

Supporting information

Conferring the ability to utilize inorganic polyphosphate on ATP-specific NAD kinase

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Figures S1, S2, S3 and S4

Table S1

Figure S1

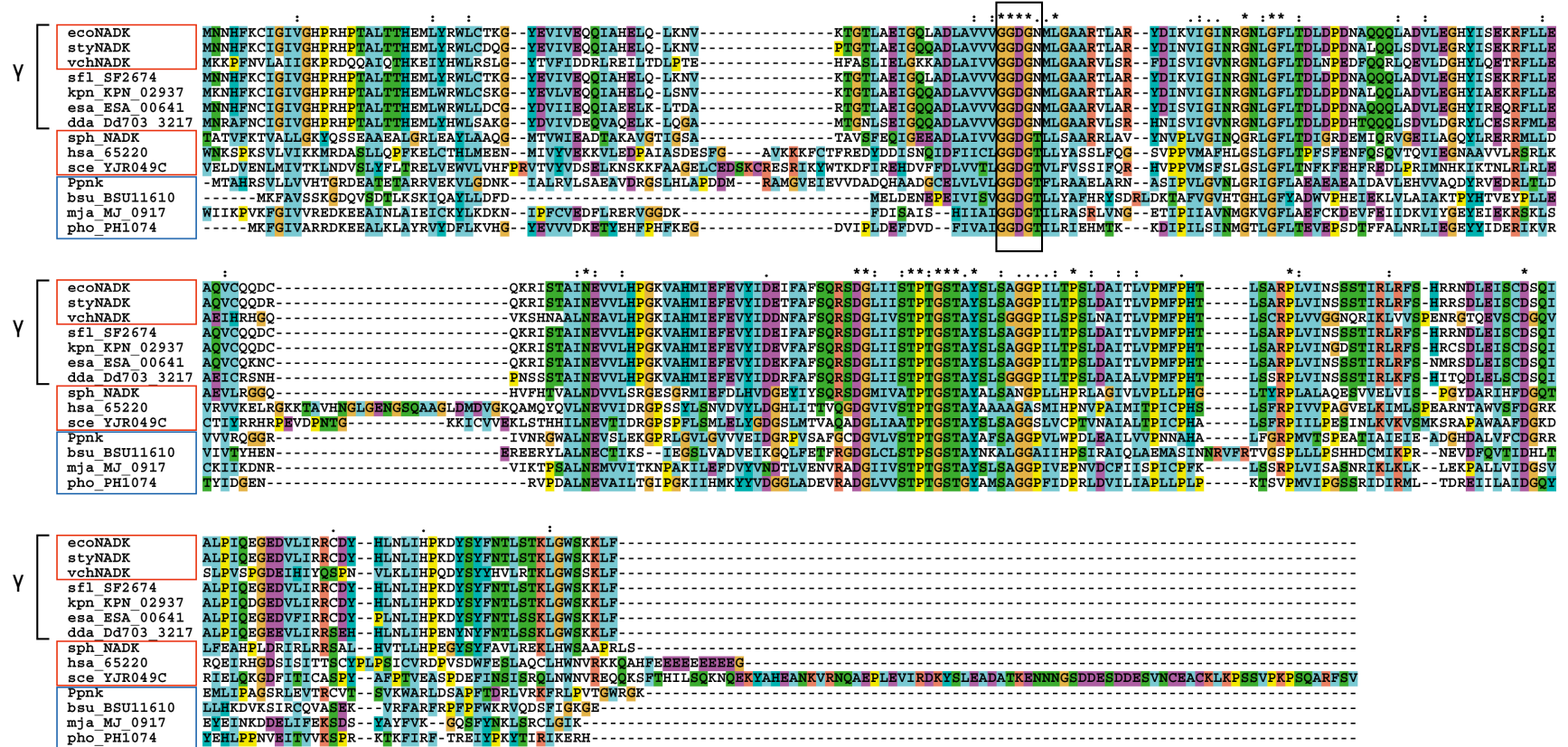


Figure S1. Multiple alignment of the primary structures of ATP-specific NADKs (boxed in red), poly(P)/ATP-NADKs (boxed in blue), and γ -proteobacterial NADK homologs (indicated by γ). ATP-specific NADKs are ecoNADK (*Escherichia coli*), styNADK (*Salmonella enterica*), vchNADK (*Vibrio cholerae*), sph_NADK (*Sphingomonas* sp. A1), hsa_65220 (human), and sce_YJR049C (*Saccharomyces cerevisiae*). Poly(P)/ATP-NADKs are Ppnk (*Mycobacterium tuberculosis*), bsu_11610 (*Bacillus subtilis*), mja_MJ_0917 (*Methanococcus jannaschii*), and pho_PH1074 (*Pyrococcus horikoshii*). γ -Proteobacterial NADK homologs are ecoNADK, styNADK, vchNADKs, sfl_SF2674K (*Shigella flexneri*), kpn_KPN_02937 (*Klebsiella pneumoniae*), esa_ESA_00641 (*Cronobacter sakazakii*), and dda_Dd703_3217 (*Dickeya dadantii*).

The motif GGDGN and GGDGT are boxed in black. The motif GGDGN is conserved in the primary structures of some γ -proteobacterial NADK homologs, while the motif GGDGT is found in those of other ATP-specific NADKs and poly(P)/ATP-NADKs. We failed to identify the key conserved residues in all ATP-specific NADKs or all poly(P)/ATP-NADKs; see also Fig. 1.

Figure S2

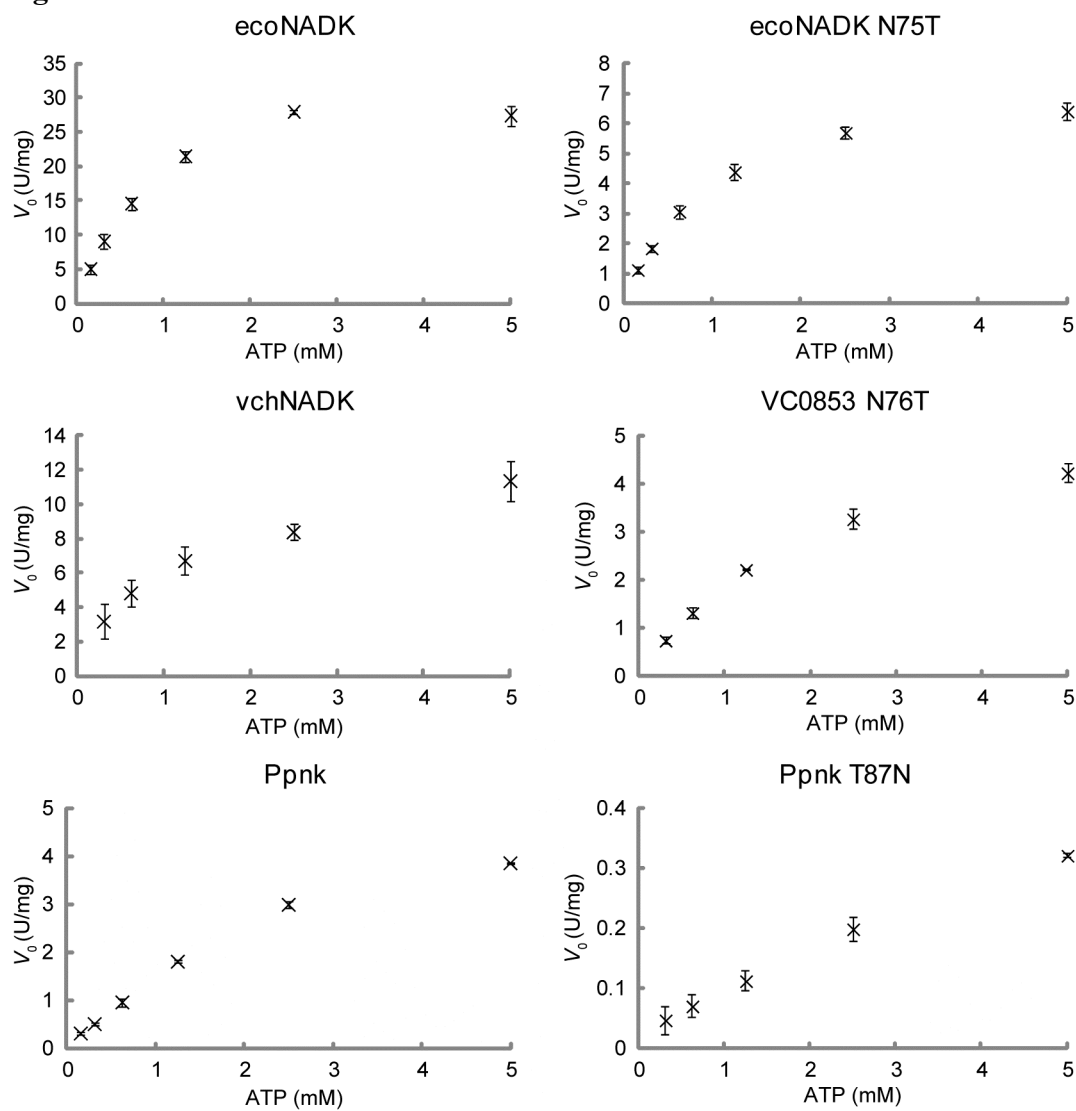


Figure S2. The data used to determine k_{cat} and K_m of each NADK toward ATP. Means and standard deviations of three independent experiments are shown.

Figure S3

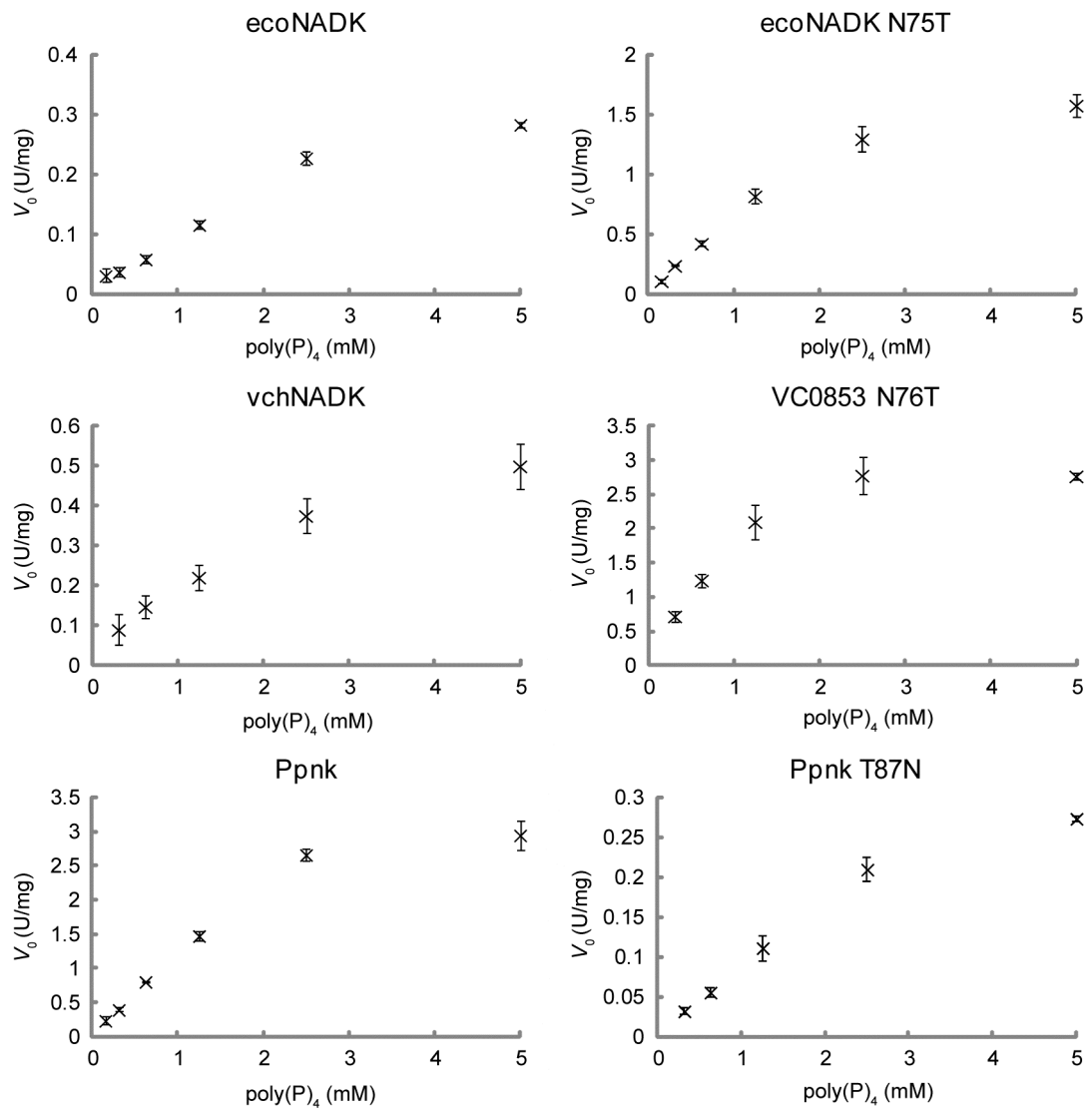


Figure S3. The data used to determine k_{cat} and K_m of each NADK toward poly(P)₄. Means and standard deviations of three independent experiments are shown.

Figure S4

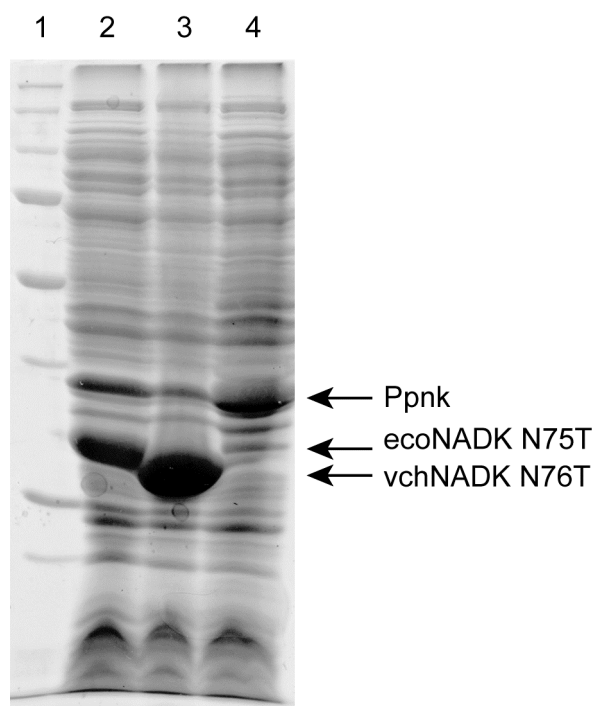


Figure S4. SDS-PAGE of extracts of cells expressing NADK. Cells were suspended in 100 mM Tris-HCl (pH 7.5) and ultrasonically disrupted at 9 kHz (Insonator model 201M; Kubota, Osaka, Japan) at 0°C for 10 min, and then centrifuged at 28,000 × *g* for 20 min at 4°C. The resultant clear supernatant was used as a cell extract, which was subjected to SDS-PAGE (12.5% gel). The remaining cells were immobilized and used for mass production of NADP⁺. Lane 1: Protein markers (Bio-Rad, Hercules, CA). Lanes 2–4: cell extracts (50 μg protein) of cells expressing ecoNADK N75T (lane 2), vchNADK N76T (lane 3), and Ppnk (lane 4).

Table S1. Amino acid residue of γ -proteobacterial NADK corresponding to Asn-75 of *eco*NADK

Order	Genera	N	T	S	F	Y	H
<i>Enterobacteriales</i>	<i>Cronobacter</i>	3	-	-	-	-	-
	<i>Dickeya</i>	4	-	-	-	-	-
	<i>Edwardsiella</i>	2	-	-	-	-	-
	<i>Enterobacter</i>	5	-	-	-	-	-
	<i>Erwinia</i>	6	-	-	-	-	-
	<i>Escherichia</i>	55	-	-	-	-	-
	<i>Klebsiella</i>	7	-	-	-	-	-
	<i>Pectobacterium</i>	3	-	-	-	-	-
	<i>Photorhabdus</i>	2	-	-	-	-	-
	<i>Pnatoea</i>	6	-	-	-	-	-
	<i>Proteus</i>	1	-	-	-	-	-
	<i>Providencia</i>	1	-	-	-	-	-
	<i>Rahnella</i>	3	-	-	-	-	-
	<i>Salmonella</i>	26	-	-	-	-	-
	<i>Serratia</i>	5	-	-	-	-	-
	<i>Shigella</i>	9	-	-	-	-	-
	<i>Xenorhabdus</i>	2	-	-	-	-	-
<i>Yersinia</i>	19	-	-	-	-	-	
<i>Pasteurellales</i>	<i>Actinobacillus</i>	4	-	-	-	-	-
	<i>Aggregatibacter</i>	4	-	-	-	-	-
	<i>Buchnera</i>	10	-	-	-	-	-
	<i>Haemophilus</i>	12	-	-	-	-	-
	<i>Mannheimia</i>	1	-	-	-	-	-
	<i>Pasteurella</i>	4	-	-	-	-	-
	<i>Sodalis</i>	1	-	-	-	-	-
	<i>Wigglesworthia</i>	-	-	2	-	-	-
<i>Vibrionales</i>	<i>Aliivibrio</i>	3	-	-	-	-	-
	<i>Listonella</i>	1	-	-	-	-	-
	<i>Photobacterium</i>	16	-	-	-	-	-
	<i>Vibrio</i>	20	-	-	-	-	-
<i>Aeromonadales</i>	<i>Aeromonas</i>	3	-	-	-	-	-

Table S1. Continued

Order	Genera	N	T	S	F	Y	H
<i>Alteromonadales</i>	<i>Alteromonas</i>	-	-	2	-	-	-
	<i>Colwellia</i>	-	-	-	-	1	-
	<i>Glaciecola</i>	1	-	-	-	1	-
	<i>Idiomarina</i>	1	-	-	-	-	-
	<i>Mariniobacter</i>	-	-	3	-	-	-
	<i>Oceanimonas</i>	1	-	-	-	-	-
	<i>Pseudoalteromonas</i>	3	-	-	-	-	-
	<i>Psychromonas</i>	-	-	-	-	1	-
	<i>Saccharophagus</i>	-	-	1	-	-	-
	<i>Shewanella</i>	24	-	-	-	-	-
<i>Oceanospirillales</i>	<i>Alcanivorax</i>	-	-	1	-	-	-
	<i>Chromohalobacter</i>	-	-	1	-	-	-
	<i>Hahella</i>	-	-	1	-	-	-
	<i>Halomonas</i>	-	-	1	-	-	-
	<i>Marinomonas</i>	-	-	3	-	-	-
<i>Pseudomonadales</i>	<i>Acinetobacter</i>	-	-	13	-	-	-
	<i>Azotobacter</i>	-	-	1	-	-	-
	<i>Cellvibrio</i>	-	-	1	-	-	-
	<i>Moraxella</i>	-	-	1	-	-	-
	<i>Pseudomonas</i>	-	-	27	-	-	-
	<i>Psychrobacter</i>	-	-	3	-	-	-
<i>Legionellales</i>	<i>Coxiella</i>	-	-	5	-	-	-
	<i>Legionella</i>	-	-	7	-	-	-
<i>Methylococcales</i>	<i>Alkalilimnicola</i>	-	-	-	-	1	-
<i>Chromatiales</i>	<i>Allochromatium</i>	-	-	-	-	1	-
	<i>Halorhodospira</i>	-	-	-	-	1	-
	<i>Halothiobacillus</i>	-	-	-	-	1	-
	<i>Methylococcus</i>	-	-	-	-	1	-
	<i>Methylomicrobium</i>	-	-	-	-	1	-
	<i>Methylomonas</i>	-	-	-	-	1	-
	<i>Nitrosococcus</i>	-	-	-	-	3	-
	<i>Thioalkalivibrio</i>	-	-	-	-	2	-
<i>Xanthomonadales</i>	<i>Frateuria</i>	-	1	-	-	-	-
	<i>Pseudoxanthomonas</i>	-	2	-	-	-	-
	<i>Stenotrophomonas</i>	-	4	-	-	-	-
	<i>Xanthomonas</i>	-	11	-	-	-	-
	<i>Xylella</i>	-	5	-	-	-	-

Table S1. Continued

Order	Genera	N	T	S	F	Y	H
<i>Thiotrichales</i>	<i>Francisella</i>	15	-	-	-	-	-
	<i>Methylophaga</i>	-	-	-	-	2	-
	<i>Thioalkalimicrobium</i>	-	-	-	-	1	-
	<i>Thiomicrospira</i>	-	-	-	-	1	-
<i>Acidithiobacillales</i>	<i>Acidithiobacillus</i>	-	-	-	-	4	-
<i>Cardiobacteriales</i>	<i>Dichelobacter</i>	-	-	-	-	1	-
others	<i>Baumannia</i>	1	-	-	-	-	-
	<i>Blochmannia</i>	2	-	-	-	-	-
	<i>Ferrimonas</i>	1	-	-	-	-	-
	<i>Gallibacterium</i>	1	-	-	-	-	-
	<i>Hamiltonella</i>	1	-	-	-	-	-
	<i>Histophilus</i>	2	-	-	-	-	-
	<i>Kangiella</i>	-	-	1	-	-	-
	<i>Riesia</i>	-	-	1	-	-	-
	<i>Ruthia</i>	-	-	1	-	-	-
	<i>Teredinibacter</i>	-	-	1	-	-	-
	<i>Tolumonas</i>	-	-	-	-	-	1
	<i>Unclassified</i>	-	-	1	-	-	-
<i>Vesicomysocius</i>	-	-	1	-	-	-	