

1 Supplemental tables and figure Qiu et al., 2013

2 Table S1 Bacterial strains and plasmids

Strain	Description	Source or reference
<i>Escherichia coli</i> WM3064	<i>thrB1004 pro thi rpsL hsdS lacZDM15 RP4-1360 (araBAD)567 dapA1341::[erm pir(wt)]</i>	W. Metcalf (1)
<i>E. coli</i> TOP10	F2 <i>mcrA Δ(mrr-hsdRMS-mcrBC) Φ80lacZDM15 Δ lacX74 deoR recA1 araD139 Δ(ara-leu)7697 galU galK rpsL (Sm^r) endA1 nupG</i>	Invitrogen
<i>E. coli</i> EC100D+	F <i>mcrA Δ(mrr-hsdRMS-mcrBC) φ80dlacZAM15 ΔlacX74 recA1 endA1 araD139 Δ(ara, leu)7697 galU galK λ- rpsL (Str^R) nupG pir⁺(DHFR)</i>	Epilcentre Technologies
<i>Shewanella putrefaciens</i> W3-18-1	Dissimilatory metal-reducing strain (Pacific Ocean marine sediments)	Murray et al., 2001 (2)
W3-18-1ΔpstI	In-frame deletion mutant of pstI gene (Sputw3181_4075)	This study
W3-18-1ΔpstIM	In-frame deletion mutant of pstI gene and pstM (Sputw3181_4074)	This study
W3-18-1Δnapα	In-frame deletion mutant of <i>nap-α</i> (<i>napDABC</i>) genes (Sputw 3181_2102-2107)	This study
W3-18-1Δnapβ	In-frame deletion mutant of <i>nap-β</i> (<i>napDAGHB</i>) genes (Sputw3181_0792-0796)	This study
W3-18-1Δnapαβ	Double mutant with in-frame deletion mutant of <i>nap-α</i> and <i>nap-β</i> genes	This study
W3-18-1ΔcymA	In-frame deletion mutant of <i>cymA</i> (Sputw3181_3916)	This study
<i>S. oneidensis</i> MR-1	Dissimilatory metal-reducing strain (Lake Oneida, NY)	Myers & Nealson, 1988 (3)
Plasmid		
pDS3.0	Suicide vector derived from pCDV224; <i>Amp^r</i> , <i>Gm^r</i> , <i>sacB</i>	Wan et al., 2004 (4)
pBBR1MCS-2	Broad-host-range vector, <i>Km^r</i> , <i>lacZ</i>	Kovach et al., 1995 (5)
pBBR1MCS-5	Broad-host-range vector, <i>Gm^r</i> , <i>lacZ</i>	Kovach et al., 1995 (5)

3 **Table S2** Primers used in this study

Primer	Nucleotide sequence
Pstlko_5O	AGAGCTCTGGGCTGAATCGGTAAC TTC
Pstlko_5I	CTATGTCGGATGCTGGCCTAATGTCGGTTACTTGGCTTGG
Pstlko_3I	TAGGCCAGCATCCGACATAGTCTCTGCCAGAATCTTGCT
Pstlko_3O	AGAGCTCGTATCCGGGCAAGCTAAACA
Pstmko_5O	AGAGCTCGACTTAAAATCGCCAGGTCT
Pstmko_5I	CTATGTCGGATGCTGGCCTA ACCAAACAGTGAGCGACTAA
Pstko_Lf	TCAAGGCATTACGTGAGCTG
Pstko_Lr	AGAGCTCGCTTTCCACTTTTCAGGATCT
Napako_5O	AGAGCTCGGGCATTAAAAAGCGCCTTA
Napako_5I	GTTCACTGAACGACATAAAACCACTCATGCGTCTCCAAGA
Napako_3I	GTTTATGTCGTTTCAGTGA ACTATCTCCAGCCAAAGGCCAA
Napako_3O	AGAGCTCATAATCGAGGGGAAAATCCG
Napako_Lf	GCTCAAGATGCACACCGTCA
Napako_Lr	AAGATGCATTGGCATCAGGC
Napβko_5O	AGAGCTCGCCGTGACGGTAAAGTTGC
Napβko_5I	GACTGGCTTAGGTTCGTCTCTCGGGTGACCTTAGGTGGT
Napβko_3I	AGAGACGACCTAAGCCAGTCCCAATAGACGATACTCCCCTG
Napβko_3O	AGAGCTCGCAGCCAAATCCACCGTAT
Napβko_Lf	ATCCAAGGTTACCGTTTCGA
Napβko_Lr	CAAATTAGTGCAGATCCCA
CymAko_5O	AGCGTTTCATCGTTACAGCA
CymAko_5I	GATAACATTAAGTTACACC TGCACGCCAGTTCATTACTCT
CymAko_3I	GGTGTA ACTTAATGTTATCCTTTTGAGGGCTTTTTGCTG

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CymAko_3O	TGTA ^{CT} CGATTGTTTCGGCG
CymA_F	ACTCGAGCTAATTTGGCAATTTGGAGA
CymA_R	GGTCGACTTATCCTTTGGATATGGGTGA
crpko_5O	AGAGCTCATAAATCAGCCAGCGTTTGC
crpko_5I	TGCATCGAGTTGATTGTCGCTCAGAGCCATGTTGATGTTCC
crpko_3I	GCGACAATCAACTCGATGCAGTCTTCAGCTTGATTTAAGCCTG
crpko_3O	AGAGCTCTTACTGTTTGCTCCGTTTCA
crp_F	AGAATTCGAGGAACATCAACATGGCTCTG
crp_R	ACTGCAGAATTTATGCTAGGCCACTTTAATGA
RT-PCR primers	
16S-F	GTTGGAAACGACTGCTAATACC
16S-R	GGTCCTTCTTCTGTAGGTAACG
RT_napA α -F	TCTTTGGGTGGGTTTGTGGC
RT_napA α -R	CGGGGAGATGGTGGGCTATT
RT_napA β -F	GGATTAGGCTTAACGGCTACGGC
RT_napA β -R	TGTGGGTAAAAACATGGCGTG

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5 **Table S3** Comparison of *c*-type cytochromes between W3-18-1 and MR-1

No.	Cytochrome	Orthologous gene		Presence in the <i>Shewanella putrefaciens</i> CN-32 strains
		W3-18-1	MR-1	
1	Diheme <i>c4</i>	Sputw3181_0043	SO_4666	Sputcn32_3908
2	CoxIIc Diheme <i>c</i>	Sputw3181_0133	SO_4606	Sputcn32_3782
3	ScyA Monoheme <i>c5</i>	Sputw3181_0189	SO_0264	Sputcn32_3726
4	SHP Monoheme <i>c</i>	Sputw3181_0209	SO_4484	Sputcn32_0359
5	Diheme <i>c</i>	Sputw3181_0210	SO_4485	Sputcn32_0360
6	Tetraheme <i>c</i>	Sputw3181_0555	Absent	Sputcn32_3388
7	Hypothetical Seven-heme <i>c</i>	Sputw3181_0577	SO_0479	Sputcn32_3364
8	PetC-Monoheme <i>c1</i>	Sputw3181_0667	SO_0610	Sputcn32_3274
9	NapB Diheme <i>c</i>	Sputw3181_0792	SO_0845	Sputcn32_3151
10	Monoheme <i>c</i>	Sputw3181_1274	SO_3420	Sputcn32_2738
11	OmcA-like Decaheme <i>c</i>	Sputw3181_1501	Frameshift	Sputcn32_2507
12	Split Tetraheme Flavocytochrome	Sputw3181_1578	SO_3056	Sputcn32_2430
13	STC Small Tetraheme <i>c</i>	Sputw3181_1675	SO_2727	Sputcn32_2333
14	BCCP Diheme <i>c</i>	Sputw3181_1721	SO_2178	Sputcn32_2287
15	FixO/CcoO Monoheme <i>c</i>	Sputw3181_2047	SO_2363	Sputcn32_1958
16	FixP/CcoP Diheme <i>c</i>	Sputw3181_2049	SO_2361	Sputcn32_1956
17	NapC Tetraheme	Sputw3181_2103	Absent	Sputcn32_1905
18	NapB Diheme <i>c</i>	Sputw3181_2104	Absent	Sputcn32_1906
19	Tetraheme Cytochrome <i>c3</i>	Sputw3181_2184	Absent	Sputcn32_1825
20	OmcA-like 11 heme <i>c</i> (UndA1 or OmcE)	Sputw3181_2622	SO_1779 (OmcA1)	Sputcn32_1479
21	MtrC/OmcB Decaheme <i>c</i>	Sputw3181_2623	SO_1778	Sputcn32_1478
22	MtrA Decaheme <i>c</i>	Sputw3181_2624	SO_1777	Sputcn32_1477
23	OmcA-like Decaheme <i>c</i>	Sputw3181_2721	SO_1659	Sputcn32_1380

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24	Decaheme Cytochrome <i>c</i>	Sputw3181_2977	Absent	Sputcn32_1187
25	FccA Tetraheme Flavocytochrome	Sputw3181_3267	SO_0970	Sputcn32_0905
26	Split-Soret Diheme	Sputw3181_3294	SO_0939	Sputcn32_0879
27	NrfA Pentaheme <i>c</i>	Sputw3181_3486	SO_3980	Sputcn32_0685
28	SoxA-like Diheme <i>c</i>	Sputw3181_3546	SO_4047	Sputcn32_0624
29	Diheme <i>c4</i>	Sputw3181_3547	SO_4048	Sputcn32_0623
30	NrfA-like Pentaheme <i>c</i>	Sputw3181_3743	Absent	Sputcn32_3604
31	NrfB Pentaheme <i>c</i>	Sputw3181_3895	Deleted	Sputcn32_0313
32	CymA Tetraheme <i>c</i>	Sputw3181_3916	SO_4591	Sputcn32_0286
33	Monoheme <i>c</i> , Acceptor for Molybdopterin	Absent	SO_0714	Absent
34	Monoheme <i>c</i>	Absent	SO_0716	Absent
35	Monoheme <i>c4</i>	Absent	SO_0717	Absent
36	TorC pentaheme <i>c</i>	Absent	SO_1233	Absent
37	Split Tetraheme flavocytochrome	Absent	SO_1413	Absent
38	IfcA Tetraheme flavocytochrome	Disrupted by transposon	SO_1421	Sputcn32_0810 (Sputw3181_3363 was disrupted by insertion of sputw3181_3364)
39	MtrAD-like Decaheme <i>c</i>	Absent	SO_1427	Absent
40	MtrF Decaheme <i>c</i>	Absent	SO_1780	Absent
41	MtrD Decaheme <i>c</i>	Absent	SO_1782	Absent
42	Hypothetical Diheme <i>c</i>	Absent	SO_2930	Absent
43	Hypothetical Diheme <i>c</i>	Absent	SO_2931	Absent
44	Split Tetraheme Flavocytochrome	Absent	SO_3300	Absent
45	Split Tetraheme Flavocytochrome	Absent	SO_3623	Sputcn32_3535

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46	Mono-heme <i>c</i>	Absent	SO_4142	Absent
47	Octaheme Tetrathionate Reductase (Otr)	Absent	SO_4144	Absent
48	MtrAD-like Decaheme	Absent	SO_4360	Absent
49	Mono-heme <i>c</i>	Absent	SO_4570	Absent
50	Triheme <i>c</i>	Absent	SO_4572	Absent

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7 **Table S4** Comparison of respiration chain components and related genes between MR-1 and
8 W3-18-1

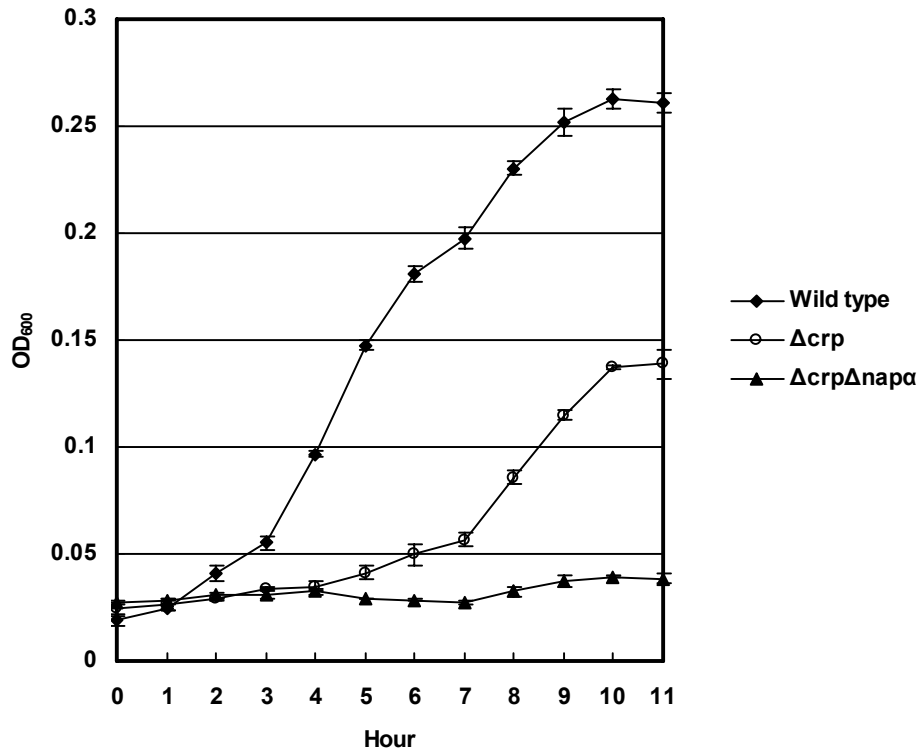
Gene or gene cluster	Functions	Locus		Presence in the CN-32 strain
		W3-18-1	MR-1	
<i>fdh-N</i> operon	Nitrate reductase-linked formate dehydrogenase, proton pump	Absent	SO_0101 - SO_0113	Absent
<i>fdh-O</i> operons	Formate dehydrogenase-O, proton pump, encoded by two contiguous <i>fdoGHI</i> gene cassettes	Sputw3181_3870 - 3873	SO_4508 - SO_4511	Sputcn32_0338 - 0335
		Sputw3181_3874 - 3877	SO_4512 - SO_4515	Sputcn32_0334 - 0331
<i>fdh-H</i>	Hydrogenase-linked formate dehydrogenase	Sputw3181_0894	SO_0988	Sputcn32_3051
<i>hydAB</i> operon	Fe-only hydrogenase, hydrogen production	Absent	SO_3920 - SO_3926	Absent
<i>hyaAB</i> operon	Ni-Fe hydrogenase, hydrogen utilization	Separated into two gene clusters: Sputw3181_1919-1924; sputw3181_2173-2178	SO_2089 - SO_2099	Separated into two gene clusters: Sputcn32_2093 – 2088; Sputcn32_1836 - 1831
<i>ndh-I</i> operon	NADH dehydrogenase I (NuoA-NuoN), proton pump	Absent	SO_1009 - SO_1021	Absent
<i>ndh-II</i>	NADH dehydrogenase II	Sputw3181_3093	SO_3517	Sputcn32_1702
		Sputw3181_2863	Deleted in MR-1	Sputcn32_1241
<i>nqrABCDEF-1</i>	Sodium ion translocating NADH dehydrogenase I	Sputw3181_3324 - 3319	SO_0902 - SO_0907	Sputcn32_0849 - 0854
<i>nqrABCDEF-2</i>	Sodium ion translocating NADH dehydrogenase II	Sputw3181_3236 - 3230	SO_1103 - SO_1109	Sputcn32_0946 - 0940
<i>rnfABCDGE</i>	Electron transfer complex RnfABCDGE	Sputw3181_2159 - 2164	SO_2508 - SO_2513	Sputcn32_1850 - 1845
<i>otr</i> operon	Octaheme tetrathionate reductase Otr	Absent	SO_4142 - SO_4144	Absent
<i>ttr</i> operon	Tetrathionate reductase TtrACB	Sputw3181_3510 - 3512	Absent	Sputcn32_0664- 0662
<i>psrABC</i>	Polysulfide reductase PsrABC	Sputw3181_3557-3559	SO_4060 - SO_4062	Sputcn32_0613- 0611
<i>sorAB</i> operon	Sulfite hydrogenase SorAB and monoheme cytochromes, sulfite oxidation	Absent	SO_0714, 0715(<i>sorA</i>), 0716 (<i>sorB</i>), 0717	Absent
<i>nap-alpha</i>	Periplasmic nitrate reductase NapDABC	Sputw3181_2103-2107	Absent	Sputcn32_1901- 1905
<i>nap-beta</i>	Periplasmic nitrate reductase NapDAGHB	Sputw3181_0792-0796	SO_0845 - SO_0849	Sputcn32_3151- 3147
<i>norZ-norR</i>	Nitric oxide reductase NorZ and regulator NorR	Sputw3181_0872 (<i>norR</i>) -0873 (<i>norZ</i>)	Absent	Sputcn32_3073 - 3072
<i>cyoABCDE</i>	Cytochrome <i>bo</i> terminal oxidase, proton pump	Sputw3181_0091-0096	Absent	Sputcn32_3862 - 3857
<i>cioAB</i>	Cyanide insensitive cytochrome <i>bd</i> terminal oxidase	Sputw3181_3246-3247	Absent	Sputcn32_0930 - 0929
<i>torECADSTR</i>	Trimethylamine N-oxide reductase	Absent	SO_1228 - SO_1234	Absent

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<i>dms</i>	Dimethyl sulfoxide reductase I, Dms-1	Absent	SO_1427 - SO_1432	Absent
	Dimethyl sulfoxide reductase II, Dms-2	Absent	SO_4357 - SO_4362	
<i>mtrABC</i>	Dissimilatory metal reductase	Sputw3181_2623-2625	SO_1776 -SO_1778	Sputcn32_1478 - 1476
<i>mtrDEF</i>	Secondary metal reductase	Absent	SO_1780 - SO_1782	Absent

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10 **Figure S1**



20 **Supplemental Figure S1** Anaerobic growth (OD_{600}) of wild type strain of *S. putrefaciens*
21 W3-18-1, Δcrp mutant, and double mutant of Δcrp and $\Delta nap\text{-}\alpha$ on nitrate (2mM) in
22 modified M1 minimum media supplemented with 50 mM of sodium lactate as the
23 electron donor and carbon source.

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