SUPPLEMENTARY INFORMATION

Hamartin confers neuroprotection against ischemia by inducing autophagy

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Experimental design of proteomic experiments. Rats were subjected to either sham ischemia or 10 min ischemia. Following 24 h of reperfusion, the CA1 and CA3 regions were microdissected and each region was subcellularly fractionated by differential centrifugation to generate a cytoplasmic and a membrane fraction. Each fraction was quantitatively analysed to define its proteome using UPLC-QTOF MS.

Purity of subcellular fractions. Microdissected hippocampal tissue was subjected to subcellular fractionation to derive a cytoplasmic (C) and a membrane (M) fraction. The purity of the fractions was assessed by immunobloting using nuclear (Histone H3), plasma membrane (Na+/K+ ATPase), and endoplasmic reticulum (ER; protein disulfide isomerase (PDI), Calnexin) markers. The figure shows representative immunoblots from n=3, demonstrating that neither the cytoplasmic nor the membrane fraction was contaminated with nuclear proteins. In addition, the immunoreactivity of the plasma membrane and organelle markers was limited to the membrane fraction.

Ontological analysis of enriched proteins in the CA1 and CA3 regions of sham ischemic animals. Using the proteomic datasets, proteins that were overexpressed in the CA1 or the CA3 region of sham ischemic rats were ontologically analyzed using the panther classification system according to both the biological process and the molecular pathway they participate. The bar graph shows the number of protein that belong to each group for the CA3 (black-filled bars) or the CA1 (white-filled bars) region. The ten biological processes and ten molecular pathways containing most proteins and those that were selectively affected either in the CA1 or CA3 region are displayed.

Ontological analysis of proteins with altered expression within the CA1 or CA3 by ischemia and reperfusion. (**a**) Bar graph shows the percentage increase of the five mostly upregulated proteins within the CA1 or CA3 regions, after ischemia and reperfusion, as determined by the proteomic analysis. (**b**,**c**) The proteins whose expression was increased (**b**) or decreased (**c**) by ischemia within the CA1 (white-filled columns) or the CA3 (black-filled columns) region were analyzed with the panther classification system, and were grouped according to both their biological process and their molecular pathway. Both bar graphs show the number of proteins belonging to each group. The ten biological processes and ten molecular pathways containing most proteins and those that were selectively affected either in the CA1 or CA3 region are displayed.

Supplementary Figure 5

Network of genes affected in the CA1 after ischemia. Diagram of a protein network, which was significantly associated with the expression level changes taking place by ischemia in the CA1 region. Proteins in red and yellow were upregulated and downregulated, respectively, by ischemia. Proteins in grey were detected by the proteomic analysis, but their expression was unaffected. Proteins in white participate in the network, but were not detected by the proteomic experiments. Images were created using the IPA software. The relationships between the displayed proteins are indicated.

Selective induction of 14-3-3 theta expression in the CA3 region after ischemia. (**a**) Representative immunoblots illustrating the protein expression levels of 14-3-3 theta in CA1 and CA3 cytoplasmic fractions from both sham ischemic and ischemic animals. (**b**,**c**) Bar graphs summarizing the immunoblotting (IB) and proteomic results for 14-3-3 theta in the CA1 (**b**) and CA3 (**c**) region. The values were corrected to total protein content, determined by Ponceau S staining and normalized such that the expression levels from sham ischemic samples were 1 and are means ± S.E.M. for n=6 per group (***p*<0.01, two-tailed t-test).

Selection criteria for the identification of individual proteins associated with the resistive properties of CA3. Panel shows the selection criteria applied to the proteomic datasets for the identification of individual proteins that were differentially regulated after ischemia in CA1 compared to CA3. (**1**) downregulated following ischemia in the CA1, but upregulated in the CA3 or (**2**) vice versa; (**3**) unaffected following ischemia in the CA1, but either upregulated or downregulated in the CA3 or (**4**) vice versa.

Hamartin induction in the CA1 following IPC and ischemia. (**a**) Experimental design showing rats subjected to either sham IPC or 2 min IPC, followed by 72 h of reperfusion, 10 min injurious ischemia and 24 h of reperfusion. In (**b–d**), the CA1 and CA3 membrane fractions were analyzed by immunoblotting using anti-hamartin antibodies. (**b**) Representative immunoblots showing hamartin expression. (**c**,**d**) Quantification of hamartin expression from the immunoblotting data for the CA1 (**c**) and CA3 (**d**) regions. The values were corrected to total protein content, determined by Sypro staining and were normalized such that expression levels from sham IPC samples was 1 and are means ± S.E.M. for n=4 per group (***p*<0.01, two-tailed t test).

Suppression efficiency of TSC1 shRNA treated cultures. (**a**) Phase contrast and fluorescent images of primary hippocampal neurons transduced with a control pLKO.1 TurboGFP vector (n=2). Merge shows the phase contrast and GFP fluorescence images. Scale bar is 50 µm. (**b**) Representative immunoblots showing hamartin expression in untransduced, control shRNA- and TSC1 shRNA-treated cultures. (**c**) Densitometric quantification of hamartin expression levels from (**b**). Tubulin was used as a loading control and expression was normalised so that hamartin immunoreactivity in untransduced cultures was 1 and are means ± S.E.M (n=4; one-way ANOVA with Bonferonni post-test, ***p*<0.01).

Increased vulnerability to OGD in primary cortical neurons in which hamartin expression was suppressed. (**a**) Phase contrast and immunofluorescent images of primary cortical neurons transduced with pLKO.1 TurboGFP vectors. Merge shows the phase contrast and GFP fluorescence images. (**b**) Representative immunoblots of hamartin expression from untreated, control shRNA - and TSC1 shRNA-treated cultures. (c) Bar graph summarizing the immunoblotting results. Expression is normalized so that hamartin immunoreactivity in untransduced cultures was 100%. (**d–h**) Cortical cultures were transduced 7 DIV with lentiviral vectors or remained untreated. At 14 DIV cultures were subjected to 2 h of either OGD or normoxia and cell death was quantified at 24 h of reperfusion. (**d**) Illustration of the experimental design. (**e**) Representative phase contrast images of cortical cultures transduced with TSC1 shRNA after 2 h OGD and 24 h of reperfusion. (**f**) Quantification of cell death by LDH assays showing % cell death in cultures treated with TSC1 shRNA, control shRNA and untransduced cultures. (**g**) Merged fluorescent images from cultures stained with Hoechst 33342 (blue) and ethidium homodimer III (red). (**h**) Quantification of cell death expressed as the percentage of neurons in (**g**) stained with ethidium homodimer III to stained with Hoechst 33342. Scale bars are 50 μ m. All data are expressed as means \pm S.E.M. from n=3 (one-way ANOVA with Bonferroni post-test, **p*<0.05, ****p*<0.001).

Rescue of hamartin expression in hippocampal cultures. Representative immunoblot showing hamartin expression from hippocampal cells treated with both TSC1 shRNA and pEZ-Lv201 human TSC1 lentiviral vectors. GFP is pEZ-LV201 eGFP. Tubulin is loading control.

Supplementary Figure 12

Attenuation of hamartin expression in CA3 cells. (**a**) Representative fluorescence images from coronal sections immunostained with an anti-GFP antibody (green) and counterstained with DAPI (blue) following unilateral injection of control pLKO.1 TurboGFP vectors (2.5 x 106 particles) in CA3, which produced ipsilateral GFP expression. (b) Representative immunoblot from CA3 whole cell homogenates showing hamartin expression and (c) bar graph summarizing the densitometric data. Data were normalised to hamartin immunoreactivity from naïve CA3 cell homogenates. (**d**) Immunofluorescent images from brain sections immunostained with an anti-hamartin antibody (red) and DAPI (blue) (n=3). For (**a**) scale bars are 0.1 mm for first row panels and 0.01 mm for lower panels. For (**d**) scale bars are 0.5 mm for first row panels and 0.1 mm for lower panels. Boxes indicate the CA3 regions magnified in the lower panels.

Reduction of resistance to ischemia in CA3 cells in which hamartin expression was suppressed. (**a**) Hematoxylin and eosin stained hippocampal sections from rats administered bilaterally with either TSC1 shRNA (n=6) or Control shRNA (n=4) and after 14 d subjected to 10 min ischemia and 7 d reperfusion. Arrow heads show the needle trace. Boxes indicate the magnified regions displayed in the middle and lower panels. Scale bar for upper panels is 1 mm and for middle and lower panels is 0.01 mm. (**b**) Quantification of the number of healthy neurons per mm surviving ischemia in the dorsal CA3 pyramidal layer, treated with TSC1 shRNA, compared to control shRNA-treated or uninfected hippocampi subjected to sham ischemia. Data are mean ± S.E.M. (one-way ANOVA with Bonferonni post-test, ***p*<0.01)

Supplementary Table 1. Enhanced proteins in the CA1 region compared to the CA3 region of sham ischemic animals.

Supplementary Table 2. Enhanced proteins in the CA3 region relative to the CA1 region of sham ischemic animals.

Supplementary Table 3. Proteins upregulated within the CA1 region following ischemia and reperfusion.

Supplementary Table 4. Proteins upregulated within the CA3 region following ischemia and reperfusion.

Supplementary Table 5. Proteins downregulated within the CA1 region after ischemia and reperfusion.

Supplementary Table 6. Proteins downregulated within the CA3 region following ischemia and reperfusion

Supplementary Table 7. Proteins composing the pathway of Figure 1a

Supplementary Table 8. Proteins composing the network of Figure 1b

Supplementary Table 9. Differentially expressed proteins in CA1 compared to CA3 region after ischemia and reperfusion.

Table showing a list of proteins shortlisted from the proteomic datasets using the criteria described in **Supplementary Fig. 7**. CA1 S is CA1 sham ischemia; CA1 I is CA1 ischemia; CA3 S is CA3 sham ischemia; CA3 I is CA3 ischemia. Red boxes show increase, yellow boxes indicate decrease, whereas, grey boxes represent no change in the expression levels between the groups.

Supplementary Results and Discussion

Protein expression profile within the CA1 and CA3 regions under sham conditions

Proteomic analysis of the cytoplasmic and membrane fractions from sham ischemic animals identified 1,066 proteins that were present in both the CA1 and CA3 regions of the hippocampus. From those, 137 (13%) had higher expression in the CA1 region compared to the CA3 region **(Supplementary Table 1)** and 139 (13%) were enriched within the CA3 relative to the CA1 region **(Supplementary Table 2)**.

The datasets were analyzed ontologically using the panther classification system (www.pantherdb.org). For each region, the proteins with enhanced expression were grouped according to the molecular pathway or the biological process in which they participate. The number of proteins present in each molecular pathway or biological process was compared between the two regions. As a result, we identified pathways and processes that had a different number of enhanced proteins in the CA1 compared to the CA3 region.

Thirteen transporter proteins showed higher expression levels in the CA3 region, whereas in the CA1 there were 2 transporter proteins with enhanced expression relative to the CA3 region (**Supplementary Fig. 3**). Interestingly, one of these 13 transporters was the Ca^{2+} transporting ATPase, which is involved in calcium homeostasis^{1, 2}. Its expression was enhanced $38\pm14\%$ in the CA3 region compared to the CA1 region ($p<0.05$). This probably indicated an enhanced innate ability of CA3 cells to regulate Ca^{2+} levels and limit excitotoxicity. Overall, in both groups a similar number of proteins was enhanced between the two regions.

The effect of ischemia and reperfusion on the proteome profile of the CA1 and CA3 regions

In the CA1 region, 1,101 proteins were found to be present in both sham ischemic and ischemic rats, and 1,062 proteins were present the CA3 region of both sham ischemic and ischemic rats.

Profile of upregulated proteins following ischemia and reperfusion

The profile of the proteins that were either upregulated or downregulated within the CA1 region following 10 min ischemia and 24 h of reperfusion was compared to the corresponding profile of the CA3 region using ontological analyses.

The number of proteins upregulated by ischemia and reperfusion was 114 (10%) in the CA1 (**Supplementary Table 3**) and 137 (13%) in the CA3 region (**Supplementary Table 4**). In the CA1 region, proteins with the highest upregulation included serine threonine protein kinase 35 (STK35; $300\pm34\%$; $p<0.05$) and glucose-6-phosphate dehydrogenase (G6PD; $127 \pm 27\%$; *p*<0.05) (**Supplementary Fig. 4a**).

In the CA3 region, proteins that were most highly upregulated as a consequence of ischemia were lon protease homolog (LONP1; $1,566\pm87\%$; $p<0.05$), Rho GTPase-activating protein 6 $(ARHGAP6; 1.326 \pm 67\%; p<0.05)$ and advillin $(261 \pm 48\%; p<0.05)$ (**Supplementary Fig. 4a**). LONP1 is a component of the proteolytic mitochondrial system which is inactivated by peroxynitrite³. It may be that CA3 cells are able to degrade abnormal proteins within the mitochondria more efficiently than the CA1 cells, preserving the vital integrity of mitochondria. ARHGAP6 is a Rho GTPase-activating protein which regulates cytoskeletal organization³. Finally, advillin is a calcium-regulated acting-binding protein that is implicated in neuronal cell morphogenesis⁴.

An ontological analysis was carried out using the panther classification to compare the profile of proteins upregulated by ischemia and reperfusion between the CA1 and the CA3 region (**Supplementary Fig. 4b**). In the CA3 region, 6 of the proteins upregulated were involved in apoptosis, with 4 being inhibitors of apoptosis**,** while 2 apoptosis inhibitors were found upregulated in the CA1 region. Additionally, Na^{+}/K^{+} ATPase was the only protein involved in homeostasis that was upregulated by ischemia in the CA3 region $(12\pm3.4\%, p<0.05)$. This protein regulates cellular homeostasis by restoring the resting potential⁵, and might constitute a mechanism which allows the CA3 region to prevent overexcitability and subsequent excitotoxicity following ischemia.

Profile of downregulated proteins following ischemia and reperfusion

Ischemia and reperfusion resulted in the significant downregulation of 81 (7%) proteins in the CA1 (**Supplementary Table 5**) and 127 (12%) proteins in the CA3 region (**Supplementary Table 6**). According to the ontological analysis, the biological processes encompassing the largest number of proteins downregulated selectively in the CA3 region were "protein modification" (29 proteins) and "signal transduction" proteins (21 proteins). In addition, the molecular pathways which had the largest number of downregulated proteins in the CA1 region were transporters (10 proteins) and hydrolases (9 proteins), whereas in the CA3 region they were nucleic acid binding proteins (17 proteins) and kinases (13 proteins) (**Supplementary Fig. 4c**).

Protein network associated with the vulnerability of CA1 to ischemia

To further identify targets for future studies we investigated the protein network most significantly associated with the events induced in the CA1 after ischemia (**Supplementary Fig. 5**). Examination of this protein network revealed important insights and complemented previous studies explaining the vulnerability of CA1 cells to ischemia^{6, 7}. In particular, many of the proteins comprising this network regulate ionic and calcium homeostasis dictating the outcome of excitotoxicity. ATP1A1, ATP1A2 and ATP1A3 are Na^{+}/K^{+} ATPases that help maintain the resting potential⁸. Interestingly, all three were found downregulated in the CA1 by ischemia probably contributing to hyperexcitability. It has been suggested that the reduction in the expression levels of the glutamate transporter, GLT-1 in astrocytes exacerbates excitotoxicity due to accumulation of synaptic glutamate as uptake is suppressed. Further evidence for excitotoxic injury was provided by the downregulation of ATP2B3, ATP2B4 and calcium/calmodulin-dependent kinase II (CaMKII). ATP2B3 and ATP2B4 are $Ca²⁺$ ATPases that are essential to rectify changes in calcium levels resulting from excitotoxicity⁹. CaMKII inhibition enhances neuronal activity and increases sensitivity to excitotoxic glutamate signaling following ischemia 10 . Impairment of all these processes leads to the generation of free radicals, well established mediators of ischemic cell death, with superoxide mediating oxidative stress in the CA1 region by ischemia⁶. Protection from reactive oxygen species can be conferred by G6PD, which can act as a reducing agent¹¹. We found GP6D to be one of the mostly upregulated proteins in the CA1 following ischemia possibly denoting that the vulnerable CA1 neurons have to cope with oxidative stress overload after ischemia and reperfusion. Ultimately, induction of apoptosis is the endexecutor of cell death. Activation of caspases has been demonstrated as a detrimental step in the CA1 region¹². Although we did not detect activated caspases in our proteomic analysis, primarily since we did not analyze the nuclear fraction, their activity after ischemia has been shown to cleave Ca^{2+} ATPases resulting in calcium overload and cell death⁹. Overall, the biological importance of this network is in agreement with current knowledge explaining the vulnerability of CA1 neurons to ischemia.

Validation of the proteomic datasets by immunoblotting and comparison with current knowledge

We carried out immunoblotting analysis to validate 14-3-3 theta expression levels in the cytoplasmic fraction (**Supplementary Fig. 6a**). In agreement with the proteomic data, 14-3-3 theta expression levels were unaffected following ischemia in the CA1 region (**Supplementary Fig. 6b**), but were significantly upregulated in the CA3 region (**Supplementary Fig. 6c**).

In addition, our proteomic datasets are in keeping with previously published proteomic studies. Gozal and colleagues¹³ used 2D electrophoresis to characterize the CA1 and CA3 proteome from rats exposed to 6 h of intermittent hypoxia. Although the effect of hypoxia rather than ischemia was studied, a significant number of proteins exhibited an expression profile similar to the one shown in our study. For instance, beta tubulin, gamma enolase, and creatine kinase B were upregulated in the CA3 region, after either ischemia or intermittent hypoxia. Similarly, triose phosphate isomerase expression was found to be higher in the CA3 compared to the CA1 of control animals in both studies. The differences that exist between our study and the Gozal et al.¹³ study may be attributed mainly to the difference in the insult (intermittent hypoxia versus ischemia) and in the analytical techniques used (2D electrophoresis versus LC-based label-free quantitation).

Results from studies showing the expressed genome of CA1 and CA3 cells after ischemia in rats¹⁴ do not correlate closely with our results, with many of the entries identified in the genomic analyses not present in our data and vice versa. These differences could be a result of the experimental limitations of the proteomics as compared to the microarray studies. However, the current study shows similarities to genomic studies of human post-mortem tissue that compared the gene expression between the CA1 and the $CA3¹⁵$. For example, cytoskeletal proteins, such as actin, were found to be enriched in the CA3 region compared to

the CA1 in both studies. In agreement with our findings, microarray studies in mice¹⁶ revealed overexpression of phosphofructokinase and Ca^{2+} transporting ATPase in the CA3 region compared to the CA1 region.

Proteins associated with the CA3 resistance to ischemia

To identify individual protein candidates associated with the ischemia-resistive properties of CA3 neurons, we hypothesized that in order for a protein to be functionally relevant to the differential response of the CA1 and the CA3 to ischemia, its expression profile should differ between the two regions after ischemia. We used the proteomic datasets and applied the selection criteria illustrated in **Supplementary Fig. 7** (see **Supplementary Table 9**)**.** Critically, hamartin was selected using these criteria.

Methodology

Employing a label-free quantification of the proteome expression levels using groups of 5 animals allowed the detection of small expression changes with significant confidence. This relied upon the ability of the system to derive individual quantitative data for a particular protein from all the detected peptides corresponding to this protein and integrating them to overall protein levels.

Although we tried to optimize our experimental design both for the proteomics experiments and the subsequent analysis, we acknowledge the associated limitations. Group sizes of 5 were a limitation imposed by the capacity of the label-free quantitation system and by the vast amount of data generated from 20 samples (5 CA1 and 5 CA3 fractions from sham ischemic and ischemic samples). In addition, in our analysis we did not consider any posttranslational modifications since this would have increased the complexity of the analysis tremendously. Interpretation of proteomic data can often give rise to false positive

identification despite the use of rigorous filtering algorithms. However, the validation of our datasets by independent experiments and by comparison with published data provided additional confidence about the accuracy of our findings. In addition, proteomic analysis favors high copy number proteins, since protein abundance is a factor that determines the proteins identified^{17, 18}. This may have resulted in the failure to detect proteins that were expressed at very low levels.

Crude microdissection of the CA1 and CA3 regions resulted to mixed cell populations present within these regions. Unfortunately, isolating single cell subset does not provide enough material to run proteomics experiments of this magnitude and requires tissue postprocessing that can introduce artefactual changes in protein expression levels. One advantage of looking at all cell types is the creation of a more complete picture of the events of interest. Our study examined only the cytoplasmic and membrane fractions, and not the nuclear

fraction, as we were unable to avoid significant contamination in our attempts to obtain a pure nuclear fraction.

Finally, all our data were generated from animals killed at a single time point following reperfusion. The reasoning is that after 24 h of reperfusion, although the CA1 cells are still viable, the ischemic cascade has already been activated, allowing the investigation of the effects of ischemia on morphologically intact cells.

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