

## **Supplementary information**

### **MtvR is a global regulatory sRNA in *Burkholderia cenocepacia***

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## Supplementary Methods

### Assessment of anti-MtvR transcripts

The time required by the anti-MtvR transcript to silence MtvR was assessed using *B. cenocepacia* J2315 transformed with plasmid pCGR16, grown for 24h (t = 0), time after which the expression of anti-MtvR was induced by the addition of 1% of L-arabinose (final concentration) to the culture. Samples were taken after 0, 5 and 10 min. RNA was extracted and processed as described in the main text.

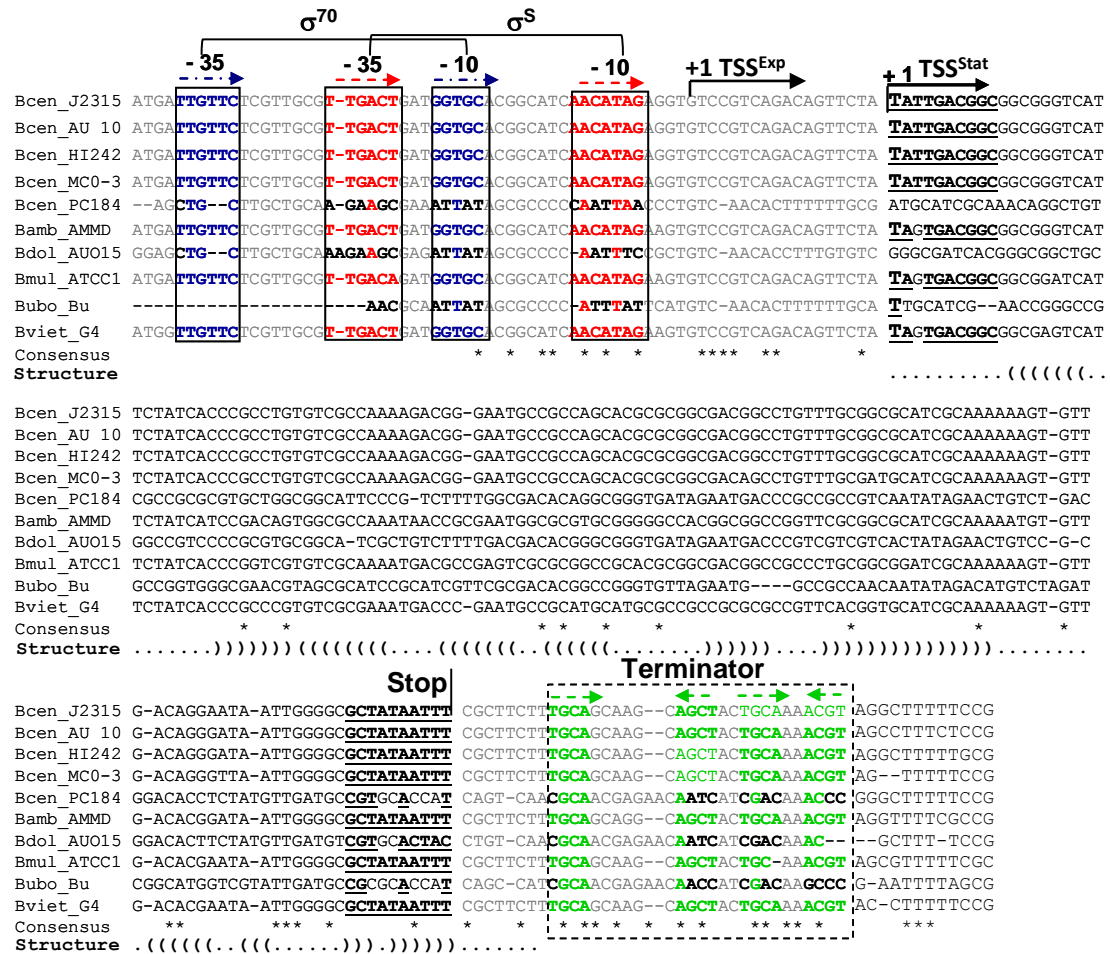
### Bioinformatic analysis

The MtvR sRNA sequence was aligned with the RNA sequence of the putative mRNA targets, predicted by the sRNATarget software (1), using the RNAHybrid web tool (2). In order to predict potential sigma-factor binding sites in putative MtvR mRNA targets, regulated by  $\sigma^{70}$  transcription factor, the BPROM software (Softberry, Inc., Mt. Kisco, NY) bioinformatics tool (minimum score of 7) and MEME suite webserver (3) were used. The alignment of the MtvR encoding sequences from Bcc strains was performed using Clustal W (4).

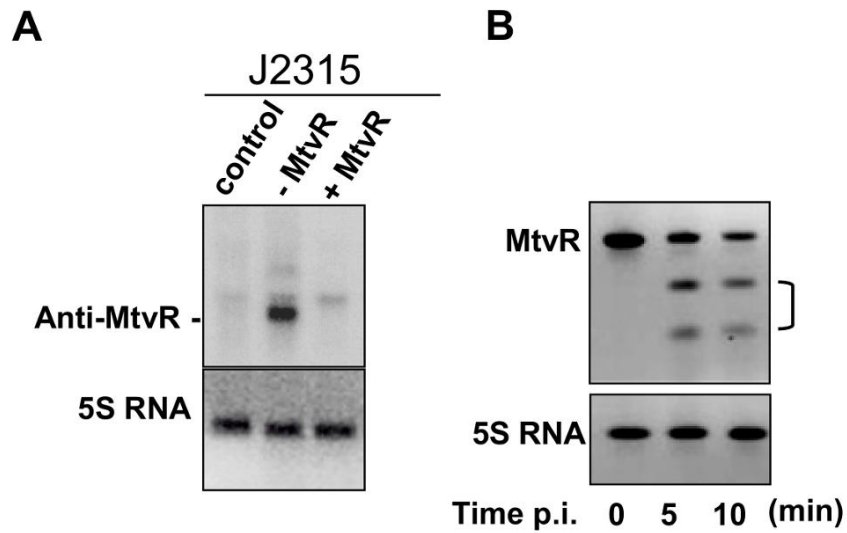
### EMSA experiments with RNA chaperones

The ability of 25 nM (final concentration) of MtvR to bind to Hfq (concentrations ranging from 0.1 to 200 nM of the hexameric form of Hfq (Hfq<sub>6</sub>)), or to Hfq2 (concentrations ranging from 0.1 to 200 nM of the trimeric form of Hfq2 (Hfq2<sub>3</sub>)), was also evaluated with EMSA assays. Non-labelled yeast tRNA (Ambion) was added in excess to each sample to minimize non-specific binding. Incubation, resolution of RNA-RNA and RNA-protein complexes and detection of band-shifts was performed as previously described (5).

## Supplementary Figures

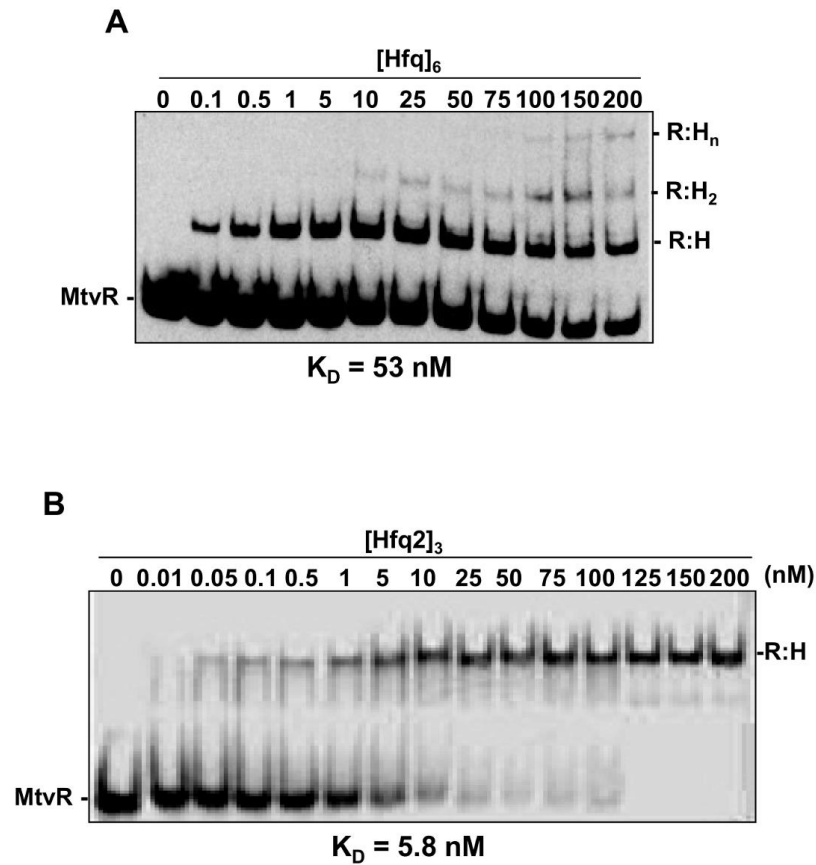


**FIG S1. MtvR is exclusive to Bcc strains.** Sequence alignment of the MtvR homologues of the indicated *Burkholderia* strains. Transcribed sequences are shown in black, non-transcribed elements are shown in grey. The putative  $\sigma^{70}$  (blue, p value =  $4.84e^{-4}$ ) and  $\sigma^S$  (red, p value =  $2.37e^{-3}$ ) regulatory sequences predicted using the MEME suite webserver are indicated. Non conserved nucleotides in regulatory sequences are shown in black. The MtvR Transcription Start Sites (TSS) are represented by capital casing with the conserved nucleotides shown in bold and underlined (exp, exponential; stat, stationary phase). The Transcription Stop Site is indicated by an underlined bold sequence, and the terminator sequences are represented in green, with the non-conserved nucleotides in black. Asterisks below the sequences indicate conserved nucleotides. Bcen, *B. cenocepacia*; Bamb, *B. ambifaria*; Bdol, *B. dolosa*; Bmul, *B. multivorans*; Bubo, *B. ubonensis*; Bviet, *B. vietnamiensis*.

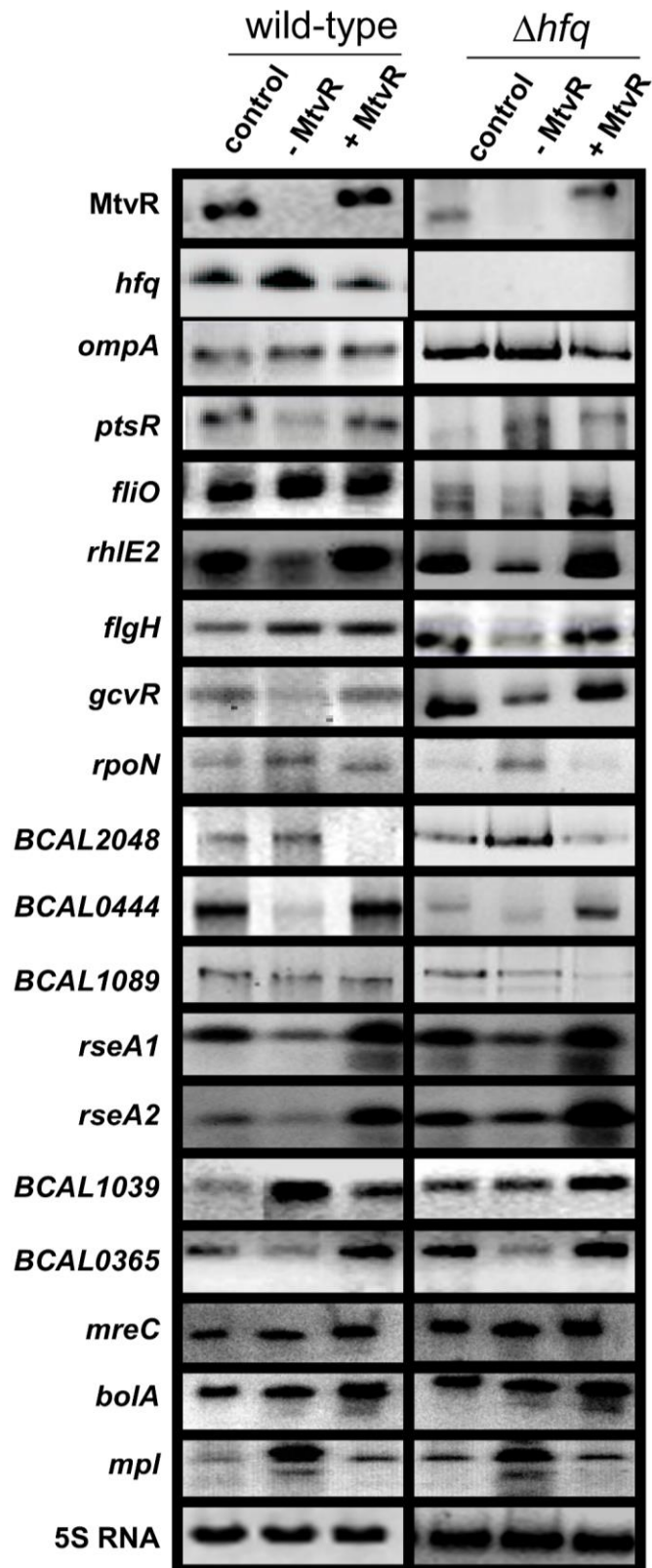


**FIG S2. *hfq* mRNA levels are affected by MtvR.** A) Northern blot analysis of the transcript designed to silence MtvR in wt, wt overexpressing the anti-MtvR RNA (-MtvR), and wt overexpressing MtvR (+ MtvR). B) Effects of expression of the anti-MtvR on the MtvR levels, after induction with 2% L-arabinose for the indicated time. Putative processing products are marked with bracket.





**FIG S4. MtvR binds to Hfq and Hfq2.** EMSA assays performed to assess the ability of Hfq (A), or Hfq2 (B) to bind to 25 nM of MtvR. The estimated  $K_D$  values are shown. Concentrations of the RNA chaperones were calculated based on their respective multimeric state, hexameric for Hfq and trimeric for Hfq2, respectively.



**FIG S5. MtvR affects multiple genes.** Northern blot analysis of the effect of MtvR on the mRNA levels of selected predicted targets. Total RNA from the wt (control) and derivative strains with the MtvR silenced (-MtvR), or overexpressing MtvR (+MtvR), and from the  $\Delta hfq$  mutant (CJ1), the CJ1 with MtvR silenced (-MtvR), and the CJ1 overexpressing MtvR (+MtvR) was probed.

## Supplementary Tables

**Table S1.** Bacterial strains and plasmids used in this work.

Strain or plasmid	Description	Reference or source
<i>B. cenocepacia</i> J2315	CF sputum isolate	(7)
<i>B. cenocepacia</i> CJ1	<i>B. cenocepacia</i> J2315 <i>hfq</i> mutant	(8)
<i>B. cenocepacia</i> CJ2	<i>B. cenocepacia</i> J2315 <i>hfq2</i> mutant	(8)
<i>B. cenocepacia</i> J2315+pCGR15	<i>B. cenocepacia</i> J2315 overexpressing MtvR	This study
<i>B. cenocepacia</i> CJ1+pCGR15	<i>B. cenocepacia</i> J2315 <i>hfq</i> mutant overexpressing MtvR	This study
<i>B. cenocepacia</i> J2315+pCGR16	<i>B. cenocepacia</i> J2315 with the sRNA MtvR silenced	This study
<i>B. cenocepacia</i> CJ1+pCGR16	<i>B. cenocepacia</i> J2315 <i>hfq</i> mutant with the sRNA MtvR silenced	This study



Plasmids		
pMLBAD	Tmp <sup>R</sup> ; used for inducible gene expression	(9)
pBBR1MCS	Cm <sup>R</sup> ; used for constitutive gene expression	(10)
pCR 2.1	Amp <sup>R</sup> ; Km <sup>R</sup> ; used <i>in vitro</i> transcription	Invitrogen
pCGR4	pET23a+ with the <i>hfq</i> encoding sequence clone	(5)
pCGR14	pCR 2.1 with the 140 bp cDNA fragment corresponding to the <i>mtvR</i> sRNA cloned in the XbaI/HindIII sites, and a Cm <sup>R</sup> cassette (obtained by PCR from plasmid 0pKD3) cloned in the HincII site	This study
pCGR14.1	pCGR14 digested and self-ligated in the HaeIII sites	This study
pCGR15	pMLBAD with the 140 bp cDNA fragment corresponding to the <i>MtvR</i> sRNA cloned in the EcoRI/XbaI sites (pBAD promoter control)	This study
pCGR16	pMLBAD with the 140 bp cDNA fragment corresponding to the <i>MtvR</i> sRNA cloned in the XbaI/HindIII sites (pBAB promoter control)	This study
pCGR20	pCR 2.1 with the 140 bp cDNA fragment corresponding to the <i>mtvR</i> sRNA cloned in the XbaI/HindIII sites (T7 promoter control)	This study
pCGR24	pCR 2.1 with the 384 bp cDNA fragment corresponding to the <i>hfq</i> 5'-UTR cloned in the XbaI/HindIII sites (T7 promoter control)	This study

pCGR25	pCR 2.1 with the 609 bp cDNA fragment corresponding to the <i>hfq</i> full mRNA (5'-UTR and CDS) with 6 histidine at the C-terminus, cloned in the XbaI/HindIII sites (T7 promoter control)	This study
pCGR33	pMLBAD with the 5'-UTR- <i>hfq</i> -LacZ DNA fragment (pBAB promoter disrupted, only replicative)	This study

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**Table S2.** Oligonucleotides and primers used in this work.

Name	Purpose	Sequence 5' –3'	Source
UF	Cloning mtvR	TTTCTAGATATTGACGGCGGCGGGU	This study
LF	Cloning mtvR	TTAAGCTTAAATTATAGCGCCCAATTA	This study
NP	Northern analysis of mtvR	CTATCACCCGCCTGTGTCGCCA	This study
OMPA	Northern blot probe for <i>ompA</i>	AAAAGCTTGTGGCCGGAAC	This study
PTSR	Northern blot probe for <i>ptsR</i>	CGCCTCCAGGTATTTCGTACA	This study
FLIO	Northern blot probe for <i>fliO</i>	ATCTCGACGATCGTCGCGCTTTCCTTC	This study
RHLE2	Northern blot probe for <i>rhLE2</i>	TACGGCCGATCCGGTGACGTAG	This study
FLGH	Northern blot probe for <i>flgH</i>	CATCGGCATCGGCGGCTGCGCCGTCATC	This study
GCVR	Northern blot probe for <i>gcvR</i>	CGAGGCGCGGGACGAGCCACGCGGAT	This study
RECQ	Northern blot probe for <i>recQ</i>	GCGACCTGGTCCTGCATCAGCGCGAT	This study
RPON	Northern blot probe for <i>rpoN</i>	GATCAGTGCGTGCGCGTCGCGCA	This study
RPSL	Northern blot probe for <i>rpsL</i>	GCCGCGACGCTGGGGGCAGTCCTGCAGGG	This study
2048	Northern blot probe for <i>BCAL2048</i>	TTCGGCTTCGACACGAACGTGCCCTT	This study
0444	Northern blot probe for <i>BCAL0444</i>	CGCCTCCAGGTATTTCGTACA	This study
1089	Northern blot probe for <i>BCAL1089</i>	CTCGGTGTGCTGGTTCGTGCGGATC	This study
HFQ	Northern blot probe for <i>hfq</i>	AAAGGGCAATTGTTACAAG	(8)
CGRO105	Northern blot probe for <i>rseA2</i>	TGCGGTAAACGCGACGCTCA	This study
CGRO106	Northern blot probe for <i>rseA1</i>	GGTGGTAATGGGCCACGCA	This study

CGRO107	Northern blot probe for <i>BCAL1039</i>	CGGCGGATGTAGCGGAACAGG	This study
CGRO108	Northern blot probe for <i>BCAL0365</i>	GCAGTGACACGAATCGGCGGG	This study
CGRO109	Northern blot probe for <i>BCAL0640</i>	GATCCCGCGCACAGCAGCAT	This study
CGRO110	Northern blot probe for <i>mpl</i>	GTGGTCGTCTTGCCGTGCGT	This study
CGRO111	Northern blot probe for <i>mreC</i>	TGGTTGGCCTGCGTGGACAG	This study
CGRO112	Northern blot probe for <i>bolA</i>	GCACGGTGAGCGACAGCGG	This study
CGRO113	MtvR mutagenesis	TTTCTAGATATTGACTTGCTGCTTTATCACCC	This study
CGRO114	MtvR mutagenesis	TTAAGCTTAAATTATAGCCGAGGAATTA	This study
CGRO115	<i>hfq</i> mutagenesis	TTTCTAGAGCACCGAGGGCAAGGGCTAG	This study
CGRO116	<i>hfq</i> mutagenesis	TTAAGCTTTTGGCTGGCTTAAAA	This study
CGRO121	Fwd primer <i>hfq</i> 5'-UTR	TTGGATCCATTGGACGAGGCTTCCGC	This study
CGRO122	Rev primer <i>hfq</i> 5'-UTR	TTGGATCCCATGGCGTACTCCATCTTTT	This study
CGRO123	Fwd primer <i>hfq</i> his-tag	TTTCTAGAGCACGTCCCGCAAGGGCTAG	This study
CGRO124	Rev primer <i>hfq</i> his-tag	TTGGATCCATTGTGGTGGTGGTGGTGGGACGAGGCTTCCGC	This study
M13FWD	LacZ amplification	GTAAAACGACGGCCAGT	Invitrogen
M13REV	LacZ amplification	AGCGGATAACAATTCACACAGGA	Invitrogen
5S	Northern blot probe for 5S rRNA	TTCGGGATGGGAAGGGGTGGGA	(8)
5R-MT	5' RACE experiments	ATACGGGGCTGCGAGAGTCGT	This study

**Table S4.** Potential MtvR targets in *B. cenocepacia* J2315 genome

<b>Gene</b>	<b>Function</b>	<b>Locus Tag</b>
<i>hfq</i>	RNA chaperone and Pleiotropic regulator	BCAL1879
<i>recQ</i> *	ATP-dependent DNA helicase RecQ	BCAL0228
<i>rpoN</i>	Nitrogen limitation response transcription regulator	BCAL0813
<i>rpsL</i> *	Ribosomal subunit involved in translational accuracy. Important for ribosome structure	BCAL0229
<i>flgH</i>	The flagellar L-ring protein precursor protects the motor/basal body from shearing forces during rotation	BCAL0570
<i>rhlE2</i>	ATP-dependent RNA helicase	BCAL2412
<i>fliO</i>	Essential component of the flagellum-specific protein export apparatus	BCAL3504
<i>gcvR</i>	<i>gcvA</i> LysR family transcriptional regulator	BCAL0382
BCAL2048	GntR regulator of a transport cluster (Fe-S), MFS type	BCAL2048
BCAL0444	GntR regulator of a transport cluster, ABC type	BCAL0444
<i>ptsR</i>	PTS regulator	BCAL0777
BCAL1089	AnsC regulator of a ABC transport cluster	BCAL1089
<i>rseA1</i>	$\sigma^E$ negative regulatory protein 1	BCAL2871
<i>rseA2</i>	$\sigma^E$ negative regulatory protein 2	BCAL0999
BCAL1039	ATP-binding cassette transporter protein	BCAL1039
BCAL0365	Major facilitator Superfamily transporter protein	BCAL0365
<i>mpl</i>	UDP-N-acetylmuramate:L-alanyl-gamma-D-glutamyl-meso-diaminopimelate ligase	BCAL3416
<i>mreC</i>	Rod-shape determining protein	BCAL0481
<i>bolA</i>	Cell shape	BCAL1984

\* mRNA predicted MtvR targets which mRNA levels were not affected by MtvR.



TARGET : gcvR  
mfe: -39.8 kcal/mol

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target 5'          C A G AAC U A CUA A A UC GAAUA U 3'
                  GGC G CGCG GGU CAUUC GUUUUU GCGA AGA GAC UCGUU
                  CCG C GCGC CCG GUAAG CAGAAA CGCU UCU CUG GGCAG
sRNA 3' UUUAAUAUCGCGGGGUAAUAAGGACAGUUGUGAAAAACGCUACGCGGCGUUUGU GCAG G GCACGA CC GG AC GUGUCCGCCACUA UA GGCGGC UUAU 5'

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TARGET : BCAL2048  
mfe: -58.2 kcal/mol

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target 5'          AU UA CUCG A GUCAUA A CC CCCC AA G 3'
                  CGCC CGC GCUGGC CA UCCC UCUU ACA GU UGA GAUG ACCC CGUCGU
                  GCGG GCG CGACCG GU AGGG AGAA UGU CG ACU UUAC UGGG GCGGCA
sRNA 3' UUUAAUAUCGCGGGGUAAUAAGGACAGUUGUGAAAAACGCUACGCGGCGUUUGUCCGGCA C CA CC A C AACCGCUG C CCC AUC CG GUUAU 5'

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TARGET : BCAL0444  
mfe: -64.4 kcal/mol

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target 5' A C A C U ACCA A U AAC U C C 3'
          UAUGGCGCCCU GUU UUC UC AUA GC CCGC GCC UGCC CG UG C GC CCCG GCG C CAG GC
          AUAUCGCGGGG UAA AAG AG UGU CG GGCG CGG GCGG GC AC G CG GGGC CGC G GUC CG
sRNA 3' UUUAA U U GAC U GAAAAACGCUA C UUUGUC CA C GCACG C C UAA AGAAAAAC U U CCCACUAUCUUACUGGGCGGCGCAGUUUAU 5'

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TARGET : ptsR  
mfe: -72.3 kcal/mol

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target 5' U A U G U A A C GA A GGAAA G G CGA U 3'
          GCGUUC CU UCGACG GUCG AGA UCGUCG CCG C CG G GGC CAU CCG CU GGC AC GGC GCCCG
          CGCGGG GA AGUUGU CGGC UUU GGCAGC GGC G GC C CCG GUA GGC GA CCG UG CCG UGGGC
sRNA 3' UUUAAUAU GUUAAUAAG C GAAAAACGCUACG G GUCC C A GA CC AG A AAA CUG U CCCACUAUCUUAC GCGGCGAGUUUAU 5'

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TARGET : BCAL1089  
mfe: -35.5 kcal/mol

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target 5' A AA A A G A GA AC A A A 3'
          AUAG UAA AUUUCU UCGACG G GA UG G AA AUGG GCC UCG UCGC
          UAUC GUU UAAGGA AGUUGU C CU GC C UUUGUC CGG AGC GGCG
sRNA 3' UUUAA GCGGG AA C GAAAAA G AC GG G C CGCAGACC CGGUAAGGGCAGAAAAACCGCUGUGUCCGCCACUAUCUUACUGGGCGGCGCAGUUUAU 5'

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Alignment of the MtvR sRNA sequence with the mRNA targets tested. mfe: minimum folding energy; the putative start codon in highlighted in red lettering.

**Table S6:** Relative fold-change of MtvR putative mRNA targets

	Wild-type		$\Delta hfq$		Wild-type vs $\Delta hfq$		
	- MtvR	+ MtvR	- MtvR	+ MtvR	MtvR	- MtvR	+ MtvR
<i>MtvR</i>	ND	+ 1.3	- 6.3	+ 1.4	+ 2.6	- 20.4	+ 1.5
<i>hfq</i>	+ 1.5	- 6.9	ND	ND	ND	ND	ND
<i>ompA</i>	NC	NC	NC	NC	- 2.0	- 1.9	- 1.3
<i>ptsR</i>	- 2.0	NC	- 3.0	- 2.6	+ 2.9	- 1.7	NC
<i>fliO</i>	NC	NC	- 1.5	+ 2.4	+ 2.7	+ 3.9	+ 1.3
<i>rhlE2</i>	- 1.5	+ 1.4	- 1.3	NC	NC	NC	NC
<i>flgH</i>	- 1.3	+ 1.3	- 1.9	NC	- 1.4	NC	NC
<i>gcvR</i>	- 1.6	NC	- 2.7	NC	- 1.7	NC	- 1.5
<i>rpoN</i>	+ 1.5	NC	+ 3.7	- 1.2	+ 2.2	NC	+ 2.4
<i>BCAL2048</i>	+ 1.3	- 7.1	+ 1.3	- 1.5	- 1.3	NC	- 4.2
<i>BCAL0444</i>	- 3.8	NC	- 2.0	+ 2.4	+ 6.2	+ 3.0	+ 2.0
<i>BCAL1089</i>	- 1.2	NC	- 1.5	- 3.7	- 1.6	NC	+ 1.7
<i>rseA1</i>	- 1.8	+ 1.2	- 2.0	NC	+ 1.2	+ 1.2	NC
<i>rseA2</i>	- 1.9	+ 3.7	- 1.2	+ 1.7	- 2.3	- 2.8	NC
<i>BCAL1039</i>	+ 3.6	NC	NC	+ 1.2	- 1.4	NC	- 1.5
<i>BCAL0365</i>	- 1.6	+ 1.9	- 2.6	+ 3.5	- 1.7	NC	NC
<i>mreC</i>	+ 1.6	+ 1.6	- 1.2	NC	- 1.9	NC	NC
<i>bolA</i>	NC	+ 1.2	NC	NC	NC	NC	NC
<i>mpl</i>	+ 4.8	+ 1.5	+ 3.1	NC	NC	NC	NC

ND, not determined; NC, no fold-change (NC:  $-1.2 < FC < 1.2$ ;  $p \leq 0.05$ ),  $SD < 0.01$  FC.



## Supplementary References

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