

<i>Caulobacter crescentus</i> strains:			
Name	Genotype	Description	Source or Reference
CB15	CB15	<i>C. crescentus</i> wild type ATCC 19089 <i>Caulobacter vibrioides</i> LOT: 3967454	(Poindexter and Cohen-Bazire, 1964)
NA1000	CB15N	Laboratory strain derived from CB15	(Evinger and Agabian, 1977)
SoA0071	CB15 $\Delta popA$	markerless in frame deletion of <i>popA</i> in CB15 by allelic exchange using plasmid pAD8	this study
SoA0144	NA1000 $\Delta cc0655$	markerless in frame deletion of <i>cc0655</i> in NA1000 by allelic exchange using plasmid pSA93	this study
SoA0145	NA1000 $\Delta tipF$	markerless in frame deletion of <i>tipF</i> in NA1000 by allelic exchange using plasmid pSA94	this study
SoA0154	NA1000 $\Delta cc0740$	markerless in frame deletion of <i>cc0740</i> in NA1000 by allelic exchange using plasmid pAD2	this study
SoA0155	NA1000 $\Delta cc0857$	markerless in frame deletion of <i>cc0857</i> in NA1000 by allelic exchange using plasmid pSA91	this study
SoA0156	NA1000 $\Delta cc0896$	markerless in frame deletion of <i>cc0896</i> in NA1000 by allelic exchange using plasmid pSA102	this study
SoA0157	NA1000 $\Delta cc1086$	markerless in frame deletion of <i>cc1086</i> in NA1000 by allelic exchange using plasmid pSA79	this study
SoA0161	NA1000 $\Delta cc3094$	markerless in frame deletion of <i>cc3094</i> in NA1000 by allelic exchange using plasmid pSA96	this study
SoA0162	NA1000 $\Delta cc3148$	markerless in frame deletion of <i>cc3148</i> in NA1000 by allelic exchange using plasmid pSA90	this study
SoA0163	NA1000 $\Delta dgcA$	markerless in frame deletion of <i>dgcA</i> in NA1000 by allelic exchange using plasmid pSA80	this study
SoA0166	CB15 $\Delta cc0655$	markerless in frame deletion of <i>cc0655</i> in CB15 by allelic exchange using plasmid pSA93	this study
SoA0167	CB15 $\Delta tipF$	markerless in frame deletion of <i>tipF</i> in CB15 by allelic exchange using plasmid pSA94	this study
SoA0168	CB15 $\Delta cc0740$	markerless in frame deletion of <i>cc0740</i> in CB15 by allelic exchange using plasmid pAD2	this study
SoA0170	CB15 $\Delta cc0896$	markerless in frame deletion of <i>cc896</i> in CB15 by allelic exchange using plasmid pSA102	this study
SoA0171	CB15 $\Delta cc1086$	markerless in frame deletion of <i>cc1086</i> in CB15 by allelic exchange using plasmid pSA79	this study
SoA0173	CB15 $\Delta dgcB$	markerless in frame deletion of <i>dgcB</i> in CB15 by allelic exchange using plasmid pAD7	(Abel et al., 2011)
SoA0175	CB15 $\Delta cc3094$	markerless in frame deletion of <i>cc3094</i> in CB15 by allelic exchange using plasmid pSA96	this study

SoA0176	CB15 $\Delta cc3148$	markerless in frame deletion of <i>cc3148</i> in CB15 by allelic exchange using plasmid pSA90	this study
SoA0177	CB15 $\Delta dgcA$	markerless in frame deletion of <i>dgcA</i> in CB15 by allelic exchange using plasmid pSA80	this study
SoA0739	NA1000 pSA129	NA1000 containing pSA129	this study
SoA0741	CB15 pSA129	CB15 containing pSA129	this study
SoA0764	NA1000 $\Delta dgcB \Delta cc0857 \Delta cc0740 \Delta dgcA \Delta pleD \Delta cc0655 \Delta cc3094 \Delta cc0896 \Delta cc1086 \Delta cc0091 \Delta pdeA \Delta cc3148$	markerless in frame deletion of <i>cc1086</i> , <i>cc0091</i> , <i>pdeA</i> and <i>cc3148</i> in NA1000 $cdG^0$ background by allelic exchange using plasmids pSA79, pSA156, pSA81, and pSA90, respectively; referred to as $rcdG^0$ strain in NA1000 background	this study
SoA0801	NA1000 pSRK-Km	NA1000 containing pSRK-Km	this study
SoA0803	CB15 pSRK-Km	CB15 containing pSRK-Km	this study
SoA0827	CB15 $cdG^0 dgcB^+$	the c-di-GMP free strain in CB15 background containing a chromosomal <i>dgcB</i> copy in the <i>dgcB</i> locus which was introduced by general transduction using $\phi CR30$ and the marker CMS21 (West et al 2002).	this study
SoA0918	NA1000 $\Delta cc0091$	markerless in frame deletion of <i>cc0091</i> in NA1000 by allelic exchange using plasmid pSA156	this study
SoA0930	CB15 $\Delta cc0091$	markerless in frame deletion of <i>cc0091</i> in CB15 by allelic exchange using plasmid pSA156	this study
SoA1015	CB15 $\Delta pleD$ pSA164	CB15 $\Delta pleD$ containing pSA164	this study
SoA1061	CB15 $\Delta pleD$ pSA129	CB15 $\Delta pleD$ containing pSA129	this study
SoA1154	NA1000 $cdG^0$ pSA129	the c-di-GMP free strain in NA1000 background containing pSA129	this study
SoA1273	NA1000 $\Delta dgcB \Delta cc0857 \Delta cc0740 \Delta dgcA \Delta cc0655 \Delta cc3094 \Delta cc0896 \Delta cc1086 \Delta cc0091 \Delta pdeA \Delta cc3148$	$rcdG^0$ strain lacking the final markerless in frame deletion of <i>pleD</i> ; referred to as $rcdG^0::pleD$	this study
SoA1372	NA1000 $cdG^0$ pSRK-Km	the c-di-GMP free strain in NA1000 background containing pSRK-Km	this study
SoA1374	NA1000 $cdG^0::ydeH$ pSRK-Km	the c-di-GMP free strain in NA1000 background containing a chromosomal copy of YdeH integrated by allelic exchange using plasmid pSA223 and pSRK-Km	this study
SoA1481	CB15 $cdG^0$ pTB4	the c-di-GMP free strain in CB15 background containing pTB4	this study
SoA1496	NA1000 $cdG^0$ pSA280	the c-di-GMP free strain in NA1000 background containing pSA280	this study
SoA1498	CB15 $cdG^0$ pSA280	the c-di-GMP free strain in CB15 background containing pSA280	this study
SoA1507	CB15 $cdG^0$ pSA129	the c-di-GMP free strain in CB15 background containing pSA129	this study
SoA1508	CB15 $cdG^0$ pSRK-Km	the c-di-GMP free strain in CB15 background containing pSRK-Km	this study

SoA1511	CB15 <i>cdG<sup>0</sup>::ydeH</i> pSRK-Km	the c-di-GMP free strain in CB15 background containing a chromosomal copy of YdeH integrated by allelic exchange using plasmid pSA223 and pSRK-Km	this study
SoA1538	NA1000 ΔMGE	FC766; NA1000 with a deletion of the mobile genetic element between position 473069-499098 of the NA1000 genome	(Marks et al., 2002)
SoA1539	NA1000 ΔMGE pTB4	NA1000 ΔMGE containing pTB4	this study
SoA1541	NA1000 ΔMGE pSRK-Km	NA1000 ΔMGE containing pSRK-Km	this study
SoA1569	CB15 Δ <i>cc0857</i>	markerless in frame deletion of <i>cc0857</i> in CB15 by allelic exchange using plasmid pSA91	this study
SoA1587	<i>rcdG<sup>0</sup>::ydeH</i>	chromosomal integration of <i>ydeH</i> in <i>rcdG<sup>0</sup></i> by allelic exchange using pSA223	this study
SoA1612	NA1000 <i>hfsA<sup>+</sup></i> pSRK-Km	markerless exchange of <i>hfsA</i> <sub>NA1000</sub> to functional <i>hfsA</i> by allelic exchange using pSA178 in NA1000 containing pSRK-Km	this study
SoA1615	NA1000 <i>hfsA<sup>+</sup></i> ΔMGE pSRK-Km	markerless exchange of <i>hfsA</i> <sub>NA1000</sub> to functional <i>hfsA</i> by allelic exchange using pSA178 in NA1000 ΔMGE containing pSRK-Km	this study
SoA1616	NA1000 <i>cdG<sup>0</sup> hfsA<sup>+</sup></i> pSRK-Km	markerless exchange of <i>hfsA</i> <sub>NA1000</sub> to functional <i>hfsA</i> by allelic exchange using pSA178 in NA1000 <i>cdG<sup>0</sup></i> containing pSRK-Km	this study
SoA1617	NA1000 <i>hfsA<sup>+</sup></i> pTB4	markerless exchange of <i>hfsA</i> <sub>NA1000</sub> to functional <i>hfsA</i> by allelic exchange using pSA178 in NA1000 containing pTB4	this study
SoA1620	NA1000 <i>hfsA<sup>+</sup></i> ΔMGE pTB4	markerless exchange of <i>hfsA</i> <sub>NA1000</sub> to functional <i>hfsA</i> by allelic exchange using pSA178 in NA1000 ΔMGE containing pTB4	this study
SoA1621	NA1000 <i>cdG<sup>0</sup> hfsA<sup>+</sup></i> pTB4	markerless exchange of <i>hfsA</i> <sub>NA1000</sub> to functional <i>hfsA</i> by allelic exchange using pSA178 in NA1000 <i>cdG<sup>0</sup></i> containing pTB4	this study
SoA1624	NA1000 pSA266	NA1000 containing pSA266	this study
SoA1626	NA1000::CMS0	NA1000 containing a kanamycin resistance marker (CMS0) integrated via plasmid pLW132 linked to <i>cc0091</i>	(West et al., 2002)
SoA1627	<i>rcdG<sup>0</sup>::ydeH</i> ::CMS0	marker CMS0 transferred in <i>rcdG<sup>0</sup>::ydeH</i> by general transduction using φCR30, the strain retained the <i>cc0091</i> deletion	this study
SoA1628	NA1000:: <i>ydeH</i> ::CMS0 Δ <i>dgcb</i> Δ <i>cc0857</i> Δ <i>cc0740</i> Δ <i>dgca</i> Δ <i>pleD</i> Δ <i>cc0655</i> Δ <i>cc3094</i> Δ <i>cc0896</i> Δ <i>cc1086</i> Δ <i>pdeA</i> Δ <i>cc3148</i>	marker CMS0 transferred in <i>rcdG<sup>0</sup>::ydeH</i> by general transduction using φCR30, the strain obtained a wild-type <i>cc0091</i> allele	this study
SoA1632	NA1000::CMS12	NA1000 containing a kanamycin resistance marker (CMS12) integrated via plasmid pLW92 linked to <i>cc1086</i>	(West et al., 2002)
SoA1633	<i>rcdG<sup>0</sup>::ydeH</i> ::CMS12	marker CMS12 transferred in <i>rcdG<sup>0</sup>::ydeH</i> by general transduction using φCR30, the strain retained the <i>cc1086</i> deletion	this study

SoA1634	NA1000:: <i>ydeH</i> ::CMS12 <i>ΔdgcB Δcc0857 Δcc0740</i> <i>ΔdgcA ΔpleD Δcc0655</i> <i>Δcc3094 Δcc0896 Δcc0091</i> <i>ΔpdeA Δcc3148</i>	marker CMS12 tranfered in <i>rcdG<sup>0</sup>::ydeH</i> by general transduction using $\phi$ CR30, the strain optained a wild-type <i>cc1086</i> allele	this study
SoA1644	NA1000::CMS37	NA1000 containing a kanamycin resistance marker (CMS37) integrated via plasmid pLW124 linked to <i>pdeA</i>	(West et al., 2002)
SoA1645	<i>rcdG<sup>0</sup>::ydeH</i> ::CMS37	marker CMS37 tranfered in <i>rcdG<sup>0</sup>::ydeH</i> by general transduction using $\phi$ CR30, the strain retained the <i>pdeA</i> deletion	this study
SoA1646	NA1000:: <i>ydeH</i> ::CMS37 <i>ΔdgcB Δcc0857 Δcc0740</i> <i>ΔdgcA ΔpleD Δcc0655</i> <i>Δcc3094 Δcc0896 Δcc1086</i> <i>Δcc0091 Δcc3148</i>	marker CMS37 tranfered in <i>rcdG<sup>0</sup>::ydeH</i> by general transduction using $\phi$ CR30, the strain optained a wild-type <i>pdeA</i> allele	this study
SoA1648	<i>rcdG<sup>0</sup></i> pBV-PA5295	<i>rcdG<sup>0</sup></i> containing pBV-PA5295	this study
SoA1650	<i>rcdG<sup>0</sup>::ydeH</i> pBV-PA5295	<i>rcdG<sup>0</sup>::ydeH</i> containing pBV-PA5295	this study
SoA1652	<i>rcdG<sup>0</sup>::pleD</i> pBV-PA5295	<i>rcdG<sup>0</sup>::pleD</i> containing pBV-PA5295	this study
UJ2827	NA1000 <i>ΔpopA</i>	markerless in frame deletion of <i>popA</i> in NA1000 by allelic exchange using plasmid pAD8	(Duerig et al., 2009)
UJ3252	NA1000 <i>ΔpilA</i>	deletion of <i>pilA</i> in NA1000	Assaf Levi
UJ3257	CB15 <i>ΔflgH</i>	markerless in frame deletion of <i>flgH</i> in CB15 by allelic exchange using plasmid pAL2	Assaf Levi
UJ4449	NA1000 <i>ΔdgcB</i>	markerless in frame deletion of <i>dgcB</i> in NA1000 by allelic exchange using plasmid pAD7	(Abel et al., 2012)
UJ4450	NA1000 <i>ΔpleD</i>	markerless in frame deletion of <i>pleD</i> in NA1000 by allelic exchange using plasmid pSA95	(Abel et al., 2012)
UJ4454	NA1000 <i>ΔpdeA</i>	markerless in frame deletion of <i>pdeA</i> in NA1000 by allelic exchange using plasmid pSA81	(Abel et al., 2012)
UJ4462	CB15 <i>ΔdgcB</i>	markerless in frame deletion of <i>dgcB</i> in CB15 by allelic exchange using plasmid pAD7	(Abel et al., 2012)
UJ4463	CB15 <i>ΔpleD</i>	markerless in frame deletion of <i>pleD</i> in CB15 by allelic exchange using plasmid pSA95	(Abel et al., 2012)
UJ4467	CB15 <i>ΔpdeA</i>	markerless in frame deletion of <i>pdeA</i> in CB15 by allelic exchange using plasmid pSA81	(Abel et al., 2012)
UJ4618	CB15 <i>ΔdgcB Δcc0857</i>	markerless in frame deletion of <i>cc0857</i> in CB15 <i>ΔdgcB</i> by allelic exchange using plasmid pSA91	this study
UJ4760	CB15 <i>ΔdgcB Δcc0857</i> <i>Δcc0740</i>	markerless in frame deletion of <i>cc0740</i> in CB15 <i>ΔdgcB Δcc0857</i> by allelic exchange using plasmid pAD2	this study
UJ4773	CB15 <i>ΔdgcB Δcc0857</i> <i>Δcc0740 ΔdgcA</i>	markerless in frame deletion of <i>dgcA</i> in CB15 <i>ΔdgcB Δcc0857 Δcc0740</i> by allelic exchange using plasmid pSA80	this study
UJ4774	CB15 pBV-MCS4	CB15 containing pBV-MCS4	this study
UJ4776	CB15 pBV-PA5295 <sub>E328A</sub>	CB15 containing pBV-PA5295 <sub>E328A</sub>	this study

UJ4778	CB15 pBV-PA5295	CB15 containing pBV-PA5295	this study
UJ4811	CB15 $\Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc3285$ pAD5	CB15 $\Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc3285$ containing pAD5	this study
UJ4816	CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD$	markerless in frame deletion of <i>pleD</i> in CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca$ by allelic exchange using plasmid pSA95	this study
UJ4849	CB15 $\Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc2462 \Delta cc3285$ pAD5	CB15 $\Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc2462 \Delta cc3285$ containing pAD5	this study
UJ4929	CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD \Delta cc0655$	markerless in frame deletion of <i>cc0655</i> in CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD$ by allelic exchange using plasmid pSA93	this study
UJ4932	CB15 pAD5	CB15 containing pAD5	this study
UJ5061	CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD \Delta cc0655 \Delta cc3094$	markerless in frame deletion of <i>cc3094</i> in CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD \Delta cc0655$ by allelic exchange using plasmid pSA96	this study
UJ5065	NA1000 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD \Delta cc0655 \Delta cc3094 \Delta cc0896$	markerless in frame deletion of <i>pleD</i> in NA1000 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta cc0655 \Delta cc3094 \Delta cc0896$ by allelic exchange using plasmid pSA95; referred to as cdG <sup>0</sup> strain in NA1000 background	this study
UJ5100	CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD \Delta cc0655 \Delta cc3094 \Delta cc0896$	markerless in frame deletion of <i>cc0896</i> in CB15 $\Delta dgcb \Delta cc0857 \Delta cc0740 \Delta dgca \Delta pleD \Delta cc0655 \Delta cc3094$ by allelic exchange using plasmid pSA102; referred to as cdG <sup>0</sup> strain in CB15 background	this study
UJ5153	CB15 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ pAD5	CB15 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ containing pAD5	this study
UJ5154	CB15 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc2462 \Delta cc3285$ pAD5	CB15 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc2462 \Delta cc3285$ containing pAD5	this study
UJ5155	CB15 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ pAD5	CB15 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ containing pAD5	this study
UJ5170	NA1000 pTB4	NA1000 containing pTB4	this study
UJ5172	NA1000 cdG <sup>0</sup> pTB4	the c-di-GMP free strain in NA1000 background containing pTB4	this study
UJ5380	NA1000 pCM4	NA1000 containing pCM4	this study
UJ5381	NA1000 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ pCM4	NA1000 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ containing pCM4	this study
UJ5458	NA1000 pOP290-2	NA1000 containing pOP290-2	this study

UJ5459	NA1000 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ pOP290-2	NA1000 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ containing pOP290-2	this study
UJ5487	NA1000 pfljL-lacZ	NA1000 containing pfljL-lacZ	this study
UJ5488	NA1000 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ pfljL-lacZ	NA1000 $\Delta cc0655 \Delta cc0740 \Delta cc0857 \Delta cc0896 \Delta cc1850 \Delta cc2462 \Delta cc3094 \Delta cc3285$ containing pfljL-lacZ	this study
UJ7134	CB15 $\Delta pleD$ pSA217	CB15 $\Delta pleD$ containing pSA217	this study
UJ7137	CB15 cdG <sup>0</sup> pSA164	the c-di-GMP free strain in CB15 background containing pSA164	this study
UJ7138	CB15 cdG <sup>0</sup> pSA217	the c-di-GMP free strain in CB15 background containing pSA217	this study
UJ7156	CB15 <i>dgcB</i> <sub>E261Q</sub>	CB15 with a mutation coding for the DgcB active site mutant DgcB <sub>E261Q</sub> . Created by allelic exchange using plasmid pIH99	this study
UJ7162	CB15 cdG <sup>0</sup> <i>dgcB</i> <sub>E261Q</sub>	the c-di-GMP free strain in CB15 background with a mutation coding for the DgcB active site mutant DgcB <sub>E261Q</sub> . Created by allelic exchange using plasmid pIH99	this study

*Escherichia coli* strains:

Name	Genotype	Description	Source or Reference
DH5 $\alpha$	(F-) F <sup>-</sup> <i>endA</i> 1 <i>hsdR</i> 17 (rK-mK plus) <i>glnV</i> 44 <i>thi</i> 1 <i>recA</i> 1 <i>gyr</i> $\Delta$ (Nal <sup>R</sup> ) <i>relA</i> 1 $\Delta$ ( <i>lacIZYA-argF</i> )U169 <i>deoR</i> ( $\Phi$ 80dlac $\Delta$ ( <i>lacZ</i> ) M15)	Used for general cloning purposes	(Woodcock et al., 1989)
S17-1	RP4-2, Tc::Mu, KM-Tn7	Used as donor strain in conjugations to transfer oriT containing plasmids to <i>C. crescentus</i>	(Simon et al., 1983)

Phages:

Name	Description	Source or Reference
$\phi$ CbK	Bacteriophage that infects <i>C. crescentus</i> uses the pili as a receptor.	(Schmidt et al., 1965)
$\phi$ CR30	Bacteriophage that infects <i>C. crescentus</i> uses the paracrystalline surface layer protein as a receptor; can be used for generalized transduction.	(Ely and Johnson, 1977)

Plasmids:			
Name		Description	Source or Reference
pAD2		pNPTS138; used for inframe deletion of <i>cc0740</i>	Anna Dürig
pAD5		pMR20; <i>popA-egfp</i> under control of <i>popA</i> promoter	(Duerig et al. 2009)
pAD7		pNPTS138; used for inframe deletion of <i>dgcB</i>	(Abel et al., 2011)
pAD8		pNPTS138; used for clean deletion of <i>popA</i>	(Duerig et al., 2009)
pAL2		pNPTS138; used for inframe deletion of <i>flgH</i>	Assaf Levi
pBV-MCS4		Gent <sup>R</sup> , vanillate inducible medium-copy number plasmid	(Thanbichler et al., 2007)
pBV-PA5295		pBV-MCS4; <i>PA5292</i> under control of the vanillate promoter	(Duerig et al. 2009)
pBV-PA5295 <sub>E328A</sub>		pBV-MCS4; <i>PA5292</i> <sub>E328A</sub> active site mutant under the control of vanillate promoter	(Duerig et al. 2009)
pCM4		pRK-lacZ 290; <i>lacZ</i> under control of the <i>fliL</i> / <i>fliG</i> promoter	C. Mohr
pfljL-lacZ		pRK-lacZ 290; <i>lacZ</i> under control of the <i>fliL</i> promoter	E. Mangan
pIH99		pNPTS138; used for the introduction of <i>dgcB</i> <sub>g781c</sub> coding for DgcB <sub>E261Q</sub> by allelic exchange	this study
pLW124		Kan <sup>R</sup> , suicide vector pBGS18T containing a homology region to <i>C. crescentus</i> for chromosomal intergration close to <i>pdeA</i>	(West et al., 2002)
pLW132		Kan <sup>R</sup> , suicide vector pBGS18T containing a homology region to <i>C. crescentus</i> for chromosomal intergration close to <i>cc0091</i>	(West et al., 2002)
pLW92		Kan <sup>R</sup> , suicide vector pBGS18T containing a homology region to <i>C. crescentus</i> for chromosomal intergration close to <i>cc1086</i>	(West et al., 2002)
pMR10		RK2 based Kan <sup>R</sup> low copy number and broad host range vector with oriT	(Roberts et al., 1996)
pNPTS138		Kan <sup>R</sup> , suicide vector with <i>sacB</i> gene and oriT	D. Alley
pOP290-2		pRK-lacZ 290; <i>lacZ</i> under control of the <i>fliF</i> promoter	K. Quon
pSA79		pNPTS138; used for inframe deletion of <i>cc1086</i>	this study
pSA80		pNPTS138; used for inframe deletion of <i>dgcA</i>	this study
pSA81		pNPTS138; used for inframe deletion of <i>pdeA</i>	(Abel et al., 2011)
pSA90		pNPTS138; used for inframe deletion of <i>cc3148</i>	this study
pSA91		pNPTS138; used for inframe deletion of <i>cc0857</i>	this study
pSA93		pNPTS138; used for inframe deletion of <i>cc0655</i>	this study
pSA94		pNPTS138; used for inframe deletion of <i>tipF</i>	this study
pSA95		pNPTS138; used for inframe deletion of <i>pleD</i>	(Abel et al., 2011)
pSA96		pNPTS138; used for inframe deletion of <i>cc3094</i>	this study

pSA102		pNPTS138; used for inframe deletion of <i>cc0896</i>	this study
pSA129		pMR10 lacking the lac Promoter	this study
pSA156		pNPTS138; used for inframe deletion of <i>cc0091</i>	this study
pSA164		pSA129; <i>pleD</i> under control of the <i>pleD</i> promoter	(Abel et al., 2011)
pSA178		pNPTS138; used for the introduction of a functional <i>hfA</i> allele by allelic exchange	this study
pSA217		pSA129; <i>pleD</i> <sub>GG368DE</sub> under control of the <i>pleD</i> promoter	this study
pSA223		pNPTS138; used for the introduction of lac promoter driven <i>ydeH-3xflag</i> by allelic exchange in the intergenic region between <i>cc3065</i> and <i>cc3066</i>	this study
pSA266		pSRK-Km; <i>ydeH-gfp</i> under control of the lac promoter	this study
pSA280		pSA129; <i>ydeH-3xflag</i> under control of the lac promoter	this study
pSRK-Km		Kan <sup>R</sup> , IPTG inducible pBBR based medium-copy number plasmid	(Khan et al., 2008)
pTB4		pSRK-Km; <i>ydeH-3xflag</i> under control of the lac promoter	this study