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Supplemental Data

An X-Linked Cobalamin Disorder Caused by Mutations

in Transcriptional Coregulator *HCFC1*

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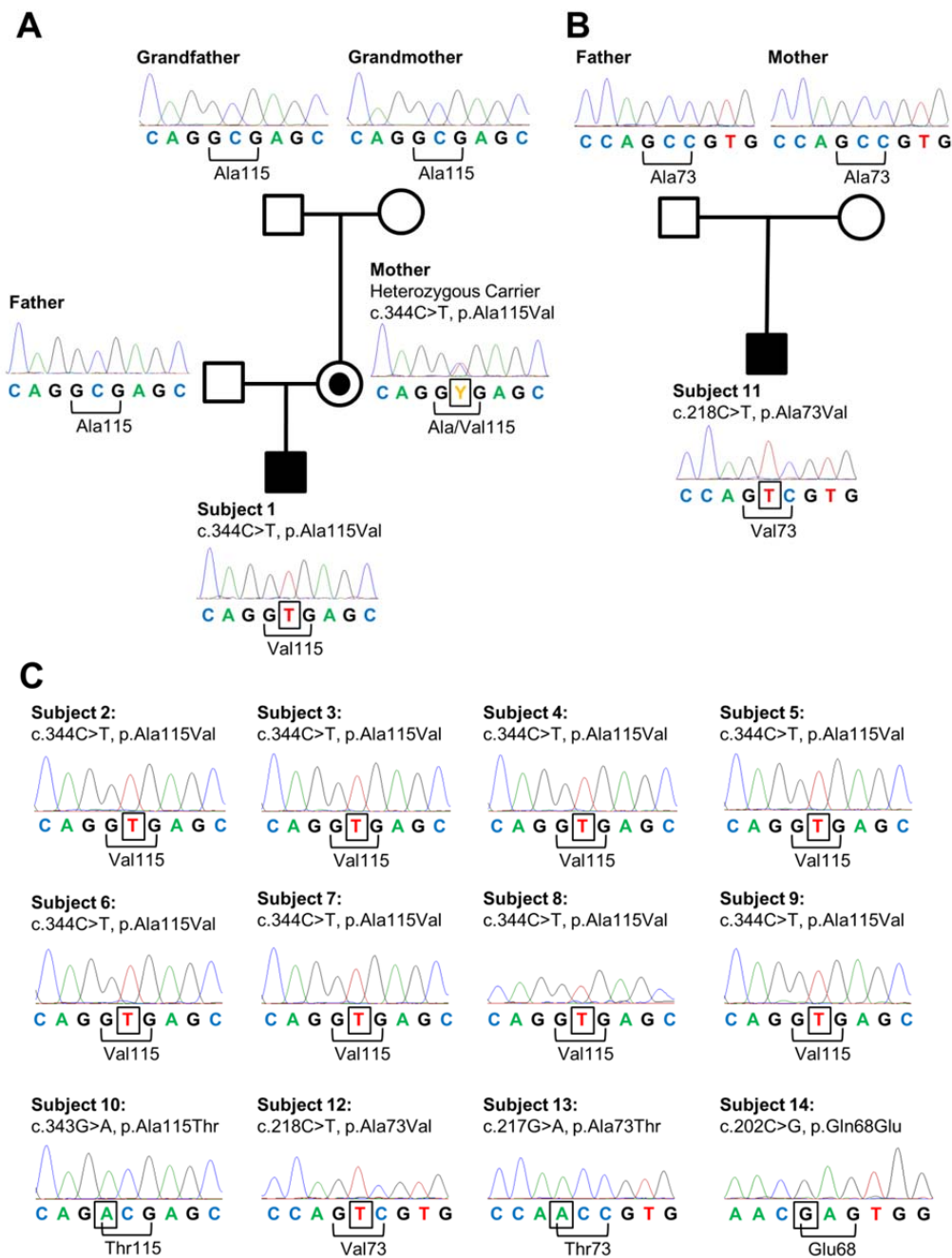


Figure S1. Pathogenic Variants in *HCFC1* Verified by Sanger Sequencing

(A) Partial chromatograms showing Sanger sequencing results of Exon 3 of *HCFC1* in the family of Subject 1. Subject 1 has a missense mutation c.344C>T (p.Ala115Val) (black box) inherited from his mother who is a heterozygous carrier with one copy each of the C and T (*de novo*) allele shown as Y (pyrimidine), due to dual peaks. (B) Partial chromatograms showing Sanger sequencing results of Exon 2 of *HCFC1* in Subject 11 and his parents. Subject 11 has a *de novo* mutation c.218C>T (p.Ala73Val) (black box). Both parents carry the normal C allele. (C) Subject 2-9 have c.344C>T (p.Ala115Val) mutation, Subject 10 has c.343G>A (p.Ala115Thr) mutation, Subject 12 has c.218C>T (p.Ala73Val) mutation, Subject 13 has c.217G>A (p.Ala73Thr) mutation, and Subject 14 has c.202C>G (p.Gln68Glu) mutation.

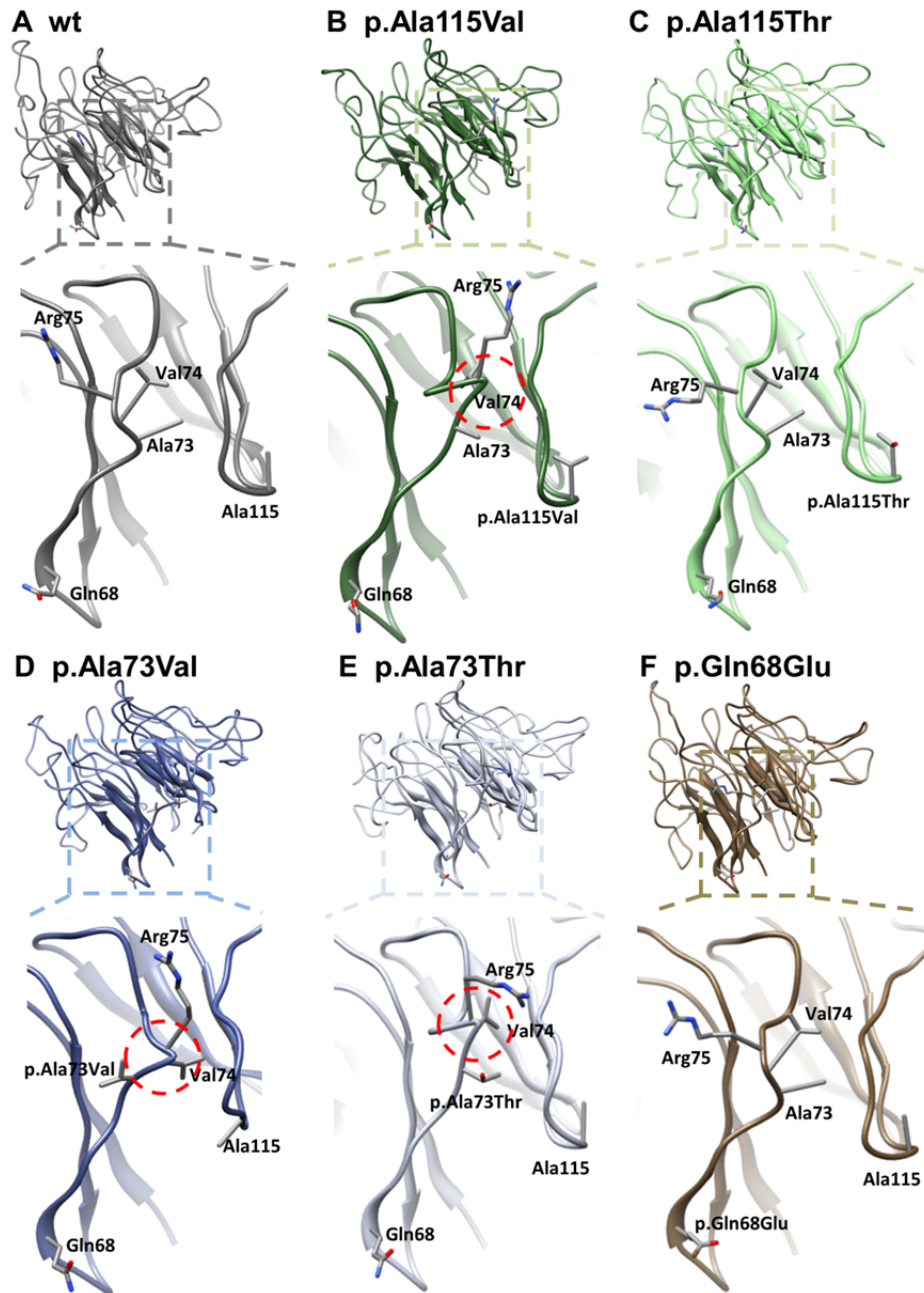


Figure S2. Protein Modeling of Wild-type and Mutant HCFC1

The entire protein is shown on the top and the expanded view of the region where the mutations have occurred is shown in the bottom. (A) Predicted structure of wild-type HCFC1 with a close-up of the region containing Gln 68, Ala73, Val74, Arg75, and Ala115 in the bottom panel. The side chains of these amino acids are also shown. The predicted structural changes due to the p.Ala115Val (B), p.Ala115Thr (C), p.Ala73Val (D), p.Ala73Thr (E) and p.Gln68Glu (F) mutations as predicted by Modeller¹. In p.Ala115Val, p.Ala73Val and p.Ala73Thr mutations, the close-up view indicates that this amino acid change generates a conformational change resulting in a kink (red dashed circle) between Val74 and Arg75. The p.Gln68Glu mutation did not generate a similar conformational change.

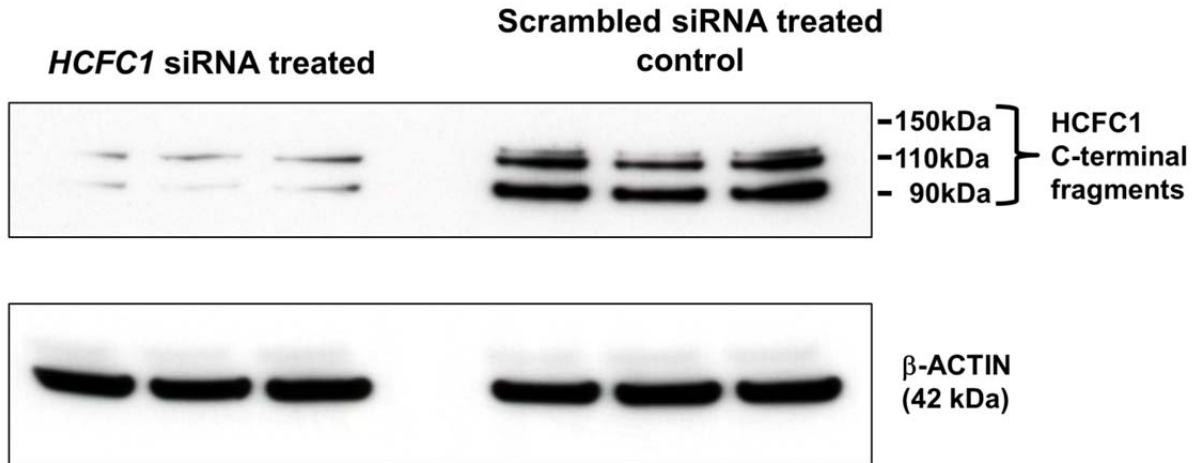


Figure S4. HCFC1 Expression in siRNA Knockdown Cells

Western blot of *HCFC1* siRNA knockdown in HEK293 cells. Three biological replicates of *HCFC1* siRNA treated cells are shown in the left and replicates of scrambled siRNA treated HEK293 cells are shown in the right. In the bottom panel, β -actin was used as a loading control.

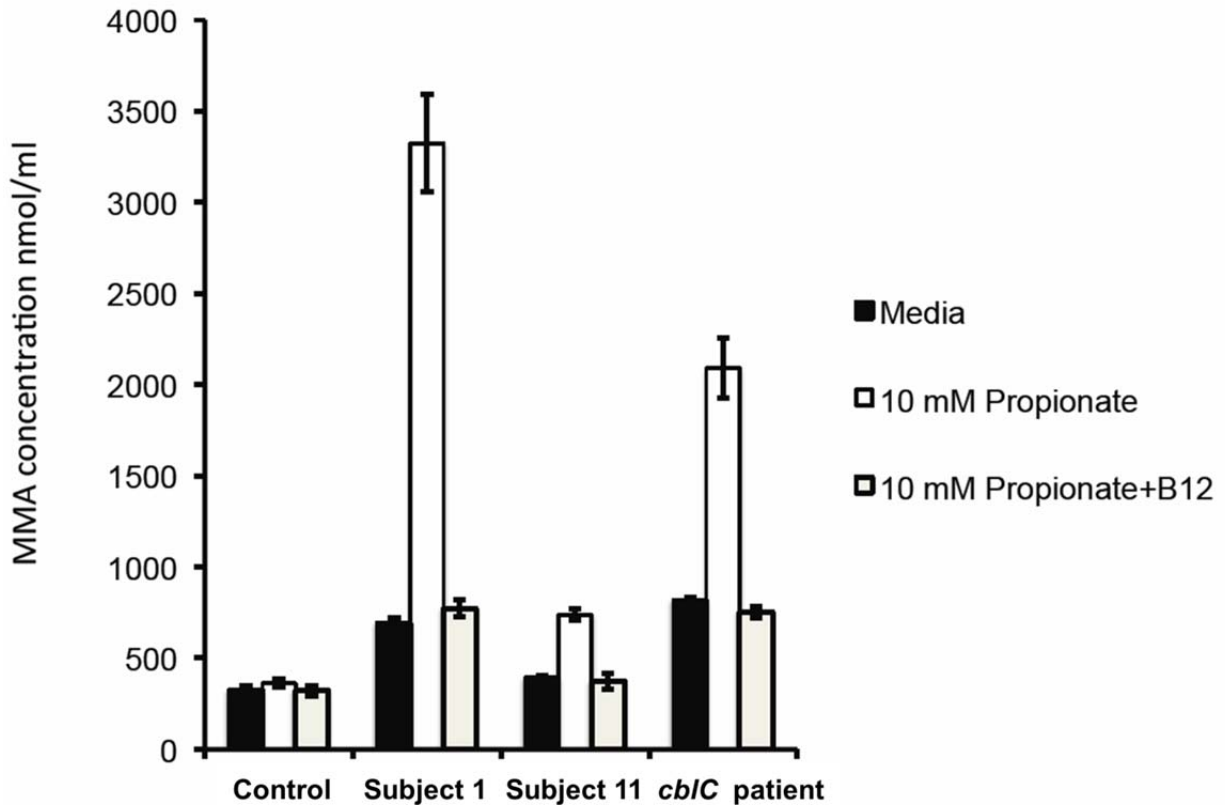


Figure S5. MMA Production by *cbIX* Fibroblasts in Cell Culture

5×10^5 cells, plated in triplicate, were incubated in complete DMEM medium (solid black bar), medium with 10 mM propionate added (white bar) or medium with 10 mM propionate and 1 μ g/ml hydroxocobalamin (grey bar) for 72 hours. The concentration of MMA (nmol/ml) was assayed in the medium. Fibroblasts from Subjects 1 and 11 show a 5.9- or 2.2-fold increase after incubation in propionic acid that was corrected with the addition of exogenous hydroxocobalamin to the medium, comparable to the response of the *cbIC* cell line, which rises 2.6 fold after identical treatment and also responds to exogenous hydroxocobalamin. Error bars, +/- 1 s.d. (n=3 replicates per cell line).

Table S1. PCR Primers

	5'-3'
Validation of WES result in Subject 1	
HCFC1 validation forward	CACCTTGAATGTTGTTCTTTGGGTCCTCG
HCFC1 validation reverse	TGACCTCTACGAACTCCAGGTAAAGCAGCC
TTN validation forward 1	CAAGTACTTTGCAATTAACGGCCACAGACC
TTN validation reverse 1	AGAGATCCTTGTGACCCTCCTGGACAACC
TTN validation forward 2	GTGTTTTTATTGCTCTCACCCATCGCAGGC
TTN validation reverse 2	AAGCAACACTTACATGGACACCTCCCTTGG
Mutation Analysis in other Subjects	
HCFC1_exon1_forward	ACCAGTCGGACTAGGGGCTTTCTTTAACTAACG
HCFC1_exon1_reverse	ATGGAGTCGTCTACCGCTGTGTGAGAAACC
HCFC1_exon2_forward	TCTTTGCTTTGAGTCTCTTCCACTCCCACC
HCFC1_exon2_reverse	AGTGGCTAGATTCCCAGTAGCTGATGAGGG
HCFC1_exon3_forward	TGGGACTACCTCCAGACAGAGAGTCCTTGG
HCFC1_exon3_reverse	GACCTCTACGAACTCCAGGTAAAGCAGCC
HCFC1_exon4_forward	ATGACAAGAGGAAAGCACGAAGTGGTATGC
HCFC1_exon4_reverse	CCAGTTGGACTCATCCTAGGGAGCTTCTGG
HCFC1_exon5&6_forward	GAGGCTTTTGCACTTTTGGATGTTGGGC
HCFC1_exon5&6_reverse	GTACCTGAAGGAGCGCTATTGCTAGCCTGG
HCFC1_exon7&8_forward	CTACAACCCTTGGAGACAAGCCCAAAGC
HCFC1_exon7&8_reverse	TTGACAACAAAAGAACCAAGGGCAACCAGGG
HCFC1_exon9_forward	TATTAATTCCCACCATGACCTCGCAGGAGC
HCFC1_exon9_reverse	CTTTGGGCTTGTCTCCAAGGGTTGTAGGG
HCFC1_exon10_forward	AAAGCGATGGGGAAACCACCCAAGATGTCC
HCFC1_exon10_reverse	GGTCTGGCAGTAGAGAAAGCTGAGCCATGC
HCFC1_exon11&12_forward	GCTGAGGTCACCAGCTTCAGGATTGTTCCC
HCFC1_exon11&12_reverse	GAAGGTGAGTGTTGAGGCAGGAGCTATGGG
HCFC1_exon13&14_forward	TTAAGCACCACTGGAACACCAGAGGAAAG
HCFC1_exon13&14_reverse	CATCCAGGTGAGCTCTCAGTCTTTGCACAT
HCFC1_exon15&16_forward	GACAGCCAAACCACGGACTATGACACAGGG
HCFC1_exon15&16_reverse	CACCTGCGAAAATCATCACTGCTGTCCC
HCFC1_exon17_forward_1	CGACATGGCTGTAGTGGCAGTGTTGGTGG
HCFC1_exon17_reverse_1	CCTCTTCCTCATCCCAGTTGCCTTTGAGG
HCFC1_exon17_forward_2	CTCACATGGTGGGTTGGAGCAGACTTGG
HCFC1_exon17_reverse_2	GCTGCTTCTTGTGACCCTCGACTGTGG
HCFC1_exon17_forward_3	AACTCATGTTGGAAGTGACAGTGGTGGCCG
HCFC1_exon17_reverse_3	CTTTTGTGCAGTTGGCCCCTCTGAGC
HCFC1_exon18_forward	GTCTTATACAGGCTGCGGTGAAGTGACACG
HCFC1_exon18_reverse	CACTTCTTCCCACAGGACCGTGGAGTCTGG
HCFC1_exon19_forward	CCTGTCTACACGGCAGTCTCTACTACTTGG
HCFC1_exon19_reverse	GAAACTTAGGCCCAAGGAAAGGTGAGG
HCFC1_exon20_forward	AGGCCTAGGAAGAAAGAAGGTGTCAGCG
HCFC1_exon20_reverse	CAGATGTGATGGGGAGGACGAGCACAGG
HCFC1_exon21&22_forward	TAACTATGGGCCAGAGGGAAAAGAGGTCAGC
HCFC1_exon21&22_reverse	CTCAACGGCCACTGAGAGTAAGCGACTGAGG
HCFC1_exon23_forward	CCCCCAAGTGGAAGGACTAAGACAAACTGG

HCFC1_exon23_reverse	GGGCTTACAGGAAACTCCTCCATCTTTGGG
HCFC1_exon24_forward	TTTCGGGCCTGGAAACCTACACTTTCCC
HCFC1_exon24_reverse	CCTTCCACTTGGGGGAAGATCTCTTTGAAGC
HCFC1_exon25&26_forward	GAAAGAGAAAGGGGAGAGGAGTGAACAGCG
HCFC1_exon25&26_reverse	TGAGTGGGTTTCTAACCTACGTCCAGAGGG
qPCR Expression Analysis	
HCFC1 left	ACACGCTGGCTTGTCTCAA
HCFC1 right	GGGATGTTGTCCTCCAGTGT
HCFC1 Universal Probe number	14
MMACHC left	GCTTCGAGGTTTACCCCTTC
MMACHC right	AGGCCAGGGTAGGTCCTG
MMACHC Universal Probe number	19
MMADHC left	GCTATTTTGCGA ACTATCTTCTCC
MMADHC right	TCCTGGGAGATAGGAAACCA
MMADHC Universal Probe number	21
MTR left	CAGAGTGCTTAACGGCACAG
MTR right	TAACAGTGGCCAGCACGAT
MTR Universal Probe number	35
SUCLG1 left	TTCACCAAACAACGCAAGTT
SUCLG1 right	TAAAAGGATCACCTCCAATGC
SUCLG1 Universal Probe number	43
ABCD4 left	ATGCAGATTCGGGTGAATG
ABCD4 right	TCTGTCCTCATGTGCTCCAC
ABCD4 Universal Probe number	40

Table S2. Enzymatic Studies in Skin Fibroblasts and Biochemical Findings in *cbIX* Patients

Subject number	<i>HCFC1</i> variant	¹⁴ C propionate incorporation ^a		¹⁴ C methylTHF incorporation ^b		¹⁴ C formate incorporation into methionine ^c		AdoCbl%	MeCbl%	Urine Hcy ^d	Propionylcarnitine ^e
		Ref. range: 10.8±3.7		Ref. range: 225±165		Ref. range: 2.2±0.6					
		basal medium	medium + OHCbl	basal medium	medium + OHCbl	basal medium	medium + OHCbl				
1	p.Ala115Val	1.0	3.2	27	52	nd	nd	3.9	1.5	nd	nd
2	p.Ala115Val	1.8	2.6	49	61	nd	nd	6.5	2.8	61	nd
3	p.Ala115Val	2.4	5.9	50	74	nd	nd	7.1	10.6	56	nd
4	p.Ala115Val	1.9	4.1	47	60	nd	nd	3.8	6.7	nd	nd
5	p.Ala115Val	3.3	6.0	48	73	nd	nd	6.5	2.6	nd	nd
6	p.Ala115Val	1.5	3.9	23	32	nd	nd	9.8	15.9	nd	nd
7	p.Ala115Val	1.4	4.1	21	39	nd	nd	3.1	2.3	nd	nd
8	p.Ala115Val	1.2*	8.0*	nd	nd	0.3	1.9	12.9	5.1	nd	nd
9	p.Ala115Val	1.4*	7.3*	nd	nd	0.4	1.6	0	1.1	nd	nd
10	p.Ala115Thr	0.6	0.9	36	52	nd	nd	3.0	0.7	nd	nd
11	p.Ala73Val	2.0	4.7	48	54	nd	nd	2.7	15.6	nd	2.16
12	p.Ala73Val	3.7*	11.4*	nd	nd	0.6	1.3	1.0	1.6	nd	nd
13	p.Ala73Thr	1.9	3.0	38	44	nd	nd	5.2	16.7	nd	nd
14	p.Gln68Glu	2.5	4.0	61	87	nd	nd	9.6	6.9	nd	nd

Ref. range: Reference range(mean ± SD) ; OHCbl: hydroxycobalamin ; THF: tetrahydrofolate; AdoCbl: adenosylcobalamin; MeCbl: methylcobalamin; tHcy: total homocysteine; Hcy: homocystine; MMA: methylmalonic acid; nl: normal; nd: not determined; und, undetectable

a: (nmol/mg protein/18 hrs), * nmol/mg protein/16hrs in patients 8, 9 and 12 (reference range 11.7 ± 4.3); b: (pmol/mg protein/18h); c: (nmol/mg protein/16h); d: μmol/L; e: nmol/mL

Table S3. Summary of Exome Variants and Test of Inheritance Models in Subject 1

Subject 1				
Total variants	76,191			
Coding variants	20,279			
Nonsynonymous, splice-site, InDel variants	9,249			
Rare variants	161			
Test of inheritance model	Dominant model	Recessive models		
	<i>de novo</i>	Compound heterozygous	Homozygous	X-linked hemizygous
	0	1 (<i>TTN</i>)	0	1 (<i>HCFC1</i>)

Table S4. Annotation of Mutation Effect and Conservation Score by SeattleSeq134

cDNA variant	Amino acid	PolyPhen-2 ^a	Grantham Score ^b	Conservation GERP ^c	Conservation phastCons ^d
c.344C>T	p.Ala115Val	0.997 (Probably Damaging)	64	5.25	0.981
c.343G>A	p.Ala115Thr	0.998 (Probably Damaging)	58	5.25	0.998
c.218C>T	p.Ala73Val	0.991 (Probably Damaging)	64	5.46	0.958
c.217G>A	p.Ala73Thr	0.731 (Possibly Damaging)	58	5.46	0.952
c.202C>G	p.Gln68Glu	0.722 (Possibly Damaging)	29	5.46	1

a: 1=Most Damaging, 0=Most Benign; b: 125=Most Damaging, 0=Most Benign; c: 6.17=Most Conserved, -12.3=Least Conserved; d: 1=Most Conserved, 0=Least Conserved.

Table S5. Human Variation Panel Controls from Coriell Institute

	Sample ID	Gender*		Sample ID	Gender*
Individuals of European Descent	NA17202	F	African Americans	NA17101	M
	NA17203	M		NA17102	M
	NA17204	F		NA17103	M
	NA17205	M		NA17104	M
	NA17206	F		NA17105	M
	NA17207	M		NA17106	M
	NA17208	F		NA17107	M
	NA17209	F		NA17108	M
	NA17210	F		NA17109	M
	NA17211	M		NA17110	F
	NA17212	M		NA17111	M
	NA17213	F		NA17112	F
	NA17214	M		NA17113	F
	NA17215	F		NA17114	M
	NA17216	F		NA17115	M
	NA17218	F		NA17116	F
	NA17219	M		NA17117	M
	NA17220	M		NA17118	F
	NA17221	F		NA17119	F
	NA17222	M		NA17120	F
	NA17223	M		NA17121	F
	NA17224	F		NA17122	F
	NA17225	M		NA17123	F
	NA17226	M		NA17124	F
	NA17227	M		NA17125	M
	NA17228	M		NA17126	F
	NA17229	F		NA17127	F
	NA17230	F		NA17128	F
	NA17231	M		NA17129	F
	NA17232	M		NA17130	F
	NA17233	F		NA17131	F
	NA17234	F		NA17132	F
	NA17235	M		NA17134	F
	NA17236	F		NA17135	F
	NA17237	M		NA17137	F
	NA17238	F		NA17138	F
	NA17239	F		NA17149	F
	NA17240	M		NA17150	F
	NA17241	M		NA17151	F
	NA17242	F		NA17152	M
	NA17243	M		NA17154	F
	NA17244	M		NA17156	F
	NA17245	F		NA17157	F
	NA17246	F		NA17164	F
	NA17247	F		NA17167	F

	NA17248	M		NA17173	F
	NA17250	F		NA17176	F
	NA17251	F		NA17181	F
	NA17252	F		NA17185	F
	NA17253	M		NA17189	F

*: M=Male, F=Female

References

1. Eswar, N., Webb, B., Marti-Renom, M.A., Madhusudhan, M.S., Eramian, D., Shen, M.-Y., Pieper, U., and Sali, A. (2006). Comparative protein structure modeling using Modeller. *Curr Protoc Bioinformatics Chapter 5*, Unit 5.6.
2. Dejosez, M., Levine, S.S., Frampton, G.M., Whyte, W.A., Stratton, S.A., Barton, M.C., Gunaratne, P.H., Young, R.A., and Zwaka, T.P. (2010). Ronin/Hcf-1 binds to a hyperconserved enhancer element and regulates genes involved in the growth of embryonic stem cells. *Genes Dev.* *24*, 1479–1484.