Supplementary Table 1: FRET-based multiplexing studies

Reference	Fluorophores Used	Methods Used for Imaging	Biological Insights
2-4	CFP/YFP/mRFP1	Experimental: Three-color,	Identification of trimeric
		Ratiometric FRET x2 on	molecular complexes; Analysis of
		same probe	caspase activities
5, 6	CFP/DsRed with	Experimental: Intensity-	Analysis of multiple caspase
	YFP/DsRed	based FRET x2	activities upon TNF α -induced cell
			death
7	CFP/YFP	Experimental: Spatial	Analysis of cAMP signaling in
		separation, Ratiometric	multiple cellular compartments
		FRET x2	simultaneously
8	CFP/YFP	Experimental: Ratiometric	Simultaneous monitoring of PKA
		FRET x2 on same probe	and PKC activities
9	CFP/YFP,	Experimental: Ratiometric	Relative timing relationships of
	mOrange/mCherry, Fura	FRET x2, Spatial separation,	CaMKII and PKC activation and
	Red	Ratiometric FRET x2 +	Annexin A4 assembly in response
		Intensiometric FRET +	to Ca ²⁺ stimulation
		Calcium Sensor	
10	mTFP1/YFP,	Experimental: Ratiometric	Dual caspase-3 sensors monitor
	mAmetrine/tdTomato	FRET x2	delay between cytoplasmic and
			nuclear caspase-3 activity
11	Sirius/mseCFP,	Experimental: Ratiometric	Relative timing of Ca ²⁺ signals and
	Sapphire/DsRed	FRET x2	caspase-3 activity in response to
			ΤΝFα
12	mSapphire/RFP	Experimental: Single	Simultaneous imaging of cGMP
	CFP/YFP	wavelength excitation,	and cAMP, or simultaneous
		Ratiometric FRET x2	imaging of Ca ²⁺ and cAMP in
			contracting cardiomyocytes
13, 14	CFP/YFP, Dyes:	Experimental and	Relative timing and position of
	Mero87/Alexa750 pair	Computational: Ratiometric	Cdc42, Rac1, and RhoA during
	-	FRET + Ratiometric dye	cellular protrusion
		change	-
15	mTagBFP/sREACh,	Experimental: Ratiometric	Simultaneous imaging of cGMP,
	CFP/YFP, Fura Red	FRET $x^2 + Ca^{2+}$	cAMP, and Ca ²⁺ using spectrally
	-		separable probes
16	CFP/YFP,	Experimental: Ratiometric	Simultaneous analysis of the
	mOrange2/mCherry	FRET x2	relative timing of Src and MMP-
	с .		MT1 activities in response to EGF
			stimulation
17	FLIM: CFP/tHcRed and	Experimental: FLIM FRET	Simultaneous analysis of K-ras
	YFP/tHcRed		and H-ras activities in different
			cellular compartments in response
			to EGF stimulation using a
			common binding domain
18	CFP/YFP and FLIM:	Experimental: Ratiometric	Simultaneous imaging of Ca ²⁺
	mTagRFP/mPlum	FRET + FLIM FRET	signals and H-ras activity in
	in ragici 17 mi tum		
	in ragici 17ini funi		response to EGF stimulation
19	FLIM: mRFP/GFP,	Experimental: FLIM FRET	
19	FLIM: mRFP/GFP,	Experimental: FLIM FRET	Analysis of CaMKII activity in
19	FLIM: mRFP/GFP, mStrawberry/GFP,	Experimental: FLIM FRET	Analysis of CaMKII activity in neuronal synapses and evaluation
19	FLIM: mRFP/GFP,	Experimental: FLIM FRET	Analysis of CaMKII activity in

- 1. McNamara, G., Gupta, A., Reynaert, J., Coates, T.D. & Boswell, C. Spectral imaging microscopy web sites and data. *Cytometry A* **69**, 863-71 (2006).
- 2. Galperin, E., Verkhusha, V.V. & Sorkin, A. Three-chromophore FRET microscopy to analyze multiprotein interactions in living cells. *Nat Methods* **1**, 209-17 (2004).
- 3. He, L., Wu, X., Simone, J., Hewgill, D. & Lipsky, P.E. Determination of tumor necrosis factor receptor-associated factor trimerization in living cells by CFP->YFP->mRFP FRET detected by flow cytometry. *Nucleic Acids Res* **33**, e61 (2005).
- 4. Wu, X. et al. Measurement of two caspase activities simultaneously in living cells by a novel dual FRET fluorescent indicator probe. *Cytometry A* **69**, 477-86 (2006).
- 5. Kawai, H. et al. Simultaneous imaging of initiator/effector caspase activity and mitochondrial membrane potential during cell death in living HeLa cells. *Biochim Biophys Acta* **1693**, 101-10 (2004).
- 6. Kawai, H. et al. Simultaneous real-time detection of initiator- and effector-caspase activation by double fluorescence resonance energy transfer analysis. *J Pharmacol Sci* **97**, 361-8 (2005).
- 7. DiPilato, L.M., Cheng, X. & Zhang, J. Fluorescent indicators of cAMP and Epac activation reveal differential dynamics of cAMP signaling within discrete subcellular compartments. *Proc Natl Acad Sci U S A* **101**, 16513-8 (2004).
- 8. Brumbaugh, J., Schleifenbaum, A., Gasch, A., Sattler, M. & Schultz, C. A dual parameter FRET probe for measuring PKC and PKA activity in living cells. *J Am Chem Soc* **128**, 24-5 (2006).
- 9. Piljic, A. & Schultz, C. Simultaneous recording of multiple cellular events by FRET. *ACS Chem Biol* **3**, 156-60 (2008).
- 10. Ai, H.W., Hazelwood, K.L., Davidson, M.W. & Campbell, R.E. Fluorescent protein FRET pairs for ratiometric imaging of dual biosensors. *Nat Methods* **5**, 401-3 (2008).
- 11. Tomosugi, W. et al. An ultramarine fluorescent protein with increased photostability and pH insensitivity. *Nat Methods* **6**, 351-3 (2009).
- 12. Niino, Y., Hotta, K. & Oka, K. Simultaneous live cell imaging using dual FRET sensors with a single excitation light. *PLoS One* **4**, e6036 (2009).
- 13. Tsukada, Y. et al. Quantification of local morphodynamics and local GTPase activity by edge evolution tracking. *PLoS Comput Biol* **4**, e1000223 (2008).
- 14. Machacek, M. et al. Coordination of Rho GTPase activities during cell protrusion. *Nature* **461**, 99-103 (2009).
- 15. Niino, Y., Hotta, K. & Oka, K. Blue fluorescent cGMP sensor for multiparameter fluorescence imaging. *PLoS One* **5**, e9164 (2010).
- 16. Ouyang, M. et al. Simultaneous visualization of protumorigenic Src and MT1-MMP activities with fluorescence resonance energy transfer. *Cancer Res* **70**, 2204-12 (2010).
- 17. Peyker, A., Rocks, O. & Bastiaens, P.I. Imaging activation of two Ras isoforms simultaneously in a single cell. *Chembiochem* **6**, 78-85 (2005).
- 18. Grant, D.M. et al. Multiplexed FRET to image multiple signaling events in live cells. *Biophys J* **95**, L69-71 (2008).
- 19. Kwok, S. et al. Genetically encoded probe for fluorescence lifetime imaging of CaMKII activity. *Biochem Biophys Res Commun* **369**, 519-25 (2008).