

SUPPLEMENTARY DATA

FIG. S1. Area of transverse section of a mature *Zea mays* leaf. The arrows indicate the vascular bundle sheath, while the arrowheads “palisade-like” MCs. This figure has been taken from a slide provided by Carolina Biological Supply (Burlington NC, catalog number 30-3514).

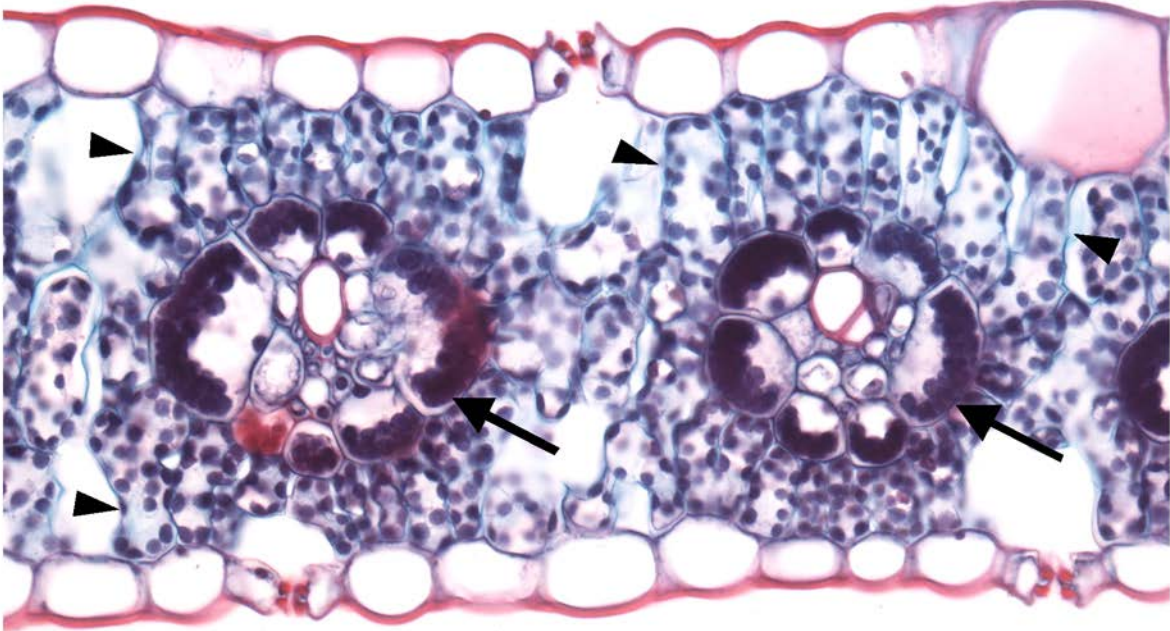


FIG. S2. Shaping MCs as they are seen in DIC optics (A, C) and in epifluorescence microscope under a set filter provided with exciter G365 and barrier LP420 (B) and another with exciter BP450-490 and barrier BP 515-565 (D). Under these filters, the cell walls do not emit any autofluorescence. Scale bars = 5 μ m.

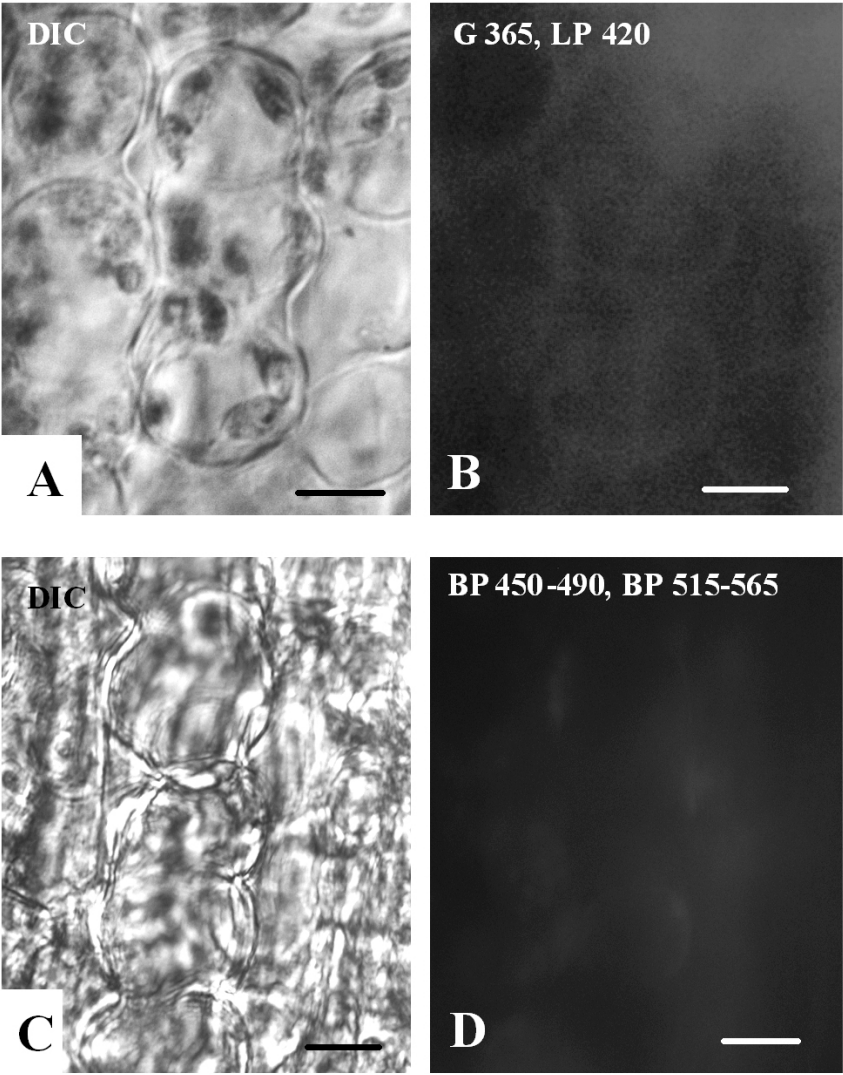


FIG. S3. TEM micrographs of shaping MCs. Scale bars: (A-D) = 500 nm; A *inset* = 2 μ m.

A: Longitudinal section of the MT-ring (arrows) lining the wall thickening in the region of forming cell isthmus. *Inset:* The shaping MC in lower magnification. The arrows point to the forming cell isthmus shown in **A** in a higher magnification. **B:** Local wall thickening deposited on a longitudinal cell wall at the site of future MC isthmus. MT groups (arrows) line the cell wall thickening on both sides. **C:** Paradermal section of a local cell wall thickening deposited at the region of a future MC isthmus. Note the co-alignment of the cellulose microfibrils (black and white lines) and of the underlying MTs (arrows). **D:** Developing intercellular space (IS) at the site of a forming MC isthmus. Arrows point to the MT-bundles lining the detached cell wall thickenings.

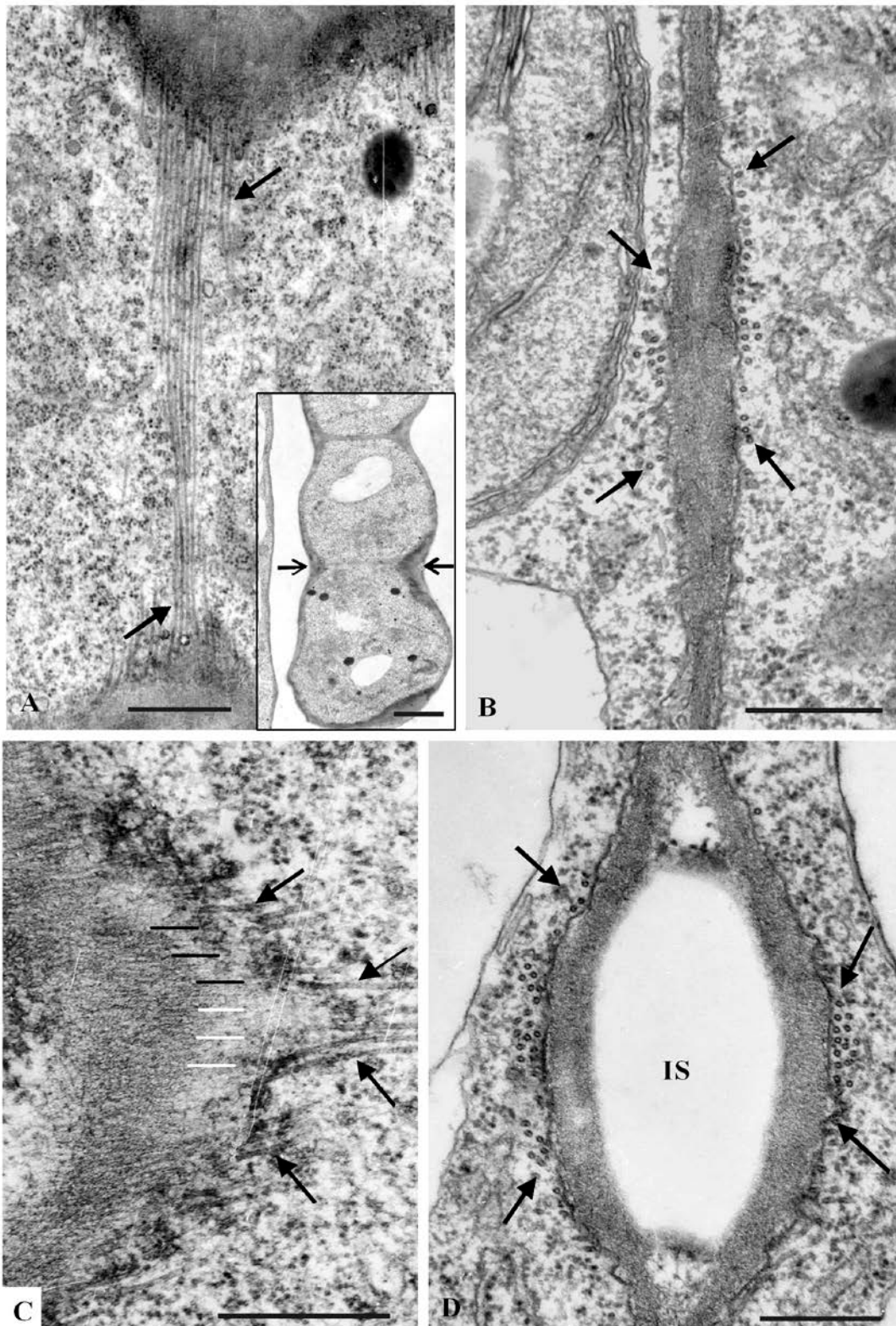


FIG. S4. **A, C**: Young (**A**) and shaping (**C**) MCs of *Triticum turgidum* as seen in epifluorescence microscope after aniline blue staining. Callose patches are localized in the area of future (arrows in **A**) and forming (arrows in **C**) MC contacts. **B, D**: The MCs seen with DIC optics. Scale bars = 10 μm .

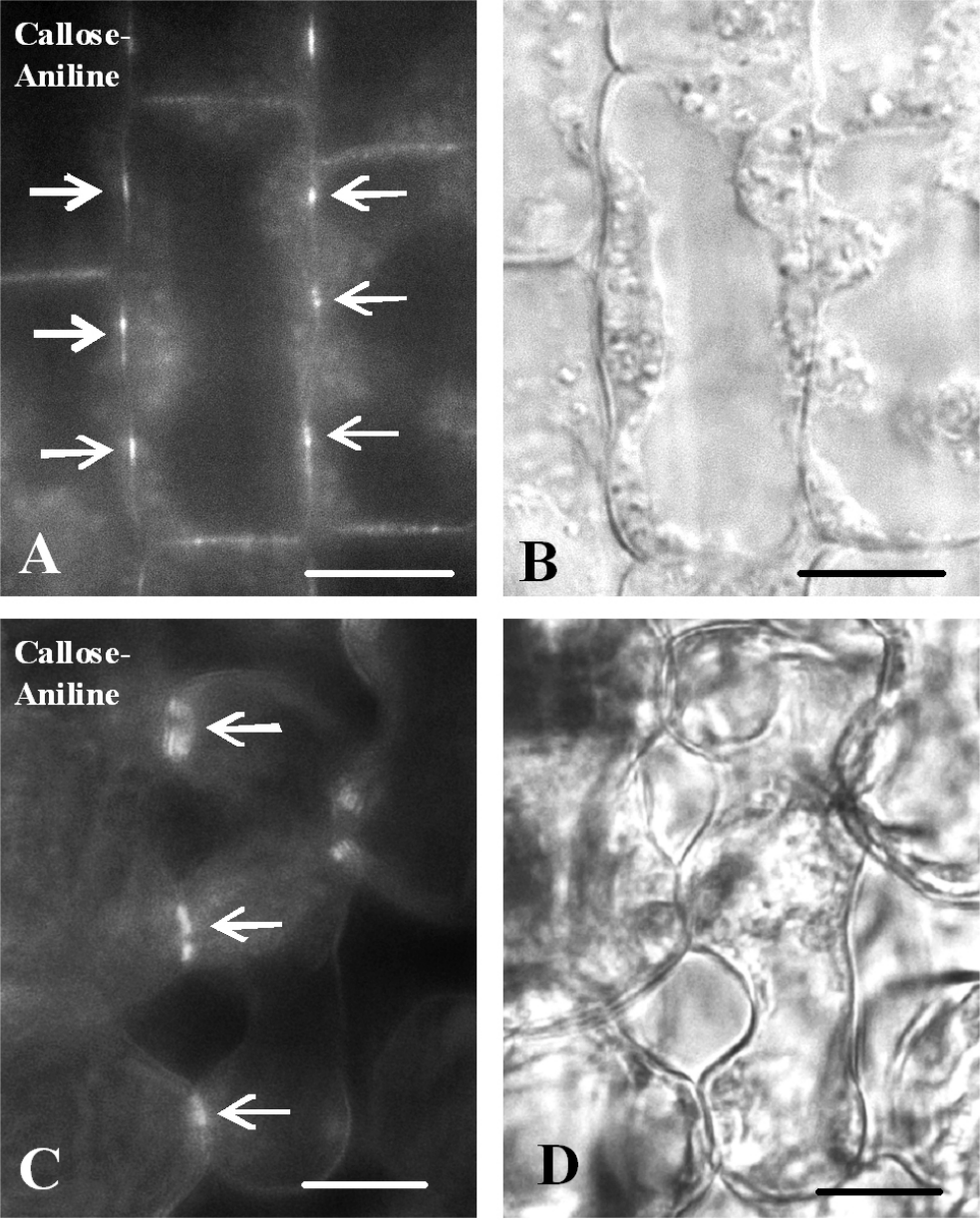


FIG. S5. Immunodetection of cell wall matrix polysaccharides in sections of nascent MCs embedded in LRW. **A₁, C₁, E, G**: Control sections. **B₁, D₁, F, H**: Sections pretreated with 1 M KOH for 1 h at 25 °C. All sections have been taken from the same leaf part. Scale bars= 10 μm. **A₁-D₂**: Immunodetection of JIM5- (**A₁, B₁**) and JIM7- (**C₁, D₁**) HGA epitopes. Arrows in **A₁** and **C₁** point to the positions where the above HGA epitopes are localized. They have been removed from cell walls after the treatment of sections with KOH (**B₁, D₁**; cf. **A₁, C₁**). **A₂, B₂, C₂, D₂**: The areas of MCs presented in **A₁, B₁, C₁** and **D₁** as appeared with DIC optics. Arrowheads point to intercellular spaces. **E, F**: Callose immunodetection in control section (**E**) and in section pretreated with KOH (**F**). In both cases, callose patches (arrows) are obvious. **G, H**. MLGs immunodetection in control section (**G**) and in section pretreated with KOH (**H**). In both cases, MLGs are localized in cell walls delimiting the developing intercellular spaces (arrowheads), while they are absent from the area of future cell contacts (arrows).

