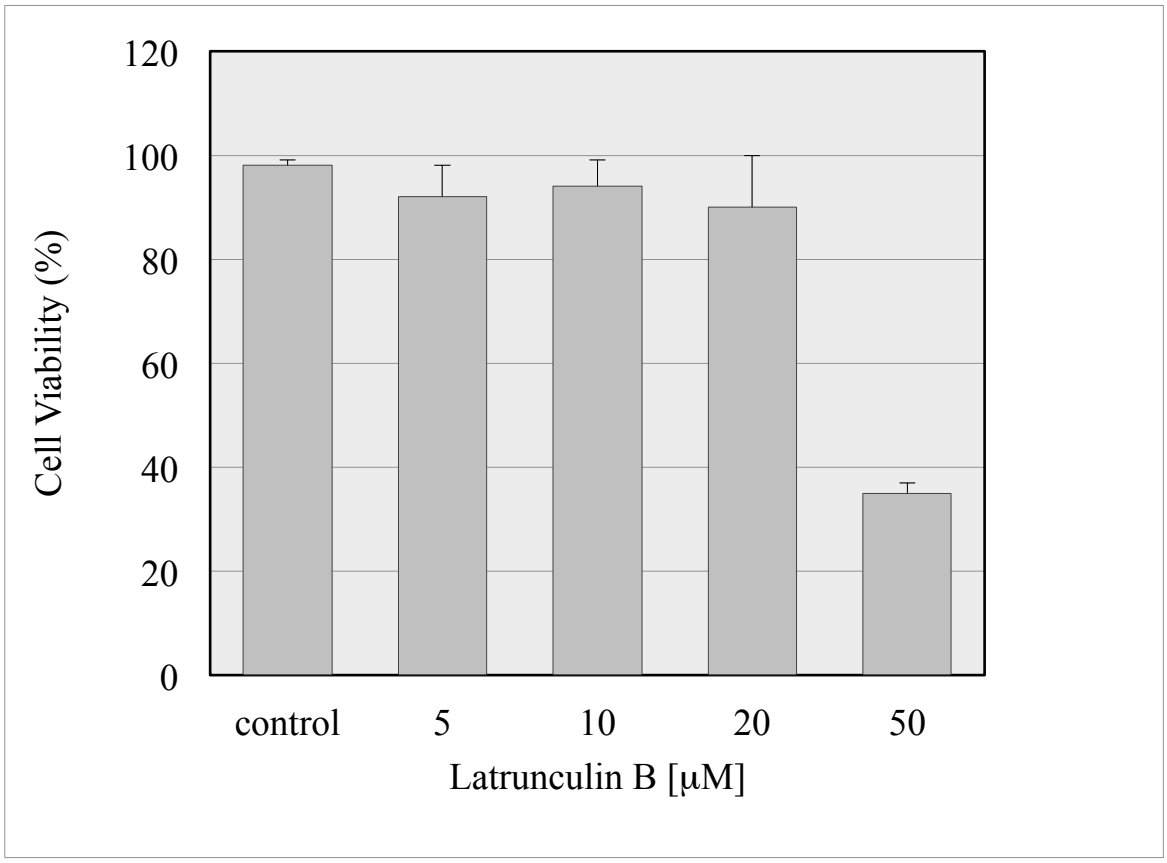
'The role of cytoskeleton networks on lipid-mediated delivery of DNA' by Coppola et al.



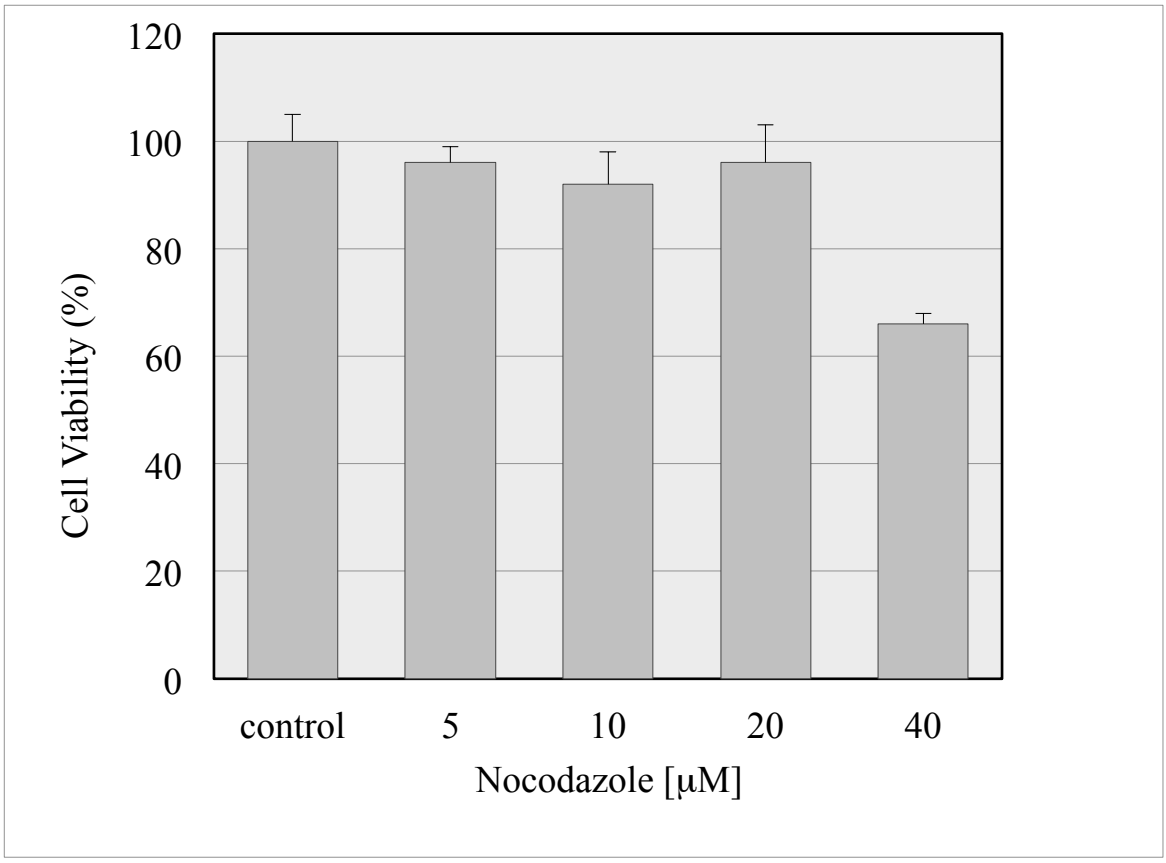


Figure S1. Cell viability of CHO cells as a function of increasing concentrations of inhibitors.

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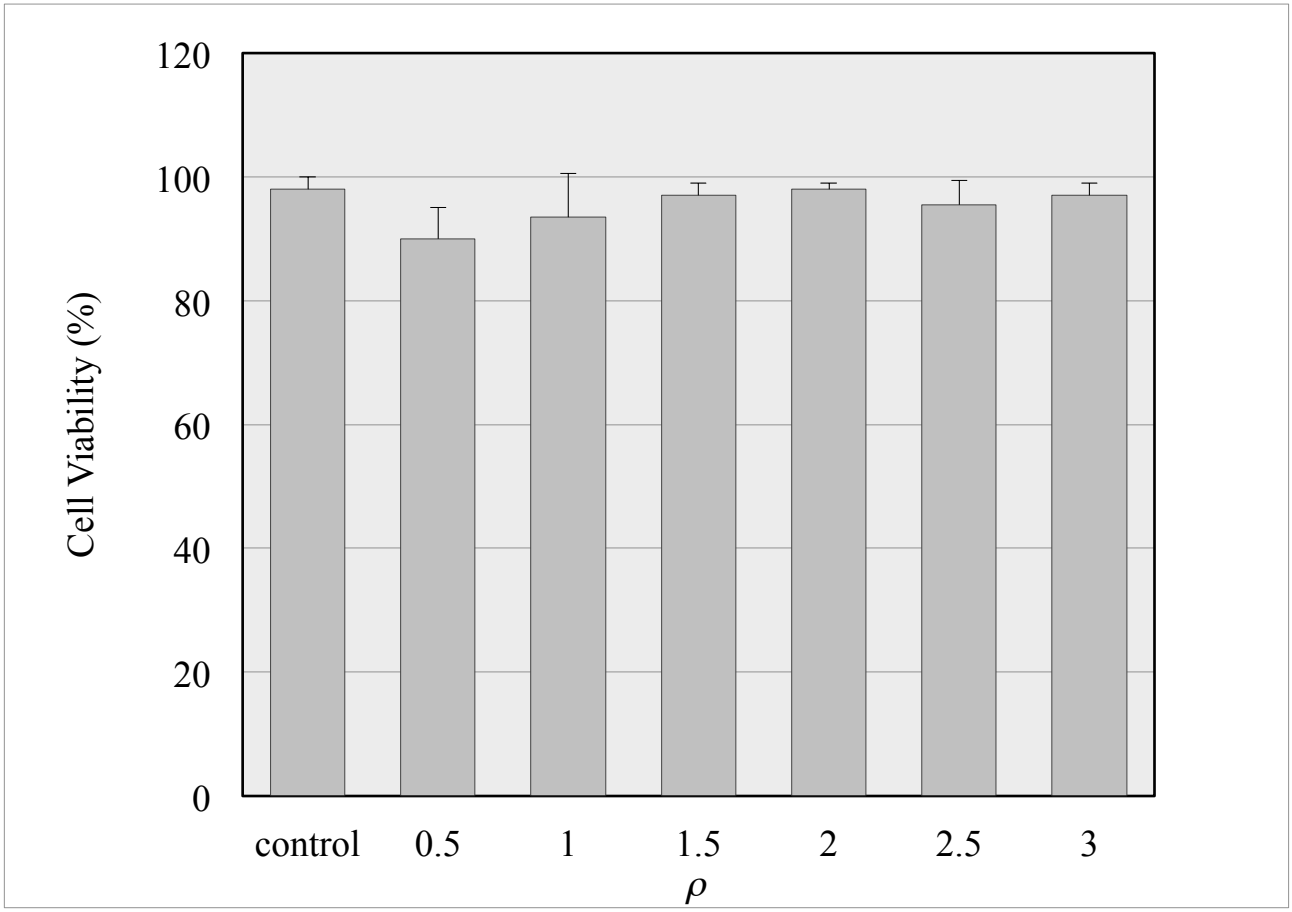

Figure S2. Cell viability of CHO cells as a function of increasing cationic lipid/anionic DNA charge ratio, ρ, in MC lipoplexes.

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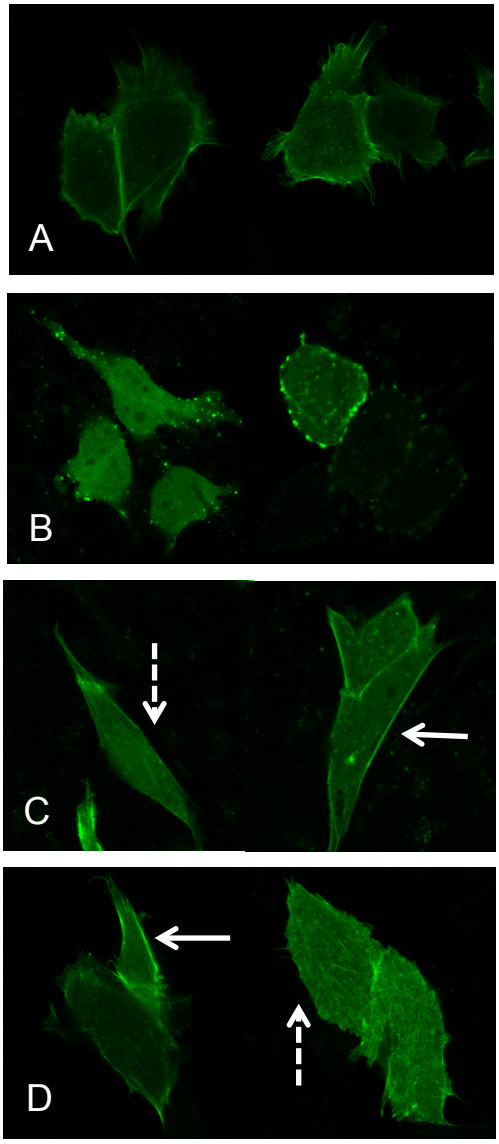
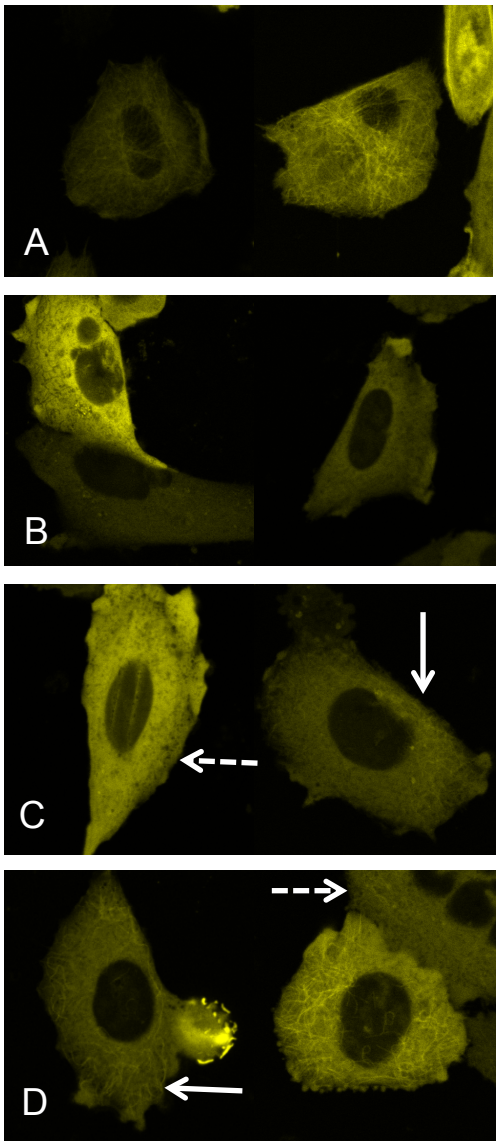
Figure S3

Actin recovery after Latrunculin B treatment



Actin cytoskeleton staining of CHO cells by Actin-Green Fluorescent Protein (panel A). Actin cytoskeleton recovery after Latrunculin B-treatment: t=15 minutes (panel B); t=4h (panel C); t=8h (panel D). Solid lines indicate newly formed microfilaments, while dotted lines indicate regions of the plasma membrane where only poorly formed, if any, actin microfilaments are visible.

Microtubule recovery after Nocodazole treatment



Tubulin cytoskeleton staining of CHO cells by Tubulin-Yellow Fluorescent Protein (panel A). Tubulin cytoskeleton recovery after Nocodazole-treatment: t=15 minutes (panel B); t=4h (panel C); t=8h (panel D). Solid lines indicate newly formed microtubules, while dotted lines indicate regions of the cytoplasm where only poorly formed, if any, microtubules can be seen.

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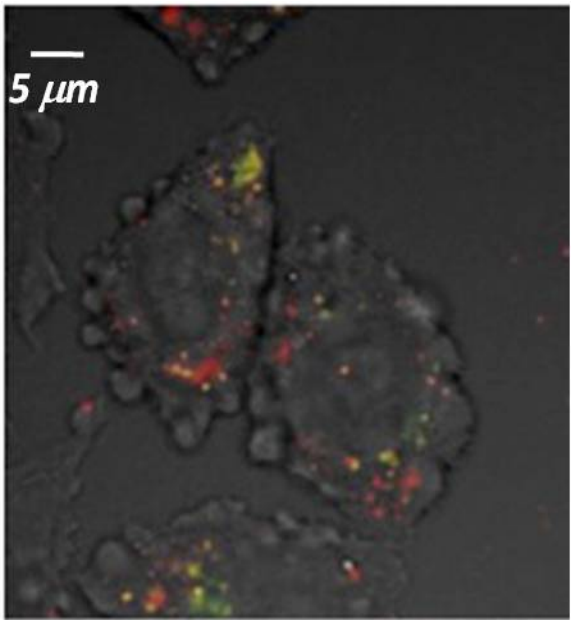

Figure S4. Representative CLSM image of LAT-treated cells more than 3 hours from lipoplex administration. A minor fraction of lipoplexes (red) was found to co-localize with lysosomes (green).

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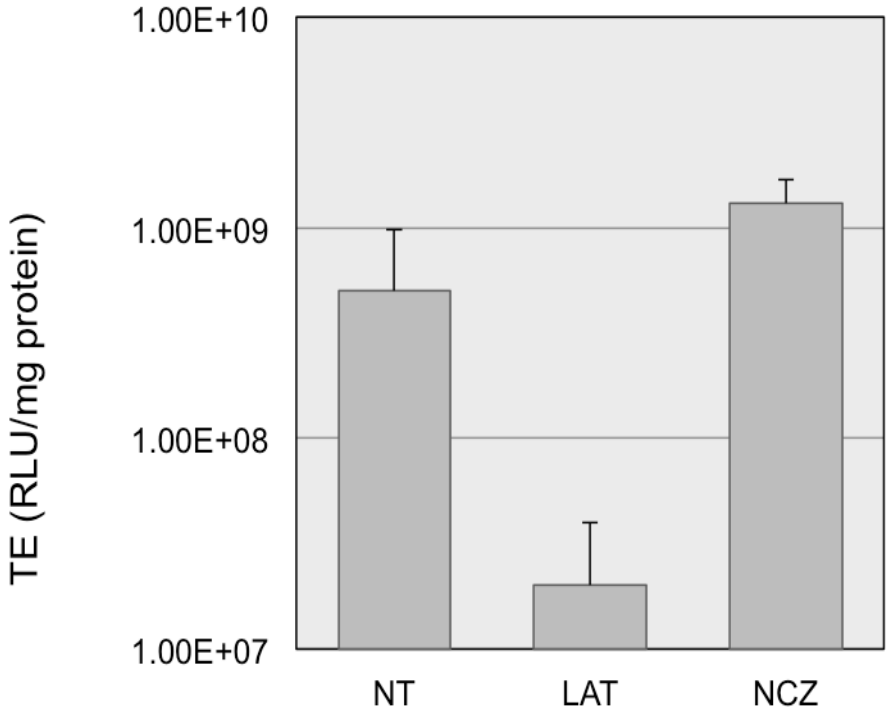

Figure S5. Transfection Efficiency (TE) of multicomponent lipoplexes in untreated (NT) CHO cells and after treatment with Latrunculin B (LAT) and Nocodazole (NCZ). Experiments were performed at the Center for Nanotechnology Innovation @NEST using a Luciferase Assay kit (Promega.Madison, WI) following the manufacturer's instructions.