

## SUPPLEMENTARY TABLE

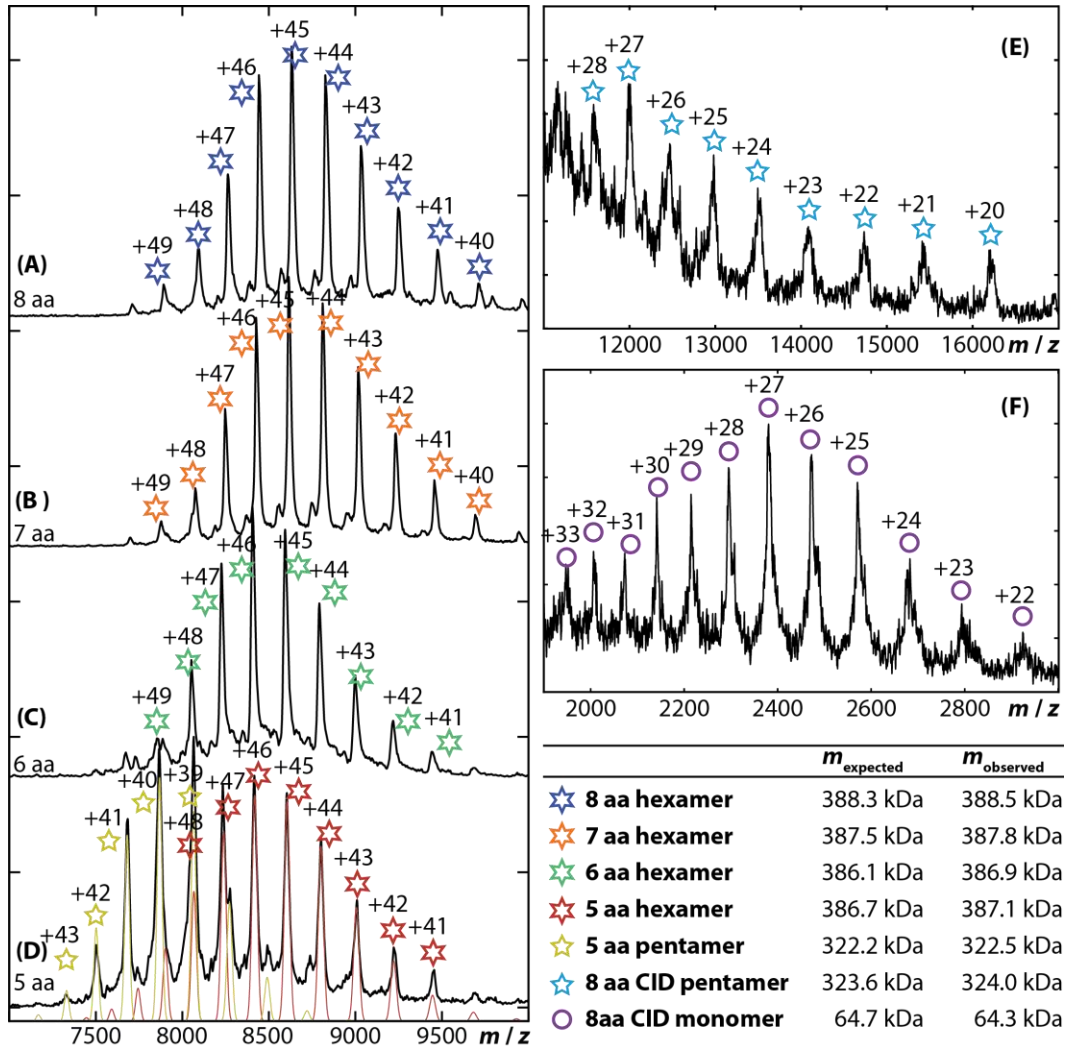
**Supplementary Table S1. Pairwise comparisons of the N2D-vs-CTD orientation in *V. cholerae* GspE<sup>EpsE</sup>, AaPiIT and PaPiIT subunits. Related to Figure 2 and 4**

		VcGspE				AaPiIT			PaPiIT				
		qC <sub>6</sub>	C <sub>2</sub>			C <sub>6</sub>	qC <sub>2</sub>			C <sub>2</sub>			
			A	B	C		D	E	F	A	B	C	
Vc GspE	qC <sub>6</sub>		<b>34.0</b>	<b>16.0</b>	<b>40.9</b>	<b>40.2</b>	<b>43.0</b>	<b>43.0</b>	<b>42.4</b>	<b>35.3</b>	<b>24.6</b>	<b>23.1</b>	
	C <sub>2</sub>	A	<b>34.0</b>		<b>31.5</b>	<b>48.4</b>	<b>26.0</b>	<b>38.8</b>	<b>29.4</b>	<b>35.6</b>	<b>20.7</b>	<b>21.5</b>	<b>18.8</b>
		B	<b>16.0</b>	<b>31.5</b>		<b>46.7</b>	<b>37.9</b>	<b>47.2</b>	<b>35.2</b>	<b>44.4</b>	<b>37.7</b>	<b>26.8</b>	<b>19.2</b>
		C	<b>40.9</b>	<b>48.4</b>	<b>46.7</b>		<b>27.6</b>	<b>14.7</b>	<b>73.4</b>	<b>17.2</b>	<b>30.0</b>	<b>30.2</b>	<b>38.0</b>
Aa PiIT	C <sub>6</sub>		<b>40.2</b>	<b>26.0</b>	<b>37.9</b>	<b>27.6</b>		<b>16.8</b>	<b>51.8</b>	<b>13.6</b>	<b>11.8</b>	<b>15.3</b>	<b>19.9</b>
	qC <sub>2</sub>	D	<b>43.0</b>	<b>38.8</b>	<b>47.2</b>	<b>14.7</b>	<b>16.8</b>		<b>66.4</b>	<b>3.9</b>	<b>20.3</b>	<b>23.7</b>	<b>31.9</b>
		E	<b>43.0</b>	<b>29.4</b>	<b>35.2</b>	<b>73.4</b>	<b>51.8</b>	<b>66.4</b>		<b>63.2</b>	<b>48.3</b>	<b>44.0</b>	<b>36.1</b>
		F	<b>42.4</b>	<b>35.6</b>	<b>44.4</b>	<b>17.2</b>	<b>13.6</b>	<b>3.9</b>	<b>63.2</b>		<b>16.5</b>	<b>20.9</b>	<b>29.1</b>
Pa PiIT	C <sub>2</sub>	A	<b>35.3</b>	<b>20.7</b>	<b>37.7</b>	<b>30.0</b>	<b>11.8</b>	<b>20.3</b>	<b>48.3</b>	<b>16.5</b>		<b>11.0</b>	<b>17.7</b>
		B	<b>24.6</b>	<b>21.5</b>	<b>26.8</b>	<b>30.2</b>	<b>15.3</b>	<b>23.7</b>	<b>44.0</b>	<b>20.9</b>	<b>11.0</b>		<b>9.2</b>
		C	<b>23.1</b>	<b>18.8</b>	<b>19.2</b>	<b>38.0</b>	<b>19.9</b>	<b>31.9</b>	<b>36.1</b>	<b>29.1</b>	<b>17.7</b>	<b>9.2</b>	

Each pairwise comparison of two subunits is based on a superposition of two CTD domains. The resultant superposition operation is applied to the entire subunit. Subsequently the two N2D domains are superimposed. The rotation angle of this second superposition is the difference in N2D-vs-CTD orientation given in the Table.

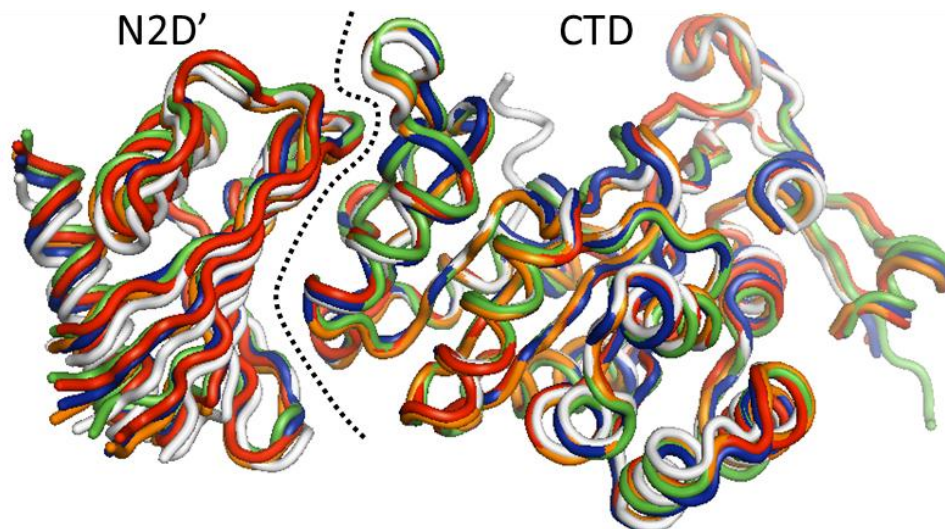
AaPiIT with C<sub>6</sub> hexamer symmetry (PDB: 2EWV (Satyshur et al., 2007)); AaPiIT with quasi-C<sub>2</sub> (qC<sub>2</sub>) hexamer symmetry (PDB: 2GSZ (Satyshur et al., 2007) ); PaPiIT with C<sub>2</sub> hexamer symmetry (PDB: 3JVV (Misic et al., 2010)).

## SUPPLEMENTARY FIGURES



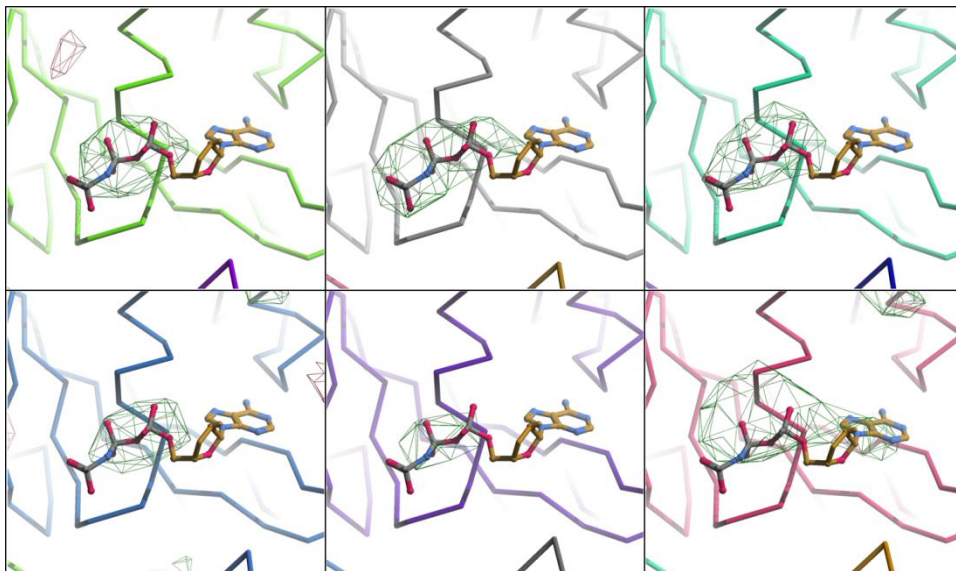
**Figure S1. Oligomerization states of  $\Delta N^1$ GspE<sup>EpsE</sup>-5aa-Hcp1 and  $\Delta N^1$ GspE<sup>EpsE</sup>-7aa-Hcp1 from *V. cholerae* Related to Figure 1**

**Native mass spectra of  $\Delta N^1$ GspE<sup>EpsE</sup>-8aa-Hcp1 (A),  $\Delta N^1$ GspE<sup>EpsE</sup>-7aa-Hcp1 (B), and  $\Delta N^1$ GspE<sup>EpsE</sup>-6aa-Hcp1 (C).** The data show that these proteins can each assemble as hexamers in solution, but that  $\Delta N^1$ GspE<sup>EpsE</sup>-5aa-Hcp1 (D) forms pentameric and hexameric complexes. Tandem mass spectra were also performed in which all ions greater than  $\sim 7500 m/z$  were isolated in the gas phase and subsequently fragmented using collision-induced dissociation. The appearance of pentamer and monomer product ions during all gas-phase CID experiments supports the assignment of hexamer precursor ions in A-D. Note that the loss of peptide ions from the precursor ions was also observed, but that fragmentation channel was less structurally informative. The measured and expected masses for all ions are reported in the table.



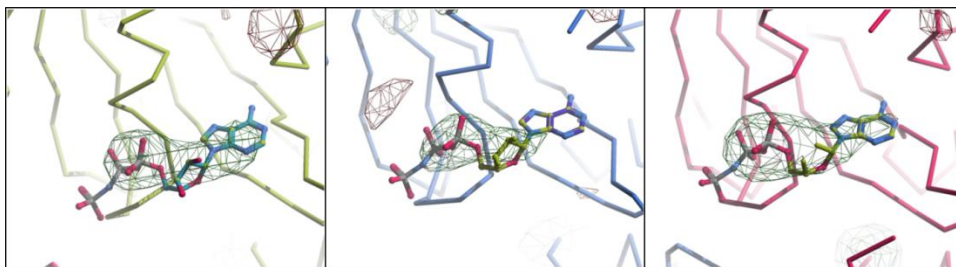
**Figure S2. The CTD•N2D' construction unit in  $\text{GspE}^{\text{EpsE}}$  hexamers and the helical  $\text{GspE}^{\text{EpsE}}$  structure. Related to Figure 2**

Superposition of the CTDs shows that the CTD•N2D' construction units are essentially the same for all *V. cholerae*  $\text{GspE}^{\text{EpsE}}$  structures. Depicted are: one CTD•N2D' unit from the  $C_6$  hexamer (orange), the three independent units from the  $C_2$  hexamer (colored blue, red and green as in Figure 2A), and the CTD•N2D' unit from the helical  $\Delta^{90}\text{GspE}^{\text{EpsE}}$  structure (grey; PDB: 1P9W (Robien et al., 2003)). The dashed line indicates the separation between the N2D' and CTD domains.

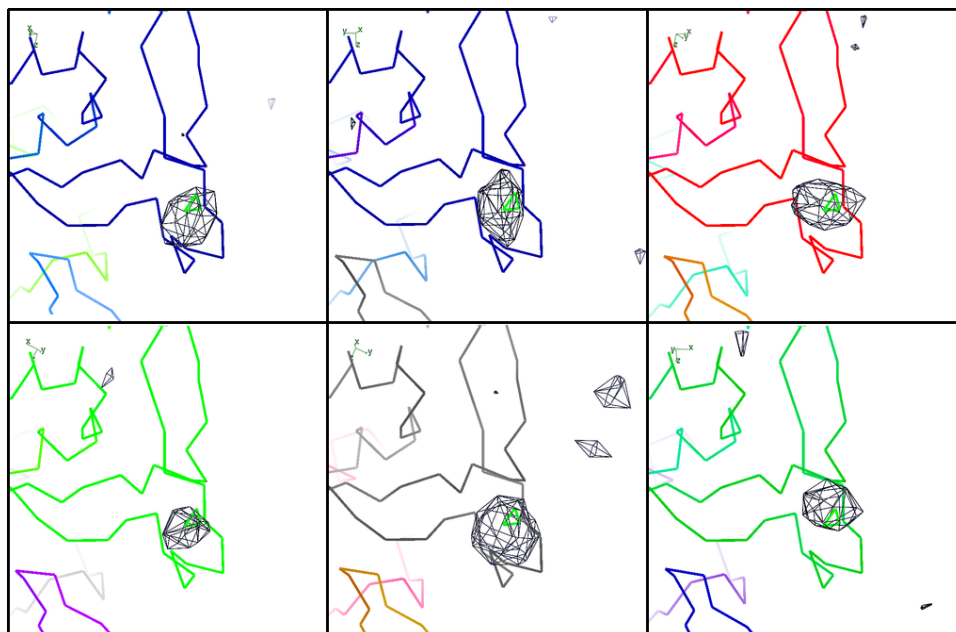


**Figure S3. Electron densities of nucleotides in *V. cholerae*  $\Delta N1$ GspE<sup>EpsE</sup>-Hcp1 fusion structures.** Related to Figure 2 (Fobs-Fcalc) difference electron densities at the 3 sigma level are shown. The phases were obtained without including nucleotide coordinates.

**Figure S3A. Electron densities at the nucleotide position in the six subunits of  $\Delta N1$ GspE<sup>EpsE</sup>-6aa-Hcp1.** The nucleotides present in the protein solution were ADP and AMPPNP. Shown in each panel are: (i) the C $\alpha$  traces of the six subunits in the asymmetric unit (in six different colors); (ii) AMPPNP coordinates from the  $\Delta 90$ GspE<sup>EpsE</sup> structure (PDB: 1P9W (Robien et al., 2003)) after superimposing the CTD of that structure onto the CTD of each of the six crystallographically independent subunits of the  $\Delta N1$ GspE<sup>EpsE</sup>-6aa-Hcp1 structure.

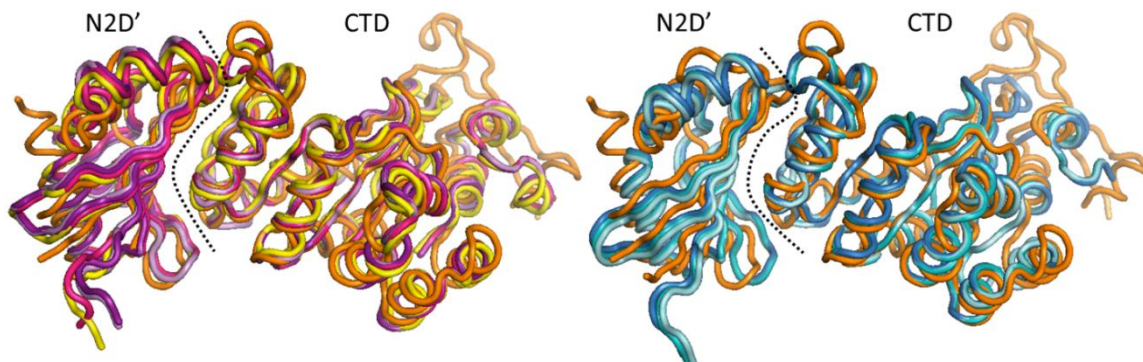


**Figure S3B. Electron densities at the nucleotide position in the  $\Delta N1$ GspE<sup>EpsE</sup>-8aa-Hcp1 structure.** The nucleotide present in the protein solution was ADP, AlCl<sub>3</sub>, and NaF. Shown in each panel are: (i) the C $\alpha$  traces of the three subunits in the asymmetric unit in three different colors; (ii) AMPPNP coordinates from the *V. cholerae*  $\Delta 90$ GspE<sup>EpsE</sup> structure (PDB: 1P9W (Robien et al., 2003)) after superimposing the CTD of that structure onto the CTD of each of the three crystallographically independent subunits from the  $\Delta N1$ GspE<sup>EpsE</sup>-8aa-Hcp1 structure.



**Figure S4. Anomalous electron difference densities of zinc sites in *V. cholerae*  $\Delta N1$ GspE<sup>EpsE</sup>-6aa-Hcp1. Related to Figure 2**

The peak heights at the zinc positions (as located in the  $\Delta 90$ GspE<sup>EpsE</sup> structure (PDB: 1P9W; (Robien et al., 2003)) in the six independent crystallographic subunits of the  $\Delta N1$ GspE<sup>EpsE</sup>-6aa-Hcp1 hexamer are: 4.4, 4.5, 4.5, 4.8, 5.3 and 6.7 sigma.



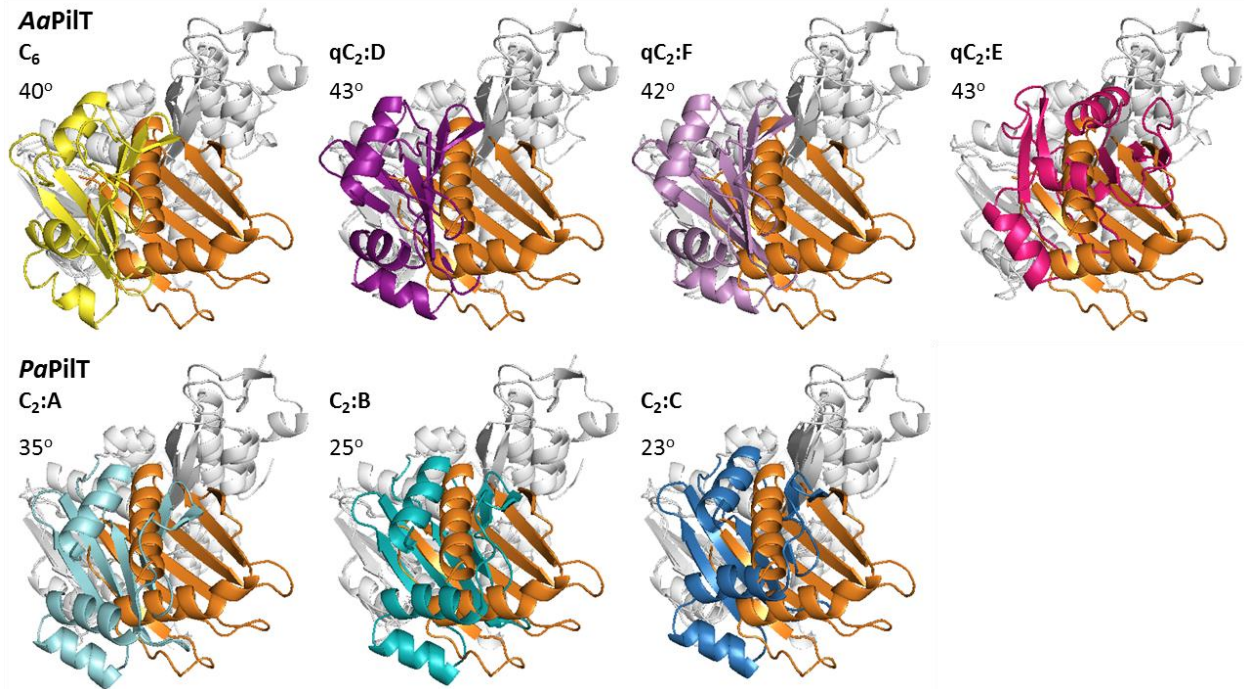
**Figure S5. The CTD•N2D' construction units in GspE<sup>EpsE</sup> and PilT hexamers.**

**Related to Figure 4**

The CTD•N2D' construction units of the T2SS GspE<sup>EpsE</sup> ATPase and of two T4PS PilT ATPases superimpose remarkably well. The dotted line indicates the separation of the N2D' and the CTD.

**Left:** superposition of the CTDs of the *AaPilT* C<sub>6</sub> hexamer (yellow;(Satyshur et al., 2007)) and *AaPilT* quasi C<sub>2</sub> hexamer (different shades of purple) onto subunit E of the GspE<sup>EpsE</sup> C<sub>6</sub> hexamer (orange).

**Right:** alignment of the CTDs of *PaPilT* (different shades of blue;(Misic et al., 2010)) to subunit E of the GspE<sup>EpsE</sup> C<sub>6</sub> hexamer (orange).

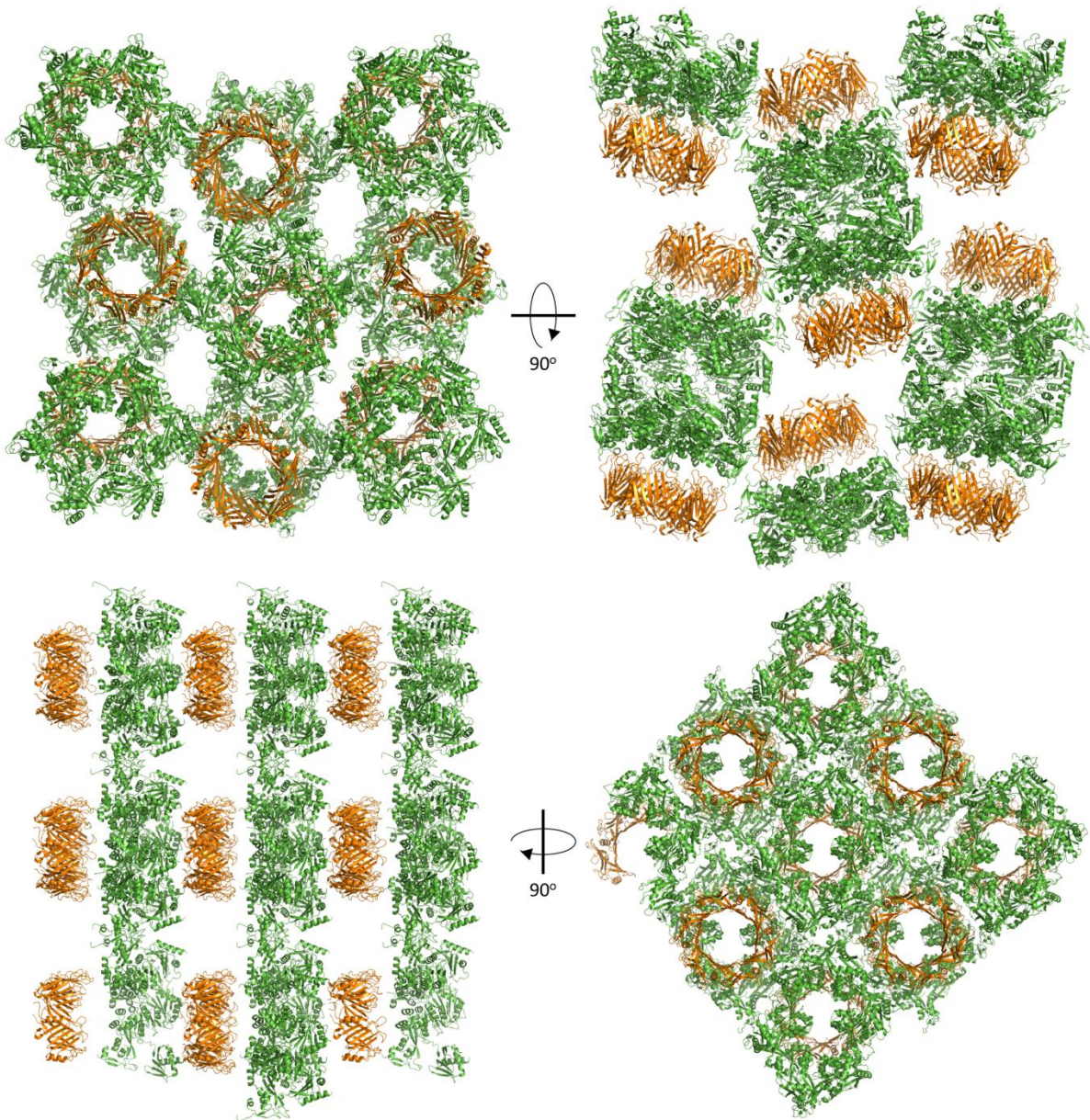


**Figure S6. The variability in N2D-vs-CTD orientations in GspE<sup>EpsE</sup> and PilT hexamers. Related to Figure4**

**Differences in orientation of the N2D-vs-CTD orientations in T2SS and T4PS ATPases, viewed with the N2D on top and the CTD below.** This direction of view is approximately orthogonal to the more canonical views of Figures 2B and 4B. The superimposed CTDs of each pair of subunits are colored grey. Subunit E of the GspE<sup>EpsE</sup> qC<sub>6</sub> hexamer (orange N2D domain) functions as reference in all figures. The difference in N2D orientation is shown in degrees in the left upper corner of each pair. Note that none of the other subunits have the “orange” N2D-vs-CTD orientation.

**Top:** superposition of the CTDs of AαPilT C<sub>6</sub> hexamer (yellow; PDB: 2EWV) and qC<sub>2</sub> hexamer (different shades of purple; PDB: 2GSZ) to subunit E of the GspE<sup>EpsE</sup> qC<sub>6</sub> hexamer (orange).

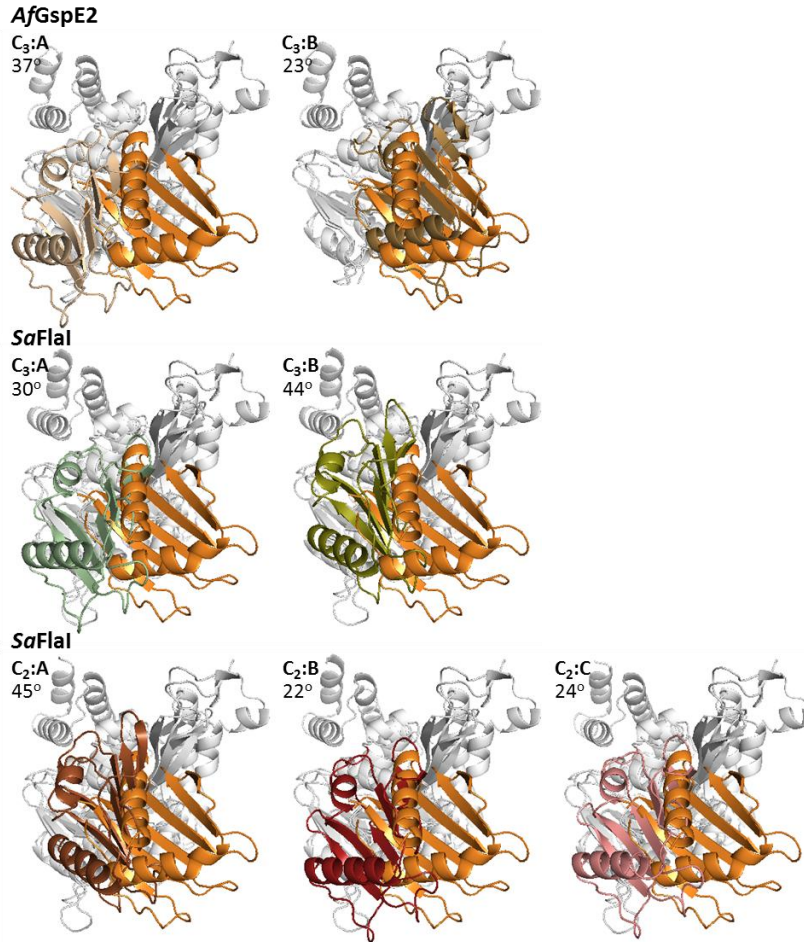
**Bottom:** superposition of the CTDs of PaPilT C<sub>2</sub> hexamer (different shades of blue; PDB: 3JVJ) to subunit E of the GspE<sup>EpsE</sup> C<sub>6</sub> hexamer (orange).



**Figure S7. Crystal packing of GspE<sup>EpsE</sup>-Hcp1 hexamers. Related to Figure1**

Crystal packing of  $\Delta^{N1}$ GspE<sup>EpsE</sup>-6aa-Hcp1 (top) and  $\Delta^{N1}$ GspE<sup>EpsE</sup>-8aa-Hcp1 (bottom) with  $\Delta^{N1}$ GspE<sup>EpsE</sup> in green and Hcp1 in orange. In the case of  $\Delta^{N1}$ GspE<sup>EpsE</sup>-8aa-Hcp1 alternating layers of Hcp1 and  $\Delta^{N1}$ GspE<sup>EpsE</sup> are clearly present.





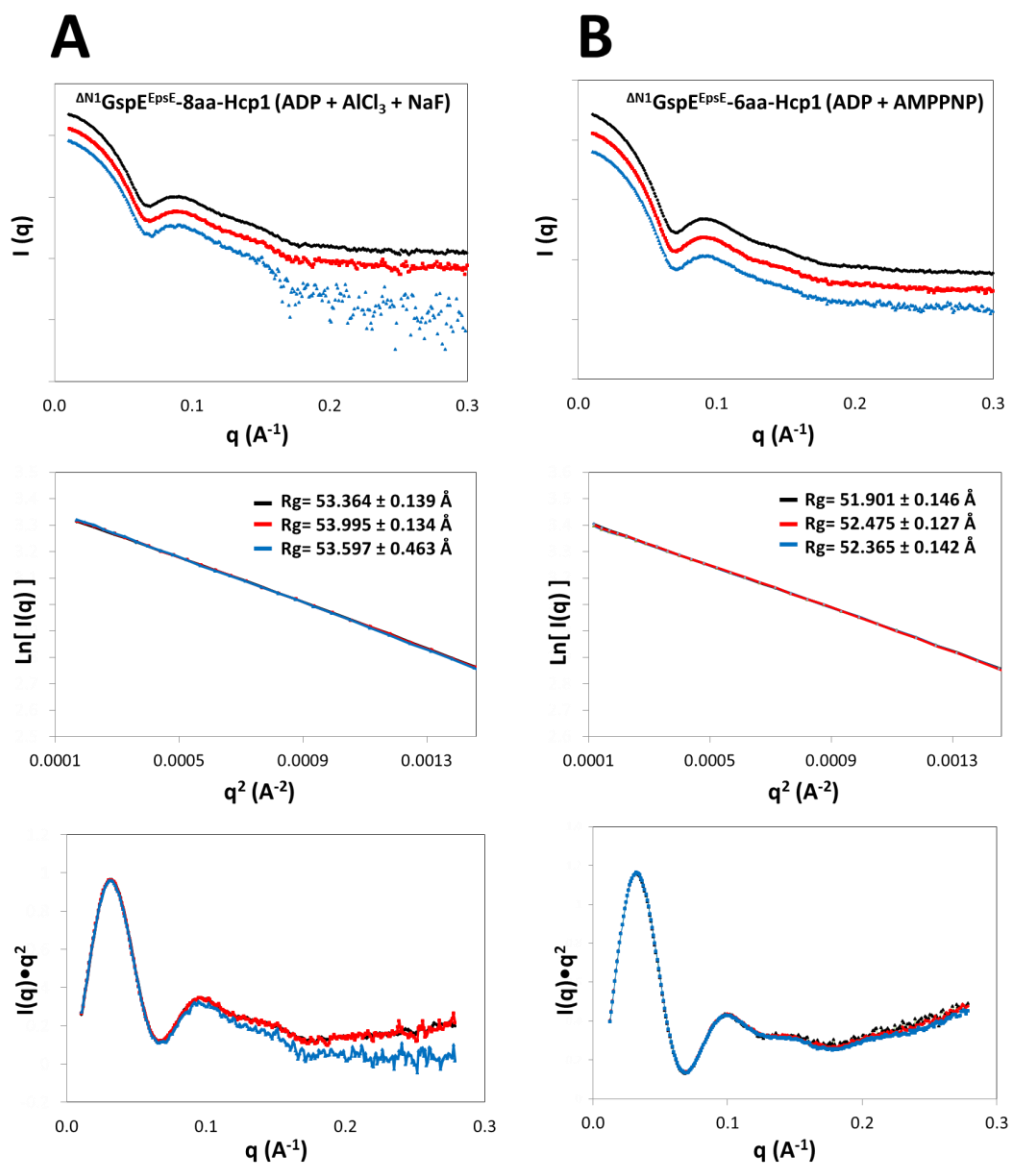
**Figure S8. The variability of the N2D-vs-CTD orientations in *V. cholerae* GspE<sup>EpsE</sup> and archaeal ATPases from *Archaeoglobus fulgidus* (AfGspE2) and from *Sulfolobus acidocaldarius* (SaFlal). Related to Figure 4**

Differences in orientation of the N2D-vs-CTD orientations in T2SS and archaeal non-T2SS ATPases, viewed with the N2D on top and the CTD below. The percentage of sequence identity per domain is given in Figure 1A. This direction of view is approximately orthogonal to the more canonical views of Figures 2B and 4B. The superimposed CTDs of each pair of subunits are colored grey. Subunit E of the GspE<sup>EpsE</sup> qC<sub>6</sub> hexamer (orange N2D domain) functions as reference in all figures. The difference in N2D orientation is shown in degrees in the left upper corner of each pair. ((Reindl et al., 2013; Yamagata and Tainer, 2007)).

**Top:** superposition of the CTDs of AfGspE2 C<sub>3</sub> hexamer (different shades of brown; PDB: 2OAP (Yamagata and Tainer, 2007)) to subunit E of the GspE<sup>EpsE</sup> qC<sub>6</sub> hexamer (orange).

**Middle:** superposition of the CTDs of SaFlal C<sub>3</sub> hexamer (different shades of green; PDB: 4I17 (Reindl et al., 2013)) to subunit E of the GspE<sup>EpsE</sup> qC<sub>6</sub> hexamer (orange).

**Bottom:** superposition of the CTDs of SaFlal C<sub>2</sub> hexamer (different shades of red; PDB: 4I17 ) to subunit E of the GspE<sup>EpsE</sup> qC<sub>6</sub> hexamer (orange).



**Figure S9. SAXS studies on  $\Delta N1$ GspE<sup>EpsE</sup>-6aa-Hcp1 and  $\Delta N1$ GspE<sup>EpsE</sup>-8aa-Hcp1. Related to Figure 3**

The SAXS data were measured at three protein concentrations for each sample: 4.8 (black), 2.4 (red) and 1.37 (blue) mg/mL at 20 °C. **Top:** The raw SAXS data. **Middle:** the linearity of the Guinier region *versus* Rg (for  $q \cdot R_g < 1.3$ ). **Bottom:** the Kratky plots (bottom). No significant concentration dependent effects were observed.

## REFERENCES

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