

Supplemental Materials

Molecular Biology of the Cell

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Figure S1. The growth phenotypes of polarisome component mutations in wt or *ptc1Δ* cells. **(A)** Serial dilutions of the overnight cultures from indicated strains were plated on YPD plates and grown for 2 days at different temperatures as shown. **(B)** Synthetic lethality between *msb3Δ msb4Δ* and *ptc1Δ*. Dissection plates are of the following diploid strains: *msb3Δ msb4Δ* crossed to *ptc1Δ*. The *msb3Δmsb4Δ* double mutant (representative spores are squared) grows slower than WT. Inferred triple mutant spores (*msb3Δmsb4Δptc1Δ*) are circled.

Figure S2. Deletion of all individual polarisome component genes (except *msb3* and *msb4*) significantly suppresses the ER inheritance defects in *ptc1Δ* mutant cells. Shown are the representative GFP fluorescence of the ER marker Hmg1-GFP and DIC images of the following strains: (top) wild type, *sph1Δ*, *pea2Δ*, *spa2Δ*, *msb3Δ*, *msb4Δ*, (bottom) *ptc1Δ*, *sph1Δptc1Δ*, *pea2Δptc1Δ*, *spa2Δptc1Δ*, *msb3Δptc1Δ*, and *msb4Δptc1Δ*. Arrowheads point to small buds. Bars, 5μm. The percentage of small buds that contained normal cortical ER in these strains is shown in Figure 1B.

Figure S3. Deletion of *PEA2* or *SPA2* affects Slt2-GFP localization at the bud tip or bud neck, while deletion of other polarisome component genes does not. Shown are Slt2-GFP fluorescence and DIC images of representative cells from more strains: *ptc1Δ*, *sph1Δ*, *spa2Δ*, *msb3Δ*, *msb4Δ* and *msb3Δmsb4Δ*, which are not shown in Figure 2. Arrowheads and arrows point to bud tips in small buds and bud necks in large buds, respectively. Bars, 5μm. The quantification results of the Slt2-GFP localization in small buds from these

strains are shown in Figure 2B.

Figure S4. Deletion of *PEA2* or *SPA2* affects Mkk1-GFP localization at the bud tip or bud neck. A plasmid expressing an Mkk1-GFP fusion from the *ADHI* promoter was transformed into the indicated strains. Cells were grown to early log phase in SC medium at 25°C and examined by fluorescence microscopy. Mkk1-GFP localization at polarized sites was quantified. The error bars in the bar graph represent the SEM from three independent experiments.

Figure S5. The effects of Latrunculin A (Lat-A) treatment on the time course of cER inheritance in *ptc1Δ* or *sec3Δ* mutant cells. Arrowheads point to small buds. *ptc1Δ* and *sec3Δ* cells expressing the ER marker Hmg1p-GFP were treated with 200μM Lat-A on the slide. Time lapse images were acquired by fluorescence microscopy immediately after Lat-A addition. Selected frames from the time series of representative cells are shown. Bar, 5μm.

Figure S6. The effects of Latrunculin A (Lat-A) treatment on Mitochondrial inheritance or cER inheritance in *ptc1Δ* mutant cells. Wild-type and *ptc1Δ* cells expressing the ER marker Hmg1p-GFP and containing a plasmid expressing the F₀ATP synthase mitochondrial targeting sequence fused to RFP (Mito-RFP) were treated with control DMSO or 200μM Lat-A for 10 min. Cells were immediately fixed, washed and then imaged by fluorescence microscopy. (A) Representative cell images. Arrowheads point to the cER staining in small buds. Arrows point to the efficient or deficient mitochondria

staining. **(B)** Percentage of small buds containing normal cER staining or normal mitochondrial formation with or without Lat-A treatment. The error bars in the bar graph are the SEM from three independent experiments.

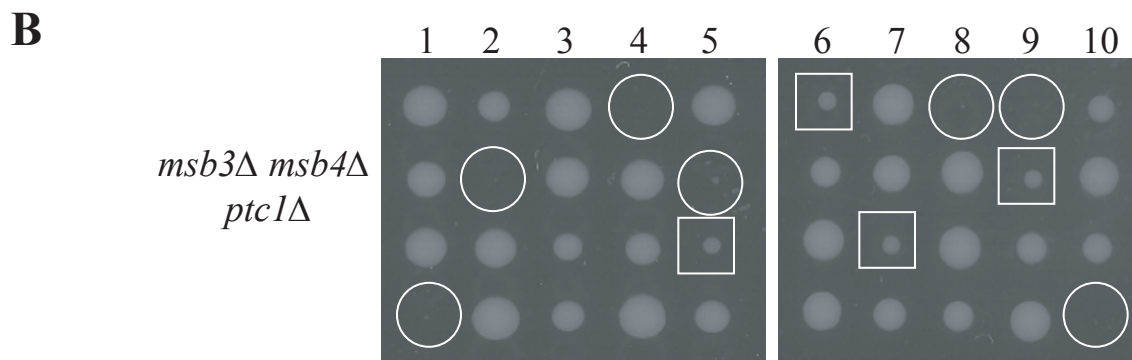
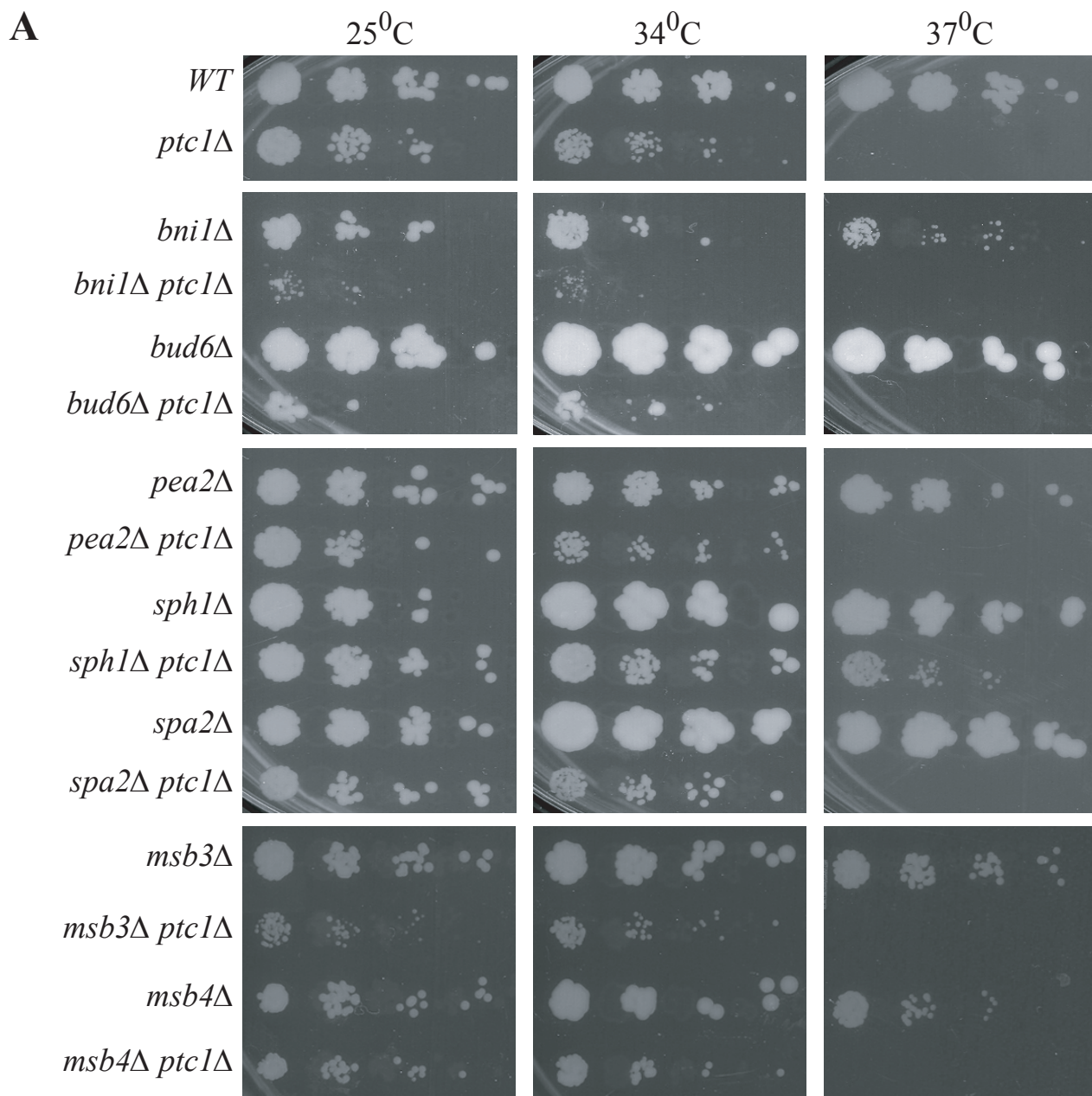


Figure S1

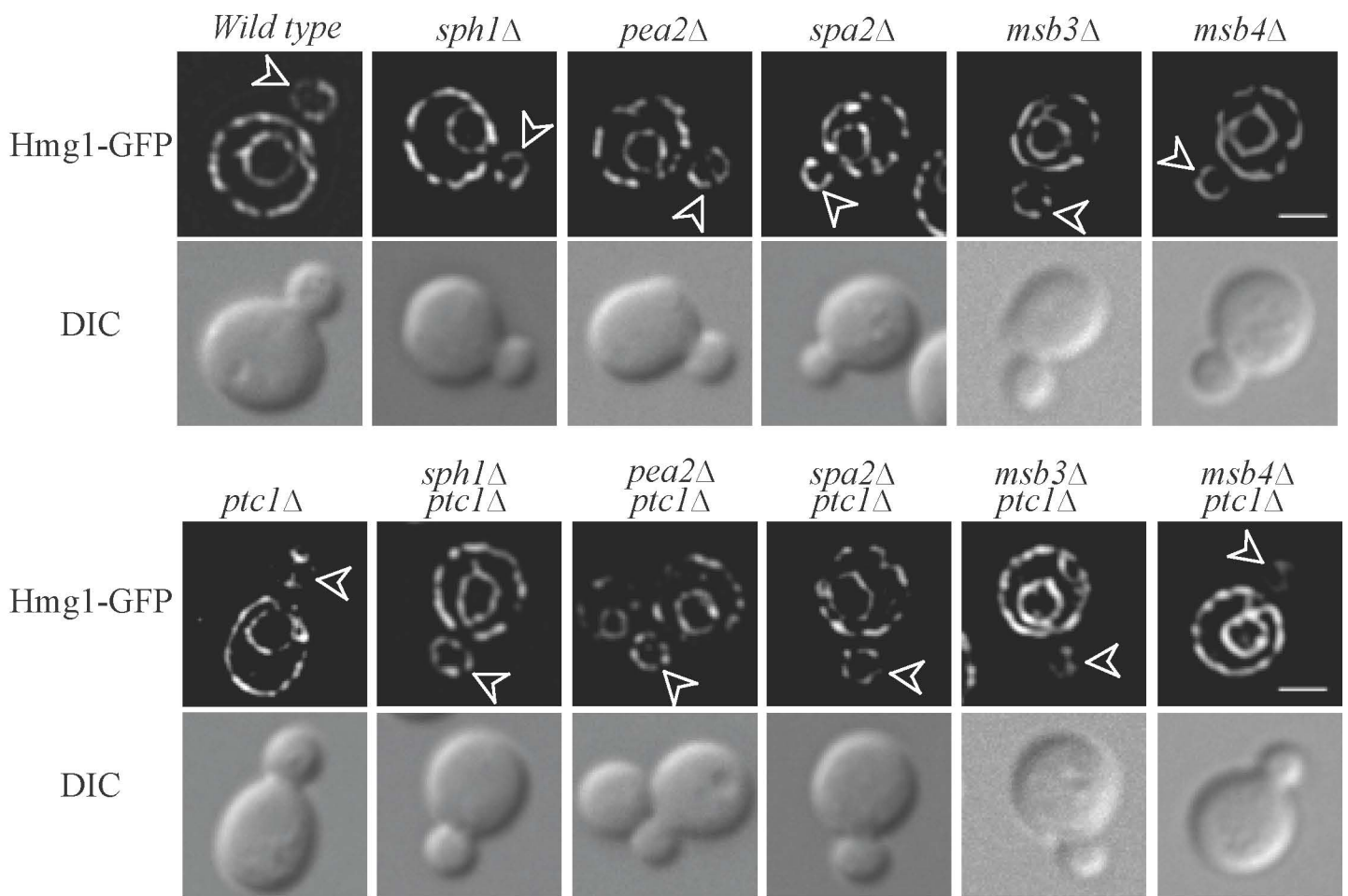


Figure S2

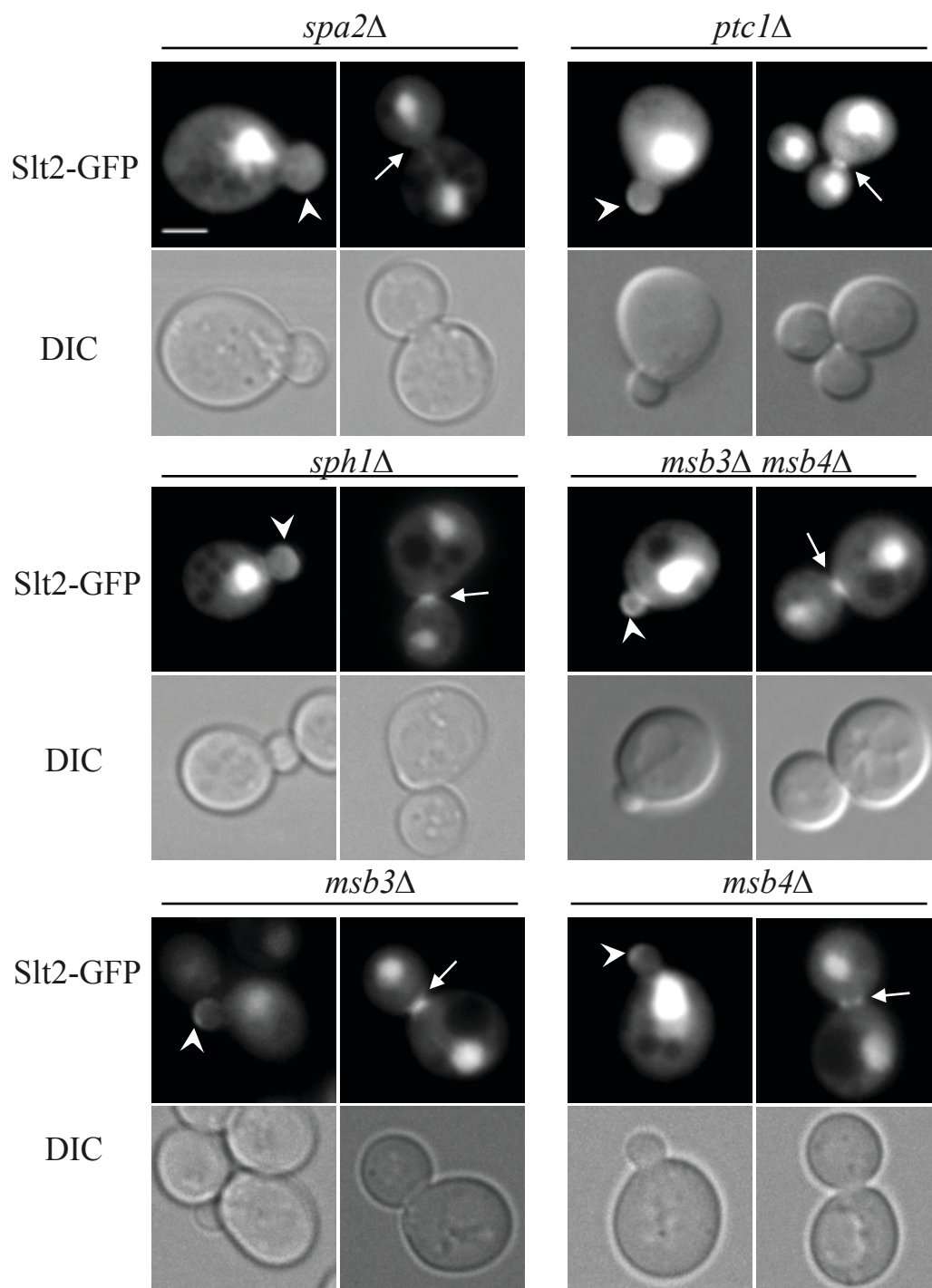


Figure S3

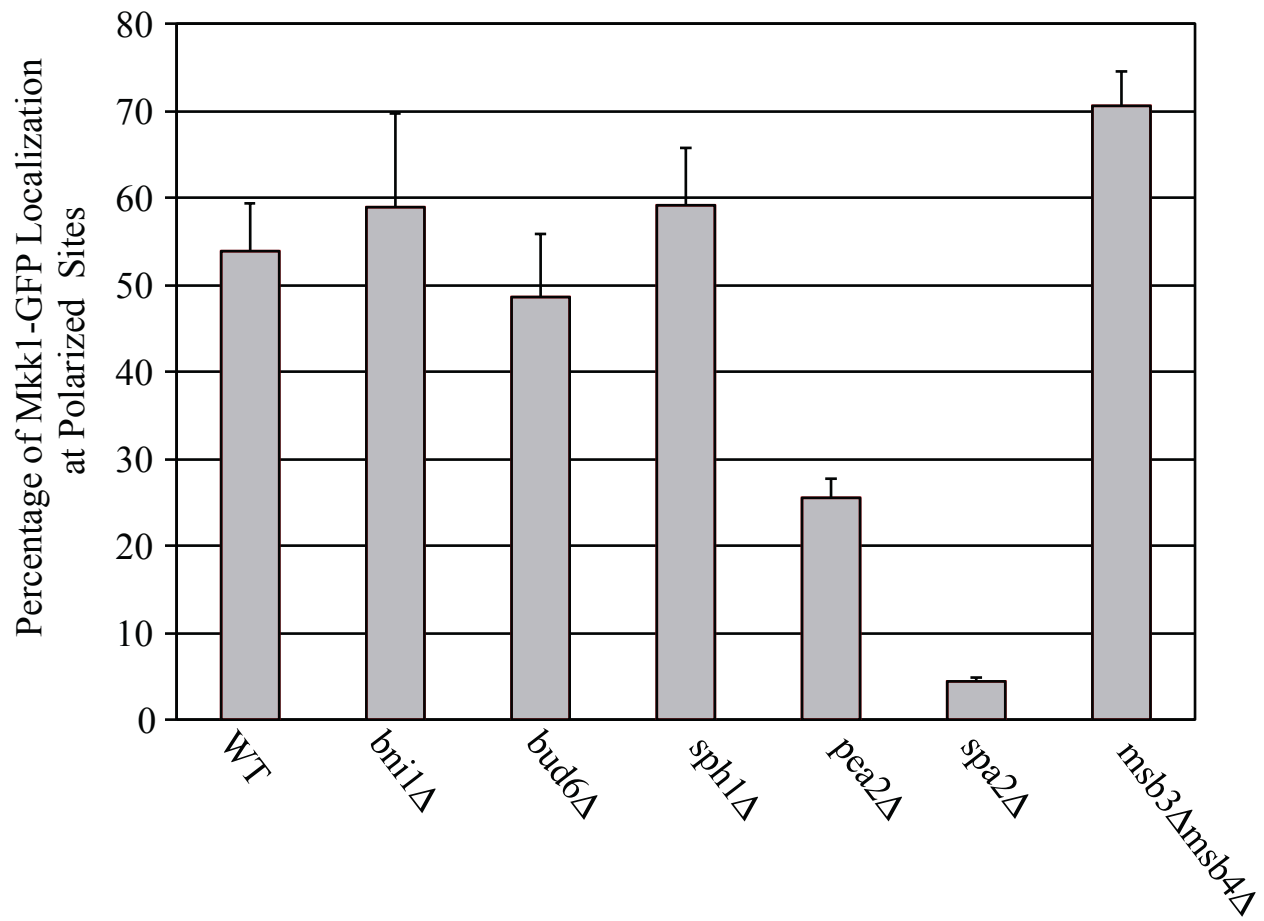


Figure S4

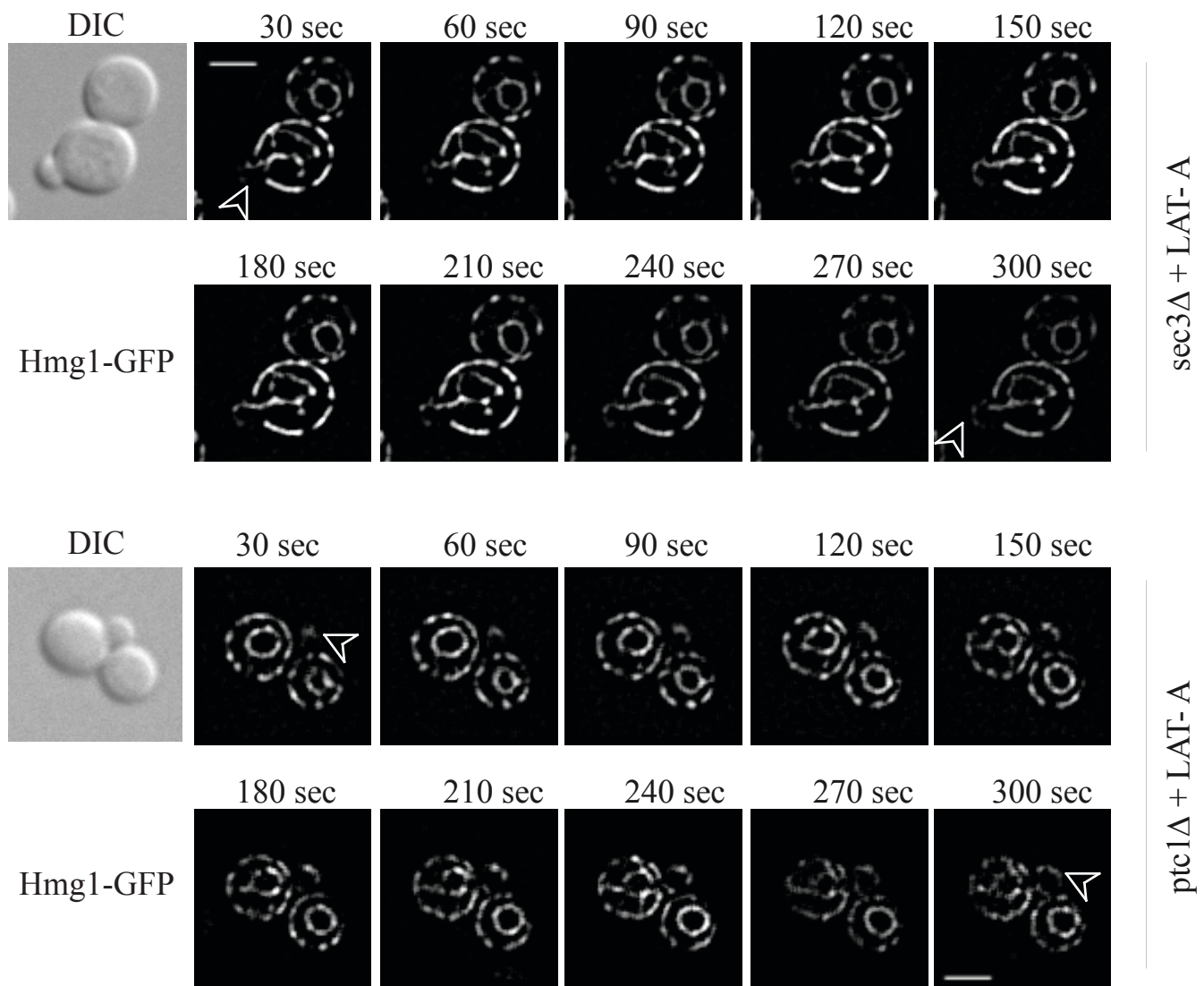


Figure S5

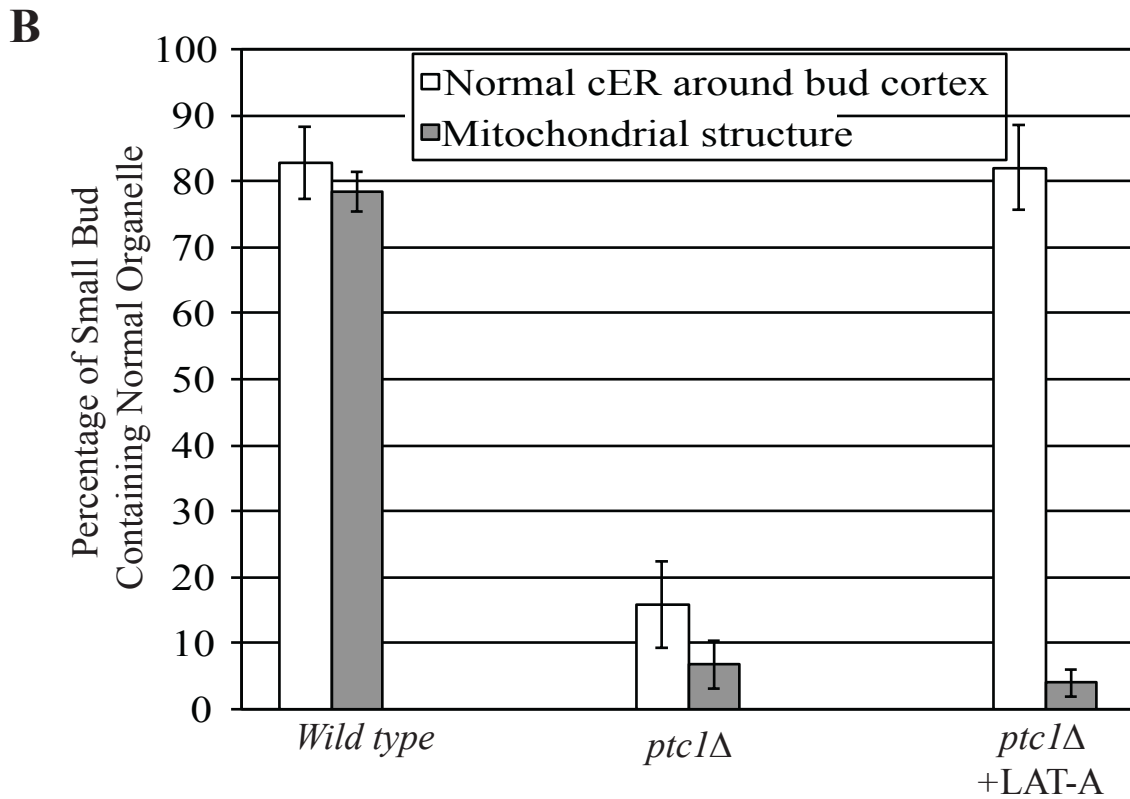
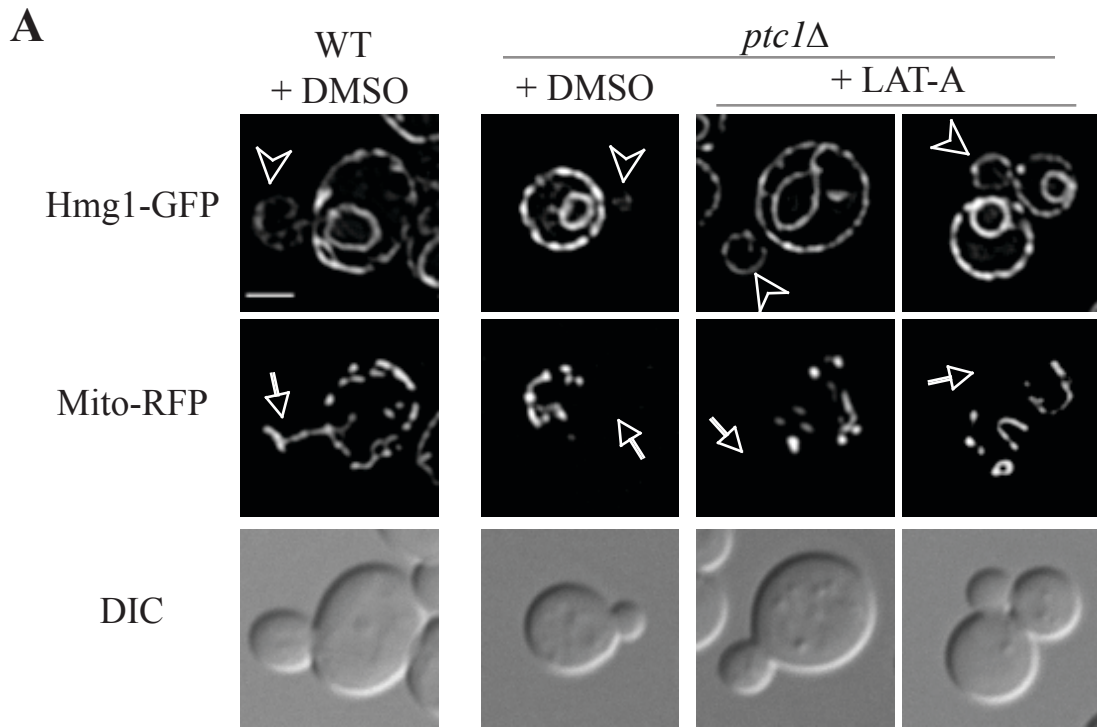


Figure S6