## **Supporting Information**

 $Mn^{2+}$  - nucleotide coordination at the myosin active site as detected by pulsed EPR.

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## *Structures of the nucleotides studied in this work*







Figure S1. The structure of ADP.AlF<sup>4</sup>  $(C_{10}H_{15}N_5O_{10}P_2AlF_4)$  with the metal ion, coordinating β phosphate and  $AlF<sub>4</sub>$ .



Figure S2. The structure of AMPPNP  $(C_{10}H_{17}N_6O_{12}P_3)$  with the metal ion, coordinating γ and β phosphates.

## Interpretation of the ligand hfi parameters observed in the complexes of high-spin transition metal ions.

Although the relationship between the observed (effective) *hfi* constant and the true oneelectron *hfi* constant in high-spin systems is well-known [1-3], the necessity to appropriately rescale the <sup>31</sup>P and <sup>14(15)</sup>N isotropic *hfi* constants of coordinated phosphates and nitrogens in the Mn.nucleotide complexes was missed in all of the high-resolution EPR works where such complexes were studied [4-10]. Therefore, as a reminder, we present here a simple derivation that demonstrates the general approach to obtaining such scaling factors. Let us consider the specific system at hand, the Mn<sup>2+</sup> center, which has the  $d_5$  electronic configuration with the total spin *S* = 5/2. This total spin represents a sum of the individual spins of each of the *d*-electrons:

$$
S = \sum_{j} S_{j}
$$
 (S1)

where  $S_j = 1/2$  are the individual spins, and the simple summation implies that the multiplet with the highest possible spin is formed. The states with positive total  $S_Z$  are described by:

2/5 = ααααα ( ) 5 1 2/3 = ααααβ +αααβα +ααβαα +αβααα + βαααα (S2) ) ( 10 1 2/1 αβαβα βααβα αββαα βαβαα ββααα αααββ ααβαβ αβααβ βαααβ ααββα + + + + + = + + + + +

and those with the negative  $S_Z$  are obtained from the corresponding positive ones by interchanging the single-electron functions  $\alpha$  and  $\beta$ . The average values of single-electron operators  $S_{Zk}$  (where the index *k* indicates any one of the five electrons) obtained from eq S2 are:

$$
\langle S_{Zk} \rangle_{5/2} = \langle 5/2 | \mathbf{S}_{Zk} | 5/2 \rangle = 1/2 = (5/2) \times 1/5
$$
  

$$
\langle S_{Zk} \rangle_{3/2} = \langle 3/2 | \mathbf{S}_{Zk} | 3/2 \rangle = 3/10 = (3/2) \times 1/5
$$
  

$$
\langle S_{Zk} \rangle_{1/2} = \langle 1/2 | \mathbf{S}_{Zk} | 1/2 \rangle = 1/10 = (1/2) \times 1/5
$$
 (S3)

*etc*. The factor of 1/5 in eq S3 equals to 2*S*, and the average single-electron values can be expressed as:

$$
\left\langle S_{Zk} \right\rangle_{Sz} = \left\langle S_Z \right\rangle / (2S) \tag{S4}
$$

Although the derivation of eq S4 was based on considering the specific case of  $S = 5$ , it is actually general and applies to the highest spin multiplets formed by any number of individual spins  $S_j = 1/2$ . In particular, it is relevant for the half-filled orbital shells of atoms and ions, *e.g.*,  $p_3$  (N<sup>0</sup>),  $d_5$  (Mn<sup>2+</sup>), or  $f_7$  (Gd<sup>3+</sup>).

The ligand isotropic *hfi* constants can be considered based on the model where only one of the five *d*-orbitals predominantly interacts with a given ligand, and thus only one unpaired electron delocalizes on that ligand and results in the isotropic *hfi* with the ligand nucleus. The isotropic *hfi* constant in such a model can either be expressed through the total electron spin (the experimentally observable effective *hfi* constant *aef*) or using the single-electron operators (the true one-electron *hfi* constant *a*):

$$
H_{iso} = a_{ef} S_{Z} I_{Z} = a S_{Zk} I_{Z}
$$
 (S5)

where a high-field approximation is assumed. Substituting the operators by the expectation values and using eq S4, one immediately obtains:

$$
a = 2S \cdot a_{ef} \tag{S6}
$$

For the anisotropic *hfi*, two contributions should be considered. The first contribution comes from the (one-electron) spin density delocalized on the ligand and is treated similar to the isotropic *hfi* (eqs S5 and S6), *e.g*.,

$$
H_{aniso}^{ligand} = T_{ZZef}^{ligand} \mathbf{S}_{\mathbf{Z}} \mathbf{I}_{\mathbf{Z}} = T_{ZZ}^{ligand} \mathbf{S}_{\mathbf{Z}k} \mathbf{I}_{\mathbf{Z}}
$$
(S7)

(and similar terms for  $T_{ZX}$  and  $T_{ZY}$ ), which results in

$$
T_{ZZ}^{ligand} = 2S \cdot T_{ZZef}^{ligand} \tag{S8}
$$

The second contribution is the through-space magnetic dipole interaction of the ligand nucleus with the spin density on the central ion. In this case, all  $N = 2S$  individual unpaired electrons interact with the ligand nucleus approximately equally:

$$
H_{aniso}^{central} = T_{ZZef}^{central} \mathbf{S}_{\mathbf{Z}} \mathbf{I}_{\mathbf{Z}} = (\sum_{k=1}^{2S} T_{ZZ}^{central} \mathbf{S}_{\mathbf{Z}k}) \cdot \mathbf{I}_{\mathbf{Z}} = T_{ZZ}^{central} (\sum_{k=1}^{2S} \mathbf{S}_{\mathbf{Z}k}) \cdot \mathbf{I}_{\mathbf{Z}}
$$
(S9)

(and similar terms for  $T_{ZX}$  and  $T_{ZY}$ ). Substituting the average values of the spin projections and using eq S4, one immediately arrives at:

$$
T_{ZZ}^{central} = T_{ZZef}^{central} \tag{S10}
$$

Taking into account the conversion factors given by eqs S6, S8, and S10 allows one to meaningfully compare the *hfi* parameters obtained for the ligands of high-spin ions with those observed for the ligands of the  $S = 1/2$  ions. In addition, using the true isotropic *hfi* constant (eq. S6) rather than the effective one results in more accurate estimates of the spin density delocalization on the ligands.

As an example, for the phosphate ligand at  $Mn^{2+}$  ion we obtained, on average,  $a_P \sim 23$ MHz (see Table 1 in the main text). Taking into account that the unpaired electron on the 3*s* orbital of phosphorus results in the *hfi* constant of about  $1.33 \times 10^4$  MHz [11] and that the *s*character of the phosphorus electronic orbitals in phosphate is about 1/4 (*sp*<sub>3</sub> orbitals), we can estimate the spin density on phosphorus as  $\rho_P \sim 0.007$ . Such a spin density will result in the anisotropic *hfi* constant  $T_{\perp}^{ligand}$  ~ (-367 MHz)×(3/4)× $\rho$ <sub>P</sub> ~ -1.9 MHz, where -367 MHz is the value of  $T_{\perp}$  that would be obtained for a pure *p*-orbital and  $\rho_{\rm P}$  =1 [11], and 3/4 is the *p*-character of the *sp*3 orbital. The effective (observable) anisotropic *hfi* from this source is thus  $T_{\perp ef}^{ligand} = T_{\perp}^{ligand}$  / 2*S* ~ -0.4 MHz. The "through-space" anisotropic *hfi* with the central ion at the distance  $R_{\text{MnP}} = 3.2$ -3.3 Å is  $T_{\perp ef}^{central} = T_{\perp}^{central} \sim -0.9$  MHz. Adding the two contributions together (with the proper account of the geometry of the Mn-O-P fragment) results in a slightly rhombic tensor  $(T_{11}, T_{22}, T_{33})_{ef}$  ~ (-1.1, -1.36, 2.46) MHz. The two smaller components obtained by this estimate are somewhat larger than the experimental value of *T*<sup>⊥</sup>*ef* = -0.9 MHz. This discrepancy can be corrected by taking into account the spin delocalization from the Mn ion to

the ligands (*e.g.*, assuming  $\rho_{Mn} = 0.7$  results in the calculated  $T_{\perp ef}$  similar to the experimental one).

The purpose of the above discussion was only to demonstrate the general approach to the analysis of the anisotropic and isotropic *hfi* for the ligands of the high-spin ions, in particular,  $Mn^{2+}$ . While this analysis is rather approximate, it is qualitatively better than simply estimating the Mn – P distance using a point dipole approximation. As follows from this analysis, the fact that the point-dipolar anisotropic *hfi* (for  $\rho_{Mn} = 1$ ) is similar to the experimental one is purely coincidental because the spin density delocalization into the phosphate orbitals gives a noticeable (~40%) contribution into the total anisotropic *hfi*.

## **References**

- 1. Kent, T. A., B. H. Huynh, and E. Munck. (1980) Iron-sulfur proteins: spin-coupling model for three-iron clusters, Proc Natl Acad Sci U S A 77, 6574-6576.
- 2. Stich, T. A., J. W. Whittaker, and R. D. Britt. (2010) Multifrequency EPR studies of manganese catalases provide a complete description of proteinaceous nitrogen coordination, J Phys Chem B 114, 14178-14188.
- 3. Owen, J., and J. H. M. Thornley. (1966) Covalent bonding and magnetic properties of transition metal ions, Rep. Prog. Phys. 29, 675-728.
- 4. Schneider, B., C. Sigalat, T. Amano, and J. L. Zimmermann. (2000) Evidence for changes in the nucleotide conformation in the active site of H<sup>+</sup>-ATPase as determined by pulsed EPR spectroscopy, Biochemistry 39, 15500-15512.
- 5. Zoleo, A., G. Lippe, S. Contessi, M. Brustolon, F. Dabbeni-Sala, and A. L. Maniero. (2007) Conformational role of the divalent metal in bovine heart mitochondrial F1-ATPase: an electron spin echo envelope modulation study, Biochemistry 46, 13443-13450.
- 6. Potapov, A., and D. Goldfarb. (2006) Quantitative Characterization of the Mn<sup>2+</sup> Complexes of ADP and ATPgS by W-band ENDOR, Appl. Magn. Reson. 30, 461-472.
- 7. Petersen, J., C. Gessner, K. Fisher, C. J. Mitchell, D. J. Lowe, and W. Lubitz. (2005)  $\text{Mn}^2$ <sup>+</sup>adenosine nucleotide complexes in the presence of the nitrogenase iron-protein: detection of conformational rearrangements directly at the nucleotide binding site by EPR and 2D-ESEEM (two-dimensional electron spin-echo envelope modulation spectroscopy), Biochem J 391, 527- 539.
- 8. Hoogstraten, C. G., C. V. Grant, T. E. Horton, V. J. DeRose, and R. D. Britt. (2002) Structural Analysis of Metal Ion Ligation to Nucleotides and Nucleic Acids Using Pulsed EPR Spectroscopy., J. Am. Chem. Soc. 124, 834-842.
- 9. Morrissey, S. R., T. E. Horton, C. V. Grant, C. G. Hoogstraten, R. D. Britt, and V. J. DeRose. (1999)  $Mn^{2+}$ -nitrogen interactions in RNA probed by electron spin-echo envelope modulation spectroscopy: Application to the hammerhead ribozyme, J. Am. Chem. Soc. 121, 9215-9218.
- 10. Buy, C., T. Matsui, S. Andrianambinintsoa, C. Sigalat, G. Girault, and J. L. Zimmermann. (1996) Binding sites for Mg(II) in H<sup>+</sup>-ATPase from Bacillus PS3 and in the alpha 3 beta 3 gamma

subcomplex studied by one-dimensional ESEEM and two-dimensional HYSCORE spectroscopy of oxovanadium(IV) complexes: a possible role for beta-His-324, Biochemistry 35, 14281-14293.

11. Morton, J. R., and K. F. Preston. (1978) Atomic parameters for paramegnetic resonance data, J. Magn.Reson. 30, 577-582.