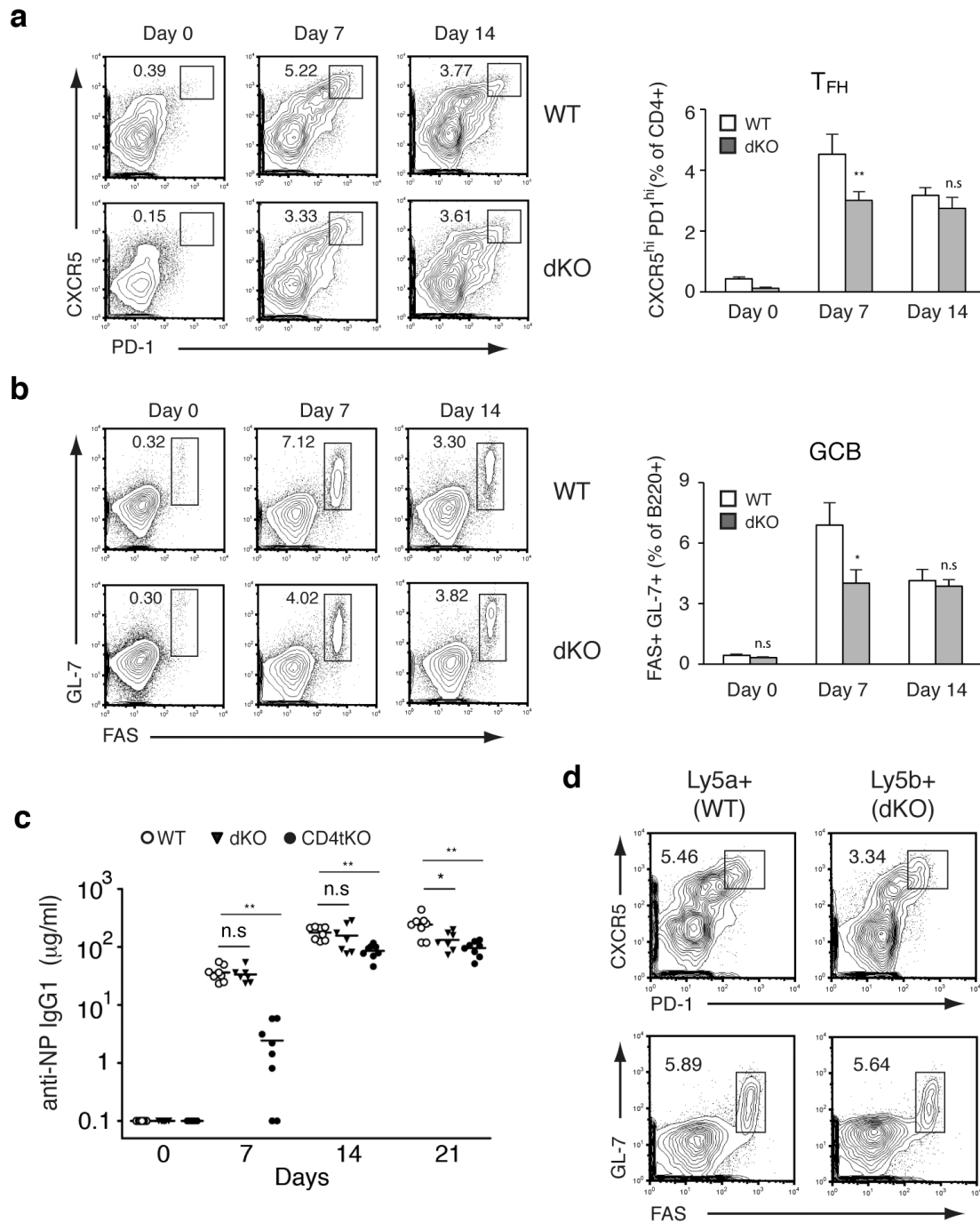


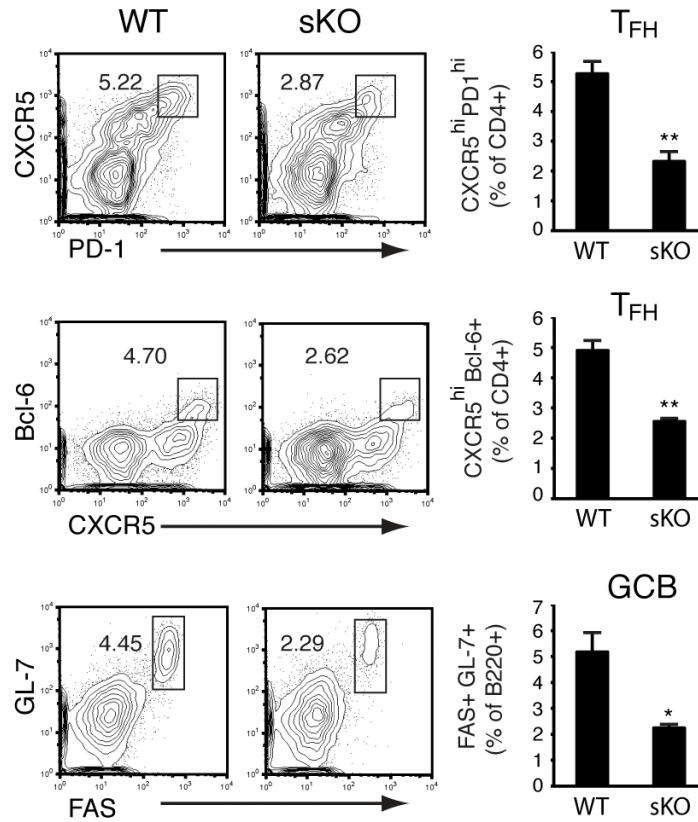
**miR-17~92 family microRNAs are critical regulators of T follicular helper cell  
differentiation**

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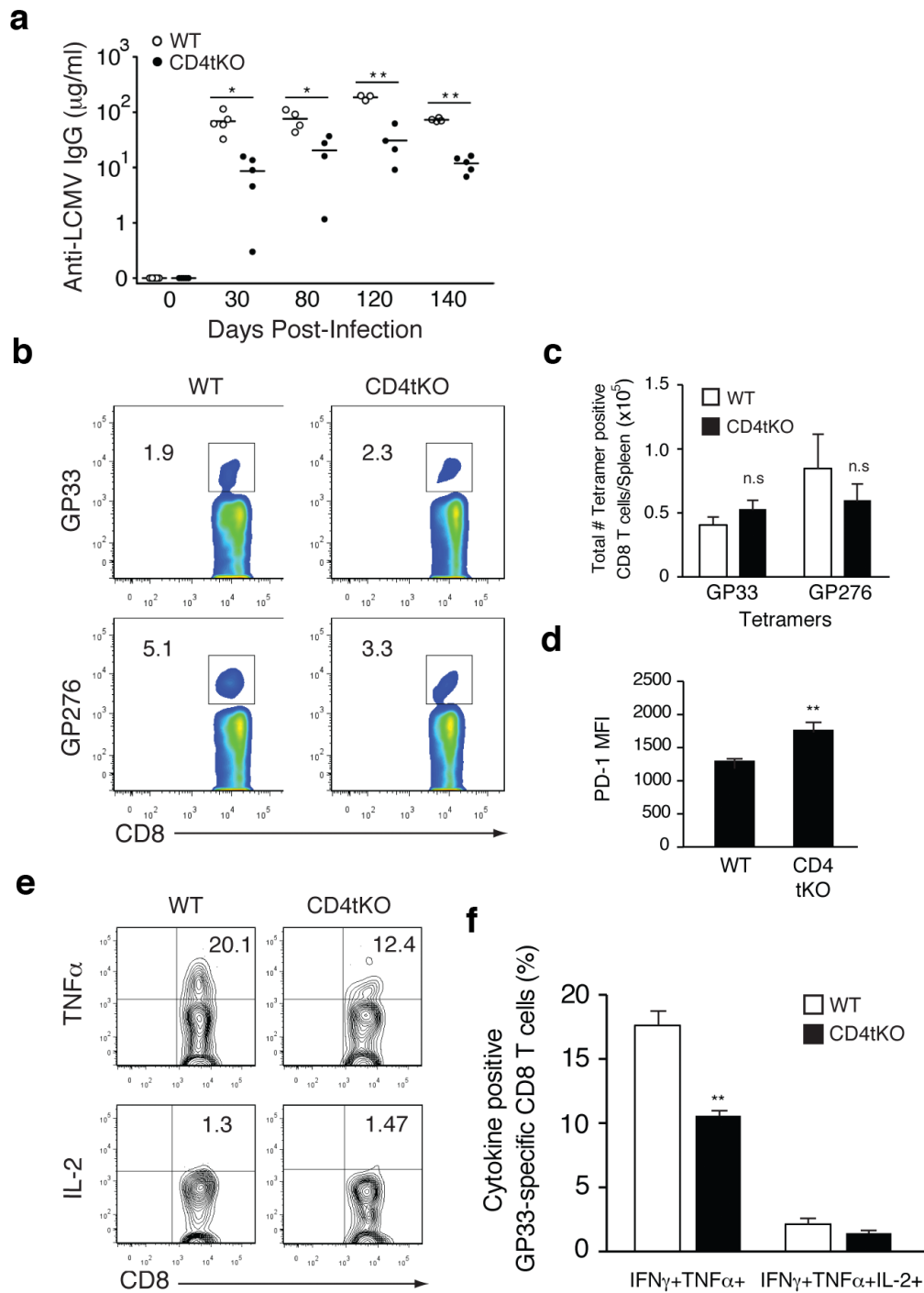
**Supplementary Figures 1-9**



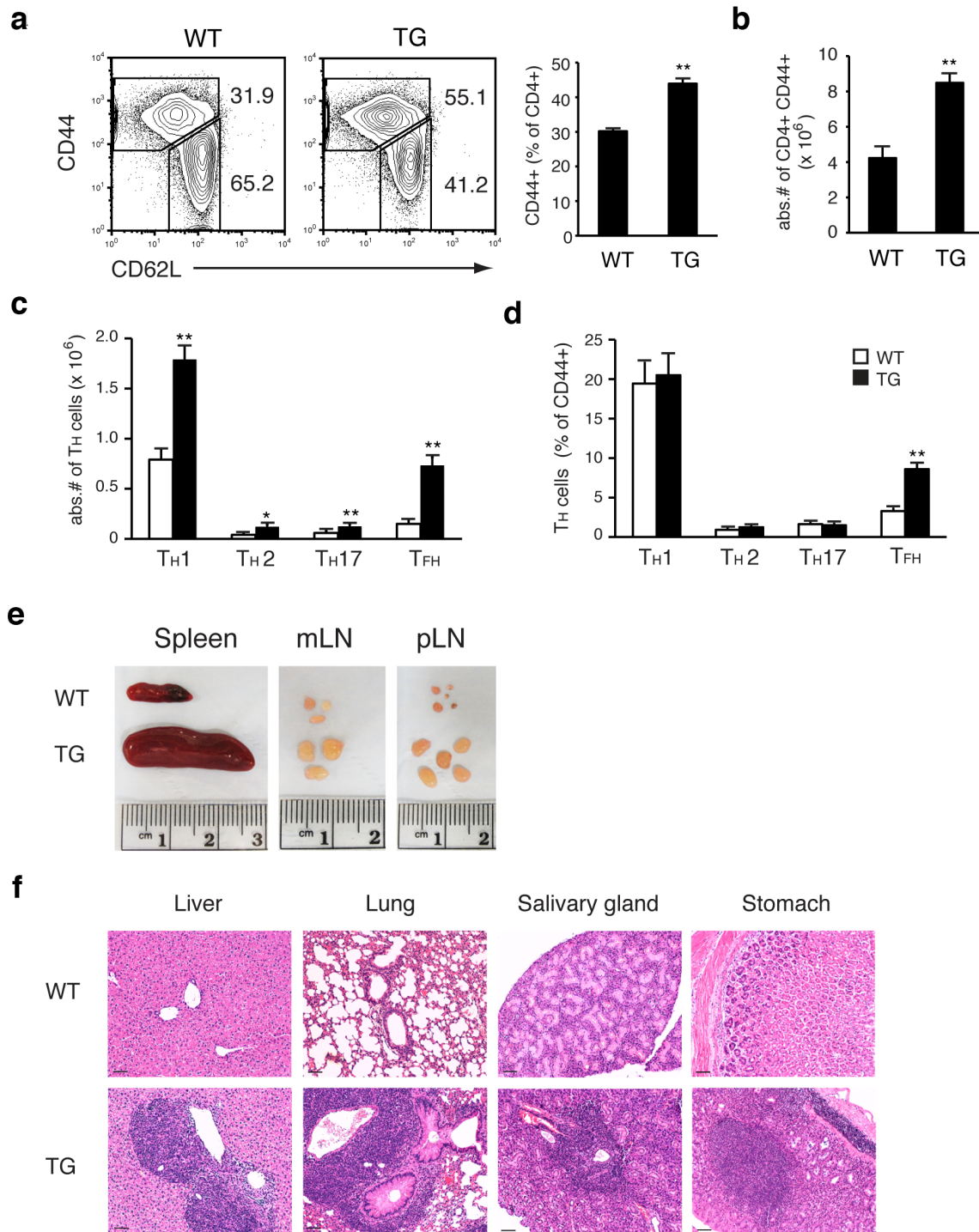
**Supplementary Figure 1. T<sub>FH</sub> differentiation, GCB formation, and antibody production in dKO mice.** (a,b) Flow cytometry analysis of T<sub>FH</sub> (a) and GCB (b) cell differentiation in WT and dKO mice at days 7 and 14 after immunization with NP-OVA+Alum+LPS (n=6 per group). (c) NP-specific IgG1 antibody production was determined by ELISA at indicated times after NP-CGG+Alum immunization (i.p). WT and CD4tKO results are also depicted in Fig. 1f. (d) Mixed bone marrow chimeras (Ly5a WT:Ly5b dKO=1:1) were immunized with NP-OVA+Alum+LPS (i.p). T<sub>FH</sub> (upper) and GCB (lower) cell differentiation was analyzed at day 7 after immunization. All graphs are shown as means ± s.e.m. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .



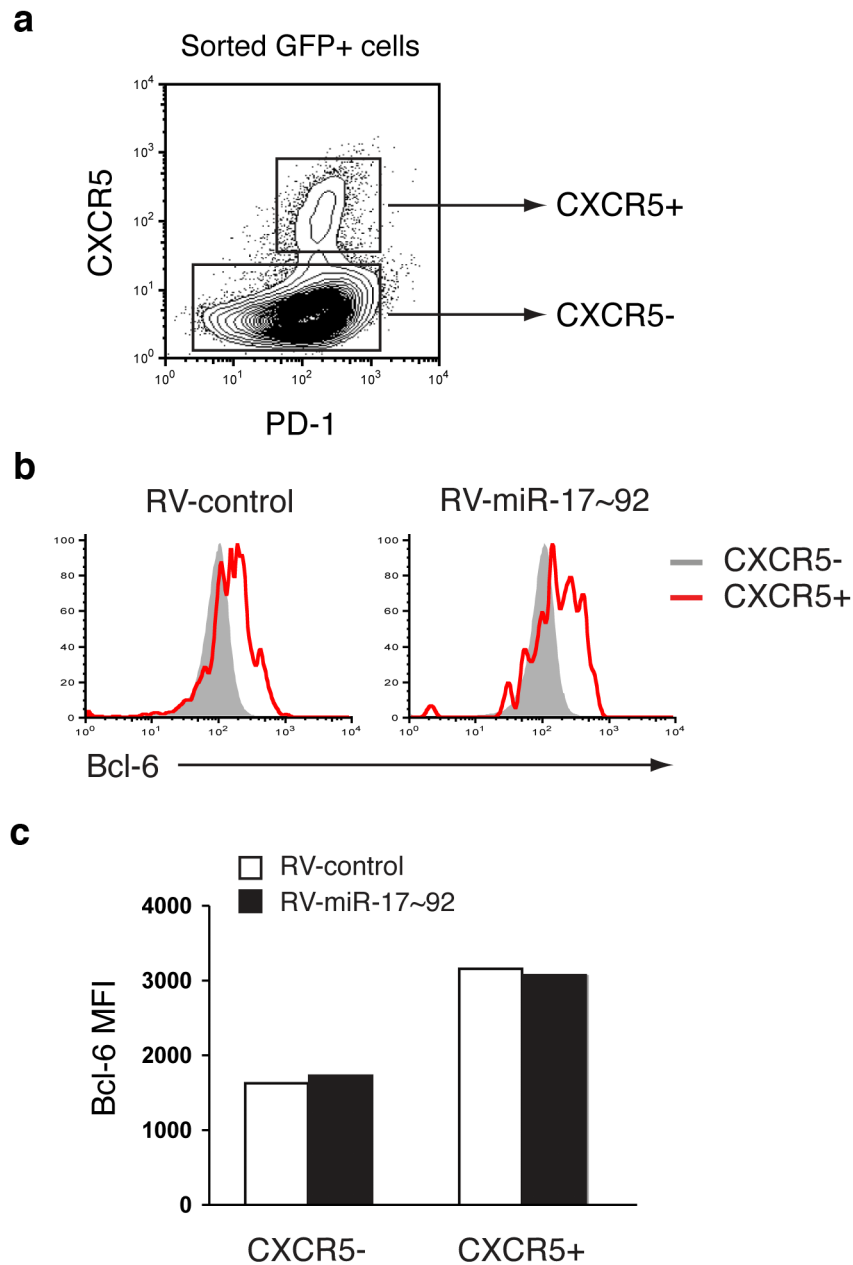
**Supplementary Figure 2. T<sub>FH</sub> and GCB differentiation in miR-17~92 single KO mice.** Flow cytometry analysis of CXCR5<sup>+</sup>PD1<sup>+</sup> T<sub>FH</sub> (upper), CXCR5<sup>+</sup>Bcl-6<sup>+</sup> T<sub>FH</sub> (middle), and FAS<sup>+</sup>GL-7<sup>+</sup> GCB (lower) cell development in WT and sKO mice (CD4Cre;miR-17~92<sup>fl/fl</sup>) at day 7 after immunization with NP-OVA+Alum+LPS (i.p.) (n=3 per group). All graphs are shown as means ± s.e.m. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .



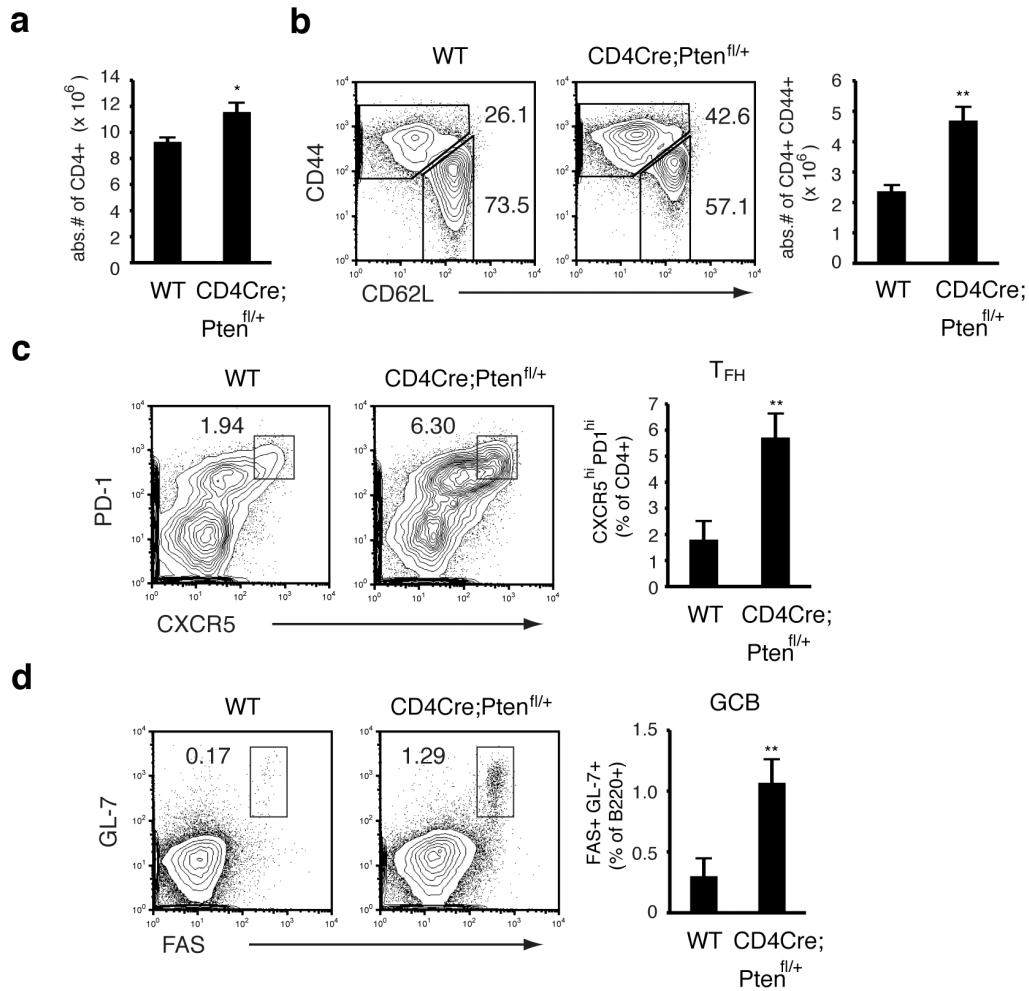
**Supplementary Figure 3. Immune responses in CD4tKO mice during chronic virus infection.** (a) Anti-LCMV IgG antibody concentration was determined by ELISA at indicated time points after LCMV C13 infection. (b,c) Frequency (b) and total numbers (c) of GP<sup>33-44</sup>- and GP<sup>276-286</sup>-specific CD8<sup>+</sup> T cells at day 30 following LCMV clone-13 infection. (d) Mean fluorescent intensity (MFI) of PD-1 expression on GP<sup>33-44</sup>-specific CD8<sup>+</sup> T cells at day 30 post-infection. (e) Flow cytometry analysis of GP<sup>33-44</sup>-specific CD8<sup>+</sup> T cell cytokine production. Graphs are gated on IFN- $\gamma$ + GP<sup>33-44</sup>-specific CD8<sup>+</sup> T cells. (f) Percentages of cytokine producing GP<sup>33-44</sup>-specific CD8<sup>+</sup> T cells at day 30 post infection. All graphs are shown as means  $\pm$  s.e.m. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .



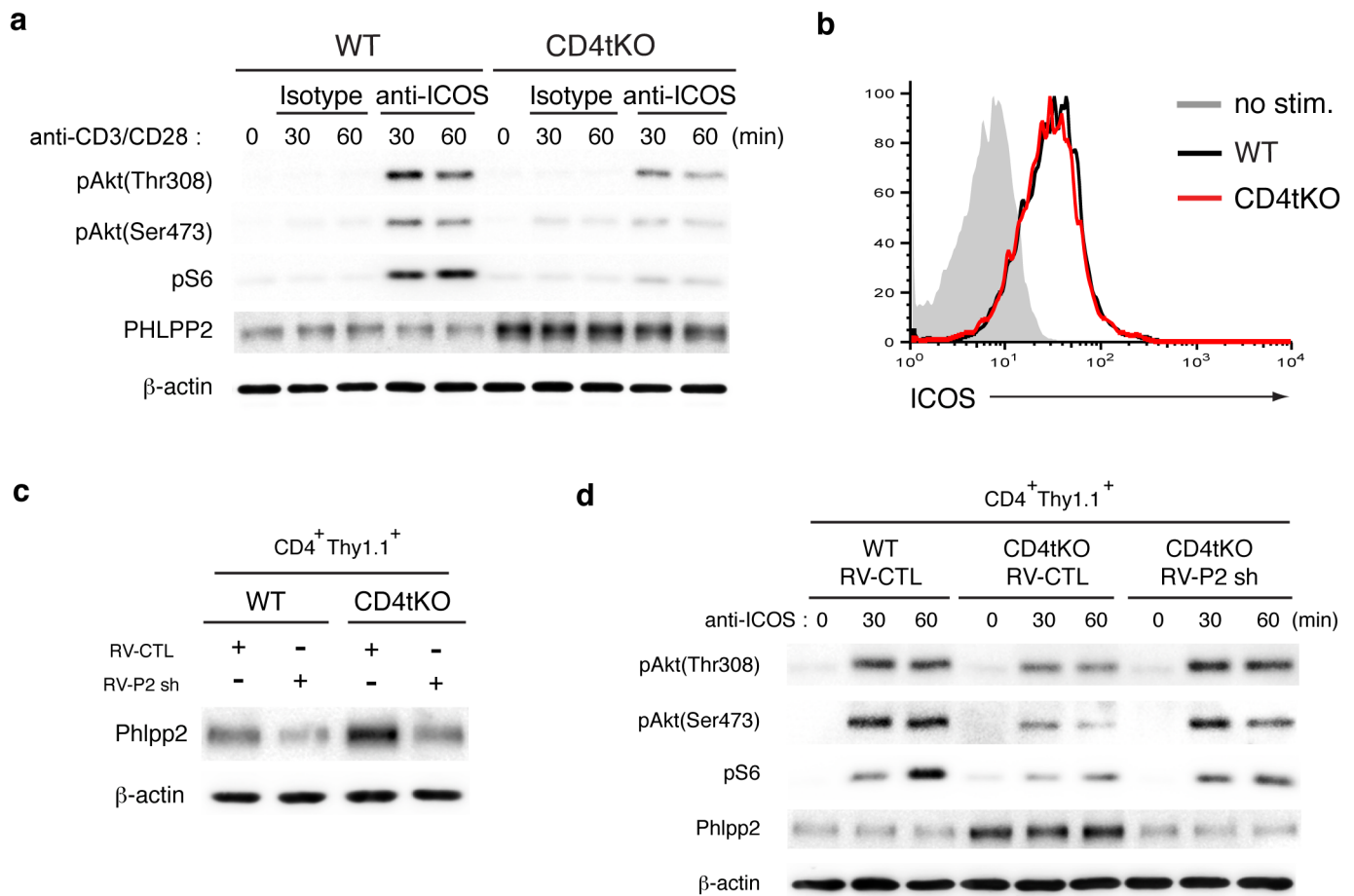
**Supplementary Figure 4. T cell-specific miR-17~92 transgenic mice developed fatal immunopathology.** (a-d) The activation status of CD4<sup>+</sup> T cells (a), CD4<sup>+</sup>CD44<sup>+</sup> T cell numbers (b), total numbers (c) and frequency (d, plotted as percentages of CD4<sup>+</sup>CD44<sup>+</sup> T cells) of T<sub>H</sub>1, T<sub>H</sub>2, T<sub>H</sub>17, and T<sub>FH</sub> cells in the spleen of 6-8 week old non-immunized mice. (e) Splenomegaly and lymphadenopathy in 6-8 month old TG mice. mLN, mesenteric lymph nodes. pLN, peripheral lymph nodes. (f) Hematoxylin and eosin staining of liver, lung, salivary gland, and stomach of 6-8 month old mice. Note mononuclear cell infiltration in non-lymphoid tissues in TG mice. Scale bar, 100  $\mu$ m. All graphs are shown as means  $\pm$  s.e.m. \*,  $p < 0.05$ ; \*\*,  $p < 0.01$ .



**Supplementary Figure 5. Bcl-6 expression in miR-17~92 over-expressing CD4<sup>+</sup> T cells.** WT naïve CD4<sup>+</sup> T cells were transduced with RV-control or RV-miR-17~92 and activated *in vitro* for T<sub>FH</sub> differentiation. GFP<sup>+</sup> cells were sorted and stained for CXCR5, PD1 and Bcl-6. Expression level of Bcl-6 in CXCR5<sup>+</sup>PD1<sup>+</sup> T<sub>FH</sub> or CXCR5<sup>+</sup>PD1<sup>+</sup> T cells (a) was examined by flow cytometry analysis and presented as histogram (b) and mean fluorescent intensity (c) (representatives of two independent experiments).

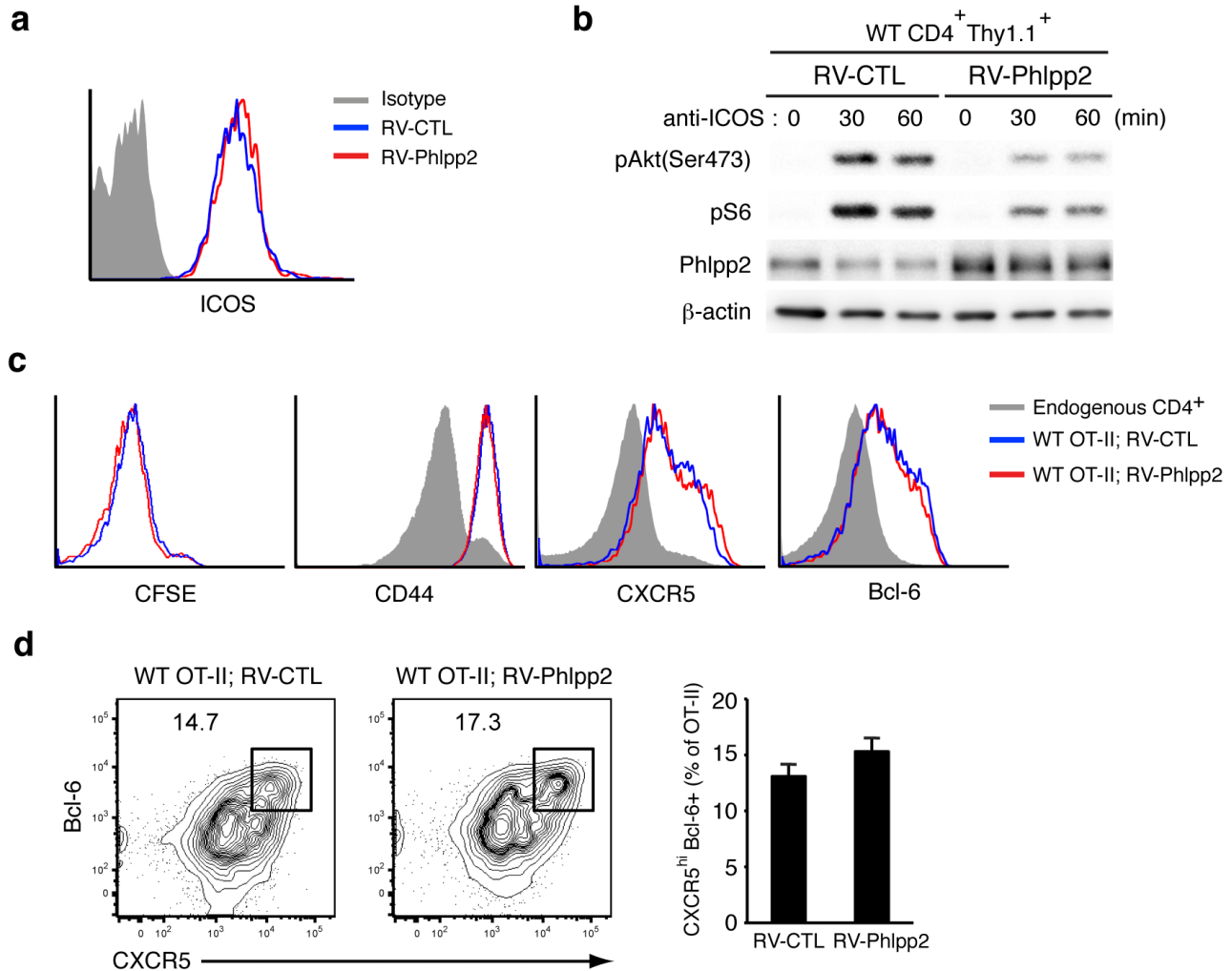


**Supplementary Figure 6. T cell-specific deletion of one copy of the *Pten* gene causes spontaneous germinal center reaction. (a-d)** Flow cytometry analysis of 8~9 month old non-immunized WT and CD4Cre;*Pten*<sup>fl/+</sup> mice (n=8 for WT, n=13 for CD4Cre;*Pten*<sup>fl/+</sup>). Total numbers of CD4<sup>+</sup> (a) and CD4<sup>+</sup>CD44<sup>+</sup> (b) T cells in the spleen are summarized in bar graphs. Flow cytometry analysis of CD4<sup>+</sup>CD44<sup>+</sup> (b), CXCR5<sup>hi</sup> PD-1<sup>hi</sup> T<sub>FH</sub> (c) and FAS<sup>+</sup>GL-7<sup>+</sup> GCB cells (d) in the spleen. (b-c), gated on CD4<sup>+</sup> T cells. (d), gated on B220<sup>+</sup> B cells. All graphs are shown as means ± s.e.m. \*, *p* < 0.05; \*\*, *p* < 0.01.

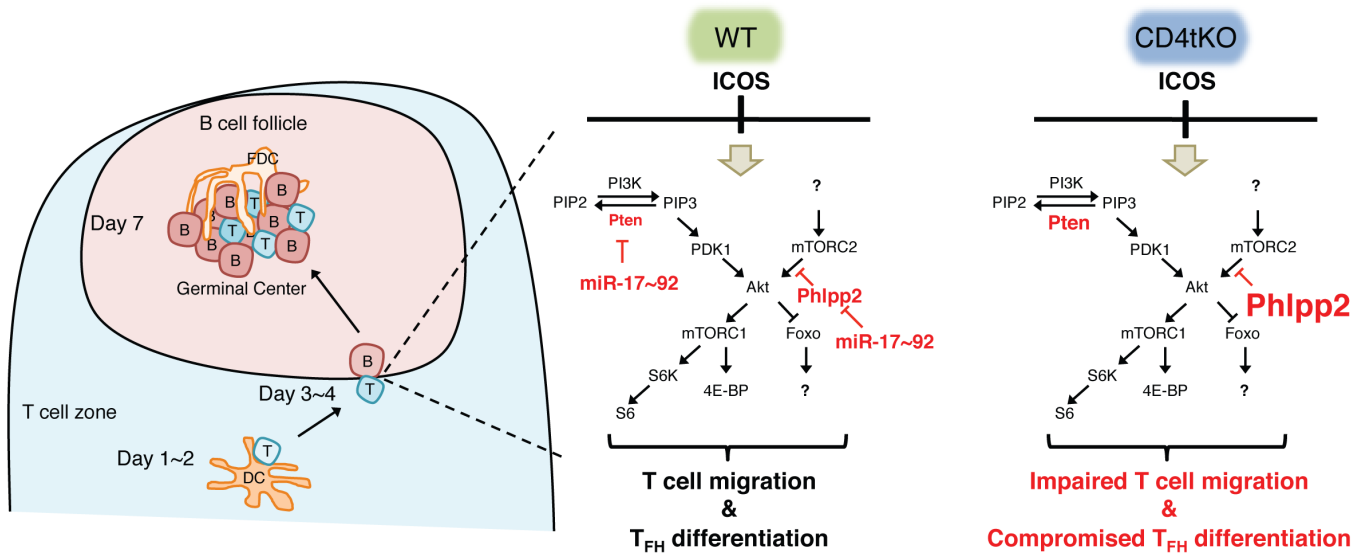


**Supplementary Figure 7. Phlpp2 knockdown restores ICOS-mediated PI3K signaling in CD4tKO CD4<sup>+</sup> T cells.** (a) WT and CD4tKO naïve CD4<sup>+</sup> T cells were activated *in vitro* with anti-CD3/CD28 for 3 days and rested in serum free media for 3 hours. Cells were treated with agonist hamster anti-ICOS or isotype antibody followed by anti-hamster IgG antibody treatment for indicated amounts of time. pAkt, pS6 and Phlpp2 levels were examined by Western blot. (b) CD4<sup>+</sup> T cells from (a) were examined for ICOS expression level on cell surface by flow cytometry. (c,d) *in vitro* activated WT and CD4tKO CD4<sup>+</sup> T cells were transduced with Thy1.1-expressing RV-CTL or RV-Phlpp2 shRNA (RV-P2 sh) retrovirus and Thy1.1<sup>+</sup> cells were isolated at day 3 after activation and infection and examined for Phlpp2 protein level by Western blot (c). Isolated Thy1.1<sup>+</sup> cells were rested for 3 hours in the absence of activation, re-stimulated with anti-ICOS, and analyzed as described in (a). Data are representatives of two (b) and three (a, c and d) independent experiments.





**Supplementary Figure 8. Phlpp2 overexpression impairs ICOS-PI3K signaling, but does not affect CD4<sup>+</sup> T cell proliferation, activation, or CXCR5 and Bcl-6 expression.** (a-b) *in vitro* activated WT naive CD4<sup>+</sup> T cells were transduced with Thy1.1-expressing RV-CTL or RV-Phlpp2 retrovirus. Thy1.1<sup>+</sup> cells were purified on day 3 after activation and infection, rested for 3 hours in serum free media. ICOS expression on cell surface was examined by flow cytometry (a). Rested cells were stimulated with anti-ICOS for indicated amounts of time, and pAkt and pS6 were examined by Western blot (b). (c-d) Thy1.1<sup>+</sup> cells were purified from RV-CTL or RV-Phlpp2 retrovirus infected WT OT-II CD4<sup>+</sup> T cells, labeled with CFSE, adoptively transferred into B6.Ly5a mice, followed by s.c. OVA+Alum immunization. OT-II CD4<sup>+</sup> T cell proliferation and expression of surface markers and Bcl-6 were examined on day 4 after immunization in draining lymph nodes. Graph is shown as means ± s.e.m.



**Supplementary Figure 9. Schematic depiction of miR-17~92 family miRNA-mediated regulation of  $T_{FH}$  differentiation.** The expression levels of both miR-17~92 family miRNAs and their target gene Phlpp2 are low in naive  $CD4^+$  T cells. They are significantly induced upon T cell activation, peak around day 2, and are downregulated afterwards. The upregulation of miR-17~92 family miRNA expression serves to limit the activation-induced Phlpp2 protein to a proper range. In CD4tKO  $CD4^+$  T cells, the Phlpp2 protein level remains high on day 3, resulting in compromised ICOS-PI3K signaling, and inability of T cells to migrate into B cell follicles and to differentiate into  $T_{FH}$  cells. Note that the font sizes of Pten, Phlpp2, and miR-17~92 indicate their relative expression levels during  $T_{FH}$  differentiation.