

**Table S2. Granule Phenotypes of Screen Hits, Related to Figure 1**

Phenotypes	Strong	Moderate	Weak
<b>Stress granules: P-body co-localized</b>	<i>ATG11, CHO2, ERG2, INO2, ISW1, KCS1, PGD1, RLF2, RTC2, SHE10, SSH1, TGS1, VMA11, VMA16, VMA2, VMA3, VMA4, XRN1, YBR225W</i>	<i>ADO1, ASH1, ATG8*, CDC73, CIN8, GCN5, KTR1, MDS3, MRPL22, MTM1, NSG2, PFK2, RIM15, RPL42A, SWD3, TAT2, VHS2, VMA21, YIP3, YMD8, YMR262W, YNL200C</i>	<i>AAT1, ATC1, ATG18, BCK1, CDH1, CMK1, ERP5, SIR2, GSH2, HST1, ICE2, IKI3, IRC14, MON1, NFT1, NRM1, PCL1, PGC1, PHO86, PRE9, PSD1, RIM21, RNH70, RPL39, SET2, SDL1, SQS1, STO1, STE3, SWC5, SWH1, TRM12, UBX2, WSS1, YIL024C, YJR115W, YLR050C, YMR259C, YPL205C, ZWF1</i>
<b>Stress granules: P-body distinct</b>	<i>HPR1*, MFT1*, THO2*, THP1*, THP2*</i>	<i>EOS1, EST1, HEL2, SAC3*, YLR169W</i>	<i>GGA1, LAT1, LSM6, MEH1, PAP2, POC4, SAM37, SWR1, YDL124W, YLR236C</i>
<b>P-bodies increased</b>	<i>KCS1, PGD1, RPL42A, SWD3, SWH1, TAT2, XRN1</i>	<i>ADO1, ASH1, CMK1, ERG2, HEL2, HST1, KTR1, NSG2, PCL9, PHO86, RLF2, RPL35B, SDL1, VHS2, YBR225W, YMD8</i>	<i>ATC1, ERP5, MDS3, MFT1*, PFK2, RPL39, SHE10, STO1, YIL001W, YLR050C, YLR169W, YOR338W, YNL200C, ZWF1</i>
<b>P-bodies decreased</b>	<i>BIO4, MLP1, PFD1, RPS28B, SAM37, TCM62, TIF3, VMS1, YMR086W, YSW1</i>	<i>EDC2, GGA1, GIM4, PAP2, YSW1</i>	<i>EGD1, GSP2, GIM5, HNT3, MSN5, SRO9, STE5, YIL014C-A</i>
<b>Intra-vacuolar accumulation</b>	<i>ATG15</i>		

Several gene deletions were quantified as hits in more than one category, hence duplicate entries above. In addition, \*-marked gene deletions exhibited protein mis-localization phenotypes as follows: Nuclear/nuclear peripheral accumulation of Pab1-GFP: *HPR1, MFT1, THO2, THP1, THP2, SAC3*.

**Table S4. Experimental Quantitation, Related to Figures 2, 3, and 4**

<b>Midlog Cdc48/Ufd1/Npl4 ts alleles (Fig 2B)</b>		
<b>Strain/conditions</b>	<b>Stress granule size (<math>\mu\text{m}_2</math> +/- st dev.)</b>	<b>Stress granule foci/cell (+/- st. dev.)</b>
WT (W303)	0.11 +/- 0.07	0.02 +/- 0.03
WT - 39°C 1hr	0.20 +/- 0.03	0.46 +/- 0.16
Cdc48-3	0.14 +/- 0.07	0.05 +/- 0.02
Cdc48-3 - 39°C 1hr	0.35 +/- 0.09	2.03 +/- 1.13
Npl4-1	0.18 +/- 0.13	0.06 +/- 0.05
Npl4-1 - 39°C 1hr	0.31 +/- 0.04	0.91 +/- 0.29
Ufd1-2	0.07 +/- 0.03	0.02 +/- 0.02
Ufd1-2 - 39°C 1hr	0.26 +/- 0.00	0.99 +/- 0.40
<b>Stationary phase IVC quantitation (Fig 3 and 4)</b>		
<b>Strain/conditions</b>	<b>Cells with Pab1-GFP IVCs (% +/- st. dev.)</b>	<b>Significance (2-tailed T-test) relative to <i>atg15Δ</i></b>
<i>atg15Δ</i> (BY background)	54.79 +/- 13.35	
<i>atg8Δ atg15Δ</i>	22.74 +/- 6.28	0.011
<i>atg11Δ atg15Δ</i>	9.47 +/- 7.18	0.0025
<i>mon1Δ atg15Δ</i>	1.06 +/- 0.73	6.1x10 <sup>-4</sup>
<i>atg15Δ</i> (W303 background) - 27°C	48.25 +/- 4.78	
<i>atg15Δ</i> - 34°C 1hr	58.56 +/- 1.81	
<i>atg15Δ</i> - 34°C 2hr	58.64 +/- 5.73	
<i>cdc48-3 atg15Δ</i> - 27°C	33.82 +/- 1.28	0.007 (27°C)
<i>cdc48-3 atg15Δ</i> - 34°C 1hr	23.08 +/- 3.53	9.6x10 <sup>-5</sup> (34°C 1hr)
<i>cdc48-3 atg15Δ</i> - 34°C 2hr	11.00 +/- 1.79	1.6x10 <sup>-4</sup> (34°C 2hr)
<i>atg15Δ</i> (RP840 background)	33.08 +/- 9.75	
<i>atg15Δ</i> (RP840 background) - 37°C 1hr	37.93 +/- 11.92	
<i>dcp2-7 atg15Δ</i>	59.26 +/- 3.93	8.5x10 <sup>-5</sup>
<i>dcp2-7 atg15Δ</i> - 37°C 1hr	70.02 +/- 7.94	8.5x10 <sup>-5</sup>
<i>xrn1Δ atg15Δ</i>	69.46 +/- 8.44	8.6x10 <sup>-4</sup>
<b>Atg19/11-GFP co-localization (Figure S5)</b>		
	<b>Cells with Edc3-mCh foci overlap (% +/- st. dev)</b>	
Atg11-GFP (BY background, early stationary phase)	5.64 +/- 0.73	
Atg19-GFP (BY background, early stationary phase)	5.99 +/- 1.49	
	<b>Cells with Pub1-mCh foci overlap (% +/- st. dev)</b>	
<b>GFP-Atg19 co-localization (Figure S6)</b>		
	<b>Cells with Pub1-mCh/GFP-Atg19 IVC overlap</b>	
GFP-Atg19 - <i>mon1Δ</i> (BY background, early stationary phase)	7.6 +/- 2.42	
GFP-Atg19 - <i>mon1Δ</i> (BY background, early stationary phase)	16.03 +/- 3.69	
GFP-Atg19 - <i>atg15Δ</i> (BY background, early stationary phase)	71.19 +/- 6.85	
<b>HeLa Cell quantitation (Figure S7)</b>		
	<b>Cells with SGs (% +/- st. dev.)</b>	<b>Cells with PBs (% +/- st. dev)</b>
Unstressed	0.00 +/- 0	38.70 +/- 7.60
Arsenite 1mM - 1hr	100 +/- 0	90.31 +/- 6.28
Bafilomycin A1 0.5 $\mu\text{M}$ - 1hr	12.18 +/- 6.35	30.98 +/- 11.18
Arsenite 1mM - 20 min recovery	98.24 +/- 2.14	95.83 +/- 8.33
Arsenite 1mM - 1hr recovery	96.95 +/- 2.83	96.90 +/- 5.37
Arsenite 1mM - 2hr recovery	68.79 +/- 18.52	85.48 +/- 11.67
Arsenite 1mM - 3hr recovery	11.13 +/- 9.11	65.93 +/- 23.11
Arsenite 1mM - 3-MA (10 $\mu\text{M}$ ) 20 min recovery	92.39 +/- 9.75	82.95 +/- 29.54
Arsenite 1mM - 3-MA (10 $\mu\text{M}$ ) 1hr recovery	73.60 +/- 11.81	71.85 +/- 25.95
Arsenite 1mM - 3-MA (10 $\mu\text{M}$ ) 2hr recovery	36.08 +/- 15.17	84.79 +/- 5.24
Arsenite 1mM - 3-MA (10 $\mu\text{M}$ ) 3hr recovery	4.84 +/- 5.31	61.01 +/- 22.19
Arsenite 1mM - Rapamycin (10nM) 20 min recovery	84.01 +/- 0.96	70.03 +/- 10.64
Arsenite 1mM - Rapamycin (10nM) 1hr recovery	71.15 +/- 19.42	57.18 +/- 10.15
Arsenite 1mM - Rapamycin (10nM) 2hr recovery	73.05 +/- 4.93	60.85 +/- 9.19
Arsenite 1mM - Rapamycin (10nM) 3hr recovery	4.55 +/- 4.66	48.82 +/- 44.09
Arsenite 1mM - Wortmannin (1.5 $\mu\text{M}$ ) 20 min recovery	100 +/- 0	97.56 +/- 6.35
Arsenite 1mM - Wortmannin (1.5 $\mu\text{M}$ ) 1hr recovery	97.56 +/- 3.45	97.56 +/- 3.45
Arsenite 1mM - Wortmannin (1.5 $\mu\text{M}$ ) 2hr recovery	77.16 +/- 15.85	73.97 +/- 20.36
Arsenite 1mM - Wortmannin (1.5 $\mu\text{M}$ ) 3hr recovery	42.89 +/- 20.77	66.26 +/- 28.43

Details of quantitation methods presented in experimental procedures.

**Table S5. Strains and Plasmids Used in This Study, Related to Experimental Procedures**

<b>Yeast strains</b>		
<b>Name</b>	<b>Properties</b>	<b>Reference</b>
yRP840	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG</i>	Hatfield et al., 1996
yRP1501	<i>MATa leu2-3112 lys2-201 trp1 ura3-52 dcp2-7::URA3</i>	Dunckley et al, 2001
yRP2065	<i>MATa his3Δ0 leu2Δ0 met15Δ0 ura3Δ0 ('BY4741')</i>	Coller and Parker, 2005
yRP2882	<i>MATa ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 bar1D cdc48-3::HIS3</i>	Hsieh and Chen, 2011 (RHC677)
yRP2883	<i>MATa ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 bar1D ufd1-2</i>	Hsieh and Chen, 2011 (RHC1122)
yRP2884	<i>MATa ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 bar1D npl4-1</i>	Hsieh and Chen, 2011 (RHC1126)
yRP2893	<i>MATa ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 bar1D ('W303')</i>	Hsieh and Chen, 2011
yRP2894	<i>MATa his4-619 leu2-3,112 ura3-52 atg15::HYGB</i>	This study
yRP2896	<i>MATa leu2-3,112 trp1 ura3-52 his4-539 cup1::LEU2/PGK1pG/MFA2pG atg15::HYGB</i>	This study
yRP2897	<i>MATa his4-539 leu2-3,112 trp1-Δ1 ura3-52 xrn1::URA3 atg15::HYGB</i>	This study
yRP2898	<i>MATa leu2-3112 lys2-201 trp1 ura3-52 dcp2-7::URA3 atg15::HYGB</i>	This study
yRP2899	<i>MATa his3Δ0 leu2Δ0 met15Δ0 ura3Δ0 atg15::KANMX</i>	Invitrogen/Resgen collection
yRP2900	<i>MATa his3Δ0 leu2Δ0 met15Δ0 ura3Δ0 atg8::KANMX atg15::HYGB</i>	This study
yRP2901	<i>MATa his3Δ0 leu2Δ0 met15Δ0 ura3Δ0 atg11::KANMX atg15::HYGB</i>	This study
yRP2902	<i>MATa his3Δ0 leu2Δ0 met15Δ0 ura3Δ0 mon1::KANMX atg15::HYGB</i>	This study
yRP2903	<i>MATa leu2 ura3 his3 met15 VMA2-GFP (HIS)</i>	Huh et al., 2003
yRP2916	<i>MATa ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 bar1D atg15::HYGB</i>	This study
yRP2917	<i>MATa ade2-1 can1-100 his3-11,15 leu2-3,112 trp1-1 ura3-1 bar1D cdc48-3::HIS3 atg15::HYGB</i>	This study
<b>Plasmids</b>		
<b>Name</b>	<b>Properties</b>	<b>Reference</b>
pRP1574	Edc3-mCh; Cen; URA3 marker	Buchan et al, 2008
pRP1657	Pab1-GFP, Edc3-mCh; Cen; URA3 marker	Buchan et al, 2008
pRP1659	Pab1-GFP, Edc3-mCh; Cen; TRP1 marker	Buchan et al, 2008
pRP1661	Pub1-mCh; Cen; URA3 marker	Buchan et al, 2008
pRP2150	Pub1-mCh; Cen; LEU2 marker	Buchan et al, 2010
pRP1944	Edc3-mCh; Pbp1-GFP; Cen; TRP1 marker	Swisher and Parker, 2010
pRP2447	GFP-Atg19 (truncated ORF); Cen; URA3 marker	Shintani et al, 2002