

**Residue substitutions near the redox center of *Bacillus subtilis* Spx affect RNA polymerase interaction, redox control and Spx-DNA contact at a conserved cis-acting element.**

Ann A. Lin, Don Walthers, and Peter Zuber

**SUPPLEMENTAL MATERIAL**

**Figure S1. Effect of R92 substitutions in Spx on the transcription of *trxB-lacZ*.** Spx R92 residue was substituted with alanine(R92A), glutamine(R92Q), or lysine(R92K), and the resulting mutant alleles were introduced in the *amyE* locus and under IPTG control in *spx* null mutant strain bearing *trxB-lacZ*. Strains were grown in DS medium until the OD<sub>600</sub> reached 0.4. Then, each culture was divided into two flasks, and 1 mM IPTG was added to one flask to induce SpxDD expression. Samples were taken at time intervals and  $\beta$ -galactosidase activities were measured. Symbols: circles, SpxDD; diamonds, Spx(R92A)DD; squares, Spx(R92Q)DD; triangles, Spx(R92K)DD. Open symbols with broken lines represent cells cultured without IPTG, and closed symbols with solid lines represent cell culture with IPTG.

**Figure S2. Effect of Spx(G52R, R91A) mutant on *trxB-lacZ* transcription and RNAP binding.** The IPTG-inducible allele encoding Spx(G52R, R91A)DD was introduced into the *amyE* locus of the *Spx* null mutant

strain bearing *trxB-lacZ*. (A) The  $\beta$ -galactosidase assay was performed as described in Fig. 3 and the highest activities of Spx mutants were selected. The ratio of mutant activity to that of the control SpxDD was calculated. (B) The production of Spx protein in the strains was evaluated by Western blot analysis using anti-Spx antibody. (C, D) Purified Spx(G52R, R91A) $\Delta$ CHA protein was used for in vitro RNAP interaction assay. The SAd-RNAP or Holo-RNAP was incubated with Spx $\Delta$ CHA or Spx(G52R, R91A) $\Delta$ CHA and followed by the anti-HA affinity chromatography. The interaction between Spx proteins and SAd-RNAP (C)/Holo-RNAP (D) was analyzed by SDS-PAG gel. Abbreviation: I, input and E, eluate.

**Figure S3. Spx/RNAP/*trxB* crosslinking result at position -11, -21, -46, -49, -52 on *trxB* promoter.** Crosslinking reactions were performed as described in Experimental procedures. The nucleotide positions indicated here are crosslinking sites on the *trxB* promoter. Bands of crosslinked products on the gel corresponding to  $\beta\beta'$ ,  $\sigma^A$ ,  $\alpha$  and Spx are indicated. The band representing presumed uncleaved radiolabeled DNA substrate is also shown.

Table S1. *B. subtilis* strains and plasmids

Strain or plasmid	Relevant genotype or characteristics	Source or reference
<b><i>B. subtilis</i> strains</b>		
<b>JH642</b>	<i>trpC2 pheA1</i> (parental strain)	James Hoch
<b>LAB545</b>	<i>trpC2 pheA1</i> SP $\beta$ <i>c2del2::Tn917::pMMN92(srfA-lacZ)</i>	(43)
<b>ORB5853</b>	<i>trpC2 pheA1 sigA</i> (L366A) <i>neo rpoC-His<sub>10</sub> cat</i>	(42)
<b>ORB6129</b>	<i>trpC2 pheA1</i> SP $\beta$ <i>c2del2::Tn917::pMMN92(srfA-lacZ)</i> <i>amyE::pSN56(Pspankhy-spx<sup>DD</sup>)</i>	(32)
<b>ORB7262</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i>	(32)
<b>ORB7276</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pSN56(Pspankhy-spx<sup>DD</sup>)</i>	(32)
<b>ORB7282</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pMMN683(Pspankhy-spxR60E<sup>DD</sup>)</i>	(32)
<b>ORB7316</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pYZ14 (Pspankhy-spxC10A<sup>DD</sup>)</i>	(32)
<b>ORB7821</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL37(Pspankhy-spxR92A<sup>DD</sup>)</i>	This study
<b>ORB7822</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL38(Pspankhy-spxR92Q<sup>DD</sup>)</i>	This study
<b>ORB7904</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL56(Pspankhy-spxC10A,R92A<sup>DD</sup>)</i>	This study
<b>ORB7906</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL58(Pspankhy-spxR60E,R92A<sup>DD</sup>)</i>	This study
<b>ORB7945</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL64(Pspankhy-spxK62E,R92A<sup>DD</sup>)</i>	This study
<b>ORB7946</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pMMN684(Pspankhy-spxK62E<sup>DD</sup>)</i>	This study
<b>ORB7979</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL65(Pspankhy-spxC10A,R60E<sup>DD</sup>)</i>	This study
<b>ORB7981</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL67(Pspankhy-spxC10A,K62E<sup>DD</sup>)</i>	This study
<b>ORB8268</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL88(Pspankhy-spxR91A<sup>DD</sup>)</i>	This study
<b>ORB8269</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL93(Pspankhy-spxF64A<sup>DD</sup>)</i>	This study
<b>ORB8270</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i> <i>amyE::pAL94(Pspankhy-spxF64Y<sup>DD</sup>)</i>	This study
<b>ORB8271</b>	<i>trpC2 pheA1 Δspx::neo thrC::pDYR9(trxB-lacZ)</i>	This study

<b>ORB8272</b>	<i>amyE</i> ::pAL95(Pspankhy-spxQ65A <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8273</b>	<i>amyE</i> ::pAL96(Pspankhy-spxN68A <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8274</b>	<i>amyE</i> ::pAL97(Pspankhy-spxN70A <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8275</b>	<i>amyE</i> ::pAL98 (Pspankhy-spxY5A <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8309</b>	<i>amyE</i> ::pAL99 (Pspankhy-spxY5F <sup>DD</sup> ) <i>trpC2 pheA1 SPβc2del2::Tn917:: pMMN92(srfA-lacZ)</i>	This study
<b>ORB8310</b>	<i>amyE</i> ::pAL88(Pspankhy-spxR91A <sup>DD</sup> ) <i>trpC2 pheA1 SPβc2del2::Tn917:: pMMN92(srfA-lacZ)</i>	This study
<b>ORB8323</b>	<i>amyE</i> ::pAL37(Pspankhy-spxR92A <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8565</b>	<i>amyE</i> ::pAL106(Pspankhy-spxR91A,R92A <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8566</b>	<i>amyE</i> ::pAL127(Pspankhy-spxR92K <sup>DD</sup> ) <i>trpC2 pheA1 Δspx::neo thrC</i> ::pDYR9( <i>trxB-lacZ</i> )	This study
<b>ORB8566</b>	<i>amyE</i> ::pAL128(Pspankhy-spxG52R,R91A <sup>DD</sup> )	This study
<b>Plasmids</b>		
<b>pAL37</b>	pDR111 with <i>spxR92A<sup>DD</sup></i>	This study
<b>pAL38</b>	pDR111 with <i>spxR92Q<sup>DD</sup></i>	This study
<b>pAL45</b>	pDR111 with <i>spxΔCHA</i>	(19)
<b>pAL46</b>	pTYB4 with <i>spxΔCHA</i>	(19)
<b>pAL54</b>	pTYB4 with <i>spx<sup>R92A</sup></i>	This study
<b>pAL56</b>	pDR111 with <i>spxC10A,R92A<sup>DD</sup></i>	This study
<b>pAL58</b>	pDR111 with <i>spxR60E,R92A<sup>DD</sup></i>	This study
<b>pAL64</b>	pDR111 with <i>spxK62E,R92A<sup>DD</sup></i>	This study
<b>pAL65</b>	pDR111 with <i>spxC10A,R60E<sup>DD</sup></i>	This study
<b>pAL67</b>	pDR111 with <i>spxC10A,K62E<sup>DD</sup></i>	This study
<b>pAL73</b>	pTYB4 with <i>spx<sup>C10A</sup>ΔCHA</i>	(19)
<b>pAL74</b>	pTYB4 with <i>spx<sup>G52R</sup>ΔCHA</i>	(19)
<b>pAL75</b>	pTYB4 with <i>spx<sup>R60E</sup>ΔCHA</i>	(19)
<b>pAL82</b>	pTYB4 with <i>spx<sup>R92A</sup>ΔCHA</i>	This study
<b>pAL88</b>	pDR111 with <i>spxR91A<sup>DD</sup></i>	This study
<b>pAL93</b>	pDR111 with <i>spxF64A<sup>DD</sup></i>	This study
<b>pAL94</b>	pDR111 with <i>spxF64Y<sup>DD</sup></i>	This study
<b>pAL95</b>	pDR111 with <i>spxQ65A<sup>DD</sup></i>	This study
<b>pAL96</b>	pDR111 with <i>spxN68A<sup>DD</sup></i>	This study
<b>pAL97</b>	pDR111 with <i>spxN70A<sup>DD</sup></i>	This study
<b>pAL98</b>	pDR111 with <i>spxY5A<sup>DD</sup></i>	This study
<b>pAL99</b>	pDR111 with <i>spxY5F<sup>DD</sup></i>	This study
<b>pAL106</b>	pDR111 with <i>spxR91A,R92A<sup>DD</sup></i>	This study
<b>pAL121</b>	pTYB4 with <i>spx<sup>R91A</sup></i>	This study
<b>pAL122</b>	pTYB4 with <i>spx<sup>R91A</sup>ΔCHA</i>	This study
<b>pAL127</b>	pDR111 with <i>spxR92K<sup>DD</sup></i>	This study
<b>pAL128</b>	pDR111 with <i>spxG52R,R91A<sup>DD</sup></i>	This study
<b>pAL129</b>	pTYB4 with <i>spx<sup>G52R,R91A</sup>ΔCHA</i>	This study
<b>pDG793</b>	<i>thrC</i> integration vector with promoter-less <i>lacZ</i> reporter	(49)
<b>pDR111</b>	<i>amyE</i> integration vector with Pspankhy promoter	(38)
<b>pDW4</b>	TOPO vector with <i>trxB</i> promoter	This study
<b>pDW10</b>	pRLG770 with <i>trxB</i> promoter	This study
<b>pDYR9</b>	pDG793 with <i>trxB(-115~+47)-lacZ</i>	(22)
<b>pMMN470</b>	pTYB4 with <i>spx</i>	(37)
<b>pMMN683</b>	pDR111 with <i>spxR60E<sup>DD</sup></i>	(32)
<b>pMMN684</b>	pDR111 with <i>spxK62E<sup>DD</sup></i>	(32)
<b>pRLG770</b>	transcription vector	(44)
<b>pSN56</b>	pDR111 with <i>spx<sup>DD</sup></i>	(13)

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<b>pSN64</b>	pTYB4 with <i>sigA</i>	(21)
<b>pTYB4</b>	<i>E. coli</i> expression vector for IMPACT™ system	New England BioRads
<b>pUC18</b>	cloning vector	
<b>pUC19</b>	cloning vector	
<b>pYZ14</b>	pDR111 with <i>spxC10A<sup>DD</sup></i>	(50)

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Table S2. oligonucleotides used in this study

<b>oligonucleotide</b>	<b>sequence</b>	<b>purpose</b>
oAL25	GATGATCGGAGCGCGAAGC	spx R92A reverse
oAL24	GCTTCGCGCTCCGATCATC	spx R92A forward
oAL27	GATGATCGGTTGGCGAAGC	spx R92Q reverse
oAL26	GCTTCGCCAACCGATCATC	spx R92Q forward
oAL62	ATCGGACGGGCAAGCAAACC	spx R91A reverse
oAL61	GGTTTGCTTGCCCGTCCGAT	spx R91A forward
oAL65	GTTCAAAAGTAGCTCAAAAAGTGA	spx F64A forward
oAL66	TCAGTTTTTGAGCTACTTTTGAAC	spx F64A reverse
oAL68	TCAGTTTTTGATATACTTTTGAAC	spx F64Y reverse
oAL67	GTTCAAAAGTATATCAAAAAGTGA	spx F64Y forward
oAL70	ACATTCAAGTTTAGCGAATACTTTTG	spx Q65A reverse
oAL69	CAAAAGTATTTCGCTAAACTGAATGT	spx Q65A forward
oAL72	ACGTTACAGCCAGTTTTTGT	spx N68A reverse
oAL71	CAAAAAGTGGCTGTGAACGT	spx N68A forward
oAL74	GATTCAACAGCCACATTCAG	spx N70A reverse
oAL73	CTGAATGTGGCTGTTGAATC	spx N70A forward
oAL76	GCTTGGTGATGTTGCTAGTGTA	spx Y5A reverse
oAL75	TACACTAGCAACATCACCAAGC	spx Y5A forward
oAL78	GCTTGGTGATGTAATAAGTGTA	spx Y5F reverse
oAL77	TACACTATTTACATCACCAAGC	spx Y5F forward
oAL88	GATCGGAGCGGCAAGCAAACC	spx R91A,R92A reverse
oAL87	GGTTTGCTTGCCGCTCCGATC	spx R91A,R92A forward
oLA106	ATGATGATCGGTTTGCAGCAAGCA	spx R92K reverse
oAL105	TGCTTCGCAAACCGATCATCAT	spx R92K forward
oDW7	GGAATTCATCTGTGACTCTATGAGCAATC	<i>trxB</i> (-200~+20) forward
oDW8	CAAGCTTCCCATCAAACGTATTCCTTAC	<i>trxB</i> (-200~+20) reverse
oAL35	TCCCCGGGAGCATAATCAGGAACATC	<i>HA</i> reverse (pTYB4)
oAL38	CTGAGGACCGAACAGATG	spx G52R forward
oAL39	CATCTGTTCCGTCCTCAG	spx G52R reverse
oDYR06-032	TAAACCATCTTTAAGCTTTTGGGCTCTTTC	<i>trxB</i> (-115~+47) reverse
oDYR07-052	GCGAATTCGGCCTTCTATAAACAGAAGGC	<i>trxB</i> (-115~+47) forward
oMMN01-135	AGAGGAGTGAAGATCCATGGTTACTACTATAC	spx forward (pTYB4)
oMMN01-137	TAACTCCCGGGGTTTGCCAAACGCTGTGCTT	spx reverse (pTYB4)
oMMN01-173	CGAGGAAGCTTAGATGTTTCATCCTACTA	spx forward (pDR111)
oMMN01-174	TACCAGCAGGTCGACAAATAAAAGAAGG	spx reverse (pDR111)
oSN03-66	CACATACCAAGCGCGACTTCATGCAGAA	spx C10A forward
oSN03-67	TTCTGCATGAAGTCGCGCTTGGTGATGTG	spx C10A reverse
oDYR06-02	[Biotin]-CTGAACTAGAGGCCAAGGCTCTTTCTTTATTAACATATAC	<i>trxB</i> template (forward) for crosslinking
oDYR06-03	ATCCCCATCAAACGTATTCCCTTAC	<i>trxB</i> template (reverse) for crosslinking
oDYR06-05	ATCCCCATCAAACGTATTCCCTTACCCATATCC*T	<i>trxB</i> -11 crosslinking probe
oDYR07-35	ATCCCCATCAAACGTATTCCCTTACCCATATCCTATAAAAATCT*C	<i>trxB</i> -21 crosslinking probe
oAL47	ATCCCCTCAAACGTATTCCCTTACCCATATCCTATAAAAATCTCATGGTGA TACGCTA*T	<i>trxB</i> -36 crosslinking probe
oAL49	ATCCCCTCAAACGTATTCCCTTACCCATATCCTATAAAAATCTCATGGTGA TACGCTATTTT*T	<i>trxB</i> -40 crosslinking probe
oAL51	ATCCCCTCAAACGTATTCCCTTACCCATATCCTATAAAAATCTCATGGTGA TACGCTATTTTTTTG*C	<i>trxB</i> -44 crosslinking probe
oAL52	ATCCCCTCAAACGTATTCCCTTACCCATATCCTATAAAAATCTCATGGTGA TACGCTATTTTTTTGCTCAA*C	<i>trxB</i> -49 crosslinking probe
oDYR06-08	ATCCCCTCAAACGTATTCCCTTACCCATATCCTATAAAAATCTCATGGTGA TACGCTATTTTTTTGCT*C	<i>trxB</i> -46 crosslinking probe
oDYR06-09	ATCCCCTCAAACGTATTCCCTTACCCATATCCTATAAAAATCTCATGGTGA TACGCTATTTTTTTGCTCAACAC*G	<i>trxB</i> -52 crosslinking probe