutamate:NA 2 cpd00067 + cpd00004 + cpd01974 <=> cpd00003 + c $(R)-2,3-Dihydroxy-3-methylpenta$ cpd00006 + cpd02535 <=> cpd000067 + cpd00005 + cpd000005 + cpd00005 + cpd (S) -2-Aceto-2-hydroxybutanoate: cpd00498 <= > cpd10162 $(R)-2,3-Dihydroxy-3-methylpenta cpd02535 => cpd00001 + cpd00508$ O-Phospho-4-hydroxy-L-threonine cpd00024 + cpd03607 $\lt =$ > cpd00023 + cpd03606 Arbutin 6-phosphate qlucohydrola cpd00001 + cpd03697 <=> cpd00415 + cpd00863	
1190 rxn08615 1191 rxn08669 1192 rxn08707 1193 rxn08764 1194 rxn08775 1195 rxn09011 1196 rxn09012	$\text{cpd}00387 + \text{cpd}15302 \leq y \leq \text{cpd}00008 + \text{cpd}00155$ glycogen synthase (ADPGIc) Glycerophosphodiester phosphodik cpd00001 + cpd02090 <=> cpd00080 + cpd00100 Heme O synthase protoheme ix fa cpd00001 + cpd00028 + cpd00350 <= > cpd00012 + cpd ketol-acid reductoisomerase (2-Ac cpd00067 + cpd00005 + cpd00498 \le = > cpd00006 + cp L-alanyl-gamma-L-glutamate pept cpd00001 + cpd15388 <=> cpd00023 + cpd00035 nucleoside-triphosphatase (dITP) cpd00001 + cpd00977 => cpd00009 + cpd00067 + cpd(nucleoside-triphosphatase (XTP) cpd00001 + cpd00530 => cpd00009 + cpd00067 + cpd(
322 cpd00491 323 cpd00492 324 cpd00497	(a) subset of <i>Bacillus subtilis</i> (<i>i</i> Bsu1103) reactions D-Mannitol 1-phosphate D-mannit C6H14O9P N-Acetyl-D-mannosamine 2-Aceta C8H15NO6 Xanthosine 5'-phosphate XanthylicC10H12N4O9P	C00644 C00645 C00655

(b) subset of *Bacillus subtilis* (*i*Bsu1103) metabolites

C00663

C00666

C00670

beta-D-Glucose 1-phosphate/beta C6H12O9F

LL-2,6-Diaminoheptanedioate | LL-2 C7H14N2O4

sn-glycero-3-Phosphocholine Glyc C8H20NO6

Figure 2: Missing information in genome-scale models.

325 cpd00501

326 cpd00504

327

cod00507

2a) Three reactions rxn003194, rxn03436 and rxn08764 from the reactions spreadsheet (Table S2-Reaction Data) reference a metabolite, cpd00498 (highlighted red), however information about this metabolite is missing from the metabolites sheet (Table S1-Compound Data) (Henry *et al.*, 2009).

2b) Expected location of the missing metabolite cpd00498 in the metabolites table (marked with a red line). Using the named reaction (not shown) it is possible to see that the name of missing metabolite is 2-aceto-2 hydroxybutanoate but no other details to this metabolite are provided.

References

- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., and Lipman, D. J. (1990). Basic local alignment search tool. *J Mol Biol*, 215(3), 403–10.
- Amid, C., Birney, E., Bower, L., Cerdeno-Tarraga, A., Cheng, Y., Cleland, I., Faruque, N., Gibson, R., Goodgame, N., Hunter, C., Jang, M., Leinonen, R., Liu, X., Oisel, A., Pakseresht, N., Plaister, S., Radhakrishnan, R., Reddy, K., Riviere, S., Rossello, M., Senf, A., Smirnov, D., Hoopen, P. T., Vaughan, D., Vaughan, R., Zalunin, V., and Cochrane, G. (2012). Major submissions tool developments at the European Nucleotide Archive. *Nucleic Acids Research*, 40(D1), D43–D47.
- Bernard, T., Bridge, A., Morgat, A., Moretti, S., Xenarios, I., and Pagni, M. (2012). Reconciliation of metabolites and biochemical reactions for metabolic networks. *Briefings in Bioinformatics*, pp, 1–133.
- Bolton, E., Wang, Y., Thiessen, P., and Bryant, S. (2008). Pubchem: Integrated platform of small molecules and biological activities. *Annual Reports in Computational Chemistry, American Chemical Society, Washington, DC*, 4.
- Camon, E. (2003). The gene ontology annotation (goa) project: Implementation of go in swiss-prot, trembl, and interpro. *Genome Research*, 13(4), 662–672.
- Caspi, R., Altman, T., Dreher, K., Fulcher, C. A., Subhraveti, P., Keseler, I. M., Kothari, A., Krummenacker, M., Latendresse, M., Mueller, L. A., Ong, Q., Paley, S., Pujar, A., Shearer, A. G., Travers, M., Weerasinghe, D., Zhang, P., and Karp, P. D. (2012). The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of Pathway/Genome Databases. *Nucleic Acids Research*, 40(D1), D742–D753.
- Henry, C. S., Zinner, J. F., Cohoon, M. P., and Stevens, R. L. (2009). *i*Bsu1103: a new genome-scale metabolic model of *Bacillus subtilis* based on SEED annotations. *Genome Biol*, 10(6), R69.
- Henry, C. S., DeJongh, M., Best, A. A., Frybarger, P. M., Linsay, B., and Stevens, R. L. (2010). High-throughput generation, optimization and analysis of genome-scale metabolic models. *Nature Biotechnology*, 28(9), 969–974.
- Juty, N., Novère, N. L., and Laibe, C. (2012). Identifiers.org and MIRIAM Registry: community resources to provide persistent identification. *Nucleic Acids Research*, 40(D1), D580–D586.
- Kanehisa, M., Goto, S., Sato, Y., Furumichi, M., and Tanabe, M. (2012). KEGG for integration and interpretation of large-scale molecular data sets. *Nucleic Acids Research*, 40(D1), D109–D114.
- Krause, S., Willighagen, E., and Steinbeck, C. (2000). Jchempaint using the collaborative forces of the internet to develop a free editor for 2d chemical structures. *Molecules*, 5(1), 93–98.
- Kuhn, S., Helmus, T., Lancashire, R. J., Murray-Rust, P., Rzepa, H. S., Steinbeck, C., and Willighagen, E. L. (2007). Chemical Markup, XML, and the world wide web. 7. cmlspect, an xml vocabulary for spectral data. *J Chem Inf Model*, 47(6), 2015–34.
- Kumar, A., Suthers, P., and Maranas, C. (2012). MetRxn: a knowledgebase of metabolites and reactions spanning metabolic models and databases. *BMC Bioinformatics*, 13(1), 6.
- Lang, M., Stelzer, M., and Schomburg, D. (2011). BKM-react, an integrated biochemical reaction database. *BMC biochemistry*, 12(1), 42.
- Matos, P. D., Alcantara, R., Dekker, A., Ennis, M., Hastings, J., Haug, K., Spiteri, I., Turner, S., and Steinbeck, C. (2010). Chemical Entities of Biological Interest: an update. *Nucleic Acids Research*, 38(Database), D249–D254.
- Sud, M., Fahy, E., Cotter, D., Brown, A., Dennis, E. A., Glass, C. K., Merrill, A. H., Murphy, R. C., Raetz, C. R. H., Russell, D. W., and Subramaniam, S. (2007). LMSD: LIPID MAPS structure database. *Nucleic Acids Research*, 35(Database), D527–D532.
- Teusink, B., Wiersma, A., Molenaar, D., Francke, C., de Vos, W. M., Siezen, R. J., and Smid, E. J. (2006). Analysis of growth of *Lactobacillus plantarum* WCFS1 on a complex medium using a genome-scale metabolic model. *J Biol Chem*, 281(52), 40041–8.
- The UniProt Consortium (2012). Reorganizing the protein space at the Universal Protein Resource (UniProt). *Nucleic Acids Research*, 40(D1), D71–D75.
- Warr, W. A. (2011). Representation of chemical structures. *WIREs Comput Mol Sci*, 1(4), 557–579.
- Wishart, D. S., Knox, C., Guo, A. C., Eisner, R., Young, N., Gautam, B., Hau, D. D., Psychogios, N., Dong, E., Bouatra, S., Mandal, R., Sinelnikov, I., Xia, J., Jia, L., Cruz, J. A., Lim, E., Sobsey, C. A., Shrivastava, S., Huang, P., Liu, P., Fang, L., Peng, J., Fradette, R., Cheng, D., Tzur, D., Clements, M., Lewis, A., Souza, A. D., Zuniga, A., Dawe, M., Xiong, Y., Clive, D., Greiner, R., Nazyrova, A., Shaykhutdinov, R., Li, L., Vogel, H. J., and Forsythe, I. (2009). HMDB: a knowledgebase for the human metabolome. *Nucleic Acids Research*, 37(Database), D603–D610.