## A. Supplementary materials

### A.1. The algorithm of Anchap

Algorithm 1 Stage I: first scan for sharing from unphased genotypes

1: {Input: multi-locus genotypes for individuals in the cohort}

- 2: for all individuals *i* in the cohort do
- 3:  $i.g \leftarrow i^{th}$  genotype across all markers
- 4: for all individuals j in the cohort, such that  $i \neq j$  do
- 5:  $j.g \Leftarrow j^{th}$  genotype across all markers
- 6: between *i.g* and *j.g* find regions (start and end) without opposing homozygotes longer than the IBD threshold
- 7: end for
- 8: end for
- 9: {Output: list of genomic regions and pairs of individuals putatively sharing IBD}

Algorithm 2 Stage II: alignment of haplotypes

	Source = Stage II. angument of naprojpes
1:	{Input: multi-locus genotypes for individuals in the cohort, list of genomic regions and
	pairs of individuals putatively sharing IBD}
2:	for all individuals $i$ in the cohort <b>do</b>
3:	sort the sequences shared with $i$ by the number of markers they cover, descending
4:	for all $s$ , shared sequences of $i$ do
5:	$i.hap.pat.s \Leftarrow$ current version of $i^{th}$ paternal haplotype, in the region of sharing s
6:	$i.hap.mat.s \Leftarrow$ current version of $i^{th}$ maternal haplotype, in the region of sharing $s$
7:	$s.g \Leftarrow$ the genotype of the individual sharing with <i>i</i> in region <i>s</i>
8:	if $s.g$ is matching <i>i.hap.pat.s</i> and <i>i.hap.mat.s</i> and no other sequences have been
	seen in the region before <b>then</b>
9:	s.g shares <i>i</i> 's paternal haplotype (arbitrarily)
10:	else if $s.g$ is matching $i.hap.pat.s$ then
11:	s.g shares $i$ 's paternal haplotype
12:	else if $s.g$ is matching $i.hap.mat.s$ then
13:	s.g shares <i>i</i> 's maternal haplotype
14:	else
15:	reject $s.g$
16:	end if
17:	use $s.g$ to recover the relevant haplotype
18:	end for
19:	end for
20:	{Output: revised list of genomic regions and pairs of individuals putatively sharing IBD,

with each region assigned to individuals' haplotypes, genotype phasing in IBD regions}

- 1: {Input: genotypes phased in IBD regions}
- 2: for all individuals i in the cohort do
- 3:  $i.hap.pat \leftarrow i^{th}$  paternal haplotype, from round 1
- 4:  $i.hap.mat \leftarrow i^{th}$  maternal haplotype, from round 1
- 5: for all individuals j in the cohort, such that  $i \neq j$  do
- 6:  $j.hap.pat \leftarrow j^{th}$  paternal haplotype, from round 1
- 7:  $j.hap.mat \leftarrow j^{th}$  maternal haplotype, from round 1
- 8: between haplotypes of i and j, find continuously matching regions longer than the IBD threshold AND with at least this many markers where alleles on both haplotypes are fully known
- 9: end for
- 10: **end for**
- 11: {Output: revised list of genomic regions and pairs of individuals putatively sharing IBD, with each region assigned to individuals' haplotypes, genotype phasing in IBD regions}

Algorithm 4 Choosing an optimal subset of individuals for re-sequencing

- 1: {Input: revised list of genomic regions and pairs of individuals putatively sharing IBD}
- 2: cov.mat ⇐ a matrix of size number of haplotypes (2x number of individuals) by number of markers, to store whether, with individuals picked so far, each locus of a haplotype has an individual sequenced who shares this haplotype at this region; initialized to 0 everywhere
- 3: picked.inds ⇐ and empty list of individuals to be picked for the study in the preferred order
- 4: while not chosen the desired number of individuals do
- 5: for all individuals i in the cohort, s.t. i not in *picked.inds* do

6: 
$$i.cov \Leftarrow 0$$

- 7: for all individuals j in the cohort, s.t. j not in *picked.inds* do
- 8:  $i.cov.j \leftarrow \text{total length of shared haplotype segments between } i \text{ and } j$ , such that cov.mat in the corresponding regions is 0

9: 
$$i.cov \leftarrow i.cov + i.cov.j$$

### 10: **end for**

- 11: **end for**
- 12: for sequencing choose next individual i, s.t.  $i = max_i i.cov$  and i not in *picked.inds*
- 13: add i to *picked.inds*
- 14: for all haplotypes hj in the cohort do
- 15: for all IBD regions r between i and hj do

```
16: cov.mat.hj.r \leftarrow 1
```

- 17: **end for**
- 18: **end for**
- 19: end while
- 20: {Ouput: preferred sequencing order}

# A.2. Example of ANCHAP



physical position on chromosome 2 [bp]

Fig. 1.—: Example of IBD detection (Stage I), alignment of IBD regions (Stage II) and phasing (Stage III) of one individual from ORCADES, chromosome 2. First we find his/her haplotype sharers across the genome, and mark the regions of putative IBD sharing as segments. The shared sequences are aligned into two groups, and marked red, and blue accordingly. The grey segments denote misaligned shared sequences. Individual 697 is a full sibling of the proband, with almost the entire chromosome shared and more distant relatives share smaller blocks.

#### A.3. Data pre-processing

The data sets were pre-processed in PLINK (Purcell et al. 2007) to eliminate low quality markers. We removed markers with call rate of less than 95%, out of Hardy-Weinberg equilibrium (p < 0.001), or those with minor allele frequency lower than 1%. We excluded individuals with more than 7% genotype markers missing, and retained only the autosomal SNPs. After pre-processing, the following numbers of samples remained: ORCADES (749 individuals, 302,379 SNPs on 22 chromosomes), CROATIA-KORCULA (945 individuals; 317,223 autosomal SNPs, including 295,574 ORCADES SNPs), CROATIA-VIS (991 individuals; 301,069 autosomal SNPs, including 291,857 ORCADES SNPs), SOCCS (958 individuals; 306,204 autosomal SNPs, including 294,703 ORCADES SNPs). We localized the SNPs on the HapMap genetic map of recombination rates (Consortium 2007).

# A.4. Regions of increased frequency of IBD



Fig. 2.—: IBD peaks on chromosomes 6 and 11, before a genetic map was used to account for extensive LD amoung the isolate founders. The peak on chromosome 6 was reduced and the one on chromosome 11 almost compeltely removed, when we used the HapMap genetic map.

Table 3 shows genetic positions of the peaks of IBD, which were marked at the horizontal axes in Figure 2 of the main article.

chromosome	position	position
	left [kb]	right [kb]
2	134144	138947
3	15484	24365
6	27145	33161
8	95306	97626
10	100639	119196
14	77965	88690
19	18379	34464

(a) peaks in IBD density for ORCADES(build 36)

chromosome	position	position
	left [kb]	right [kb]
1	185353	189270
2	47210	59831
6	25952	33936
9	78602	81335
9	99130	104709

(b) peaks in IBD density for CROATIA-VIS(build 35)

chromosome	position	position				
	left [kb]	right [kb]				
1	90094	101013				
1	167719	177243	C	chromosome	position	position
2	54456	63368			left [kb]	right [kb]
12	77079	90001		2	134028	139092
18	64446	66196		6	25535	33096

(c) peaks in IBD density for CROATIA-KORCULA (build 36)

(d) peaks in IBD density for SOCCS (build 36)

Fig. 3.—: Locations of IBD peaks in four cohorts under study

Figure 4 shows genes present in the two top peaks on chromosomes 2 and 6.

			4.80 Mb		Forward strand
	134.50 Mb	135.50 Mb	136.50 Mb	137.50 Mb	138.50 Mb
Chromosome bands	q21.2	q21.3		q22.1	
Ensembl/Havana	LMGAT2	LTMEM163 LACMSD LCCNT2 LYSK4 LZRANB3	LaC011999.9 LAC064850.4 R3HDM1 LOARS LOEXD2 LCT LCT LCT	L <sub>THSD7B</sub>	LHNMT LACO
	134.50 Mb	135.50 Mb	136.50 Mb	137.50 Mb	138.50 Mb
	Reverse strand		4.80 Mb		
	There are currently 68 tra	cks turned off.			
	Arabica EnaEMDI Hama	continue version E4 26n (NCB126) Chrom	accesso 2, 124 144 000 129 047 000		

(a) peak on chromosome 2, between 134144 and 138947 kb

			_		6.02 Mb		Forward strand
		28.00 Mb		29.00 Mb	30.00 Mb	31.00 Mb	32.00 Mb 33.00 I
Chromosome bands		p22.1	-			p21.33	p21.32
Assembly excepti							an a
Ensembl/Havana	1.1				•••••	· · · · · · · · · · · · · · · · · · ·	
	LAL021808.	2-1 <sup>L</sup> AL121944.14-1		L662890.3-1	LAL645939.6-3	LAL662873.3 LPS	ORS1C3 LAL645922.11-1 LAL665
	LZNF204	LOR2W6P LZ9	3745.1-1	LAL645937.	6-1 L645929.4	LAL662797.7	LHCP5 LAL645922.11-4 LHLA-7
	LZNF391	LAL121944.14-2		LAL6459	37.6-2 LHCG9	LHLA-N DPCR1	LAL663061.8-1 BTNL2
	LAL03111	8.22 <sup>L</sup> AL022393	4	LAL672	167.6 <sup>L</sup> C6orf12	LTMPOP1	G27 LST1 LAL645922.11-2 LALA-DO
	LVN1R11P	LZNF187	LRPS	AP2 LAL662	781.4 <sup>L</sup> HLA-J	LHCG19P DDR1	AL671883.3-1 L645922.11-3 LAP2
	<sup>L</sup> RP11-209A2.1	LRPLP2P1 LOF	2E1P	ZNF463P	AL645927.3	ILA-L <sup>L</sup> PTMAP1 <sup>L</sup> POL	R2LP AIF1 C2 AL662884.11 PSMB8
	LXXba	c-BPG3418.2	LNC	L5BP	LAL645936.5	AL662795.5 SFTA2	RPL3P2 LSTK19P AL662796.6-1
	LXXE	ac-BPG34I8.3	PX6	LKRT18P1	<sup>L</sup> AL645939.6-1	<sup>L</sup> RPL7P4	WASF5P MSH5 PPT2 AL845554.2
	HIST1H4I LR	P1-15D7.1 PGBD1		LOR2AD1P	L645939.6-4	GNL1 VARSL	XXbac-BPG248L24.10 LAL662789.11
	HIST1H2BJ	RP1-97D16.1 ZS	CAN23	COR2P1P	AL645939.6-2	PRR3 GTF2H4	DHFRP2 LSM2 HNRPA1P2 HLA
	HIST1H2AG	CRP1-97D16.4	SPX5	SAR1P1	GPR53P C6orf12	2 HLA-E	FGFR3P HSPA1L MTCO3P1
	"HIST1H2BK	"HIST1H4PS1	SCAND3	OR2J2	OR2I1P HLA-80	"UBQLN1P	"ZDHHC20P2 CYP21A2 "HLA-DQB3
	HIST1H2AH	°HIST1H2BPS2		°OR2J4P	*TMEM183AP1	"MICC "MDC1	"HLA-S LY6G6C FKBPL HLA-DRB5
	PRSS16	KP1-193B12.5		TRIMOT LOOK	ZDHHC20P1	SUCLAZP C6orf205	RPLISP4 TINXB THLA-DRA THLA
	PUNIZILZ	- KP3-408B20.4		Centino L			
	ZINPTO	Lopowop		LTNE244			
		LOP1E12P		LOP2W1		LARCE1 LCDS	MICE LISPATA CONTIN LILADOA
		LZSCAN12L1		LOR2B3		Land Land	CR1 LNCR3 LCFB LPBX2 LHIA-DOB
		LRP1-31316.8		LOR2J1	LHCG9P5	RPP21 FLOT1 TCI	19 LMCCD1 LRDBP LPSMB9
		LRP1-265C24	5	LOR2J3		M26 MRPS18B PO	J5F1 LBAT2 LSKIV2L LTAP1
		LAL022393.	,	LOR5V1		140 LC6orf136 L	HLA-C BAT3 DOM3Z BR
		LAL022393.	6	LOR5L	II LHCG4P8	LDHX16	LMICA LBATS LSTK19
		LHIST1H2BL		LOR	12D2 LHCG4P7	LKIAA1949	LBAT1 LHSPA1B
		LHIST1H2AI LZSC	N12	LOI	R10C1 <sup>L</sup> HLA-16	LNRM .	LATP6V1G2 LAGPAT1
		LHIST1H3H LZKSC	AN3	Loi	R11A1 MCCD1P1		LNFKBIL1 LRNF5
		LHIST1H2AJ		L.C.	R2H1 HCG4P6		LTA VARS AGER
		LHIST1H2BM		L. L.	MASIL HLA-K		LTNF C6orf48
		LHIST1H4J ZNF32	3		UBD HLA-21		LTB NEU1 GPSM3
		HIST1H4K			HCG4P5		APOM NOTCH4
		HIST1H2AK			OR2H2 MICD		C6orf47
		HIST1H2BN			MOG BAT1P2		BAT4
		-HIST1H2AL			-MCCD1P	2	-CSNK2B
		-HIST1H1B			-GABBR1 -HCG4P3		LY6G6E
		-HIST1H3I			-ZFP57 -ZNRD1		-C6orf25
		LHISTING			LHIA.G		LCUC1
		LHIST1H2AM			L <sub>HI A-A</sub>		LC6orf26
		LHIST1H2BO			LRNF39		C6orf27
		OR2B2			LTRIN	A10	LSLC44A4
		LOR2B6			LTRI	W15	LEHMT2
		LZNF165					LZBTB12
		ZNF435					
		LZNF192					
		ZNF193					
		LZKSCAN	4				
		LNKAPL.					
Assembly excepti			HAP REF:	c6_COX:28688544-3	33420241 (fwd)		
			HAI	P REF: c6_QBL:2888	5510-33451440 (fwd)		
		28.00 Mb		29.00 Mb	30.00 Mb	31.00 Mb	32.00 Mb 33.00 P
	Reverse strand				6.02 Mb		
	There are currently Archive EnsEMBL	68 tracks turned off. Iomo saniens version 54	36p (NCB	(36) Chromosome	6: 27.145.000 - 33.161.0	00	

(b) peak on chromosome 6, between 27145 and 33161 kb  $\,$ 

Fig. 4.—: Genes in the regions of increased IBD

### A.6. Parameter tuning and performance metrics

Below we describe tuning of the parameters of ANCHAP, which include:

- IBD threshold (Stage I)
- IBD region margins (Stage I)
- alignment parameters: overlap threshold and matching threshold (Stage II)
- number of markers phased for both individuals in a putative IBD region (Stage III)

Tuning is informed by the following performance metrics:

- evaluation against reference recent IBD the results between the reference individuals were evaluated against the regions of true recent IBD. The total number of markers in true regions and in resulting regions is TP, in true regions but not in the resulting regions is FN, not in true regions but in the resulting regions FP, and neither in the true regions nor in the resulting regions TN. From these, sensitivity and false discovery rate can be computed.
- sensitivity TP/(TP + FN)
- false discovery rate FP/(FP + TP)
- inconsistency rate how many of the alleles of haplotype sharers were homozygotes not consistent with homozygotes of the majority of the haplotype sharers, divided by the number of haplotype sharers.
- percentage of aligned sequences in the first round of an algorithm. Out of all detected IBD regions in the first round, what proportion of them were aligned into one of the gametes.

### A.6.1. Reference sharing in ORCADES study

The evaluation of the algorithms was possible thanks to parent offspring pairs genotyped in the ORCADES study. There are 58 individuals with both parents genotyped, and at 80% of their heterozygous loci they could be phased. There are 160 with at least one parent genotype and they could be phased at 70% of heterozygous loci.

To obtain the reference IBD information, we extracted IBD regions between the 58 reliably phased reference individuals. We required alleles with identical alleles in a region larger than 2 cM and containing at least 100 SNPs. The length of IBD regions between the reference individuals is shown in Figure 5a. The density of IBD sharing across the genome is shown in Figure 5c.

We also counted IBS sequences shorter than 2 cM. Contrary to our estimates about the expected lengths of IBD since re-settling of the island around 50 generations ago, there are many such segments. The lengths of IBS segments shorter than 2 cM are presented in Figure 5b.

### A.6.2. IBD threshold in Stage I

In Stage I of ANCHAP we would like to phase the individuals widely with as few phasing errors as possible. Genotypes would be widely phased if many haplotype sharers are widely detected. There would be few phasing errors if there is no falsely detected IBD sharing. Therefore the sensitivity and false discovery rate of IBD detection, evaluated on the reference phased individuals, are meaningful metrics which will reflect the quality of phasing.

On the other hand, when there is more than one haplotype sharers, and some falsely detected IBD region, the alignment stage of Anchap may eliminate the falsely detected



(a) How long are the regions of reference sharing? Horizontal axis - region length in cM on log-scale. Vertical axiscumulative proportion of of IBD segments in the IBD regions.



(b) Length of IBD segments in reference sharing. Most of the segments are much shorter than 2 cM.



(c) How is the reference sharing distributed across the chromosome? Is it influenced by fluctuations in the genetic or physical map? Top: density of IBD between the 58 reference individuals in ORCADES. Middle: genetic map. Bottom: physical map.

Fig. 5.—: Reference data for our comparison: sequences shared between the individuals which can be reliably phased



Fig. 6.—: Sensitivity and false positive rate of IBD regions as recovered by Stage I of ANCHAP with different IBD thresholds.

sharing.

The plot of sensitivity and false discovery rates for different IBD thresholds in the first round are shown in Figure 6.

#### A.6.3. IBD region margins

At each border of the putative sharing regions we trimmed 100 markers. In the experiments with the reference data, after trimming 100 markers at each side 94% of detected sharing regions will not contain any spurious sharing at the borders.

### A.6.4. Stage II - alignment parameters

In Stage II of ANCHAP haplotype sharers are split into two groups, and regions of falsely assumed sharing may be discarded. The algorithm starts with the longest and therefore most certain IBD regions, reconstructs a draft of the phase, and then matches the remaining sharers against the preliminary phased genotypes. Errors may occur in the preliminarily reconstructed haplotypes, and therefore few inconsistencies between the draft of the haplotypes and the aligned sequences may be allowed.

There are two parameters necessary for this part of the algorithm. The overlap threshold specifies the minimal number of markers of overlap between the draft of phase of an individual and the new IBD region. The matching threshold specifies how many alleles may be mismatching between the draft of the phase and a genotype of the putative IBD sharer.

Right values of parameters will result in a good split of haplotype sharers into two groups and consequently to low phasing error, and a good proportion of the genotypes will be phased. A good proportion of the putative IBD sequences would be aligned. The genotypes of IBD sharers who are all classified as sharing the same haplotype should also be consistent between each other. There should be no opposing homozygotes between such genotypes, and therefore the inconsistency ratio should be low.

In Table 1 we evaluate the impact of different values of the overlap threshold and the matching threshold. For each pair of values, we evaluate the percentage of the putative IBD regions successfully aligned, and the inconsistency ratio.

overlap threshold	matching threshold	percentage of haplotype	inconsistent homozygotes
		sharers aligned	among haplotype
			sharers, normalised
5	0	0.07	8.90E-06
5	0.01	0.67	5.11E-04
5	0.02	0.74	1.00E-03
5	0.05	0.82	2.12E-03
5	0.1	0.86	3.45E-03
5	0.2	0.91	5.55E-03
5	0.5	0.50	3.74E-03
10	0	0.07	8.92E-06
10	0.01	0.67	5.14E-04
10	0.02	0.73	1.00E-03
10	0.05	0.86	2.01E-03
10	0.1	0.86	3.46E-03
10	0.2	0.90	5.56E-03
10	0.5	0.50	3.74E-03
20	0	0.07	8.92E-06
20	0.01	0.66	5.19E-04
20	0.02	0.73	1.01E-03
20	0.05	0.81	2.15E-03
20	0.1	0.86	3.49E-03
20	0.2	0.90	5.59E-03
20	0.5	0.49	3.77E-03
50	0	0.07	8.92E-06
50	0.01	0.64	5.35E-04
50	0.02	0.71	1.05E-03
50	0.05	0.78	2.21E-03
50	0.1	0.83	3.55E-03
50	0.2	0.87	5.67 E-03

Table 1:: Experiments with parameters for Stage II of ANCHAP. According to these parameters it is decided whether two diplotype segments are aligned, i.e. whether they share the same gamete. Marked in gray is the value of the parameter used.

### A.6.5. Stage III parameters

In Stage III we look for haplotypes matching continuously in regions which are at least 2cM long. In addition we require that both of the compared haplotypes are phased another parameter specifies a minimum number of markers phased in both of the haplotypes - by default it is set to 100. In Table 2 we show the accuracy of IBD detection when this threshold is varied. For sensitivity the threshold has a negligible impact, while false discovery rate increases significantly when the threshold is set to less than 100.

length thresh	sensitivity	false discovery rate
10	0.84	0.031
20	0.84	0.031
50	0.84	0.027
100	0.84	0.016
200	0.81	0.011

Table 2:: Experiments with values of the parameter of the second round scan - lengththresh. This parameter specifies how many markers in the region of putative IBD need to be phased. Marked in gray is the value of the parameter used.

# A.7. Challenges to haplotype alignment

The quality of haplotype reconstruction, as measured by the switch error, is influenced by accuracy of sharing detection and of the algorithm that splits haplotype sharers into two groups. ANCHAP's greedy algorithm first chooses the longest shared sequences, as they carry most information about the haplotype, and tries to align the haplotypes from the remaining shared regions. When the proband's haplotype is not phased in the region, the assignment to a haplotype is arbitrary, and these arbitrary decisions may not be propagated between genetic regions.

### A.8. Tuning SLRP

Table 3 shows experiments with empirical and default parameter values for SLRP. We compared the default values with values obtained empirically. The expected IBD length in centimorgans was computed from the IBD regions between the reference individuals in ORCADES, after they were phased. The expected IBS but not IBD was calculated from IBS segments between the reference individuals, longer than 20 markers. Because we defined IBD as matching of haplotypes within a region longer than 2 cM, out of the output of SLRP we filtered out the results shorter than this threshold.

SLRP setting	ExpectedIBS (cM)	ExpectedIBD (cM)	sensitivity	false discovery rate
default	1	10	0.76	0.0076
empirical	0.42	9.17	0.77	0.0106

Table 3:: Tuning SLRP. Only counting the IBD regions longer than 2cM. Sharing between the 58 Orkney individuals was evaluated. Data from chromosome 2. Marked in gray is the value of the parameter used.

### A.9. Tuning fastIBD

In Table ?? we show experiments with varying the scale parameter in fastIBD. We filtered out regions shorter than 2 cM, in accordance with our definition of IBD.

#### A.10. Comparison of Anchap, SLRP and fastIBD

In the article we evaluated IBD regions as inferred by different regions against the IBD segments between the reference individuals. Here we additionally show density of IBD across the genome (Figure 7) and comparison of lengths of detected IBD segments (Figure 9).

fastIBD setting	scale	sensitivity	false discovery rate	sensitivity	false discovery rate
		$<\!2\mathrm{cM}$ pruned	<2cM pruned		
minimum advised	1	0.270	0.000	0.271	0.002
	2	0.631	0.010	0.635	0.023
	2.5	0.744	0.018	0.750	0.036
	2.6	0.767	0.018	0.774	0.037
	2.7	0.783	0.019	0.790	0.043
	2.8	0.802	0.021	0.808	0.046
	2.9	0.805	0.024	0.812	0.051
	3	0.825	0.024	0.832	0.056
	3.1	0.832	0.027	0.839	0.066
	3.2	0.837	0.028	0.844	0.070
	3.3	0.845	0.030	0.853	0.073
	3.4	0.849	0.032	0.855	0.079
	3.5	0.857	0.036	0.865	0.088
maximum advised	4	0.870	0.045	0.879	0.118
merge 10 runs	3	0.868	0.044	0.873	0.106

Table	e 4::	Tuning	fast	IBD.	Sharir	ıg b	etween	the	58	Orkney	indivi	duals	was	evaluated.	Data
from	chro	mosom	e 2.	Mark	ed in g	gray	is the	valu	e of	f the pai	amete	r use	d.		



Fig. 7.—: Genome-wide view of haplotype sharing as recovered by the compared methods. SLRP and fastIBD are more conservative in IBD detection, and have less apparent IBD peaks.



Fig. 8.—: Lengths of detected IBD segments [cM]. IBD regions detected by ANCHAP are generally shorter, as the method does not account for switch errors in phasing after the first round.

# B. Evaluation of the selection procedure



Fig. 9.—: Evaluation of ANCHAP's selection procedure for choosing subjects for resequencing studies. We estimated the IBD imputation potential when the samples are chosen randomly or based on kinship. Individuals were chosen randomly 10 times, and the results were averaged.

### REFERENCES

- I. H. Consortium. A second generation human haplotype map of over 3.1 million snps. Nature, 449(7164):851–861, Oct 2007. doi: 10.1038/nature06258. URL http://dx.doi.org/10.1038/nature06258.
- K. Palin, H. Campbell, A. F. Wright, J. F. Wilson, and R. Durbin. Identity-by-descent-based phasing and imputation in founder populations using graphical models. *Genet Epidemiol*, Oct 2011. doi: 10.1002/gepi.20635. URL http://dx.doi.org/10.1002/gepi.20635.
- S. Purcell, B. Neale, K. Todd-Brown, L. Thomas, M. A. R. Ferreira, D. Bender, J. Maller, P. Sklar, P. I. W. de Bakker, M. J. Daly, and P. C. Sham. Plink: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet, 81(3):559–575, Sep 2007. doi: 10.1086/519795. URL http://dx.doi.org/10.1086/519795.