

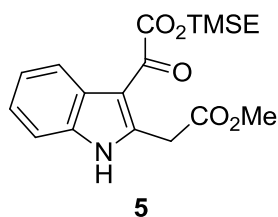
Table of Contents

Title page

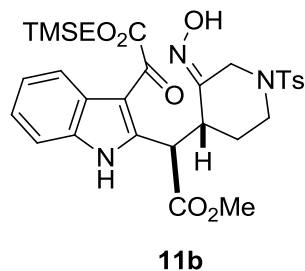
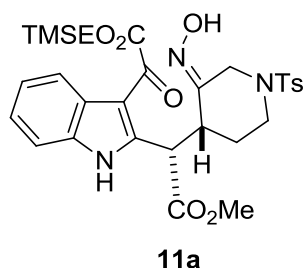
Experimental details for compounds **5, 11-21, 23-29, 3** S2-S24

NMR spectra of compounds **5, 11-21, 23-29, 3** S25-S82

General Methods. All non-aqueous reactions were carried out in oven- or flame-dried glassware under an argon atmosphere. All chemicals were purchased from commercial vendors and used as is, unless otherwise specified. Anhydrous tetrahydrofuran (THF) and dichloromethane (CH_2Cl_2) were obtained from a solvent purification system (Glass Contour). Reactions were magnetically stirred and monitored by thin layer chromatography (TLC) with 250 μm EMD 60 F₂₅₄ precoated silica gel plates. Flash column chromatography was performed using Silicycle silica gel P60 (230-400 mesh). Sonication reactions were performed in a Branson 1510 sonicator. FT-IR spectral data were recorded on a Thermo Nicolet FT-IR spectrometer equipped with a Diamond ATR accessory. ^1H and ^{13}C NMR spectral data were recorded on Bruker DPX-300, AMX-360 or DRX 400 MHz spectrometers. Chemical shifts are reported relative to chloroform (δ 7.26), methanol (δ 3.31), or DMSO (δ 2.50) for ^1H NMR and chloroform (δ 77.2), methanol (δ 49.0), or DMSO (δ 39.5) for ^{13}C NMR. Nominal mass spectra were recorded on Applied Biosystems 150EX. High resolution mass spectra were recorded on a Waters LCT Premier time-of-flight (TOF) mass spectrometer. X-Ray data was collected on a Bruker SMART APEX CCD area detector system.



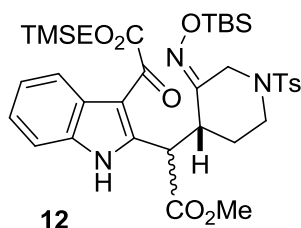
Synthesis of Keto Diester 5. To a stirred solution of the readily prepared indole ester **4**⁷ (3.50 g, 18.5 mmol) in Et₂O (310 mL) at 0 °C was added oxalyl chloride (1.84 mL, 21.4 mmol). The resultant orange solution was stirred at rt for 16 h and then cooled to 0 °C. 2-Trimethylsilylethanol (8.0 mL, 55.9 mmol) was added, followed by slow addition of triethylamine (6.4 mL, 47 mmol) over 5 min. The resulting red suspension was stirred at 0 °C for 1.5 h and then diluted with water. The organic layer was separated and the aqueous layer extracted with Et₂O. The combined organic layers were washed with water and brine, dried over MgSO₄, and concentrated *in vacuo* to give a viscous red oil which was purified by flash chromatography on silica gel (gradient 20% to 40% EtOAc/hexanes) to yield indole-3-oxoacetate **5** (6.03 g, 90%). ¹H NMR (300 MHz, CDCl₃) 10.50 (s, 1H), 7.83-7.78 (m, 1H), 7.45-7.40 (m, 1H), 7.30-7.23 (m, 2H), 4.56-4.50 (m, 2H), 4.32 (s, 2H), 3.82 (s, 3H), 1.22-1.16 (m, 2H), 0.10 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) 182.5; 171.3, 166.5, 142.3, 135.4, 126.1, 124.0, 123.4, 120.3, 112.3, 110.5, 65.2, 53.1, 32.9, 17.8, -1.1; LRMS-ES+ *m/z* (relative intensity) 362 (MH⁺, 80); HRMS-ES+ (C₁₈H₂₄NO₅Si) calcd 362.1424 (MH⁺), found 362.1416.



Synthesis of Nitrosoalkene Michael Adducts 11a/b. To a $-78\text{ }^{\circ}\text{C}$ solution of indole **5** (8.18 g, 22.6 mmol) was added LiHMDS (47.5 mL, 47.5 mmol, 1.0 M in THF) and the resulting solution was stirred for 30 min. A solution of α -chlorooxime **7** (9.6 g, 31.7 mmol) in THF (69 mL) was added via cannula over 10 min. The reaction mixture was stirred at $-78\text{ }^{\circ}\text{C}$ for 2 h and then diluted with $\text{NH}_4\text{Cl}_{(\text{aq})}$. The organic layer was separated and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over MgSO_4 and concentrated *in vacuo* to give a residue which was purified by flash chromatography on silica gel (gradient 20% to 50% EtOAc/hexanes) to yield diastereomeric Michael adducts **11a** and **11b** (14.01 g, 99%, ~1.2 : 1 by ^1H NMR) which were carried on to the next step without separation. ^1H NMR (300 MHz, CDCl_3) δ 10.31 (s, 0.5H), 10.23 (s, 0.5H), 8.90 (s, 0.5H), 8.66 (s, 0.5H), 7.79-7.58 (m, 3H), 7.39-7.17 (m, 5H), 5.59 (d, $J = 5.9$ Hz, 0.5H), 5.13 (d, $J = 8.6$ Hz, 0.5H), 4.93 (d, $J = 14.5$ Hz, 0.5H), 4.60-4.50 (m, 2H), 3.69-4.47 (m, 4H) 3.37 (p, $J = 5.7$ Hz, 0.5H), 3.20 (d, $J = 14.6$ Hz, 0.5 H); 3.10 (q, $J = 7.3$ Hz, 0.5H), 2.97-2.85 (m, 1H), 2.44-2.28 (m, 3H), 2.08-2.03 (m, 0.5H), 1.78-1.76 (m, 0.5H), 1.41-1.31 (m, 0.5H), 0.11-0.07 (m, 9H); ^{13}C NMR (75 MHz, CDCl_3) δ 183.6, 183.0, 172.6, 171.9, 166.5, 166.5, 152.8, 152.7, 144.61, 144.57, 143.5, 143.4, 135.83, 135.80, 130.4, 130.3, 128.06, 127.96, 125.8, 125.6, 124.4, 124.2, 123.4, 123.2, 120.0, 119.6, 112.9, 112.7, 111.2, 111.1, 65.33, 65.29, 53.3, 53.2, 45.4, 44.9, 44.2, 43.4, 42.9, 42.7,

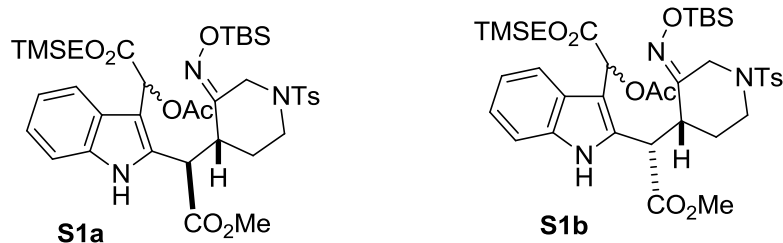
42.3, 41.9, 27.9, 27.8, 22.0, 21.9, 17.8, 14.6, 14.2, -1.1; LRMS-ES+ m/z (relative intensity) 666 ($M+K^+$, 100); HRMS-ES+ ($C_{30}H_{41}N_4O_8SSi$) calcd 645.2414 ($M+NH_4^+$), found 645.2445.

For characterization of the isomers, a small amount of the mixture was separated by flash chromatography on silica gel (gradient 2 to 10% Et_2O/CH_2Cl_2) to afford **11a** (less polar) as an orange foamy solid. 1H NMR (400 MHz, $CDCl_3$) δ 9.96 (s, 1H, NH), 7.91 (s, 1H), 7.62(d, $J = 8.0$ Hz, 3H), 7.38 (m, 1H), 7.26-7.19 (m, 4H), 5.52 (d, $J = 6.1$ Hz, 1H), 4.87 (d, $J = 14.6$ Hz, 1H), 4.50 (t, $J = 8.2$ Hz, 2H), 3.64 (s, 3H), 3.30 (m, 1H), 3.18 (d, $J = 14.7$ Hz, 1H), 2.82 (m, 1H), 2.43 (m, 1H), 2.34 (s, 3H), 2.02 (dd, $J = 3.4, 12.8$ Hz, 1H), 1.38 (m, 1H), 1.15 (t, $J = 8.7$ Hz, 2H), 0.07 (s, 9H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 183.2, 171.5, 166.0, 153.1, 144.2, 135.3, 133.0, 130.0, 127.7, 125.3, 123.9, 122.9, 119.6, 112.4, 111.1, 64.9, 52.9, 45.0, 43.0, 42.6, 42.0, 27.5, 21.6, 17.5, -1.4. **11b** (more polar, brown foamy solid), 1H NMR (300 MHz, $CDCl_3$) δ 9.84 (s, 1H), 7.97 (s, 1H), 7.61-7.57 (m, 3H), 7.29-7.23 (m, 2H), 1.17-7.14 (m, 3H), 5.00 (d, $J = 8.4$ Hz, 1H), 4.50-4.39 (m, 3H), 3.67 (s, 3H), 3.45-3.38 (m, 3H), 2.94 (m, 1H), 2.82 (m, 1H), 2.34 (s, 3H), 1.69 (m, 2H), 1.09-1.05 (m, 2H), 0.00 (s, 9H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 182.9, 172.8, 166.2, 152.9, 144.5, 143.5, 135.7, 133.3, 130.3, 128.1, 125.8, 124.4, 123.4, 120.2, 122.6, 111.2, 65.3, 53.3, 45.0, 44.1, 43.3, 42.0, 28.0, 22.0, 17.8, -1.1.



Synthesis of α -Ketoester **12.** To a solution of ester oxime mixture **11a/b** (5.66 g, 9.01 mmol) and imidazole (2.47 g, 36.3 mmol) in CH_2Cl_2 (240 mL) was added TBSCl (4.10 g, 27.2 mmol). The resulting suspension was stirred for 12 h at rt and then diluted with 1 M HCl. The organic layer was separated and the aqueous layer was extracted with CH_2Cl_2 . The combined organic layers were dried over MgSO_4 and concentrated *in vacuo* to give a residue which was purified by flash chromatography on silica gel (30% EtOAc/hexanes) yielding *O*-TBS-oximes **12** (5.98 g, 89%) as an orange solid. ^1H NMR (300 MHz, CDCl_3) δ 9.91 (s, 0.5H), 9.79 (s, 0.5H), 7.77-7.61 (m, 3H), 7.41-7.21 (m, 5H) 5.56 (d, $J = 5.7$, 0.5H), 5.06-4.97 (m, 1H), 4.84 (d, $J = 15.0$, 0.5H), 4.54-4.45 (m, 2H), 3.69-3.60 (m, 4H), 3.41 (dt, $J = 7.1, 12.6$ Hz, 0.5H), 3.30 (d, $J = 15.0$, 0.5H), 3.18 (d, $J = 14.9$, 0.5H), 3.00-2.92 (m, 1H), 2.85-2.70 (m, 0.5H), 2.42 (s, 1.5H) 2.32 (s, 1.5H), 2.06-1.98 (m, 0.5H), 1.85-1.70 (m, 0.5H), 1.65-1.55 (m, 0.5H), 1.40-1.35 (m, 1H), 1.20-1.13 (m, 2H), 0.99 (m, 9H), 0.31-0.20 (m, 6H), 0.10-0.07 (m, 9H); ^{13}C NMR (75 MHz, CDCl_3) δ 183.4, 182.8, 172.6, 171.0, 166.3, 166.1, 157.9, 156.8, 144.40, 144.37, 143.6, 143.4, 135.5, 135.4, 133.9, 133.6, 130.20, 130.15, 128.0, 127.9, 126.0, 125.7, 124.4, 124.2, 123.5, 123.2, 120.6, 120.0, 112.4, 112.2, 111.3, 65.1, 53.2, 53.0, 45.2, 44.1, 43.5, 43.3, 42.9, 28.1, 27.7, 26.34, 26.28, 21.93, 21.85, 18.3, 18.2, 17.79, 17.76, -1.1, -4.2, -4.6, -4.8; LRMS-ES+ m/z (relative intensity) 780 ($\text{M}+\text{K}^+$, 100); HRMS-ES+ ($\text{C}_{36}\text{H}_{52}\text{N}_3\text{O}_8\text{SSi}_2$) calcd 742.3014

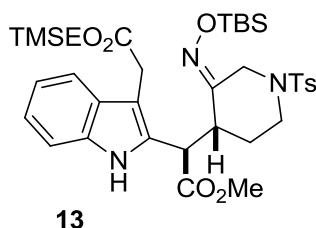
(MH⁺), found 742.3005.



Synthesis of Acetates S1a/b. To a stirred solution of α -ketoesters **12** (21.5 g, 29 mmol) in MeOH (55 mL) and THF (425 mL) cooled to 0 °C was added NaBH₄ (1.32 g, 34.7 mmol). The resulting solution was stirred for 1 h at 0 °C, and then diluted with NH₄Cl (aq). The organic layer was separated and the aqueous layer was extracted with CH₂Cl₂. The combined organic layers were washed with water and brine and then dried over MgSO₄. Removal of the solvent *in vacuo* provided the indole-3-hydroxyl acetate products as a white powder.

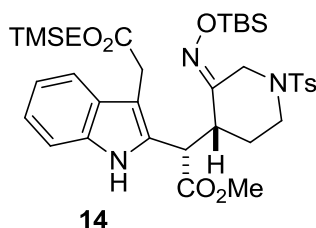
Without further purification, this mixture was dissolved in a 1:1 (v/v) mixture of Ac₂O:pyridine (292 mL) and the solution was stirred at rt for 12 h. After removing the volatiles *in vacuo*, **S1a** and **S1b** were separated by flash chromatography on silica gel (gradient 15% to 40% EtOAc/hexanes). **S1a** (more polar isomer, orange foamy solid, 12.60 g, 55%, ~3:1 mixture of acetoxy diastereomers): ¹H NMR (400 MHz, CDCl₃) δ 9.07 (s, 0.75H), 9.03 (s, 0.25H), 7.75-7.71 (m, 1H), 7.64 (d, *J* = 8.2 Hz, 2H), 7.32-7.13 (m, 5H), 6.21 (s, 0.25H), 6.18 (s, 0.75H), 4.97-4.88 (m, 1H), 4.52 (d, *J* = 5.9 Hz, 0.25H), 4.48 (d, *J* = 5.5 Hz, 0.75H), 4.28-4.20 (m, 1H), 4.09-4.02 (m, 1H), 3.67-3.55 (m, 4H), 3.41-3.31 (m, 2H), 3.00-2.93 (m, 1H), 2.39-2.36 (m, 3H), 2.14 (s, 3H), 1.95-1.70 (m, 3H), 1.60-1.40 (m, 2H), 0.98 (s, 9H), 0.29-0.18 (m, 6H), 0.00-0.02

(m, 9H); ^{13}C NMR (90 MHz, CDCl_3) δ 171.0, 170.8, 170.6, 170.5, 169.1, 157.3, 144.0, 135.3, 135.2, 133.7, 133.5, 132.2, 131.7, 129.8, 127.6, 126.2, 126.1, 122.7, 120.3, 119.3, 111.2, 108.0, 107.8, 67.7, 64.2, 53.4, 52.5, 44.8, 43.2, 42.9, 42.7, 42.4, 42.1, 28.0, 26.0, 25.9, 21.5, 20.9, 20.7, 17.9, 17.3, -1.56, -1.59, -4.8, -5.1; **S1b** (less polar isomer, orange foamy solid, 8.29 g, 36%, ~2:1 mixture of acetoxyl diastereomers): ^1H NMR (300 MHz, CDCl_3) δ 8.73-8.71 (m, 1H), 7.76 (d, $J = 7.6$ Hz, 1H), 7.66 (d, $J = 8.2$ Hz, 1H), 7.35-7.14 (m, 5H), 6.24-6.22 (m, 1H), 5.02 (t, $J = 7.1$ Hz, 1H), 4.34-4.07 (m, 3H), 3.66-3.53 (m, 4H), 3.15-3.07 (m, 2H), 2.72 (d, $J = 11.3$, 3.2 Hz, 1H), 2.45 (s, 3H), 2.20-2.18 (m, 3H), 1.58-1.39 (m, 2H), 1.00-0.89 (m, 12H), 0.26 (s, 3H), 0.20 (s, 3H), 0.02-0.04 (m, 9H); ^{13}C NMR (75 MHz, CDCl_3) δ 173.1, 173.0, 170.9, 170.8, 169.3, 169.2, 156.7, 144.4, 136.1, 136.0, 133.8, 133.6, 132.4, 132.2, 130.2, 128.1, 126.7, 123.5, 123.4, 121.1, 120.2, 111.5, 108.54, 108.50, 68.4, 68.0, 64.7, 64.5, 53.9, 53.0, 52.9, 45.4, 44.4, 44.1, 42.8, 42.6, 42.5, 28.5, 28.0, 26.4, 22.0, 21.2, 21.1, 18.4, 17.7, 17.6, -1.2, -4.8, -4.9 (2C); LRMS-ES+ (mixture of diastereomers **S1a/b**) m/z (relative intensity) 824 ($\text{M}+\text{K}^+$, 25); HRMS-ES+ ($\text{C}_{38}\text{H}_{56}\text{N}_3\text{O}_9\text{SSi}_2$) calcd 786.3276 (MH^+), found 786.3286.



Synthesis of Indole Diester 13. To a solution of acetate **S1a** (2.94 g, 3.75 mmol) in *t*-BuOH (80 mL) was added 10% Pd/C (1.20 g). The resulting mixture was

evacuated and backfilled with H₂ from a balloon and TEA (10.0 mL) was then added. The resulting mixture was warmed to 30 °C and stirred for 4 days until all the starting material was consumed as judged by TLC. The reaction mixture was then diluted with EtOAc, filtered through a pad of Celite and concentrated *in vacuo* to give a residue which was purified by flash chromatography on silica gel (gradient 10% to 25% EtOAc/hexanes) to afford indole **13** (2.47 g, 91%) as an off-white foam. ¹H NMR (360 MHz, CDCl₃) δ 8.74 (br s, 1H), 7.62 (d, *J* = 7.2 Hz, 2H), 7.57 (d, *J* = 7.8 Hz, 1H), 7.28-7.24 (m, 3H), 7.17 (dd, *J* = 7.1, 7.5 Hz, 1H), 7.09 (dd, *J* = 7.3, 7.4 Hz, 1H), 4.82 (d, *J* = 15.1 Hz, 1H), 4.33 (d, *J* = 6 Hz, 1H), 4.11-4.07 (m, 2H), 3.64 (s, 3H), 3.60 (d, *J* = 4.4 Hz, 2H), 3.58-3.55 (m, 1H), 3.36 (d, *J* = 15.2 Hz, 1H), 3.30-3.25 (m, 1H), 2.90 (td, *J* = 4.0, 11.9 Hz, 1H), 2.36 (s, 3H), 1.89-1.84 (m, 1H), 1.59-1.54 (m, 1H), 0.94(s, 9H), 0.21(s, 3H), 0.15(s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 171.7, 171.6, 157.2, 144.0, 135.4, 133.7, 130.0, 129.9, 129.5, 128.9, 127.7 (2C), 122.3, 119.7, 118.9, 111.0, 107.6, 63.2, 52.5, 44.7, 43.0, 42.7, 42.3, 30.8, 27.6, 26.0, 21.6, 18.0, 17.4, -1.4, -4.8, -5.0; LRMS-ES+ *m/z* (relative intensity) 766 (M+K⁺, 75); HRMS-ES+ (C₃₆H₅₇N₄O₇SSi₂) calcd 745.3487 (M+NH₄⁺), found 745.3478.

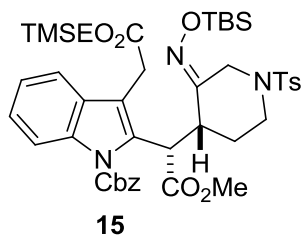


Synthesis of Indole Diester 14. Following a similar procedure described for preparation of **13**, acetate **S1b** (3.95 g, 5.03 mmol) was reduced by catalytic

hydrogenation to afford indole diester **14** as an off white foamy solid (3.16 g, 86%):

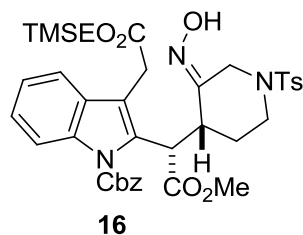
^1H NMR (400 MHz, CDCl_3) δ 8.19 (s, 1H), 7.65 (d, $J = 8.5$ Hz, 2H), 7.57 (d, $J = 7.7$ Hz, 1H), 7.32 (d, $J = 8.0$ Hz, 1H), 7.26 (d, $J = 7.6$ Hz, 1H), 7.17 (dd, $J = 7.0, 7.1$ Hz, 1H), 7.11 (dd, $J = 7.1, 7.2$ Hz, 1H), 5.03 (d, $J = 14.4$ Hz, 1H), 4.17-4.13 (m, 2H), 4.06 (d, $J = 10.7$ Hz, 1H), 3.64 (app. s, 2H), 3.62 (s, 3H), 3.57 (m, 1H), 3.06-3.00 (m, 2H), 2.66 (td, $J = 3.8, 11.5$ Hz, 1H), 2.44 (s, 3H), 1.46 (m, 1H), 1.39 (m, 1H), 0.97 (s, 9H), 0.23 (s, 3H), 0.16 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 173.0, 171.4, 156.5, 144.0, 135.7, 133.4, 129.8, 128.0 (2C), 127.8, 122.7, 120.1, 119.2, 111.0, 108.3, 63.3, 52.5, 45.2, 43.8, 42.5, 42.0, 30.6, 27.8, 26.1, 25.9, 21.7, 18.1, 17.5, -1.4, -5.1, -5.2; HRMS-ES $^+$ ($\text{C}_{36}\text{H}_{54}\text{N}_3\text{O}_7\text{SSi}_2$) calcd 728.3221 (MH^+), found 728.3224.

Epimerization of Ester 13 to Ester 14. To a solution of diester **13** (4.45 g, 6.12 mmol) in THF (81 mL) cooled to -78°C was added dropwise KHMDS solution (0.5 M in toluene, 13.5 mL, 6.73 mmol, 1.1 equiv.). After the addition was complete, the cooling bath was removed and the reaction mixture was stirred at rt for 30 min. To the resultant olive green solution was added glacial acetic acid (0.40 mL) followed by saturated NH_4Cl (aq). The mixture was extracted with EtOAc. The organic phase was washed with brine, dried over Na_2SO_4 and concentrated. The residue was purified by flash chromatography on silica gel (30% EtOAc/hexanes) to afford diester **14** as a slightly yellow foam (3.23 g, 73%), which was identical to ester **14** prepared from hydrogenation of **S1b**.



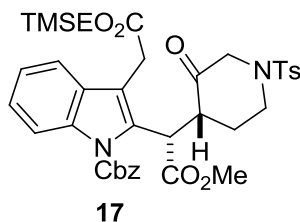
Synthesis of *N*-Cbz Derivative **15.** A stirred solution of indole **14** (1.05 g, 1.45 mmol) in acetonitrile (40 mL) was heated to 90 °C. Dibenzyl dicarbonate (2.07 g, 7.24 mmol) was added, followed immediately by DMAP (0.53 g, 4.33 mmol). After gas evolution stopped (~1 min), the reaction mixture was heated at 90 °C for another 2 min, and cooled to rt. The solvent was removed *in vacuo* to give a pale brown residue, which was purified by flash chromatography on silica gel (5 to 20 % EtOAc/hexanes) to afford **15** as a white foam (1.23 g, 99%, ~2:1 Cbz rotamers). ¹H NMR (400 MHz, CDCl₃) δ 8.11 (d, *J* = 7.6 Hz, 0.7H), 8.00 (m, 0.3H), 7.64 (d, *J* = 6.9 Hz, 2H), 7.57 (d, *J* = 6.4 Hz, 0.7H), 7.46-7.26 (m, 9H), 5.68 (d, *J* = 10.1 Hz, 0.3H), 5.17 (d, *J* = 11.7 Hz, 0.7H), 5.52-5.46 (m, 0.7H), 5.29 (d, *J* = 13.1 Hz, 0.7H), 5.16 (d, *J* = 9.3 Hz, 1.7H), 4.14-4.10 (m, 2H), 3.90 (d, *J* = 15.9 Hz, 0.3H), 3.65-3.53 (m, 2.7H), 3.48 (s, 3H), 3.11 (m, 1H), 2.90 (d, *J* = 13.2 Hz, 0.3H), 2.68 (d, *J* = 13.2 Hz, 0.7H), 2.60 (m, 0.3H), 2.49-2.44 (m, 3H), 2.17 (m, 0.7H), 1.40 (m, 0.3H), 1.12 (m, 1.7H), 0.98 (m, 9H), 0.22 (s, 2H), 0.19-0.17 (m, 4H), 0.02 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 171.7, 171.1, 170.8, 170.1, 155.8, 155.6, 151.8, 151.3, 143.8, 135.9, 135.6, 134.8, 133.4, 132.5, 132.2, 129.8, 129.4, 129.2, 129.0, 128.8, 127.9, 125.3, 123.5, 119.3, 118.9, 118.6, 117.2, 115.8, 69.3, 68.6, 63.6, 63.4, 52.1, 45.7, 42.8, 42.4, 42.0, 41.3, 40.7, 30.9, 30.2, 28.6, 27.5, 26.2, 25.6, 21.7, 18.3, 17.5, -1.5, -5.1; LRMS-ES+ *m/z* (relative intensity) 862 (MH⁺, 90); HRMS-ES⁺ (C₄₄H₆₀N₃O₉SSi₂) calcd 862.3589 (MH⁺), found

862.3585.

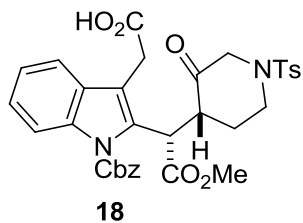


Synthesis of Oxime 16. To a stirred solution of *O*-TBS-oxime **15** (12.9 g, 14.9 mmol) in THF (600 mL) was added AcOH (6.6 mL) followed by TBAF (22.1 mL, 22.1 mmol, 1.0 M in THF). The resulting solution was stirred at rt overnight and then diluted with NH₄Cl (aq). The organic layer was separated and the aqueous layer was extracted with EtOAc. The combined organic layers were dried over MgSO₄ and concentrated *in vacuo* to give a residue which was purified by flash column chromatography on silica gel (30% EtOAc/hexanes) to yield free oxime **16** as a pale foam (11.1 g, 100%, ~2:1 Cbz rotamers). ¹H NMR (400 MHz, CDCl₃) δ 8.10 (m, 0.7H), 8.0-7.8 (m, 1.3H), 7.65 (d, *J* = 7.1 Hz, 2H), 7.54 (m, 0.7H), 7.45 (m, 3.5H), 7.35-7.26 (m, 6H), 5.68 (m, 0.3H), 5.57 (d, *J* = 11.5 Hz, 0.7H), 5.47 (s, 0.7H), 5.18 (m, 1.4H), 5.07 (m, 1H), 4.13 (m, 2H), 3.87 (m, 0.3H), 3.65-3.59 (m, 3H), 3.48 (s, 3H), 3.09 (m, 1H), 2.91 (0.3H), 2.68 (d, *J* = 13.1 Hz, 0.7H), 2.60 (m, 0.3H), 2.48-2.44 (m, 3H), 2.19 (m, 0.7H), 1.64 (m, 0.4H), 1.40 (m, 0.3H), 1.17 (1.60H), 0.94 (t, *J* = 8.8 Hz, 2H), 0.01 (s, 9H); ¹³C NMR (75 MHz, CDCl₃) δ 172.0, 171.5, 170.8, 170.5, 152.9, 151.8, 151.4, 143.9, 135.8, 135.5, 134.7, 133.0, 132.3, 132.0, 129.8, 129.4, 129.2, 129.1, 129.0, 128.8, 128.6, 127.9, 127.8, 127.1, 125.3, 124.9, 123.5, 123.2, 119.3, 118.6, 117.2, 116.1, 115.9, 69.5, 68.7, 65.4, 64.5, 63.8, 63.4, 52.6, 52.3, 45.7, 42.4, 41.8, 41.0, 40.7, 40.5, 31.0, 30.7, 30.3, 28.5, 27.6, 21.6, 21.1, 19.2, 17.3, 13.8,

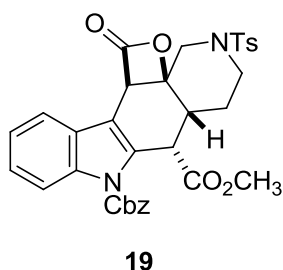
-1.5; LRMS-ES+ m/z (relative intensity) 748 (MH^+ , 75); HRMS-ES+ ($C_{38}H_{46}N_3O_8SSi_2$) calcd 748.2724 (MH^+), found 748.2690.



Synthesis of Ketone 17. Oxime **16** (11.14 g, 14.9 mmol) was added to a mixture of levulinic acid and 1 M HCl (334 g, 9:1 v/v) and the mixture was stirred at 30 °C for 4.5 h. The reaction mixture was diluted with water (1L), and was extracted with dichloromethane. The organic layer was washed with sat. $NaHCO_3$, water and brine, dried over Na_2SO_4 , and concentrated. The residue was purified by flash column chromatography on silica gel (gradient, 20 to 30% EtOAc/hexanes) to afford ketone **17** as a slightly pink foam (10.04 g, 92%). 1H NMR (400 MHz, $CDCl_3$) δ 8.06 (d, $J = 7.6$ Hz, 1H), 7.62 (d, $J = 8.1$ Hz, 2H), 7.55-7.26 (m, 10H), 5.54 (d, $J = 11.8$ Hz, 1H), 5.20 (d, $J = 11.6$ Hz, 1H), 4.91 (d, $J = 6.6$ Hz, 1H), 4.18-4.07 (m, 2H), 4.21 (d, $J = 13.6$ Hz, 1H), 3.62 (s, 2H), 3.48 (s, 3H), 3.32 (q, $J = 9.3$ Hz, 1H), 3.20 (d, $J = 13.6$ Hz, 1H), 2.48 (s, 3H), 2.46 (m, 1H), 1.46-1.42 (m, 2H), 0.95 (t, $J = 8.6$ Hz, 2H), 0.02 (s, 9H); ^{13}C NMR (75 MHz, $CDCl_3$) δ 201.6, 171.2, 170.2, 151.5, 144.3, 135.4, 134.7, 132.2, 131.7, 130.0, 129.3, 129.1, 128.9, 128.0, 125.4, 123.7, 119.3, 117.4, 115.9, 68.9, 63.8, 56.1, 52.4, 48.8, 45.3, 40.0, 30.8, 27.5, 21.7, 17.4, -1.5; LRMS-ES+ m/z (relative intensity) 750 ($M+NH_4^+$, 75); HRMS-ES+ ($C_{38}H_{48}N_3O_9SSi$) calcd 750.2881 ($M+NH_4^+$), found 750.2878.

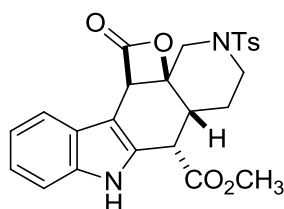


Synthesis of Keto Acid 18. To a stirred solution of TMSE-ester **17** (3.95 g, 5.38 mmol) in CH₂Cl₂ (138 mL) was added TFA (34 mL). The resulting solution was stirred at rt for 9 h and the solvent was removed *in vacuo* to give a residue which was purified by flash column chromatography on silica gel (40% EtOAc/hexanes + 1% AcOH) to give keto acid **18** (3.39 g, 100%). ¹H NMR (300 MHz, CDCl₃) δ 8.07 (d, *J* = 6.9 Hz, 1H), 7.62 (d, *J* = 8.0 Hz, 2H), 7.50-7.16 (m, 10H), 5.54 (d, *J* = 11.8 Hz, 1H), 5.21 (d, *J* = 11.1 Hz, 1H), 4.87 (d, *J* = 6.2 Hz, 1H), 4.00 (13.4 Hz, 1H), 3.68 (s, 2H), 3.46 (s, 3H), 3.36-3.29 (m, 1H), 3.18 (d, *J* = 13.9 Hz, 1H), 2.48 (s, 3H), 1.41 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 201.6, 175.3, 171.1, 151.4, 144.3, 137.9, 135.4, 134.5, 132.2, 131.9, 129.9, 129.3, 129.1, 128.8, 128.2, 127.9, 125.5, 125.3, 123.7, 119.1, 116.6, 115.9, 68.9, 56.0, 52.4, 48.7, 45.2, 40.0, 30.0, 27.5, 21.6; LRMS-ES+ *m/z* (relative intensity) 633 (MH⁺, 100); HRMS-ES+ (C₃₃H₃₆N₃O₉S) calcd 650.2172 (M+NH₄⁺), found 650.2142.

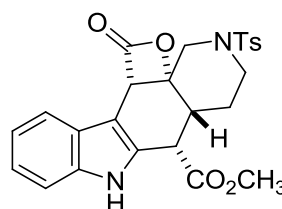


Synthesis of β-Lactone 19. To a stirred suspension of 4-PPY (1.09 g, 7.35 mmol), 2-bromo-*N*-propylpyridinium triflate (2.57 g, 7.35 mmol) and DIPEA (1.7 mL,

9.8 mmol) in CH₂Cl₂ (79 mL) at rt was added a solution of keto acid **18** (3.00 g, 4.90 mmol) and glacial acetic acid (0.35 mL, 6.1 mmol) in CH₂Cl₂ over 1 h *via* a syringe pump. The resultant orange solution was stirred for another 3 h at rt. Solvent was then removed *in vacuo* to give a residue which was purified by flash chromatography on silica gel (gradient, 30-40% EtOAc/hexanes) affording β -lactone **19** as an off white solid (2.71 g, 93%). FTIR (film) 1835 cm⁻¹; ¹H NMR (400 MHz, CDCl₃, ~97:3 mixture of diastereomers determined by ¹H NMR, only the major diastereomer peaks are reported) δ 8.02 (d, *J* = 7.2 Hz, 1H), 7.66 (d, *J* = 7.8 Hz, 2H), 7.60 (d, *J* = 7.4 Hz, 1H), 7.45 (m, 2H), 7.42-7.31 (m, 7H), 5.46 (d, *J* = 10.8 Hz, 1H), 5.32 (d, *J* = 12.0 Hz, 1H), 4.76 (s, 1H), 4.41 (d, *J* = 5.7 Hz, 1H), 3.88 (d, *J* = 12.0 Hz, 1H), 3.50 (m, 1H), 3.50 (s, 3H), 2.95 (d, *J* = 11.8 Hz, 1H), 2.55 (dd, *J* = 7.3, 14.5 Hz, 1H), 1.60 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 170.7, 165.4, 151.8, 144.5, 136.6, 134.5, 132.3, 130.1, 129.9, 129.0, 128.9, 128.8, 127.8, 126.9, 125.8, 123.8, 118.8, 115.6, 110.6, 75.6, 69.4, 53.0, 52.2, 50.9, 44.8, 43.3, 38.5, 25.1, 21.7; LRMS-ES+ *m/z* (relative intensity) 615 (MH⁺, 100). HRMS-ES+ (C₃₃H₃₄N₃O₈S) calcd 632.2067 (M+NH₄⁺), found 632.2053.



20

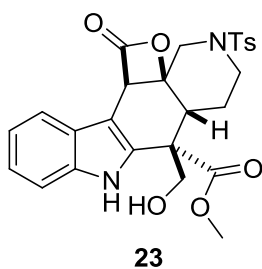


21

Synthesis of NH-Indole Esters 20 and 21. 10% Pd/C (0.54 g) was suspended in a solution of *N*-Cbz β -lactone **19** (2.71 g, 4.41 mmol) in EtOAc (280 mL). One drop

of glacial acetic acid was added to the mixture, followed by three evacuation-backfill cycles with hydrogen gas from a balloon. The reaction mixture was stirred under a balloon of H₂ for 1.5 h at rt. The mixture was filtered through a pad of Celite, concentrated and the residue was purified by flash chromatography on silica gel (gradient, 2 to 5% Et₂O/CH₂Cl₂) to afford indole **20** (1.99 g, 94%) as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 9.50 (br s, 1H), 7.66 (d, *J* = 8.0 Hz, 2H), 7.62 (d, *J* = 7.6 Hz, 1H), 7.42 (d, *J* = 7.8 Hz, 1H), 7.36 (d, *J* = 8.0 Hz, 2H), 7.25-7.18 (m, 2H), 4.86 (s, 1H), 4.21 (d, *J* = 4.5 Hz, 1H), 4.09 (d, *J* = 11.6 Hz, 1H), 3.86 (s, 3H), 3.72 (dd, *J* = 2.0, 9.6 Hz, 1H), 2.78 (d, *J* = 11.6 Hz, 1H), 2.71 (ddd, *J* = 4.5, 12.6 Hz, 1H), 2.46 (s, 3H), 2.30 (td, *J* = 2.5, 12.0 Hz, 1H), 1.58 (m, 1H), 1.40 (qd, *J* = 4.5, 13.0 Hz, 1H); ¹³C NMR (75 MHz, CDCl₃) δ 171.5, 166.1, 144.5, 136.1, 132.6, 130.1, 129.1, 127.9, 125.7, 122.8, 120.7, 118.3, 111.7, 101.4, 53.6, 53.0, 51.7, 45.4, 39.5, 39.1, 24.5, 21.7; HRMS (*m/z*): [M + NH₄]⁺ calcd for C₂₅H₂₈N₃O₆S, 498.1693; found, 498.1663.

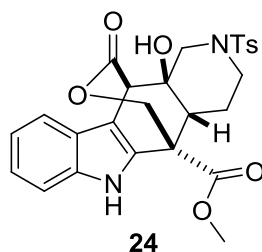
Indole **21** (65 mg, 3 %) was obtained as a white solid. ¹H NMR (300 MHz, DMSO-*d*₆) δ 7.69 (d, *J* = 8.1 Hz, 2H), 7.61 (d, *J* = 7.8 Hz, 1H), 7.45 (d, *J* = 8.1 Hz, 2H), 7.40 (d, *J* = 8.0 Hz, 1H), 7.15 (dd, *J* = 7.1, 7.4 Hz, 1H), 7.08 (dd, *J* = 7.4, 7.5 Hz, 1H), 4.84 (s, 1H), 4.03-4.00 (m, 2H), 3.51 (br s, 4H), 2.94 (dd, *J* = 3.6, 12.6 Hz, 1H), 2.88 (d, *J* = 11.6 Hz, 1H), 2.46-2.43 (m, 1H), 2.41 (s, 3H), 1.95 (br d, *J* = 10.1 Hz, 1H), 1.08 (qd, *J* = 4.3, 12.9 Hz, 1H); ¹³C NMR (75 MHz, DMSO-*d*₆) δ 170.9, 166.9, 143.9, 136.2, 132.6, 130.0, 127.5, 125.4, 121.6, 119.3, 117.9, 111.5, 100.2, 76.8, 52.4, 52.1, 50.7, 44.8, 42.8, 38.8, 30.7, 28.7, 21.0; HRMS (*m/z*): [M + H]⁺ calcd for C₂₅H₂₅N₂O₆S, 481.1428; found 481.1395.



Synthesis of α -Hydroxymethyl Ester **23.** To a solution of indole β -lactone **20** (255 mg, 0.531 mmol) in THF (25 mL) cooled to $-78\text{ }^{\circ}\text{C}$ was added a solution of LiHMDS (1.0 M in THF, 1.60 mL, 1.60 mmol) dropwise with stirring. The resulting orange red solution was stirred at $-78\text{ }^{\circ}\text{C}$ for another 30 min. A solution of freshly distilled monomeric formaldehyde¹⁵ in THF ($\sim 0.5\text{ M}$, 10.6 mL, 5.3 mmol) was added dropwise. The resulting brownish red solution was stirred at $-78\text{ }^{\circ}\text{C}$ for 5 min, then warmed to $-40\text{ }^{\circ}\text{C}$ and stirred for another 15 min. The reaction mixture was then quenched at $-40\text{ }^{\circ}\text{C}$ by addition of glacial acetic acid (0.10 mL, 1.6 mmol). The bright yellow solution was diluted with dichloromethane, quickly washed with ice cold water and dried over Na_2SO_4 . After concentration of the solution, the residue was purified by column chromatography on silica gel (5% $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$, then 50% $\text{EtOAc}/\text{hexanes}$) to afford α -hydroxymethyl ester **23** as an off-white solid (160 mg, 59%). FT-IR (ATR) 3403, 1824 cm^{-1} ; ^1H NMR (400 MHz, CDCl_3) δ 9.43 (s, 1H, NH), 7.62-7.66 (m, 4H), 7.34-7.42 (m, 4H), 7.16-7.25 (m, 2H), 4.86 (s, 1H), 3.97-4.02 (m, 3H), 3.83 (s, 3H), 3.68 (app. d, $J = 15.6\text{ Hz}$, 1H), 2.74 (d, $J = 15.7\text{ Hz}$, 1H), 2.57 (dd, $J = 6.4, 15.9\text{ Hz}$, 1H), 2.46 (s, 3H), 2.26 (td, $J = 4.2, 15.7\text{ Hz}$, 1H), 2.13 (t, $J = 8.2\text{ Hz}$, 1H), 1.43-1.58 (m, 2H); ^{13}C NMR (100 MHz, CDCl_3) δ 172.4, 167.1, 144.7, 136.6, 132.9, 131.6, 130.2, 128.0, 125.7, 123.2, 120.8, 118.6, 111.8, 101.0, 69.5, 55.4, 54.1, 53.1, 51.7, 45.3, 40.7, 26.8, 21.8. ESI MS (m/z): $[\text{M} + \text{H}]^+$ 511.3; HRMS-ES (m/z):

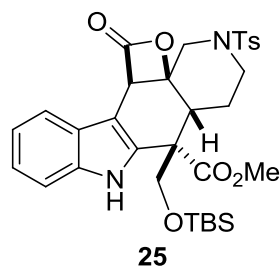
S17

$[M + H]^+$ calcd for $C_{26}H_{27}N_2O_7S$, 511.1539; found, 511.1538.

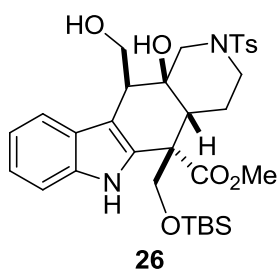


Synthesis of ϵ -Lactone **24.** To a solution of β -lactone **23** (200 mg, 0.39 mmol) in dichloromethane (2.6 mL) at rt was added triethylamine (10.6 mL, excess). The resulting slightly yellow solution was stirred at rt for 2 h, evaporated to dryness and the residue was purified by flash column chromatography on silica gel (5% Et_2O/CH_2Cl_2 , then 50% $EtOAc$ /hexanes) to afford ϵ -lactone **24** as an off white solid (65 mg, 33%). The recovered β -lactone **23** was subjected to another translactonization under the same conditions. After 2 runs, 40 mg of β -lactone **23** was recovered and ϵ -lactone **24** was isolated as an off white foam (92 mg, BRSM, 58%). FT-IR (ATR) 3407, 1725 cm^{-1} ; 1H NMR (400 MHz, $CDCl_3$) δ 9.61 (s, 1H, NH), 7.61 (d, $J = 8.2$ Hz, 2H), 7.53 (d, $J = 7.5$ Hz, 1H), 7.41 (d, $J = 8.0$ Hz, 1H), 7.25 – 7.27 (m, 4H), 7.21 (t, $J = 7.4$ Hz, 1H), 7.15 (t, $J = 7.5$ Hz), 4.90 (d, $J = 11.7$ Hz, 1H), 4.37 (d, $J = 11.7$ Hz, 1H), 4.16 (s, 1H), 3.93 (s, 3H), 3.65 (br s, 1H), 3.58 (d, $J = 13.5$ Hz, 1H), 3.52-3.58 (m, 1H), 3.01 (m, 1H), 2.81 (dd, $J = 3.8, 13.6$ Hz, 1H), 2.59 (d, $J = 13.6$ Hz, 1H), 2.37 (s, 3H), 1.40 (m, 1H), 0.76 (m, 1H); ^{13}C NMR (100 MHz, $CDCl_3$) δ 171.0, 167.8, 144.2, 135.9, 134.4, 132.0, 130.2, 127.4, 124.9, 122.8, 120.9, 118.0, 113.7, 112.1, 107.0, 76.0, 73.6, 53.7, 53.0, 52.8, 52.2, 50.6, 42.4, 23.8, 21.7; ESI MS (m/z): $[M + H]^+$ 511.3; HRMS (m/z): $[M + NH_4]^+$ calcd for $C_{26}H_{30}N_3O_7S$, 528.1799; found,

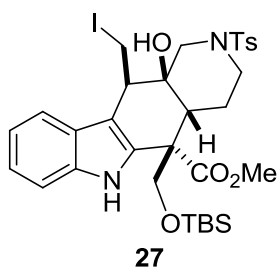
528.1797.



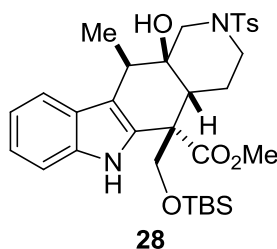
Synthesis of *O*-TBS β -Lactone **25.** To a solution of hydroxymethyl compound **23** (100 mg, 0.196 mmol) in CH_2Cl_2 (15 mL) at 0 °C was added 2,6-lutidine (226 μL , 1.95 mmol) and freshly distilled TBSOTf (225 μL , 0.98 mmol) with stirring. The colorless reaction mixture was stirred at 0 °C and monitored by TLC. Once the reaction was complete (~40 min), the solvent was removed *in vacuo*. The residue was purified by flash chromatography on silica gel (gradient, 5% to 30% EtOAc/hexanes) to afford *O*-TBS ether **25** as a white foam (105 mg, 86%). ^1H NMR (400 MHz, CDCl_3) δ 9.32 (br s, 1H, NH), 7.66-7.63 (m, 3H), 7.40 (d, $J = 8.0$ Hz, 1H), 7.36 (d, $J = 7.7$ Hz, 2H), 7.23 (m, 1H), 7.19 (dd, $J = 7.5, 7.6$ Hz, 1H), 4.84 (s, 1H), 4.03 (d, $J = 10.1$ Hz, 1H), 4.00 (d, $J = 12.4$ Hz, 1H), 3.88 (d, $J = 9.2$ Hz, 1H), 3.81 (s, 3H), 3.70 (m, 1H), 2.74 (d, $J = 11.6$ Hz, 1H), 2.55 (dd, $J = 4.7, 12.1$ Hz, 1H), 2.46 (s, 3H), 2.27 (td, $J = 3.0, 11.8$ Hz, 1H), 1.55-1.47 (m, 2H), 0.80 (s, 9H), -0.17 (s, 3H), -0.33 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 172.5, 167.4, 144.8, 136.4, 132.7, 132.4, 130.4, 128.2, 125.7, 123.1, 120.7, 118.7, 111.7, 100.4, 70.3, 55.8, 54.4, 53.0, 51.9, 45.5, 41.3, 27.1, 26.1, 25.9, 22.0, 18.4, -5.6; HRMS (m/z): $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{32}\text{H}_{41}\text{N}_2\text{O}_7\text{SSi}$, 625.2404; found 625.2390.



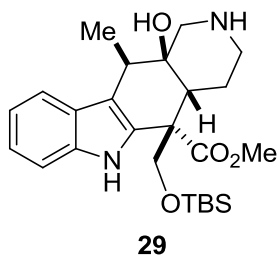
Synthesis of Indole Diol Ester 26. To a solution of *O*-TBS β -lactone **25** (120 mg, 0.193 mmol) in THF at rt was added LiBH_4 (22 mg, 1.0 mmol). The reaction mixture was stirred at rt for 12 h, diluted with $\text{NH}_4\text{Cl}_{(\text{aq})}$ and extracted with CH_2Cl_2 . The extract was dried over Na_2SO_4 and the solvent was removed *in vacuo* to give a residue which was purified by flash chromatography on silica gel (2% to 5% $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$) affording diol **26** as a white solid (90 mg, 75%). ^1H NMR (400 MHz, CDCl_3) δ 8.76 (br s, 1H), 7.69-7.66 (m, 3H), 7.39 (d, $J = 7.9$ Hz, 1H), 7.34 (d, $J = 7.7$ Hz, 2H), 7.22 (dd, $J = 7.6, 7.8$ Hz, 1H), 7.14 (dd, $J = 7.1, 7.4$ Hz, 1H), 4.87 (d, $J = 12.2$ Hz, 1H), 4.44 (d, $J = 12.9$ Hz, 1H), 4.23 (d, $J = 8.7$ Hz, 1H), 4.08 (d, $J = 9.5$ Hz, 2H), 3.71 (s, 3H), 3.67 (m, 1H), 2.45 (s, 3H), 2.25 (dd, $J = 10.5, 10.8$ Hz, 1H), 2.17 (d, $J = 11.8$ Hz, 1H), 1.92 (d, $J = 12.2$ Hz, 1H), 1.59 (m, 1H), 1.40 (m, 1H), 0.87 (s, 9H), -0.05 (s, 3H), -0.11 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 173.4, 144.3, 137.0, 135.0, 133.2, 130.2, 128.1, 125.8, 122.3, 120.2, 119.2, 111.9, 103.8, 74.5, 71.8, 61.0, 55.8, 54.9, 52.5, 47.6, 46.1, 37.4, 26.4, 26.2, 22.0, 18.5, -5.2, -5.5; HRMS (m/z): $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{32}\text{H}_{45}\text{N}_2\text{O}_7\text{SSi}$, 629.2717; found 629.2717.



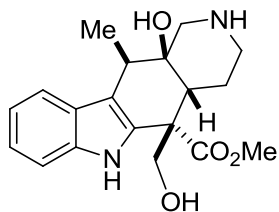
Synthesis of Iodo Alcohol 27. To a solution of PPh_3 (229 mg, 0.87 mmol) and I_2 (222 mg, 0.87 mmol) in CH_2Cl_2 (9 mL) at rt was added imidazole (99 mg, 1.48 mmol). The resulting yellow suspension was stirred at rt for 10 min. A solution of diol **26** (122 mg, 0.195 mmol) in CH_2Cl_2 (15 mL) was added dropwise, and the bright yellow suspension was stirred at rt for 5 h. The mixture was diluted with CH_2Cl_2 and washed with 5% $\text{Na}_2\text{S}_2\text{O}_3$. The aqueous layer was extracted with CH_2Cl_2 . The combined organic layers were washed with brine, dried over Na_2SO_4 , and the solvent was removed *in vacuo*. The residue was purified by flash column chromatography on silica gel (2% $\text{Et}_2\text{O}/\text{CH}_2\text{Cl}_2$) to afford iodo alcohol **27** as a white foamy solid (133 mg, 93%). ^1H NMR (400 MHz, CDCl_3) δ 8.80 (br s, 1H), 7.67 (d, $J = 8.1$ Hz, 2H), 7.63 (d, $J = 7.7$ Hz, 1H), 7.37-7.33 (m, 3H), 7.20 (dd, $J = 7.1, 7.5$ Hz, 1H), 7.14 (dd, $J = 7.4, 7.4$ Hz, 1H), 4.17-4.09 (m, 2H), 4.00-3.96 (m, 1H), 3.90-3.88 (m, 2H), 3.68 (s, 3H), 3.30 (s, 1H), 2.44 (s, 3H), 2.30 (m, 1H), 2.18 (m, 1H), 2.08 (dd, $J = 3.1, 11.5$ Hz, 1H), 1.63 (m, 1H), 1.43 (m, 1H), 0.84 (s, 9H), -0.08 (s, 3H), -0.10 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 172.9, 144.0, 136.5, 132.7, 132.3, 132.1, 129.9, 128.7, 127.9, 126.0, 122.0, 119.6, 111.4, 108.9, 72.9, 71.0, 54.8, 54.3, 52.3, 45.6, 36.4, 25.8, 21.6, 18.2, 4.8, 1.1, -5.6, -5.7; HRMS (m/z): $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{32}\text{H}_{44}\text{IN}_2\text{O}_6\text{SSi}$, 739.1734; found, 739.1764.



Synthesis of Methyl Compound 28. 10% Pd/C (158 mg) was suspended in a stirred solution of iodide **27** (29 mg, 0.039 mmol) in *t*-BuOH/EtOAc (1:1 v/v, 30 mL). The reaction mixture was evacuated and backfilled with H₂ three times from a balloon and stirred under a H₂ atmosphere at rt. After 3.5 h, the reaction was evacuated and back filled with argon, and another portion of 10% Pd/C (32 mg) was added and the reaction mixture was stirred under H₂ at rt for another 12 h. The Pd/C was filtered off through a pad of Celite, and the solvent was removed *in vacuo* to afford a yellow foamy solid. This material was purified by flash column chromatography on silica gel (CH₂Cl₂, then 2% Et₂O/CH₂Cl₂) to afford methyl compound **28** as a white solid (23 mg, 96%). ¹H NMR (400 MHz, CDCl₃) δ 8.51 (br s, 1H), 7.72 (d, *J* = 7.9 Hz, 1H), 7.67 (d, *J* = 8.1 Hz, 2H), 7.34-7.32 (m, 3H), 7.17 (dd, *J* = 7.1, 7.6 Hz, 1H), 7.08 (dd, *J* = 7.2, 7.4 Hz, 1H), 4.13 (d, *J* = 9.4 Hz, 1H), 4.00 (d, *J* = 9.4 Hz, 1H), 3.96 (d, *J* = 11.8 Hz, 1H), 3.72 (s, 3H), 3.66 (q, *J* = 7.0 Hz, 1H), 3.09 (s, 1H), 2.45 (s, 3H), 2.26 (t, *J* = 10.4 Hz, 1H), 2.04 (d, *J* = 13.0 Hz, 1H), 2.00 (dd, *J* = 3.7, 12.6 Hz, 1H), 1.63 (qd, *J* = 4.4, 12.9 Hz, 1H), 1.57 (d, *J* = 6.9 Hz, 3H), 0.80 (s, 3H), -0.07 (s, 3H), -0.12 (s, 3H); ¹³C NMR (75 MHz, CDCl₃) δ 173.3, 143.9, 137.1, 133.0, 130.0, 129.9, 129.8, 127.9, 126.7, 121.7, 120.5, 119.1, 111.4, 111.2, 72.5, 70.7, 55.1, 53.4, 52.3, 48.2, 46.0, 30.5, 26.5, 25.8, 25.7, 21.7, 18.1, 12.8, -5.7, -5.8; HRMS (*m/z*): [M + H]⁺ calcd for C₃₂H₄₅N₂O₆SSi, 613.2768; found, 613.2772.

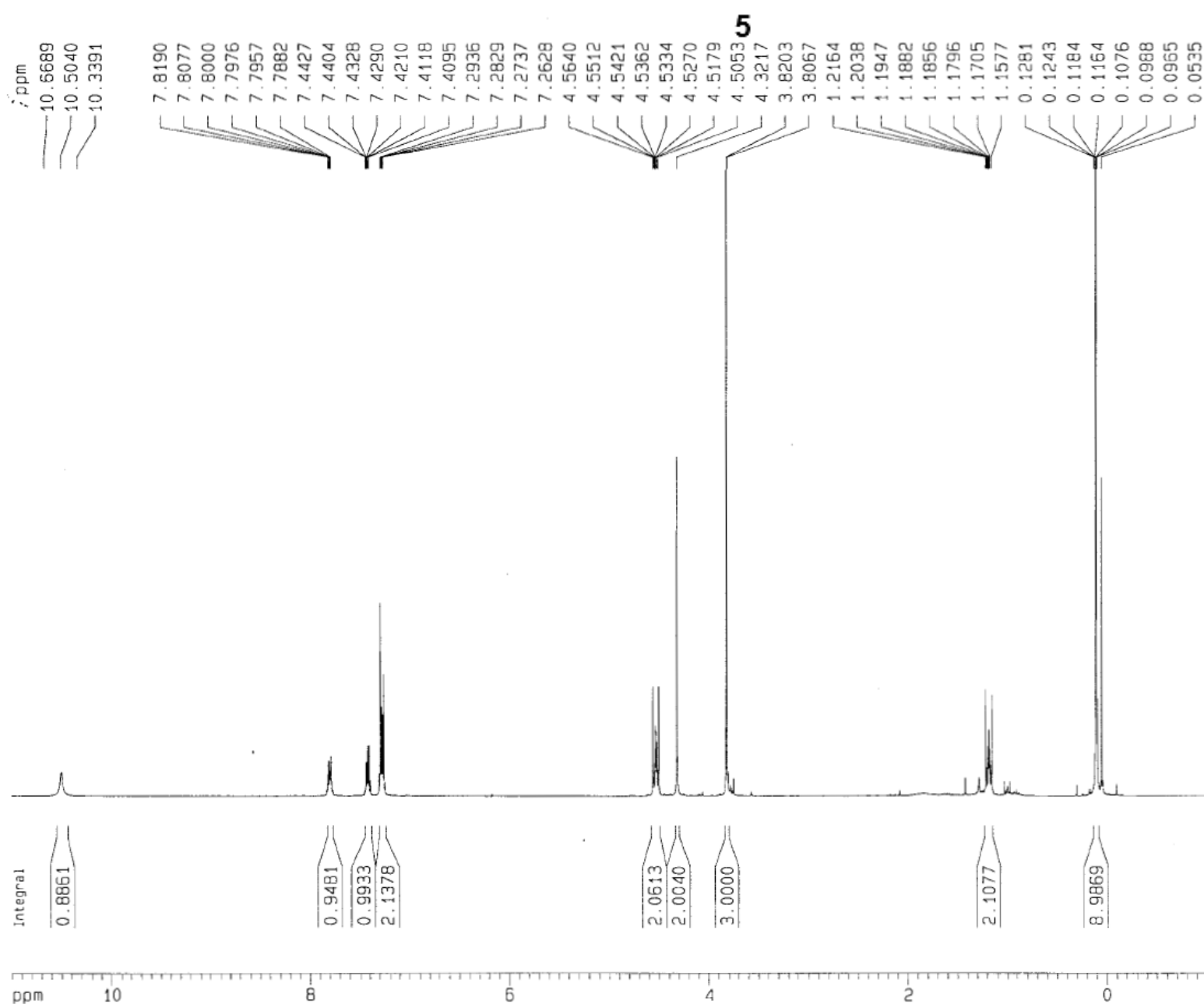
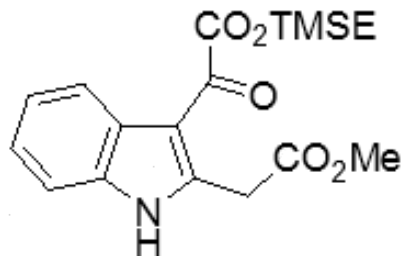


Synthesis of Piperidine 29. Magnesium turnings (399 mg, excess) were added to a solution of compound **28** (21.6 mg, 0.035 mmol) in anhydrous methanol, and the mixture was sonicated at rt until all the magnesium turnings dissolved (~45 min). The reaction mixture was then poured into $\text{NH}_4\text{Cl}_{(\text{aq})}$ at 0 °C, and extracted with CHCl_3 . The organic extract was dried over Na_2SO_4 , and evaporated to dryness *in vacuo* to give a white foamy solid. This material was purified by flash chromatography on silica gel (2% to 5% $\text{MeOH}/\text{CHCl}_3$, then 1% NEt_3 in 5% $\text{MeOH}/\text{CHCl}_3$) to afford piperidine **29** as a white solid (15.6 mg, 96%). ^1H NMR (400 MHz, CD_3OD) δ 7.90 (br s, 1H), 7.66 (d, $J = 7.8$ Hz, 1H), 7.33 (d, $J = 8.0$ Hz, 1H), 7.08 (dd, $J = 7.2, 7.4$ Hz, 1H), 6.98 (dd, $J = 7.2, 7.3$ Hz, 1H), 4.34 (d, $J = 9.3$ Hz, 1H), 4.20 (d, $J = 9.4$ Hz, 1H), 3.77 (s, 3H), 3.42 (q, $J = 6.7$ Hz, 1H), 3.28 (m, 1H), 3.03 (q, $J = 7.2$ Hz, 1H), 2.94 (br d, $J = 12.6$ Hz, 1H), 2.69 (m, 1H), 2.50 (d, $J = 12.7$ Hz, 1H), 2.29 (m, 1H), 1.49 (d, $J = 6.7$ Hz, 3H), 0.83 (s, 9H), -0.02 (s, 3H), -0.10 (s, 3H); ^{13}C NMR (75 MHz, CD_3OD) δ 175.1, 138.6, 131.7, 127.9, 122.2, 120.8, 119.4, 112.7, 112.0, 79.6, 72.4, 71.6, 56.9, 54.0, 52.7, 47.8, 45.4, 32.1, 26.4, 19.2, 14.5, -5.4, -5.5; HRMS (m/z): $[\text{M} + \text{H}]^+$ calcd for $\text{C}_{25}\text{H}_{39}\text{N}_2\text{O}_4\text{Si}$, 459.2674; found, 459.2676.



(±)-**3**

Synthesis of (±)-Alstilobanine A (3). A solution of *O*-TBS indole **29** (16.5 mg, 0.036 mmol) in CHCl₃ (3.3 mL) at 0 °C was treated with a solution of hydrogen chloride in MeOH (1.25 M, 3.3 mL). The resultant colorless solution was stirred at rt for 2 h, and the volatiles were removed under high vacuum to afford (±)-alstilobanine A (**3**) hydrochloride salt as a white solid (12.2 mg, 100%). ¹H NMR (400 MHz, CD₃OD) δ 7.55 (d, *J* = 7.9 Hz, 1H), 7.34 (d, *J* = 8.0 Hz, 1H), 7.08 (dd, *J* = 7.4, 7.5 Hz, 1H), 6.98 (dd, *J* = 7.4, 7.5 Hz, 1H), 4.14 (d, *J* = 10.9 Hz, 1H), 4.04 (d, *J* = 10.8 Hz, 1H), 3.78 (s, 3H), 3.54 (d, *J* = 12.5 Hz, 1H), 3.26 (q, *J* = 7.1 Hz, 1H), 3.09 (m, 1H), 2.93 (m, 1H), 2.87 (d, *J* = 12.6 Hz, 1H), 2.52 (m, 1H), 2.06 (m, 1H), 1.87 (m, 1H), 1.43 (d, *J* = 6.7 Hz, 3H); ¹³C NMR (75 MHz, CD₃OD) δ 174.9, 138.9, 130.9, 127.2, 122.5, 119.6, 119.5, 113.3, 112.2, 71.0, 68.7, 55.2, 53.0, 50.5, 42.4 (2C), 34.4, 22.6, 15.7; HRMS (*m/z*): [M + H]⁺ calcd for C₁₉H₂₅N₂O₄, 345.1809; found, 345.1809.



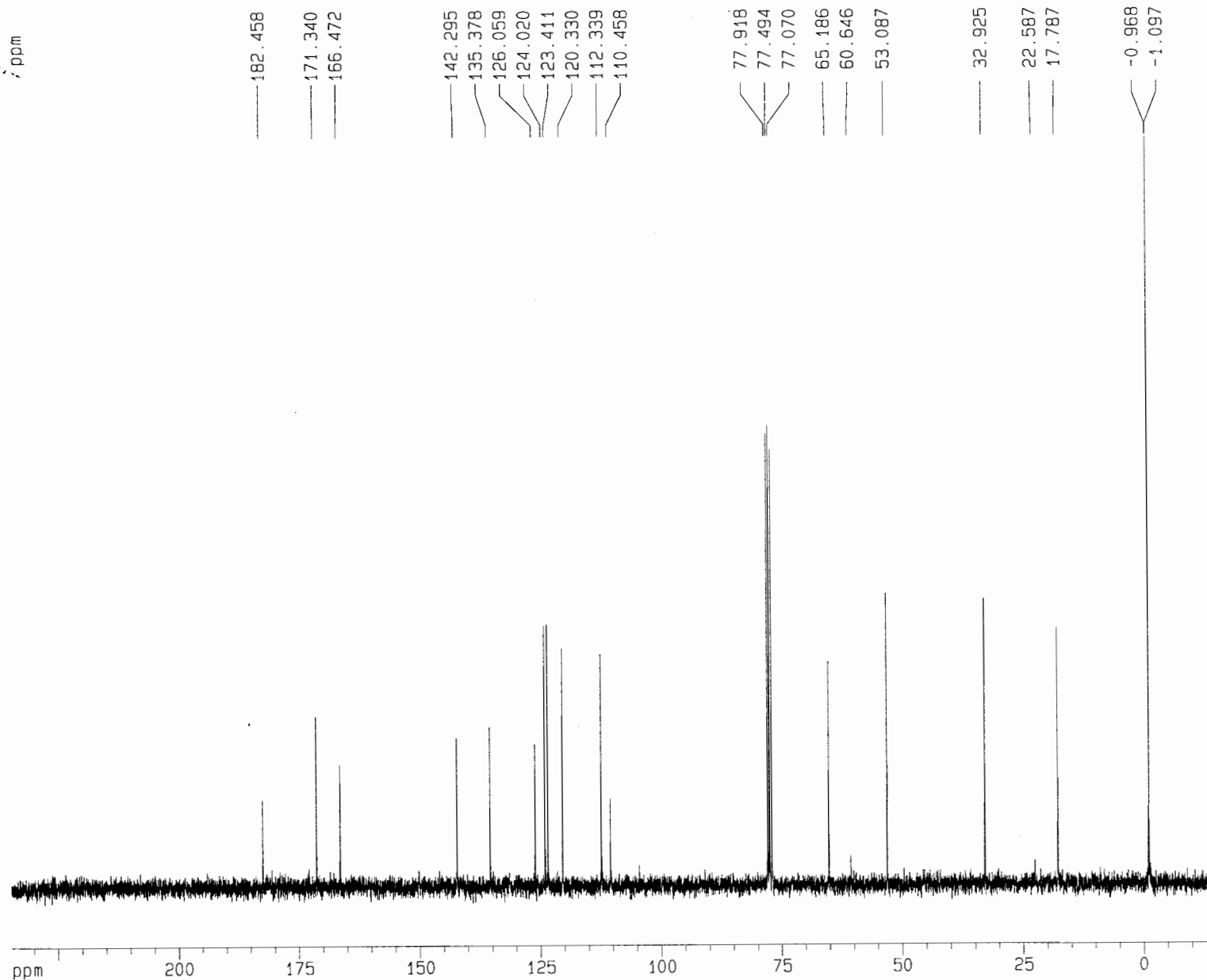
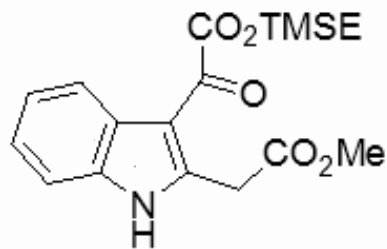
Current Data Parameters
 NAME 093009mm4170
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090930
 Time 16.33
 INSTRUM spect
 PROBHD 5 mm GNP 1H/1
 PULPROG zg30
 TD 24690
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 6172.839 Hz
 FIDRES 0.250014 Hz
 AQ 1.9999400 sec
 RG 143.7
 DW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 11.70 usec
 PL1 0.00 dB
 SFO1 299.8718518 MHz

F2 - Processing parameters
 SI 32768
 SF 299.8700000 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3298.57 Hz
 F2P -1.000 ppm
 F2 -299.87 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 179.92200 Hz/cm



Current Data Parameters
 NAME 093009mm4170
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20090930
 Time 16.44
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 172
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.573639 Hz
 AQ 0.8716788 sec
 RG 2048
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 D12 0.0000200 sec

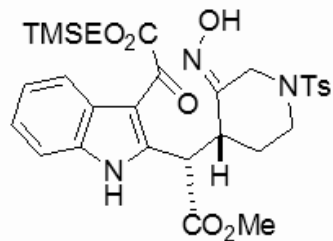
==== CHANNEL f1 =====
 NUC1 13C
 P1 5.40 usec
 PL1 -6.00 dB
 SF01 75.4106357 MHz

==== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 20.00 dB
 PL13 20.00 dB
 SF02 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4023410 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

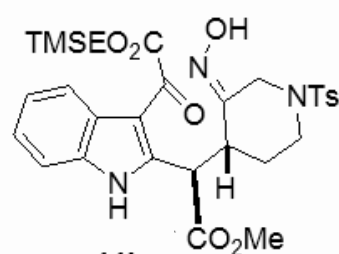
1D NMR plot parameters
 CX 20.00 cm
 F1P 234.651 ppm
 F1 17693.24 Hz
 F2P -14.638 ppm
 F2 -1103.75 Hz
 PPMCM 12.46446 ppm/cm
 HZCM 939.84955 Hz/cm

Diastereomeric products mixture of the Nitrosoalkene Michael addition

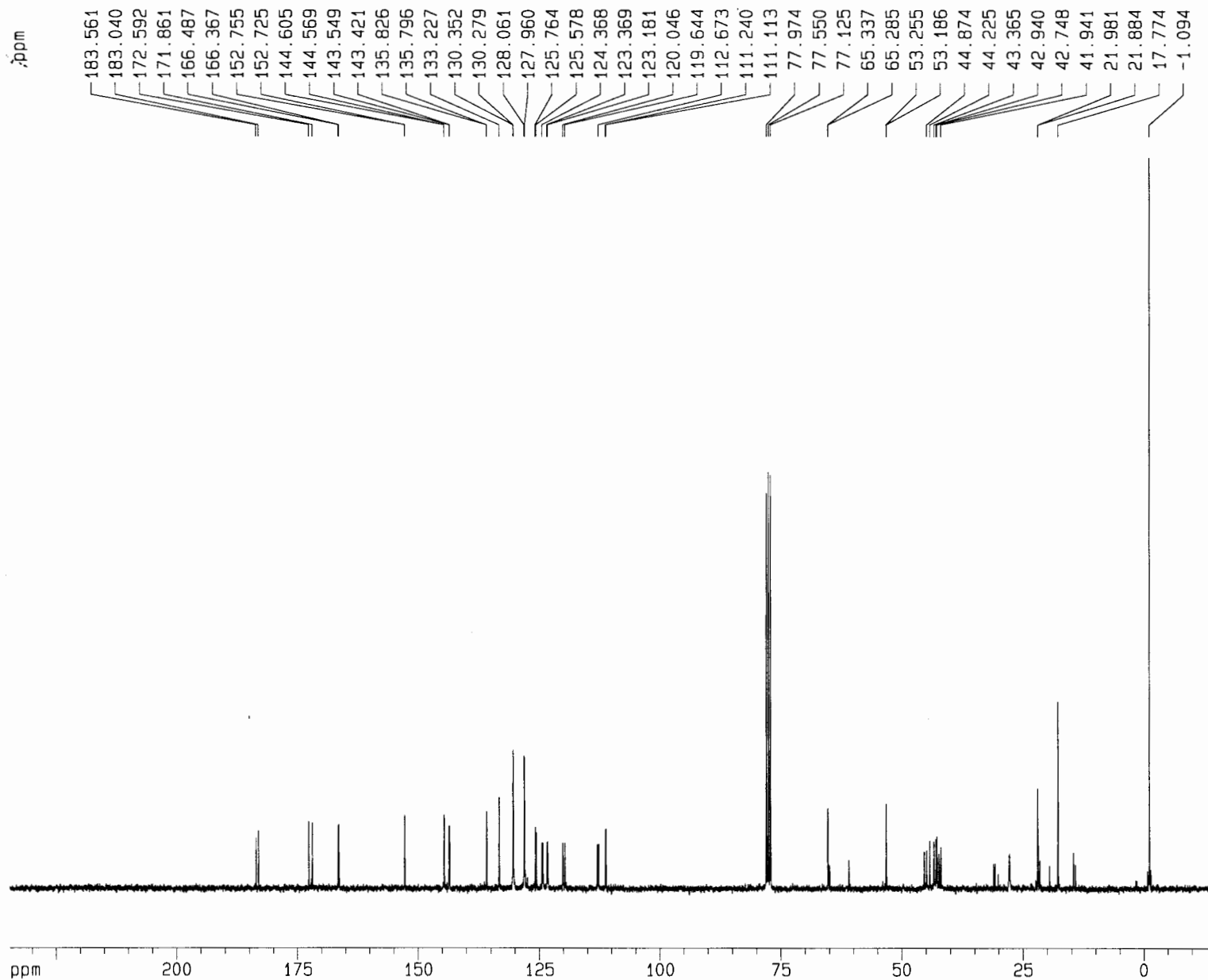


11a

+



11b



Current Data Parameters
NAME 012610mm42B1
EXPNO 2
PROCNO 1

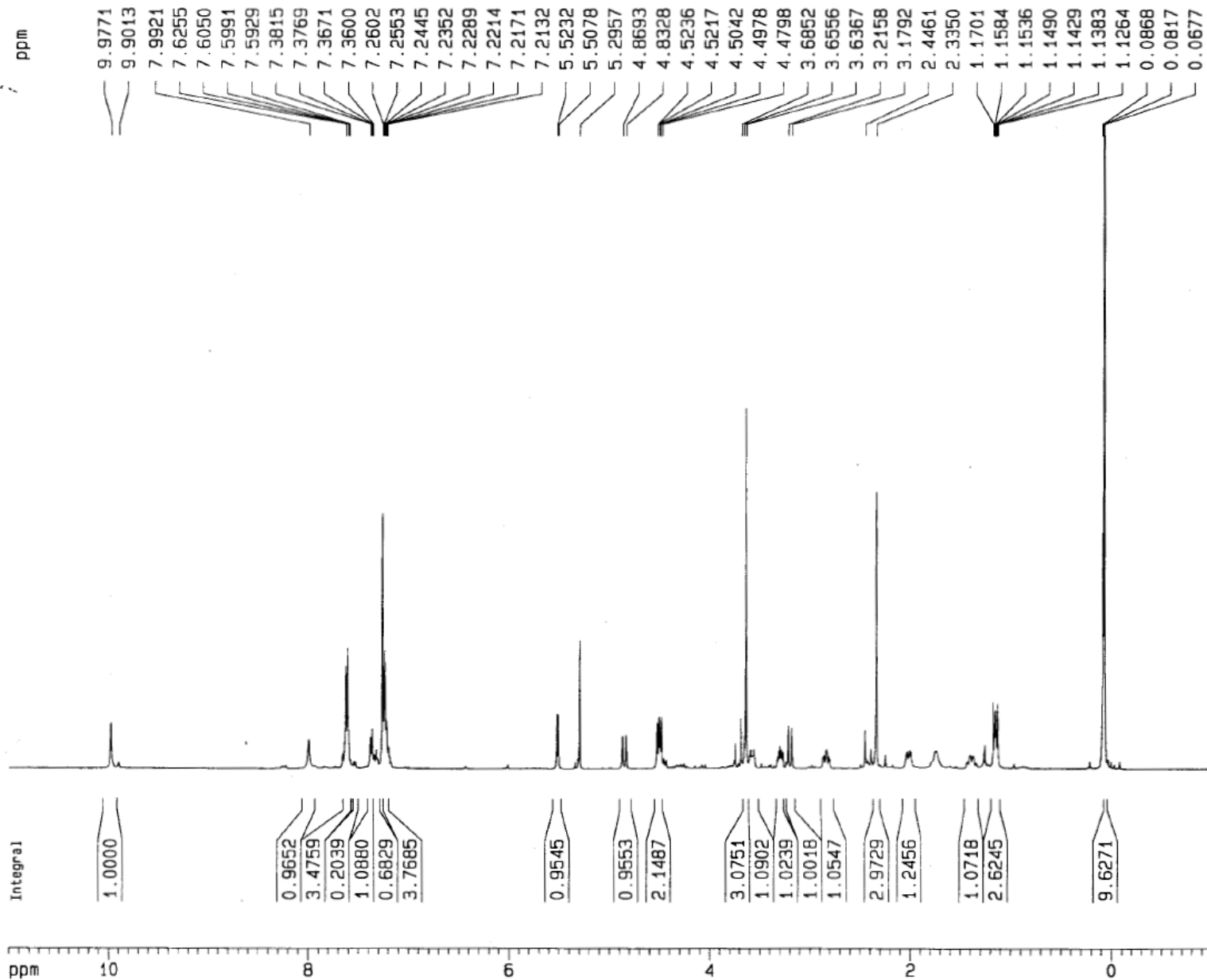
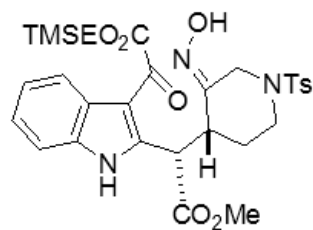
F2 - Acquisition Parameters
Date_ 20100126
Time 9.40
INSTRUM spect
PROBHD 5 mm GNP 1H/1
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 1024
DS 4
SWH 18796.992 Hz
FIDRES 0.573639 Hz
AQ 0.8716788 sec
RG 512
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
D11 0.0300000 sec
D12 0.0002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 5.40 usec
PL1 -6.00 dB
SF01 75.4106357 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 20.00 dB
PL13 20.00 dB
SF02 299.8711995 MHz

F2 - Processing parameters
SI 32768
SF 75.4023410 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
F1P 234.651 ppm
F1 17693.24 Hz
F2P -14.638 ppm
F2 -1103.75 Hz
PPMCM 12.46446 ppm/cm
HZCM 939.84967 Hz/cm



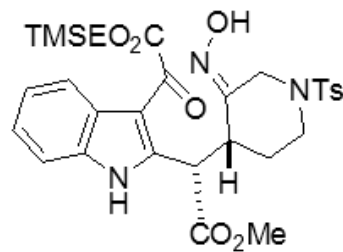
Current Data Parameters
 NAME 092612FYG-11a
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120926
 Time 9.02
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 322.5
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

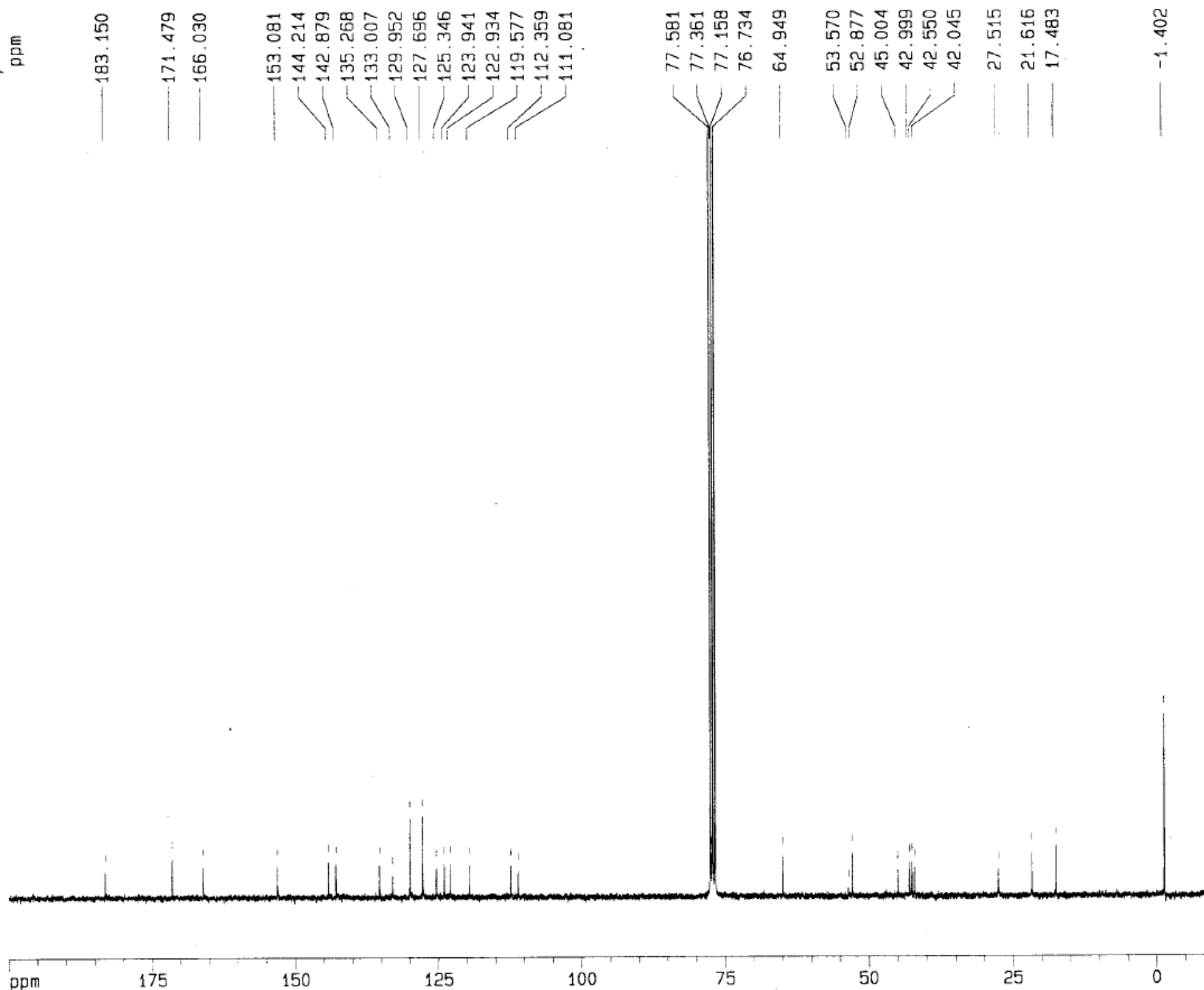
===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300089 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



11a



Current Data Parameters

NAME 092512FYG-11a
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20120925
 Time 18.45
 INSTRUM spect
 PROBHD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 3930
 DS 4
 SWH 18832.393 Hz
 FIDRES 0.287360 Hz
 AQ 1.7400308 sec
 RG 16384
 DW 26.550 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 d12 0.0000200 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.75 usec
 PL1 0.00 dB
 SF01 75.4760200 MHz

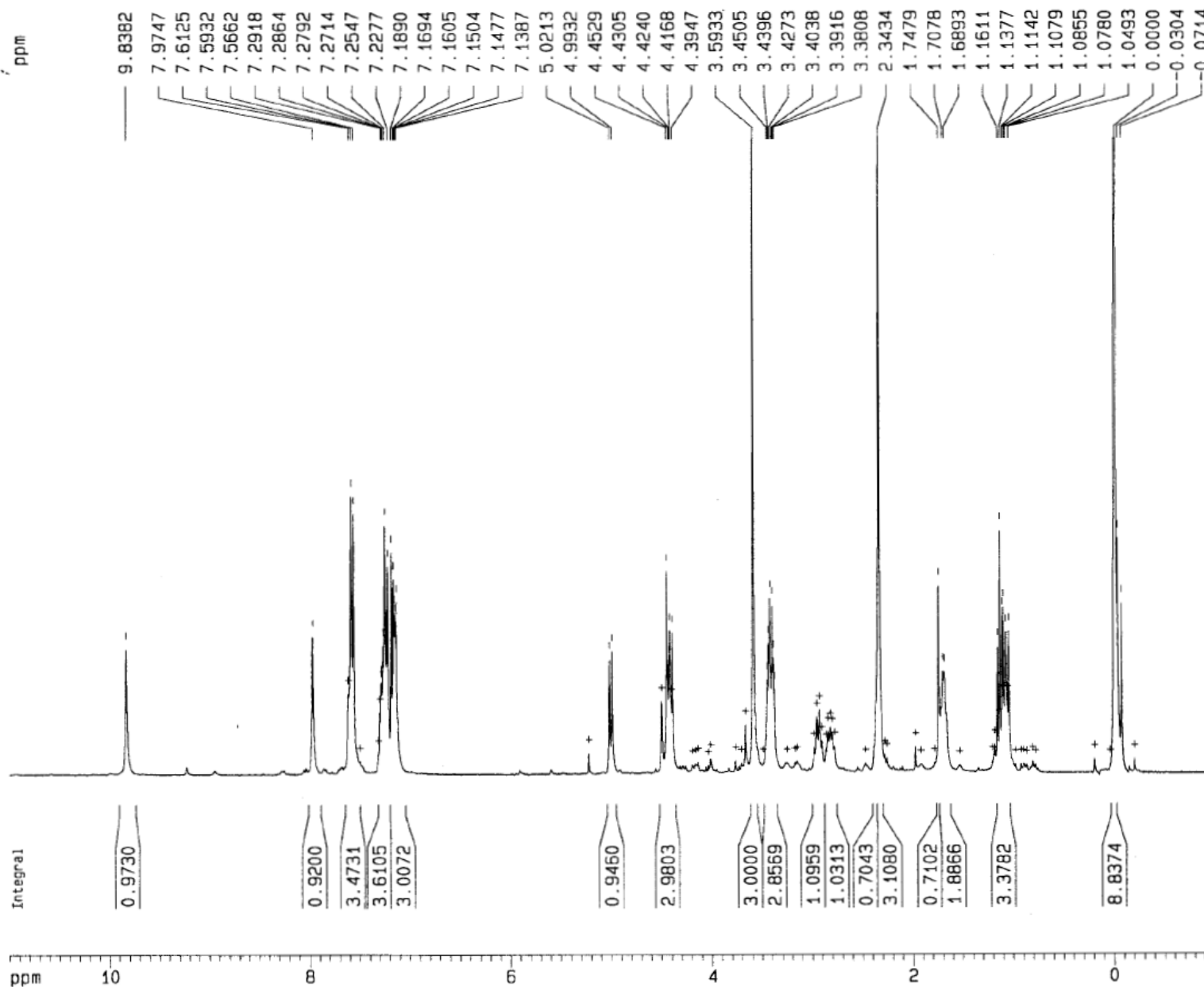
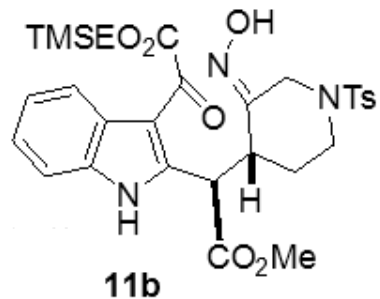
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 110.00 usec
 PL2 0.00 dB
 PL12 17.50 dB
 PL13 17.50 dB
 SF02 300.1312005 MHz

F2 - Processing parameters

S1 32768
 SF 75.4677399 MHz
 WDM EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters

CX 20.00 cm
 F1P 200.000 ppm
 F1 15093.55 Hz
 F2P -10.000 ppm
 F2 -754.68 Hz
 PPMCM 10.50000 ppm/cm
 HZCM 792.41132 Hz/cm



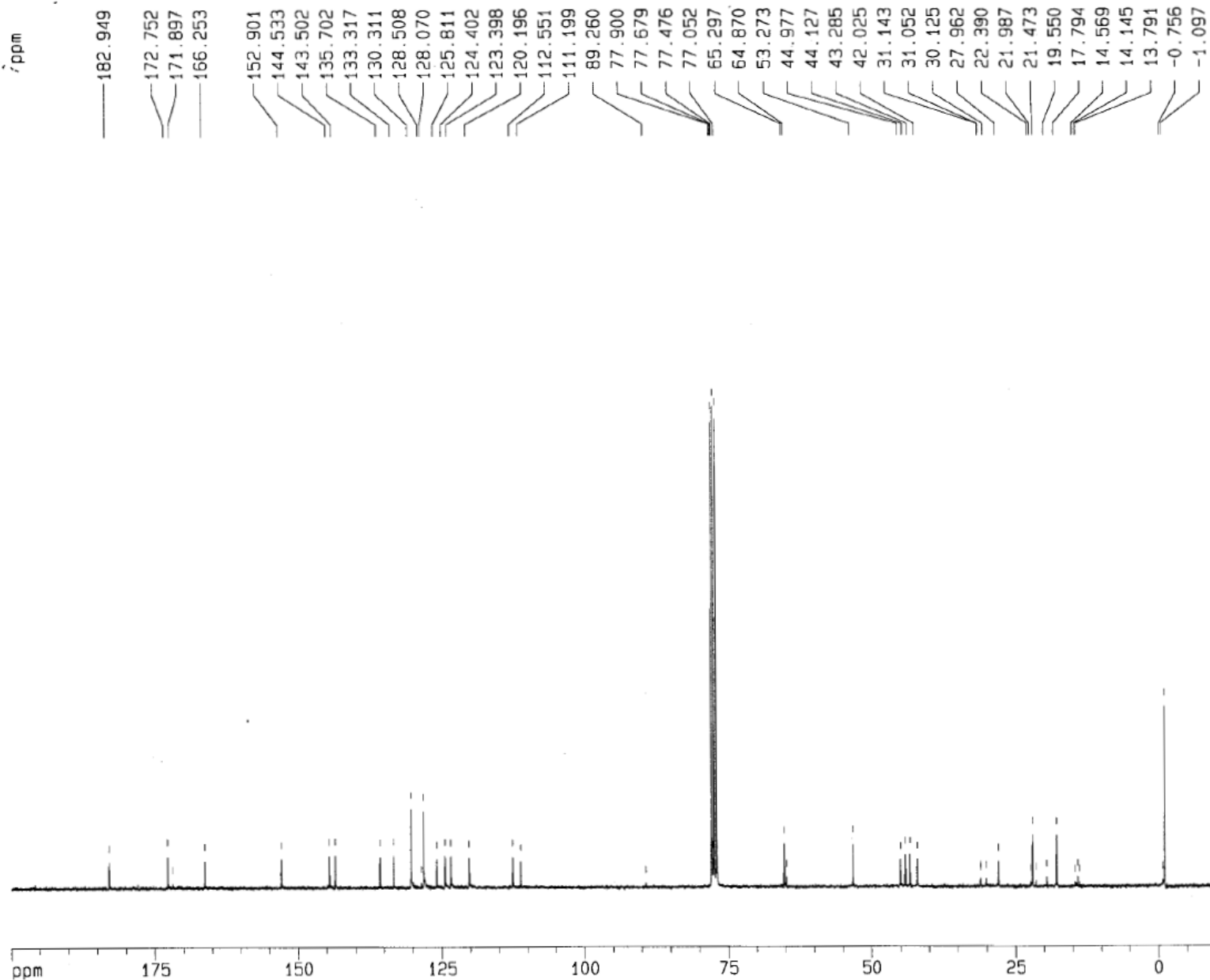
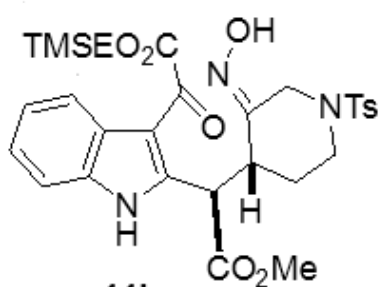
Current Data Parameters
 NAME 033111FYQI-NAM
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20110331
 Time 15.24
 INSTRUM spect
 PROBHD 5 mm Multinu
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 6172.839 Hz
 FIDRES 0.094190 Hz
 AQ 5.3084660 sec
 RG 322.5
 DW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 9.60 usec
 PL1 -6.00 dB
 SF01 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300277 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3301.43 Hz
 F2P -1.000 ppm
 F2 -300.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 180.07802 Hz/cm



Current Data Parameters

NAME 092412FYG-11b
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20120924
Time 23.14
INSTRUM spect
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 3584
DS 4
SWH 18796.992 Hz
FIDRES 0.286819 Hz
AQ 1.7433076 sec
RG 2048
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 5.25 usec
PL1 -6.00 dB
SF01 75.4106357 MHz

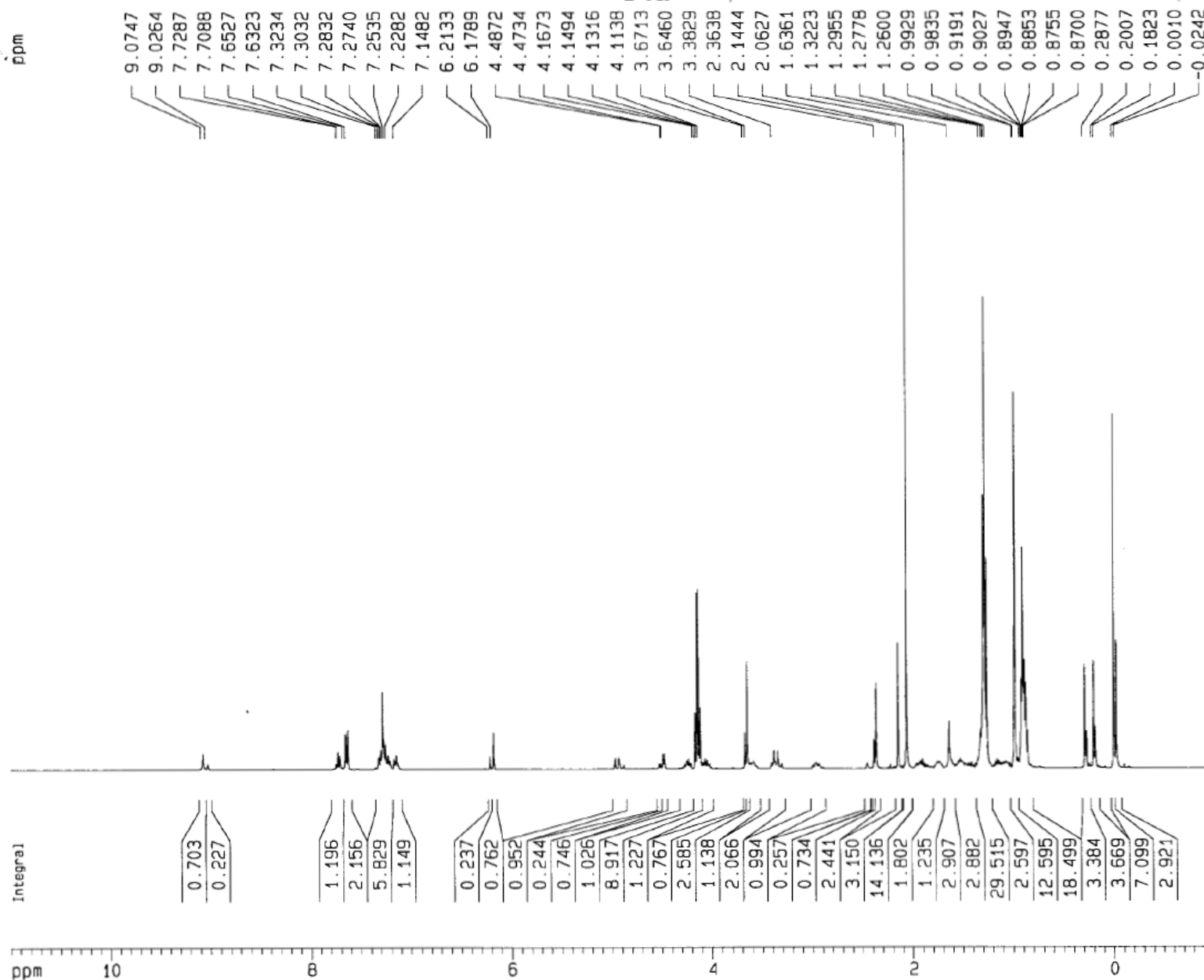
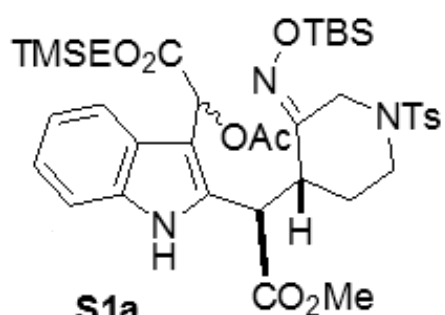
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 19.70 dB
PL13 19.70 dB
SF02 299.8711995 MHz

F2 - Processing parameters

SI 32768
SF 75.4023410 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
F1P 200.000 ppm
F1 15080.47 Hz
F2P -10.000 ppm
F2 -754.02 Hz
PPMCM 10.50000 ppm/cm
HZCM 791.72461 Hz/cm



Current Data Parameters

NAME 012710mm4284
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20100127
 Time 10.11
 INSTRUM spect
 PROBHD 5 mm BBI 1H-B
 PULPROG zg30
 TD 33110
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.250019 Hz
 AQ 1.9998940 sec
 RG 64
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====

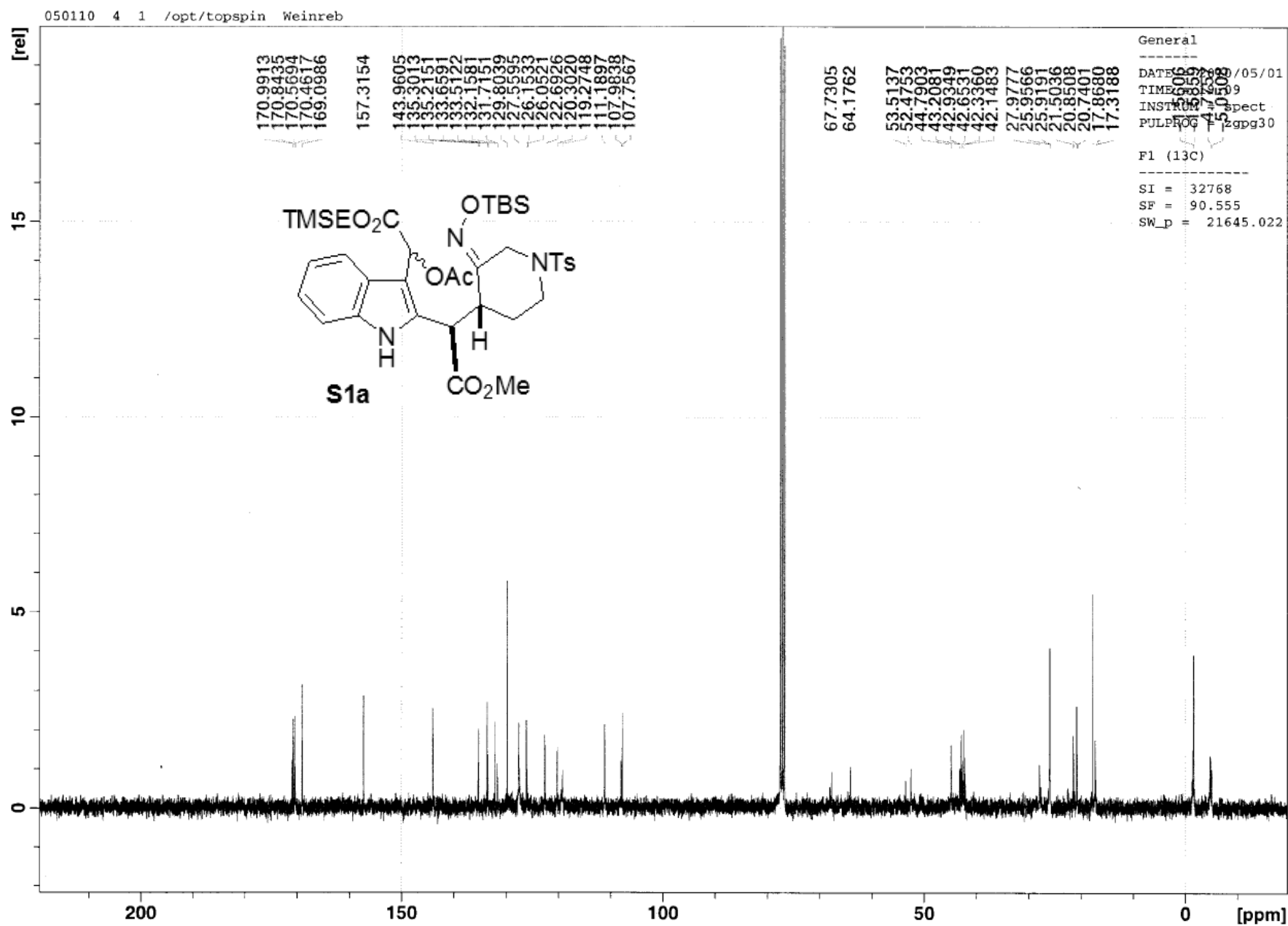
NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

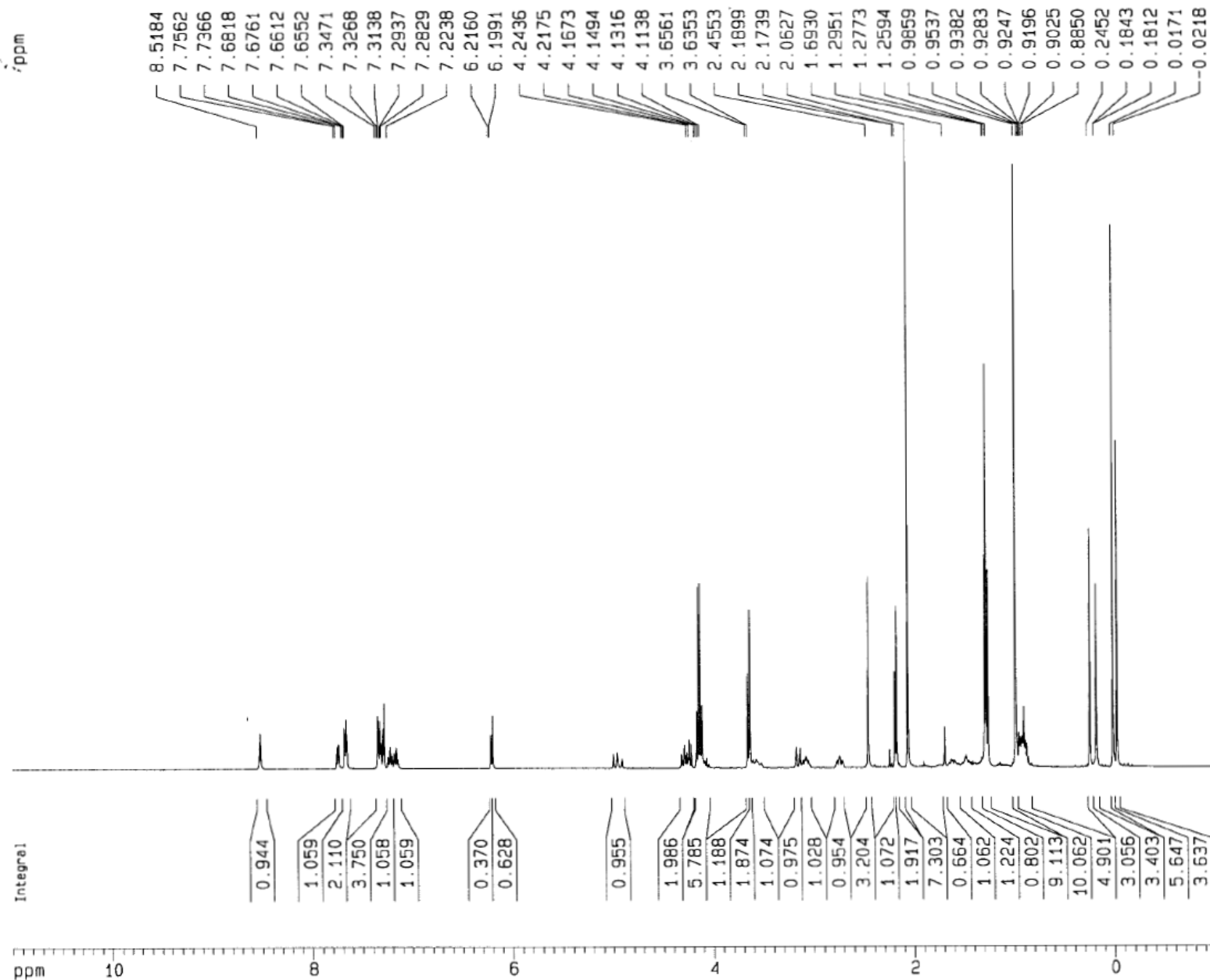
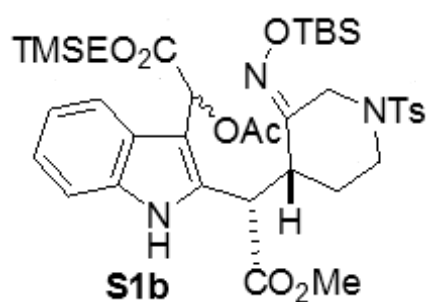
F2 - Processing parameters

SI 32768
 SF 400.1300000 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm





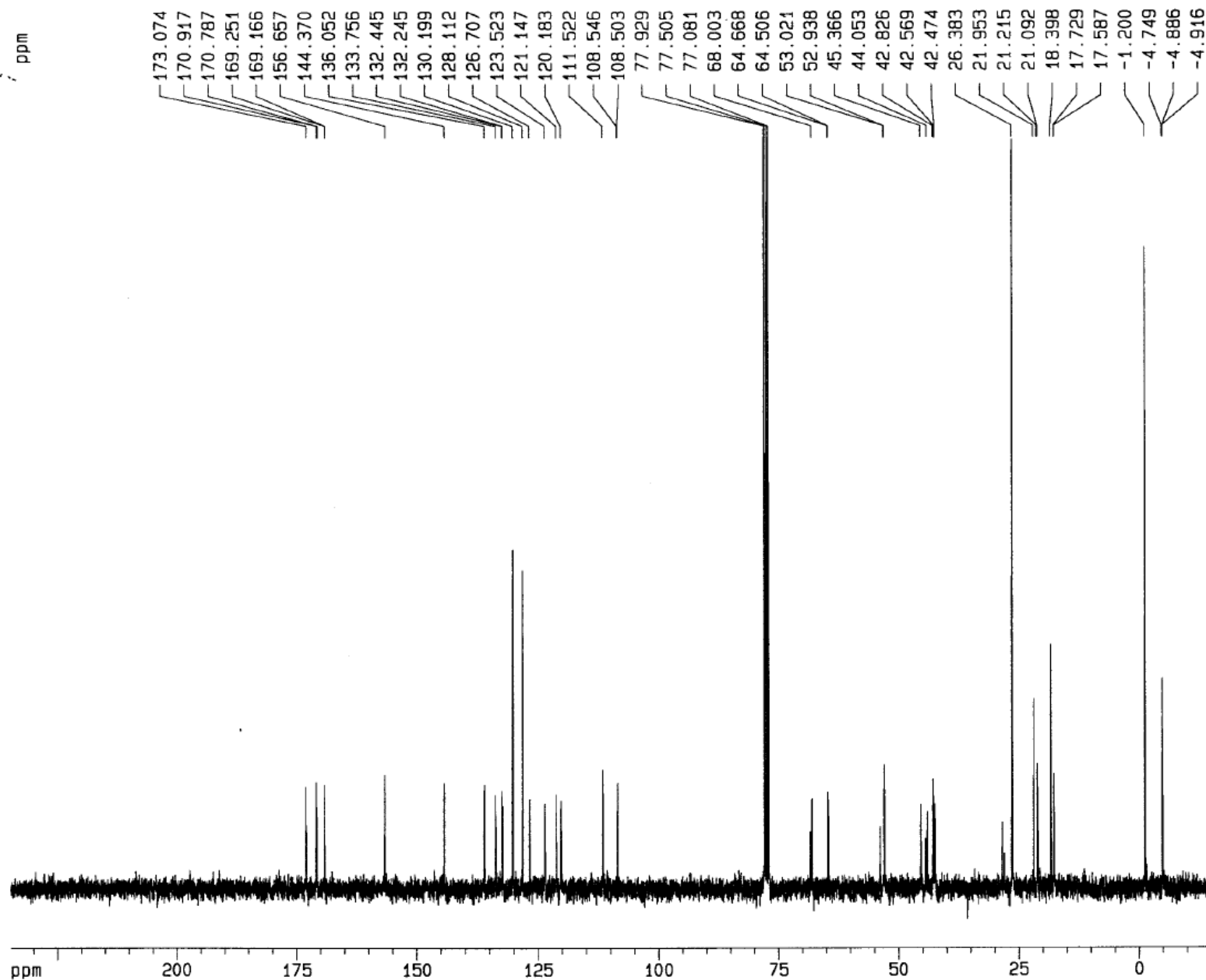
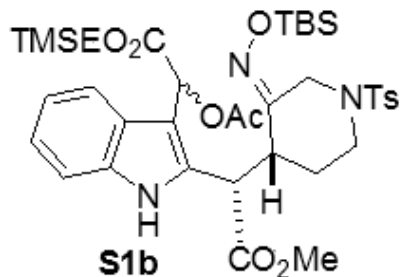
Current Data Parameters
 NAME 012710mm4284
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20100127
 Time 10.05
 INSTRUM spect
 PROBH0 5 mm BBI 1H-B
 PULPROG zg30
 TD 33110
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.250019 Hz
 AQ 1.9998940 sec
 RG 57
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300000 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
NAME 050110
EXPNO 2
PROCNO 1

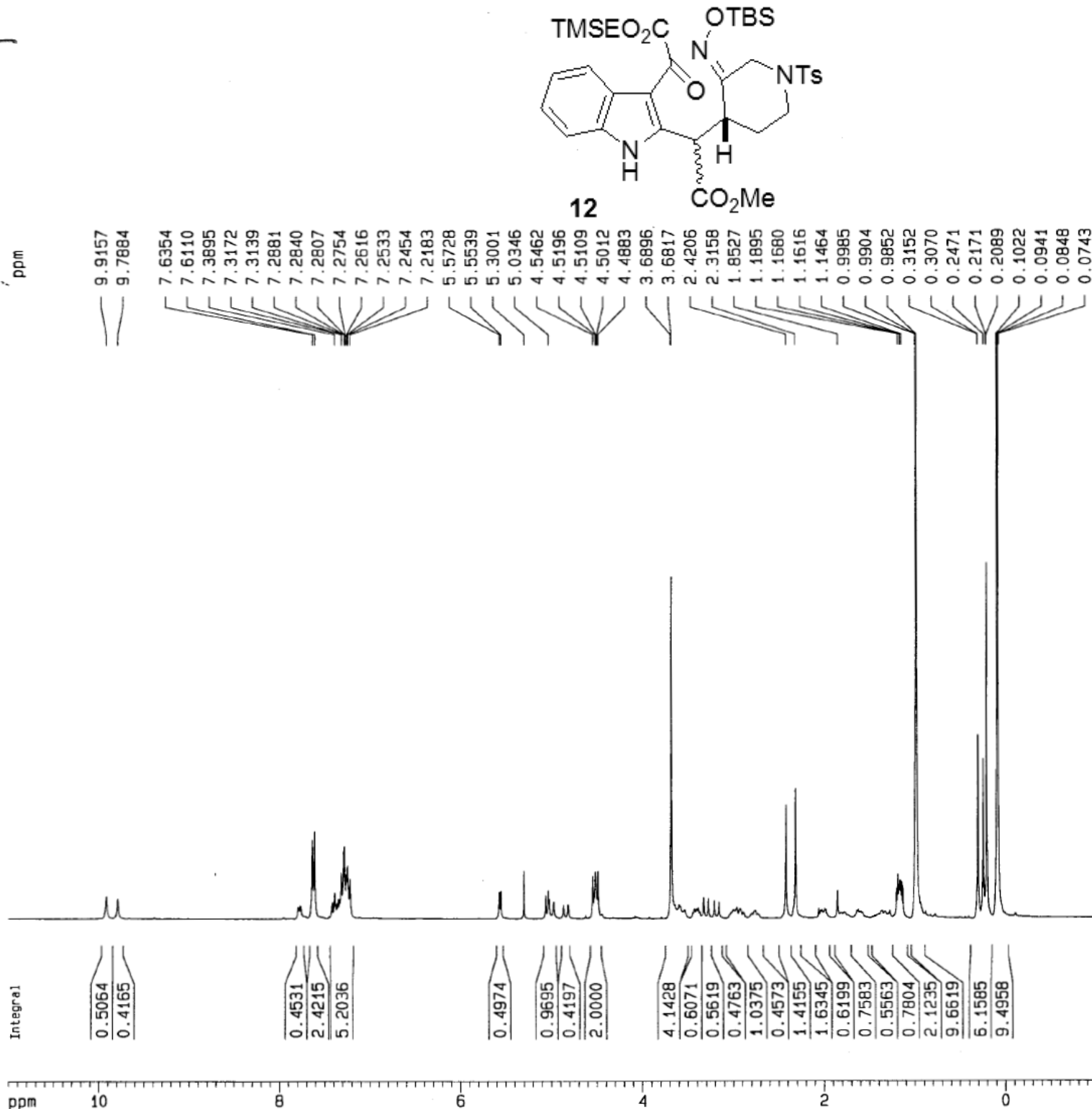
F2 - Acquisition Parameters
Date_ 20100501
Time 17.37
INSTRUM spect
PROBHD 5 mm Multinu
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 599
DS 4
SWH 18832.393 Hz
FIDRES 0.287360 Hz
AQ 1.7400308 sec
RG 16384
DW 26.550 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

----- CHANNEL f1 -----
NUC1 13C
P1 11.80 usec
PL1 0.00 dB
SFO1 75.4760200 MHz

----- CHANNEL f2 -----
CPDPRG2 waltz16
NUC2 1H
PCPD2 110.00 usec
PL2 0.00 dB
PL12 17.50 dB
PL13 17.50 dB
SFO2 300.1312005 MHz

F2 - Processing parameters
SI 32768
SF 75.4677190 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
F1P 234.765 ppm
F1 17717.16 Hz
F2P -14.778 ppm
F2 -1115.23 Hz
PPMCM 12.47712 ppm/cm
HZCM 941.61951 Hz/cm



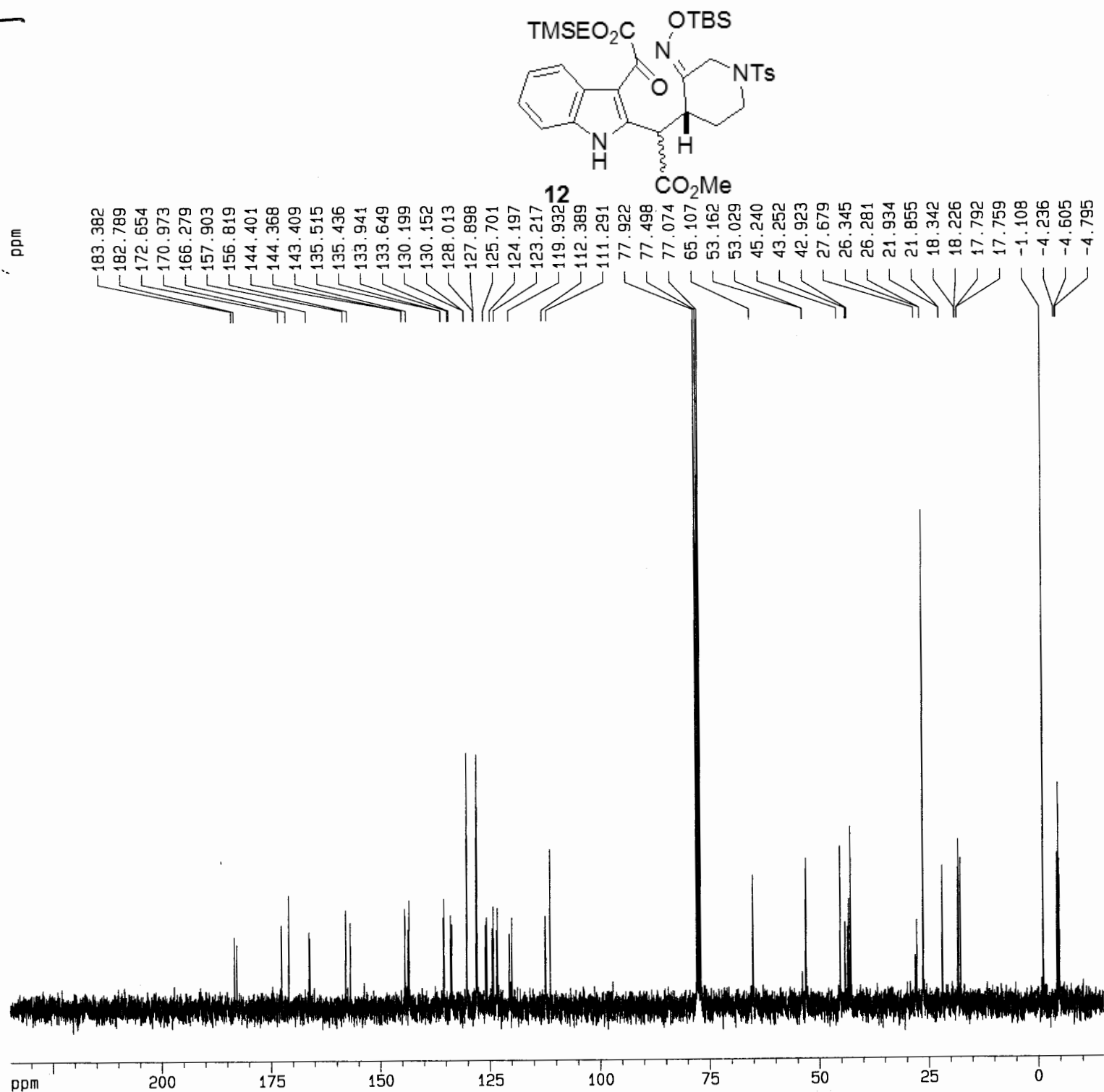
Current Data Parameters
 NAME 043010
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20100430
 Time 15.47
 INSTRUM spect
 PROBHD 5 mm Multinu
 PULPROG zg30
 TD 24590
 SOLVENT CDC13
 NS 8
 DS 2
 SWH 6172.839 Hz
 FIDRES 0.250014 Hz
 AQ 1.9999400 sec
 RG 114
 DW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 9.50 usec
 PL1 -6.00 dB
 SFO1 300.1318534 MHz

F2 - Processing parameters
 SI 32768
 SF 300.1300000 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3301.43 Hz
 F2P -1.000 ppm
 F2 -300.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 180.07800 Hz/cm



Current Data Parameters
 NAME 043010
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20100430
 Time 15.56
 INSTRUM spect
 PROBHD 5 mm Multinu
 PULPROG zgpg30
 TD 32768
 SOLVENT CDC13
 NS 500
 DS 4
 SMH 18832.393 Hz
 FIDRES 0.574719 Hz
 AQ 0.8700404 sec
 RG 16384
 DW 26.550 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 d11 0.03000000 sec
 d12 0.00002000 sec

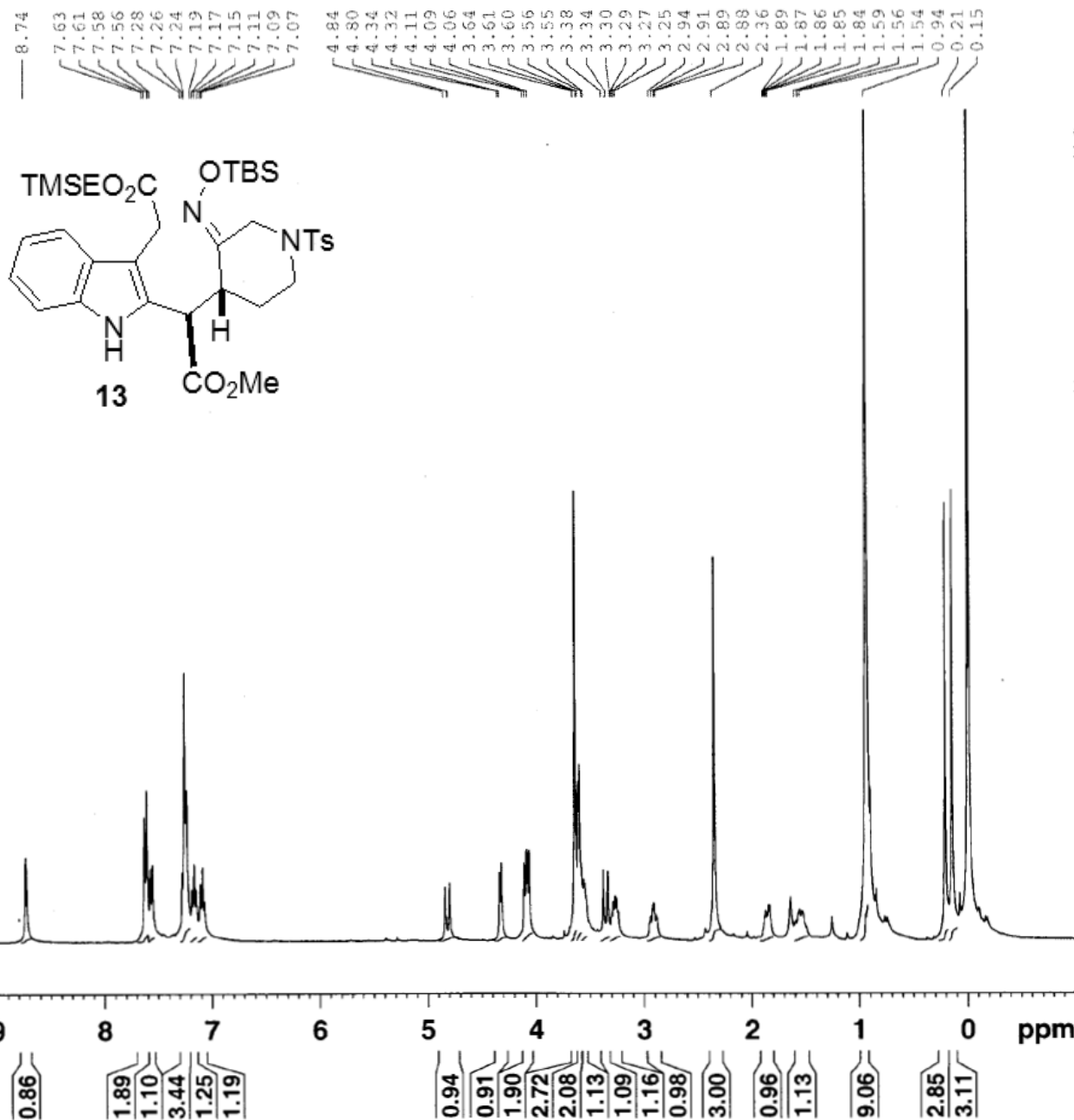
===== CHANNEL f1 =====
 NUC1 13C
 P1 11.80 usec
 PL1 0.00 dB
 SF01 75.4760200 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 110.00 usec
 PL2 0.00 dB
 PL12 17.50 dB
 PL13 17.50 dB
 SF02 300.1312005 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677190 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters

CX 20.00 cm
 F1P 234.765 ppm
 F1 17717.16 Hz
 F2P -14.778 ppm
 F2 -1115.24 Hz
 PPMCM 12.47712 ppm/cm
 HZCM 941.61963 Hz/cm

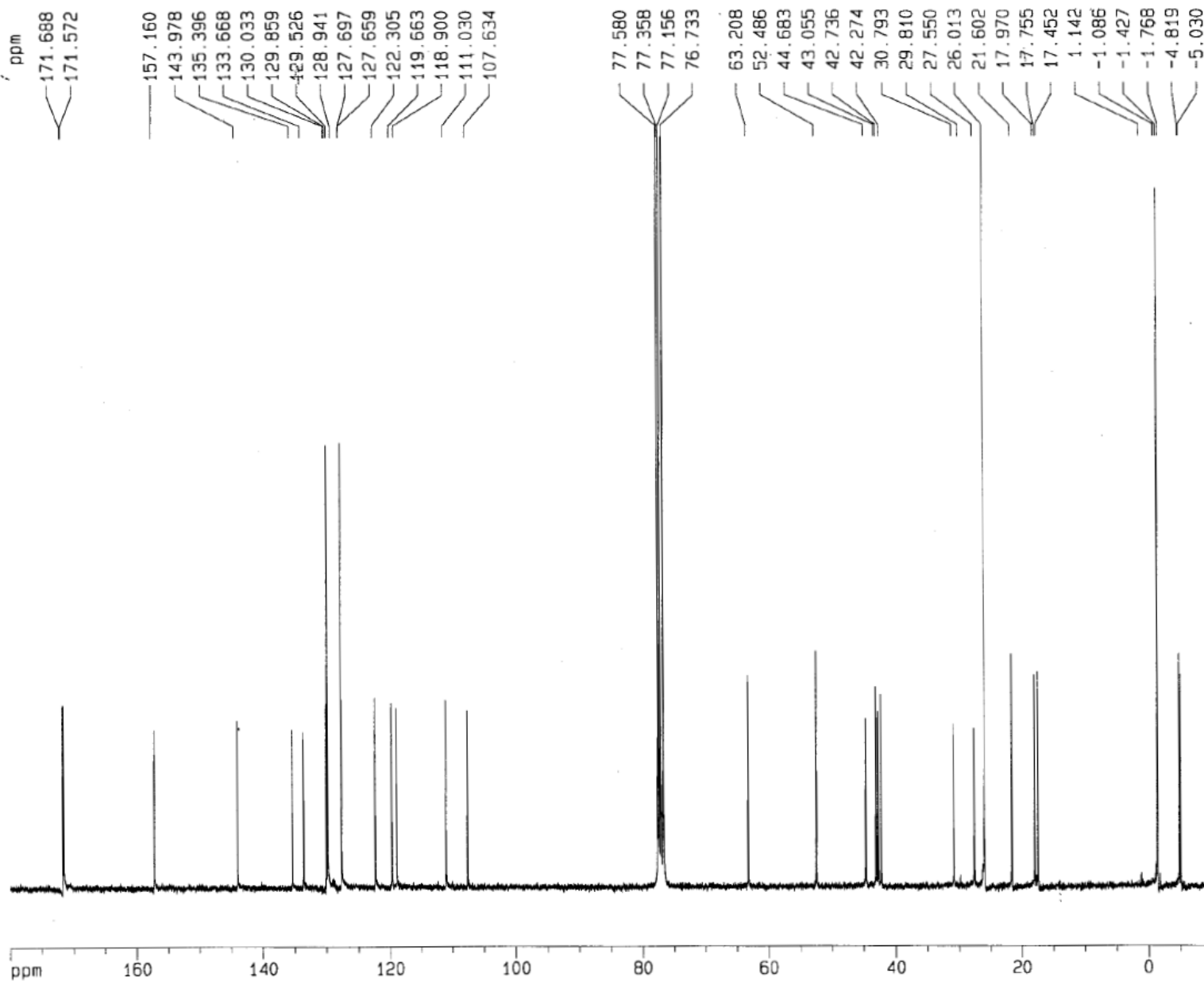
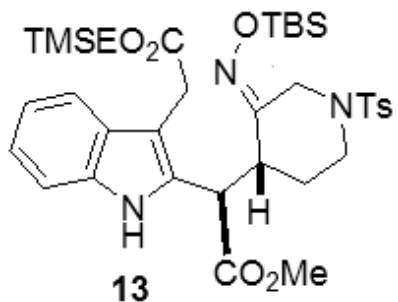


Current Data Parameters
 NAME 092212FYQ-Compound13
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120922
 Time 23.19
 INSTRUM spect
 PROBHD 5 mm QNP 1H/15
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 7440.476 Hz
 FIDRES 0.113533 Hz
 AQ 4.4040694 sec
 RG 181
 DW 67.200 usec
 DE 6.00 usec
 TE 295.7 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.93 usec
 PL1 -3.00 dB
 SFO1 360.1322240 MHz

F2 - Processing parameters
 SI 32768
 SF 360.1300145 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



Current Data Parameters
NAME 092212FY0cpd13
EXPNO 2
PROCNO 1

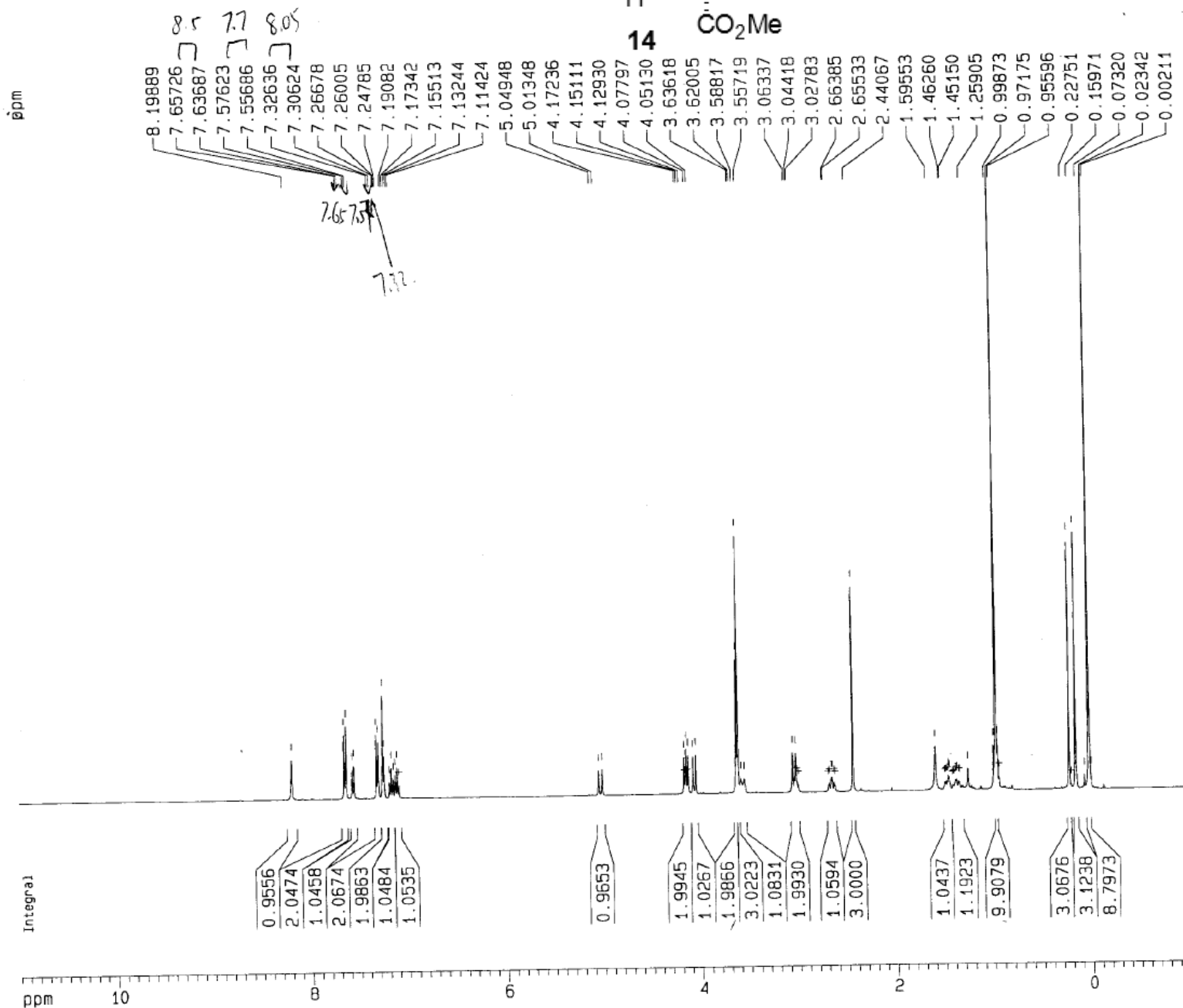
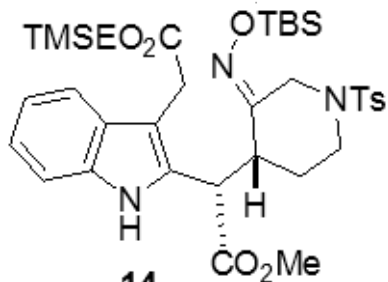
F2 - Acquisition Parameters
Date_ 20120922
Time 22.08
INSTRUM spect
PROBHD 5 mm GNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 7168
DS 4
SWH 18796.992 Hz
FIDRES 0.286819 Hz
AQ 1.7433076 sec
RG 1024
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 5.25 usec
PL1 -6.00 dB
SF01 75.4106357 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 19.70 dB
PL13 19.70 dB
SF02 299.8711995 MHz

F2 - Processing parameters
SI 32768
SF 75.4023654 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
F1P 180.000 ppm
F1 13572.43 Hz
F2P -10.000 ppm
F2 -754.02 Hz
PPMCM 9.50000 ppm/cm
HZCM 716.32251 Hz/cm



Current Data Parameters
 NAME 071911FJ11-019
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20110719
 Time 9.33
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 90.5
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

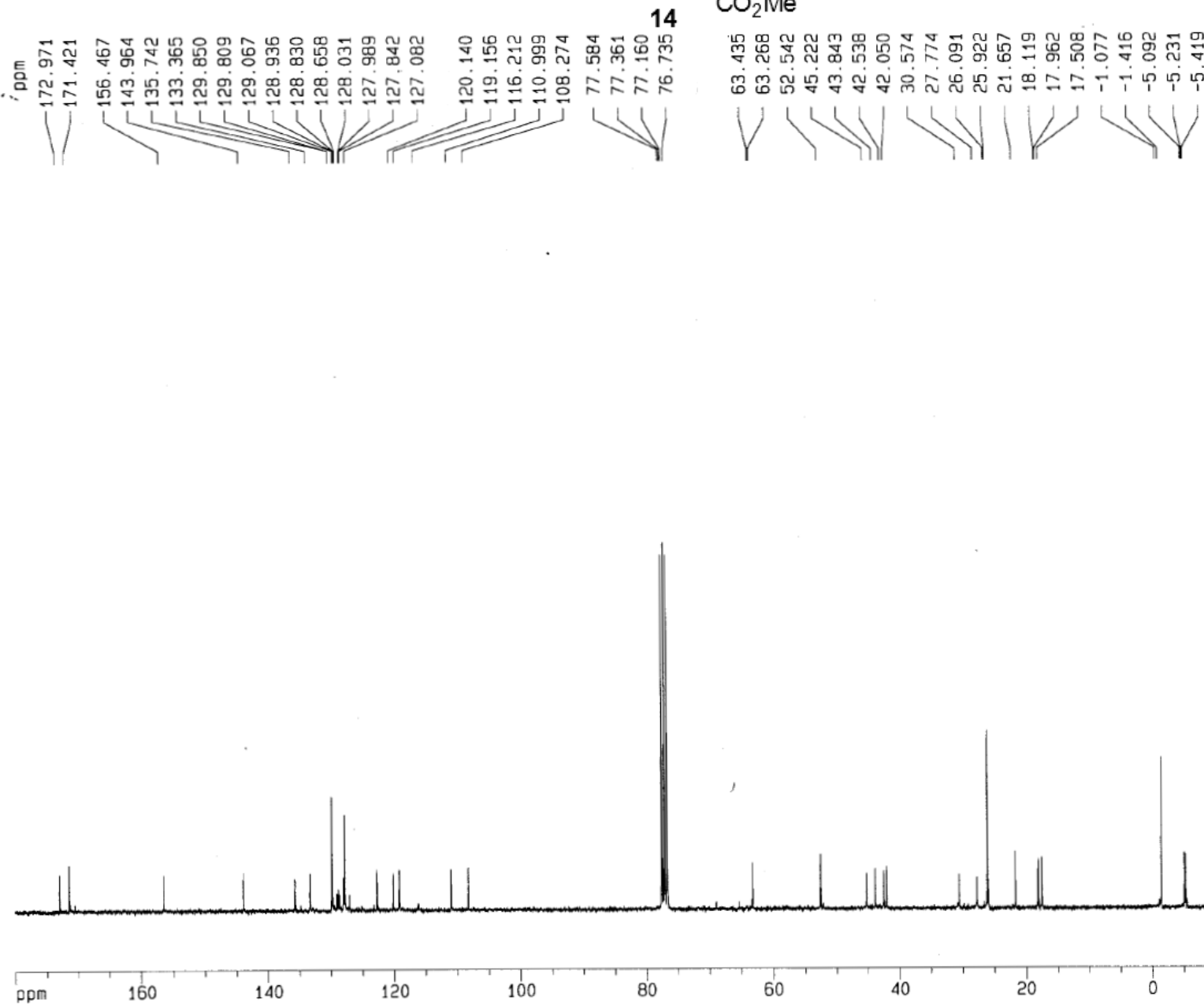
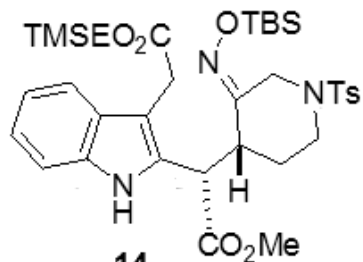
----- CHANNEL f1 -----
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters

SI 32768
 SF 400.1300086 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters

NAME 092212FYGcpd14
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20120922
Time 23.19
INSTRUM spect
PROBHD 5 mm GNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 1024
DS 4
SWH 18796.992 Hz
FIDRES 0.286819 Hz
AQ 1.7433076 sec
RG 1024
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
D11 0.0300000 sec
D12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 5.25 usec
PL1 -6.00 dB
SF01 75.4106357 MHz

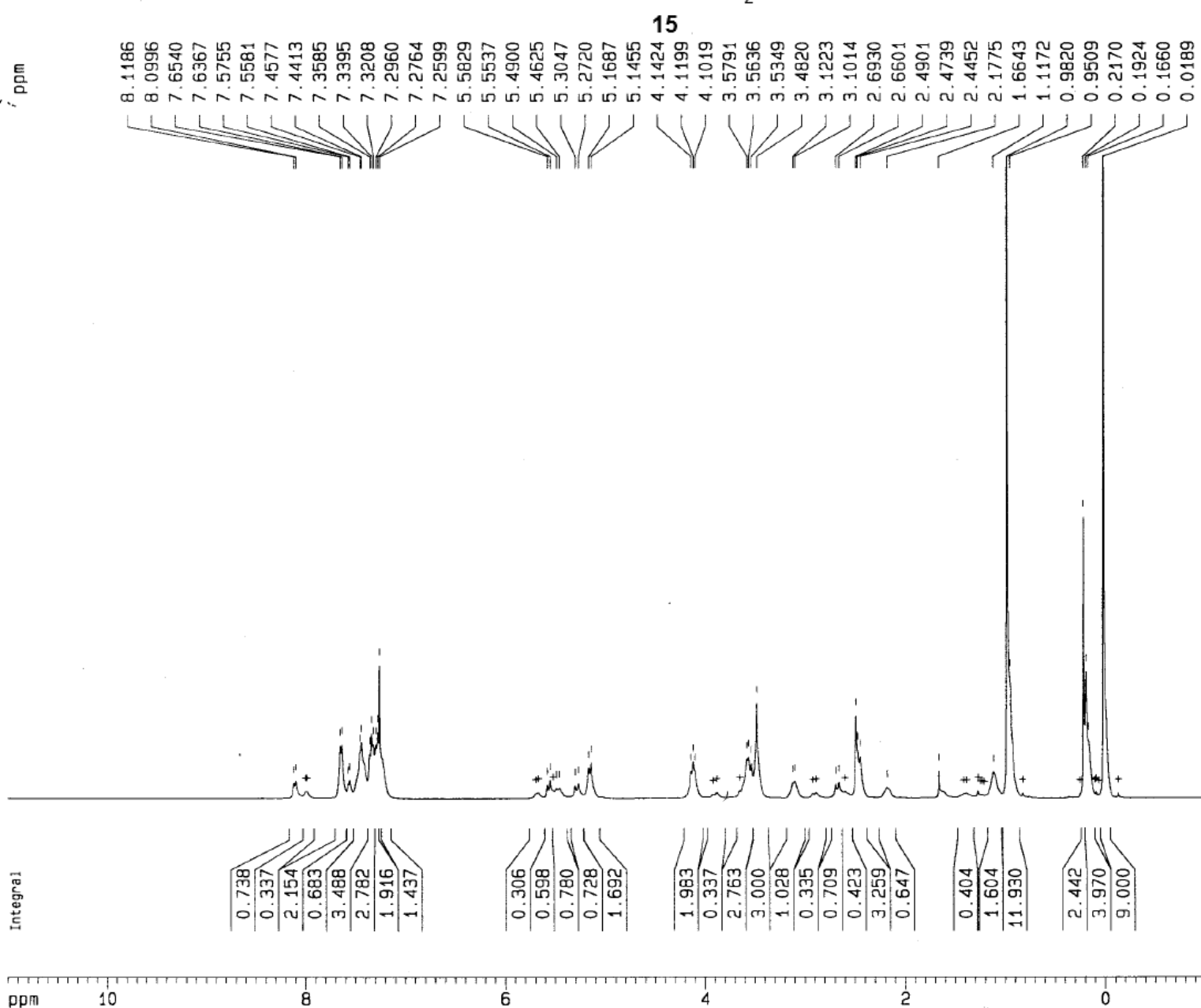
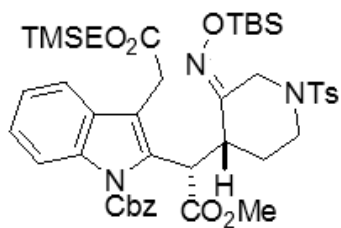
===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 19.70 dB
PL13 19.70 dB
SF02 299.8711995 MHz

F2 - Processing parameters

SI 32768
SF 75.4023660 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
F1P 180.000 ppm
F1 13572.43 Hz
F2P -10.000 ppm
F2 -754.02 Hz
PPMCM 9.50000 ppm/cm
HZCM 716.32251 Hz/cm



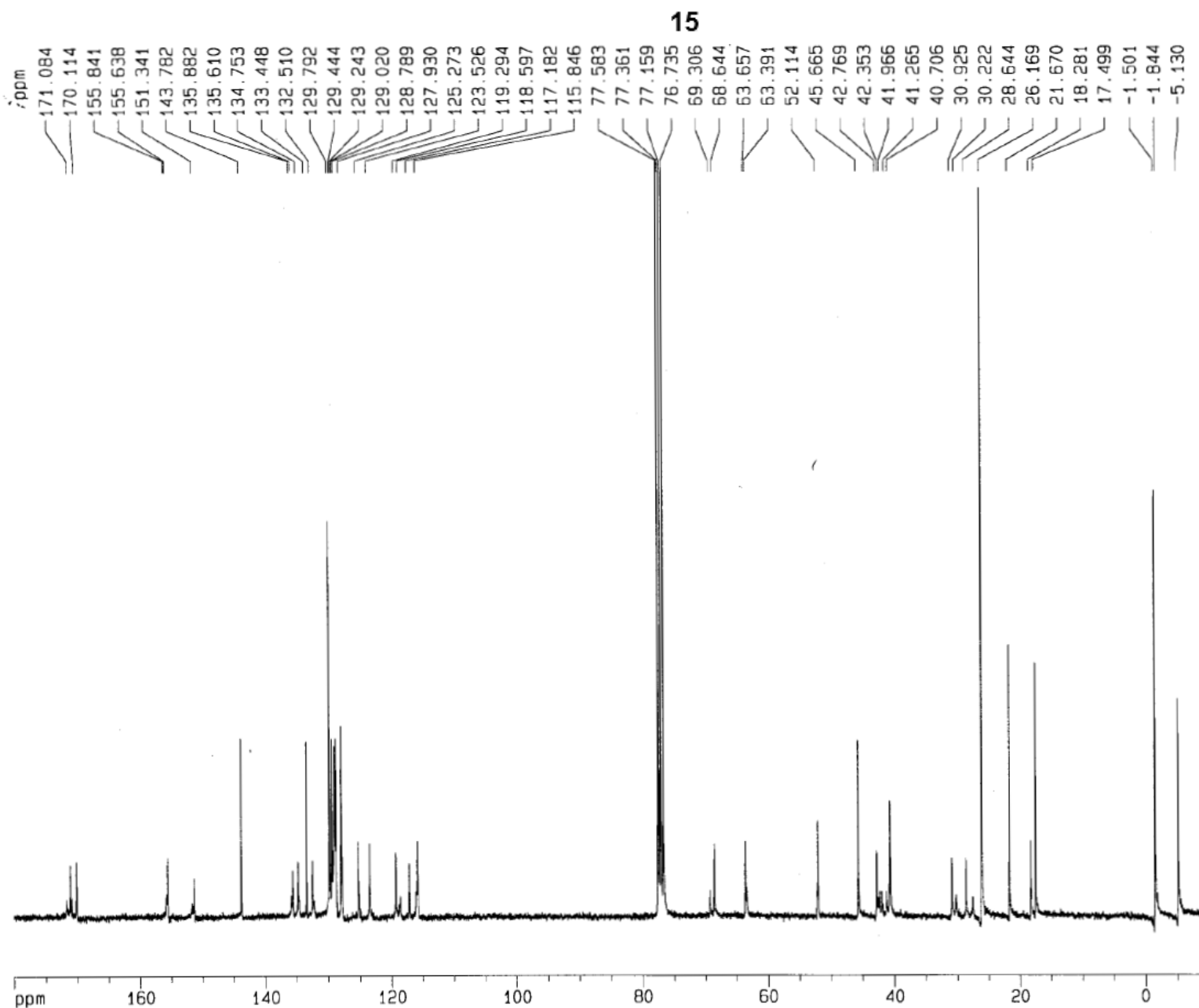
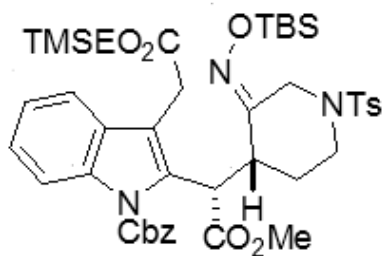
Current Data Parameters
 NAME 092512FY0-15
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120925
 Time 18.02
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 57
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
 NAME 092412FYQ-15
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120925
 Time 6.27
 INSTRUM spect
 PROBHD 5 mm GNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 6656
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 1024
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 D12 0.0000200 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SF01 75.4106357 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SF02 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4023671 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 180.000 ppm
 F1 13572.43 Hz
 F2P -10.000 ppm
 F2 -754.02 Hz
 PPMCM 9.50000 ppm/cm
 HZCM 716.32251 Hz/cm

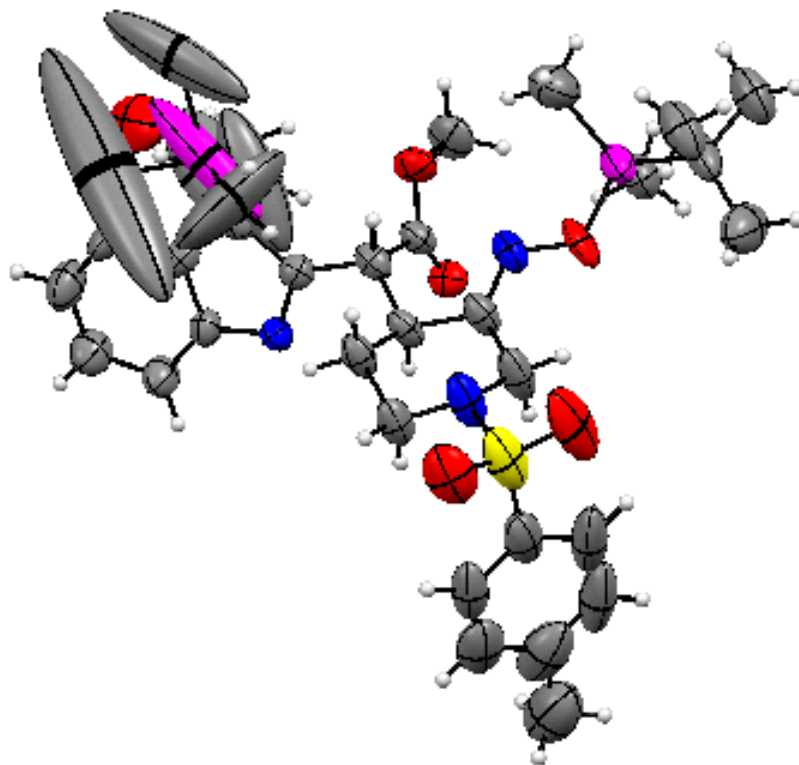
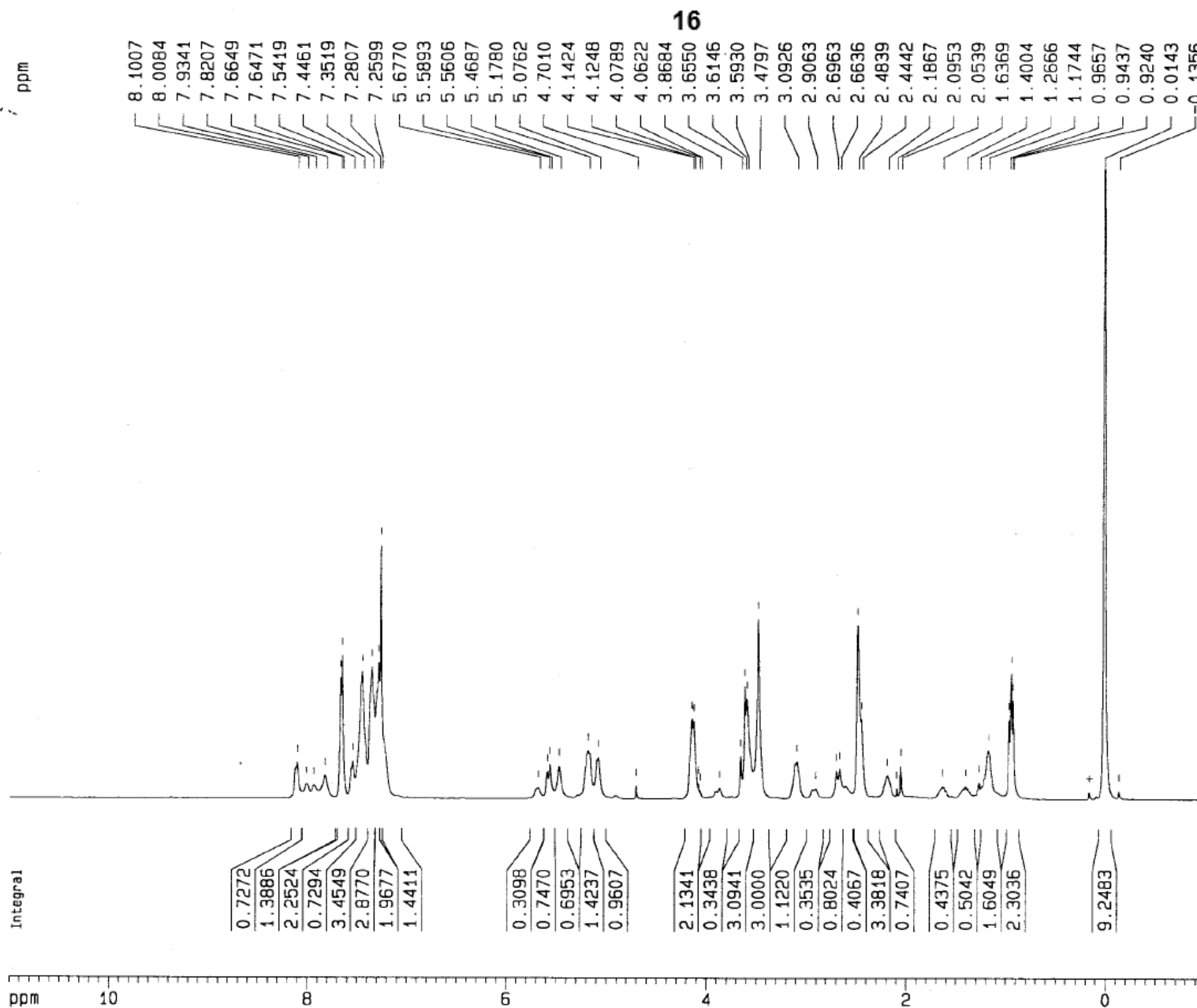
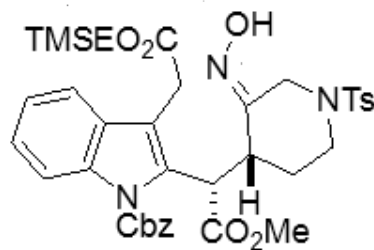


Fig. 1 ORTEP of *N*-Cbz derivative **15**



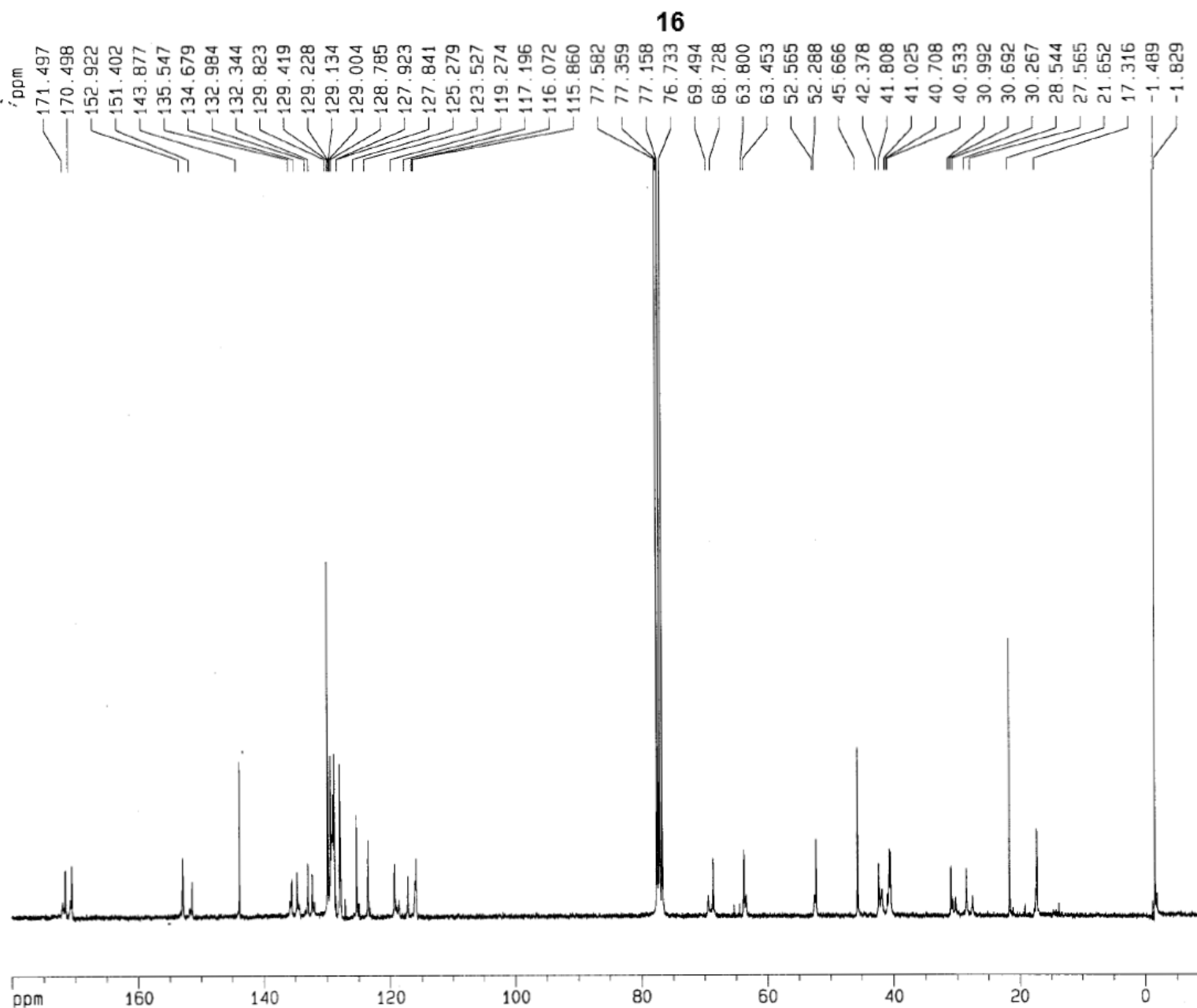
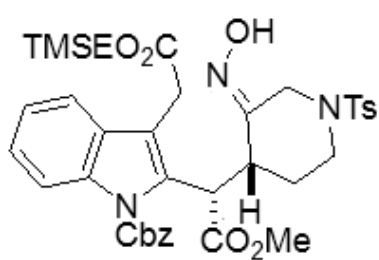
Current Data Parameters
 NAME 092512FYQ-16
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120925
 Time 18.29
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 50.8
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

==== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
 NAME 092512FYG-16
 EXPNO 1
 PROCNO 1

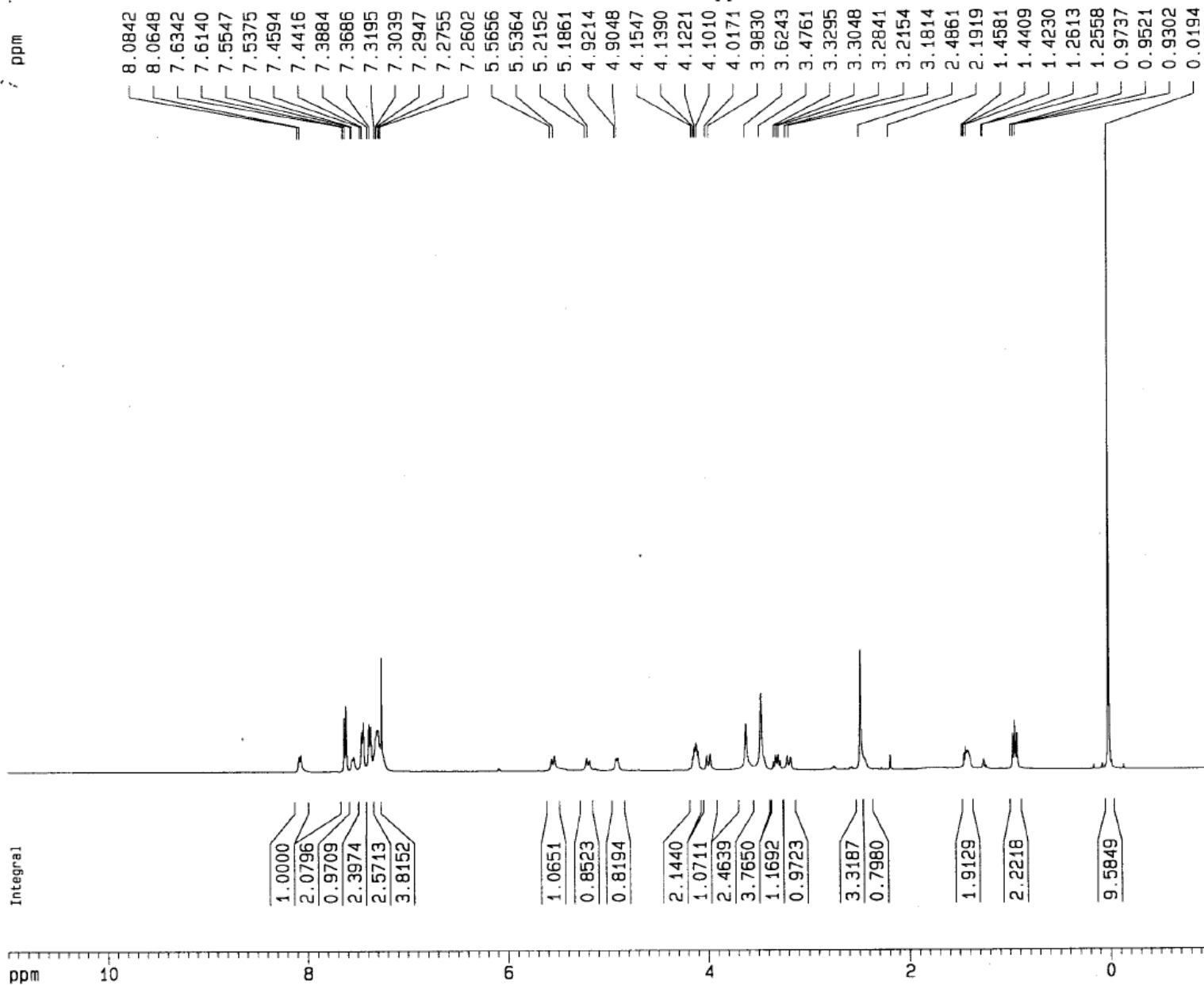
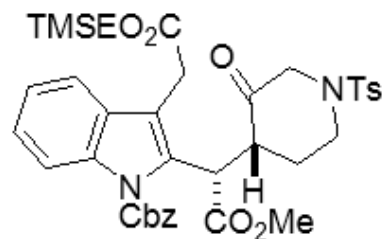
F2 - Acquisition Parameters
 Date_ 20120926
 Time 7.49
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 8192
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 4096
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec

----- CHANNEL f1 -----
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SFO1 75.4106357 MHz

----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SFO2 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4023694 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 180.000 ppm
 F1 13572.43 Hz
 F2P -10.000 ppm
 F2 -754.02 Hz
 PPMCM 9.50000 ppm/cm
 HZCM 716.32251 Hz/cm



Current Data Parameters
 NAME 092612FYQ-17
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20120926
 Time 9.06
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 114
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====

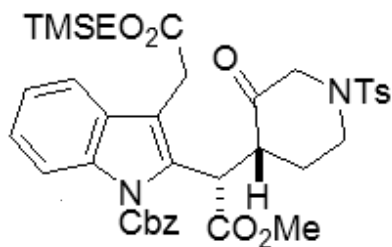
NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters

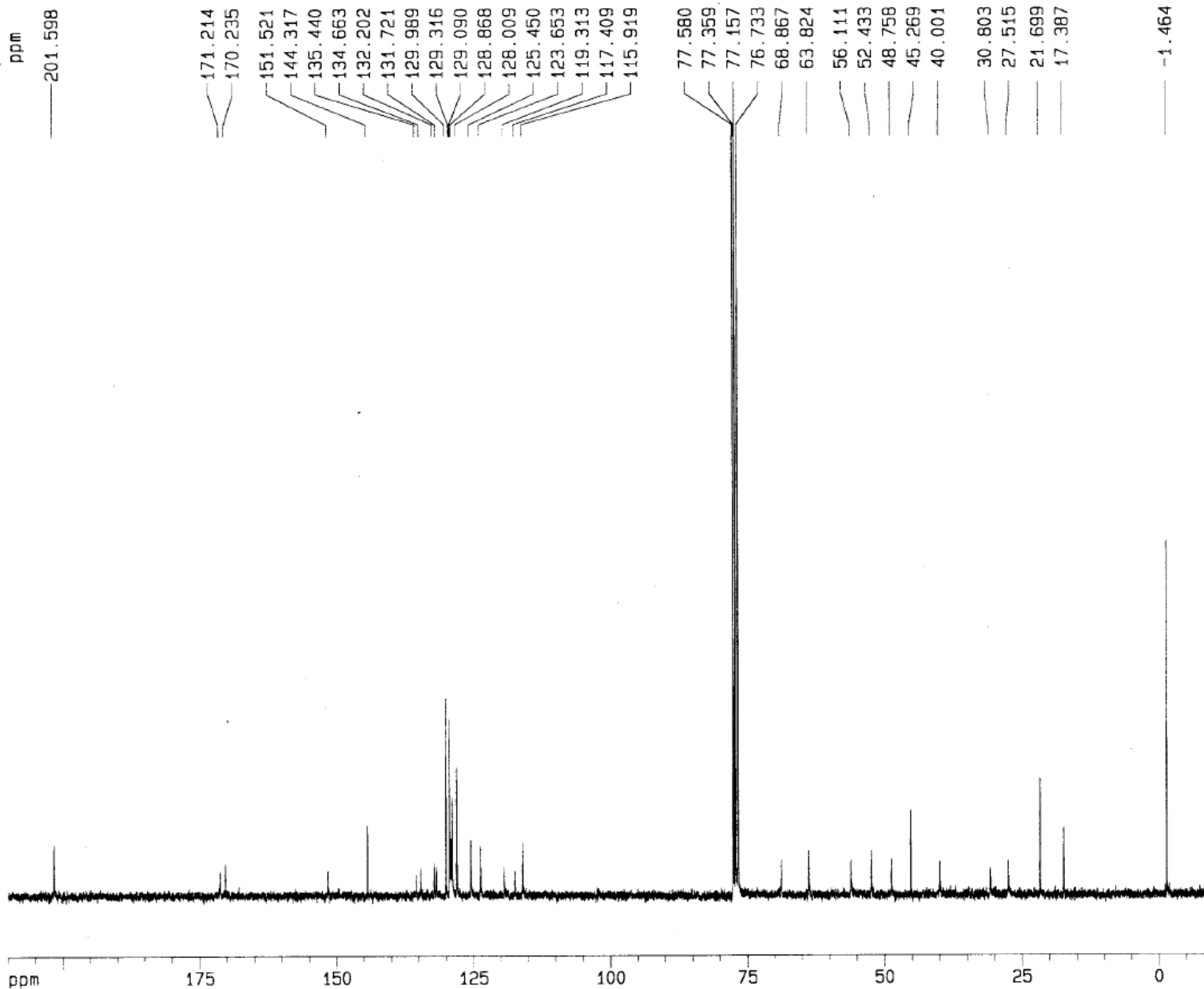
SI 32768
 SF 400.1300091 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters

CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



17



Current Data Parameters
 NAME 092512FYG-17
 EXPNO 2
 PROCNO 1

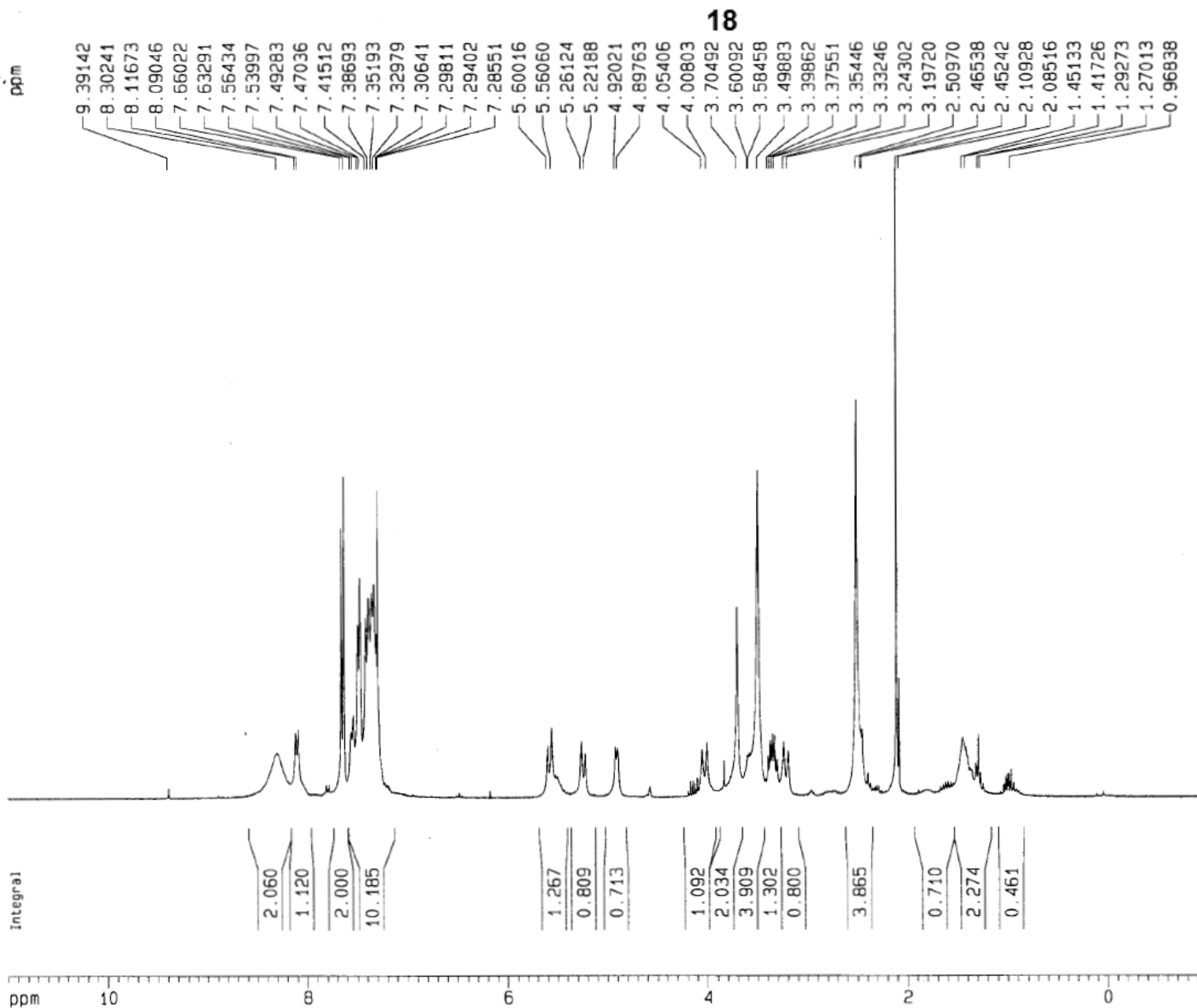
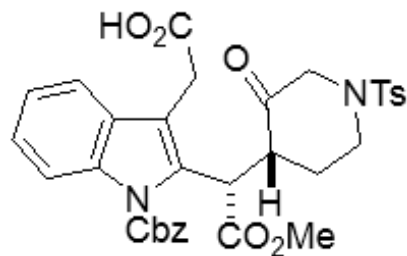
F2 - Acquisition Parameters
 Date_ 20120925
 Time 21.56
 INSTRUM spect
 PROBHD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 18832.393 Hz
 FIDRES 0.287360 Hz
 AQ 1.7400308 sec
 RG 13004
 DW 26.550 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 d12 0.0002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.75 usec
 PL1 0.00 dB
 SF01 75.4760200 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 110.00 usec
 PL2 0.00 dB
 PL12 17.50 dB
 PL13 17.50 dB
 SF02 300.1312005 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4677428 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 210.000 ppm
 F1 15848.23 Hz
 F2P -10.000 ppm
 F2 -754.68 Hz
 PPMCM 11.00000 ppm/cm
 HZCM 830.14514 Hz/cm



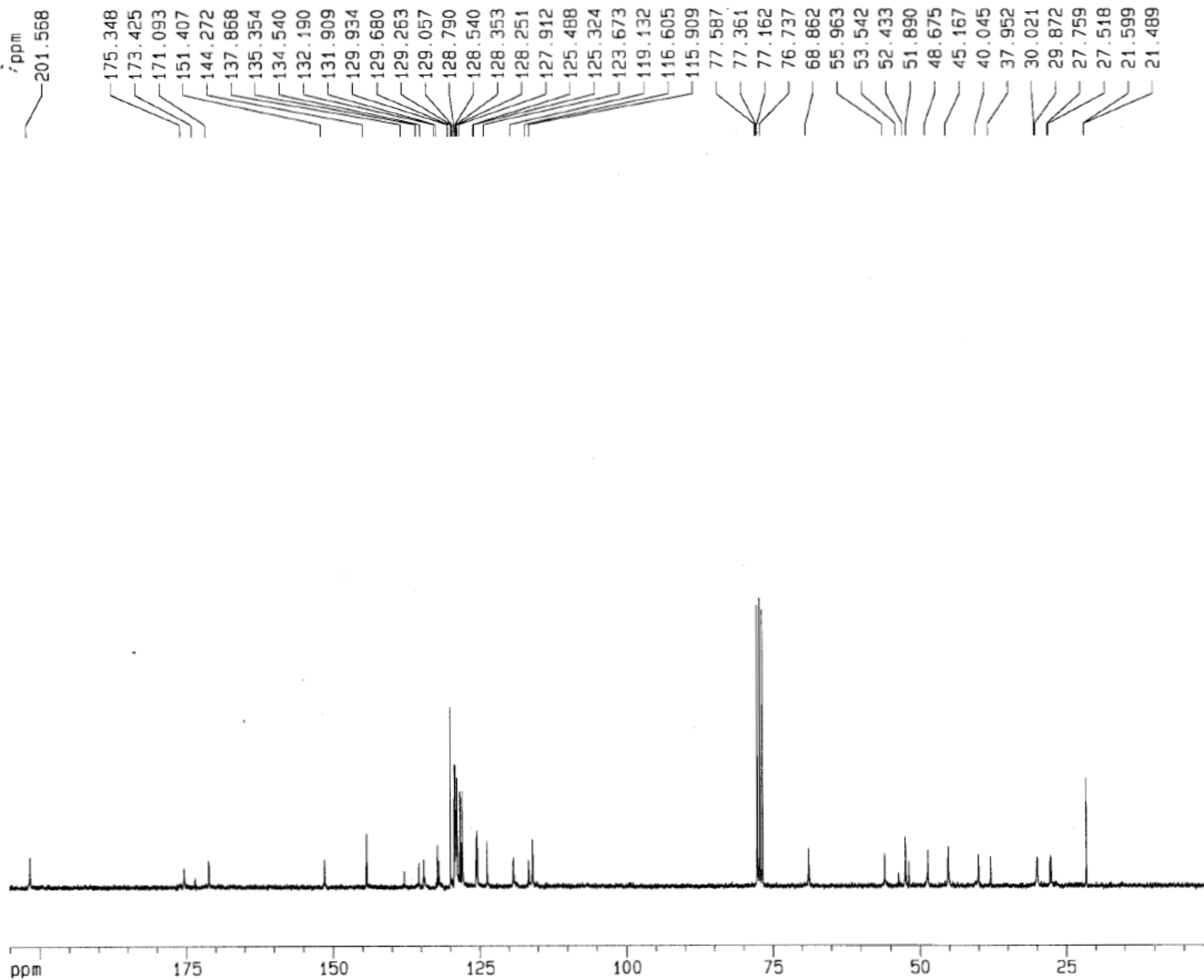
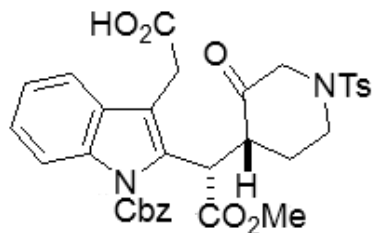
Current Data Parameters
 NAME 032310mm5085
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20100323
 Time 6.57
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zg30
 TD 24690
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 6172.839 Hz
 FIDRES 0.250014 Hz
 AQ 1.9999400 sec
 RG 228.1
 DW 81.000 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 11.70 usec
 PL1 0.00 dB
 SFO1 299.8718518 MHz

F2 - Processing parameters
 SI 32768
 SF 299.8700000 MHz
 WDW no
 SSB 0
 LB 0.00 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 3298.57 Hz
 F2P -1.000 ppm
 F2 -299.87 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 179.92200 Hz/cm



Current Data Parameters

NAME 092512FYQ-18
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters

Date_ 20120925
Time 23.10
INSTRUM spect
PROBHD 5 mm GNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 769
DS 4
SWH 18796.992 Hz
FIDRES 0.286819 Hz
AQ 1.7433076 sec
RG 4096
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec

----- CHANNEL f1 -----
NUC1 13C
P1 5.25 usec
PL1 -6.00 dB
SFO1 75.4106357 MHz

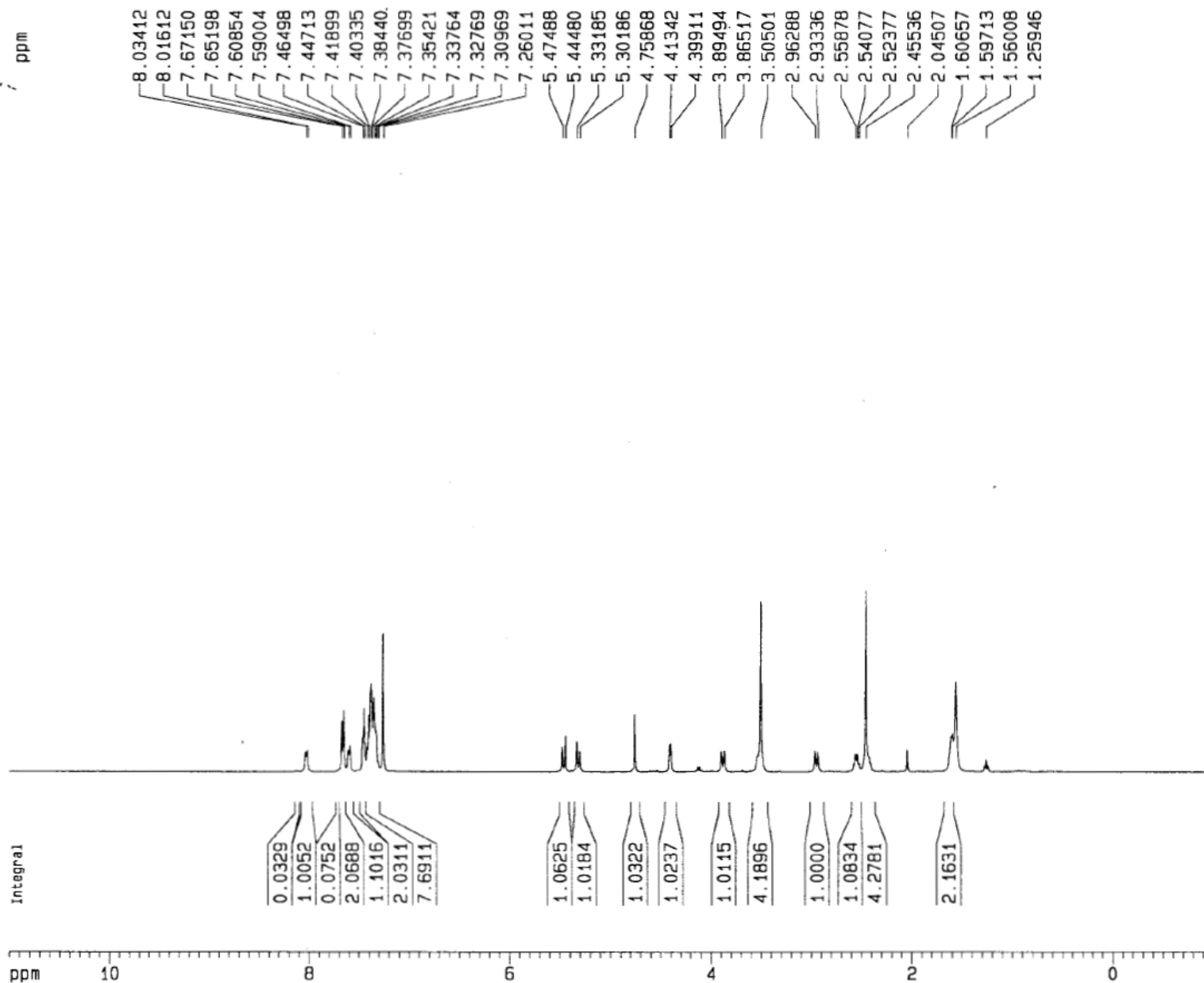
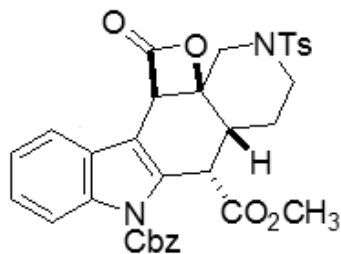
----- CHANNEL f2 -----
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 19.70 dB
PL13 19.70 dB
SFO2 299.8711995 MHz

F2 - Processing parameters

SI 32768
SF 75.4023746 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
F1P 205.000 ppm
F1 15457.49 Hz
F2P 0.000 ppm
F2 0.00 Hz
PPMCM 10.25000 ppm/cm
HZCM 772.87439 Hz/cm



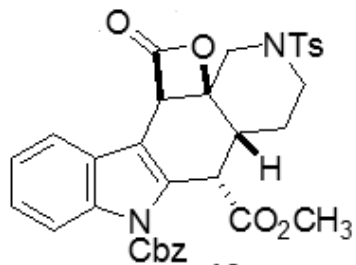
Current Data Parameters
 NAME 090112FJIV101
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120901
 Time 8.24
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 645.1
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

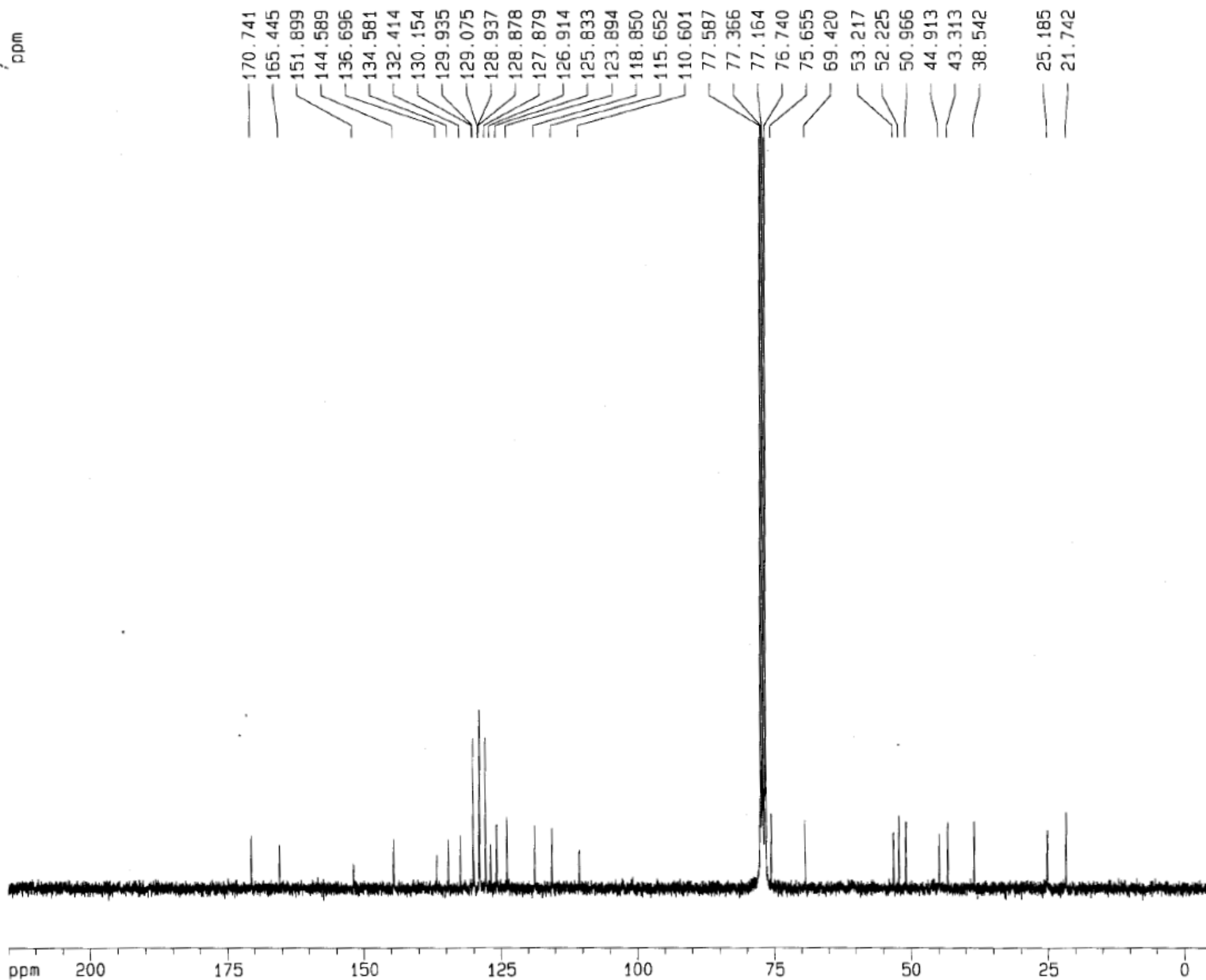
===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300094 MHz
 WDN EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



19



Current Data Parameters
NAME 092612FY0-19C
EXPNO 1
PROCNO 1

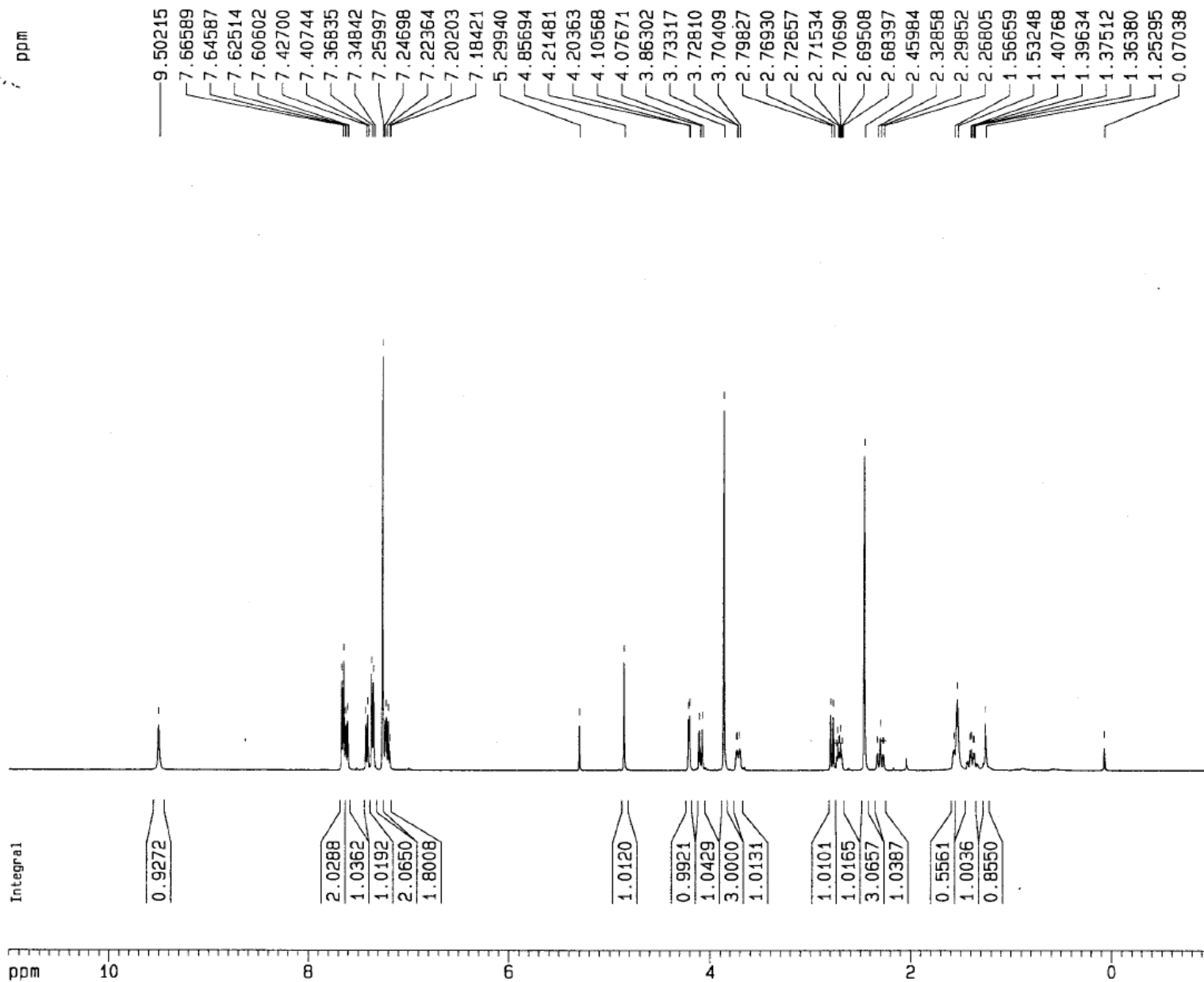
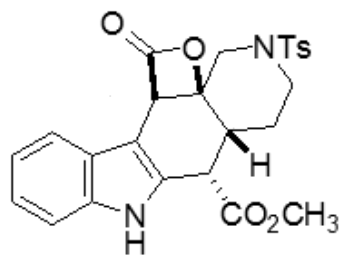
F2 - Acquisition Parameters
Date_ 20120927
Time 6.39
INSTRUM spect
PROBHD 5 mm QNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 7168
DS 4
SWH 18796.992 Hz
FIDRES 0.286819 Hz
AQ 1.7433076 sec
RG 812.7
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
D11 0.03000000 sec
D12 0.00002000 sec

----- CHANNEL f1 -----
NUC1 13C
P1 5.25 usec
PL1 -6.00 dB
SF01 75.4106357 MHz

----- CHANNEL f2 -----
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 19.70 dB
PL13 19.70 dB
SF02 299.8711995 MHz

F2 - Processing parameters
SI 32768
SF 75.4023625 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
F1P 215.000 ppm
F1 16211.51 Hz
F2P -5.000 ppm
F2 -377.01 Hz
PPMCM 11.00000 ppm/cm
HZCM 829.42596 Hz/cm



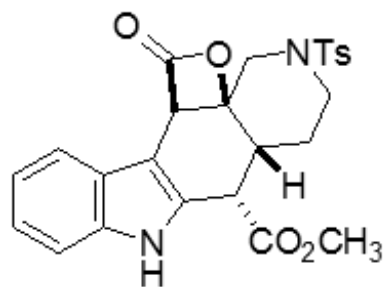
Current Data Parameters
 NAME 092612FY0-20
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120926
 Time 22.45
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 574.7
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

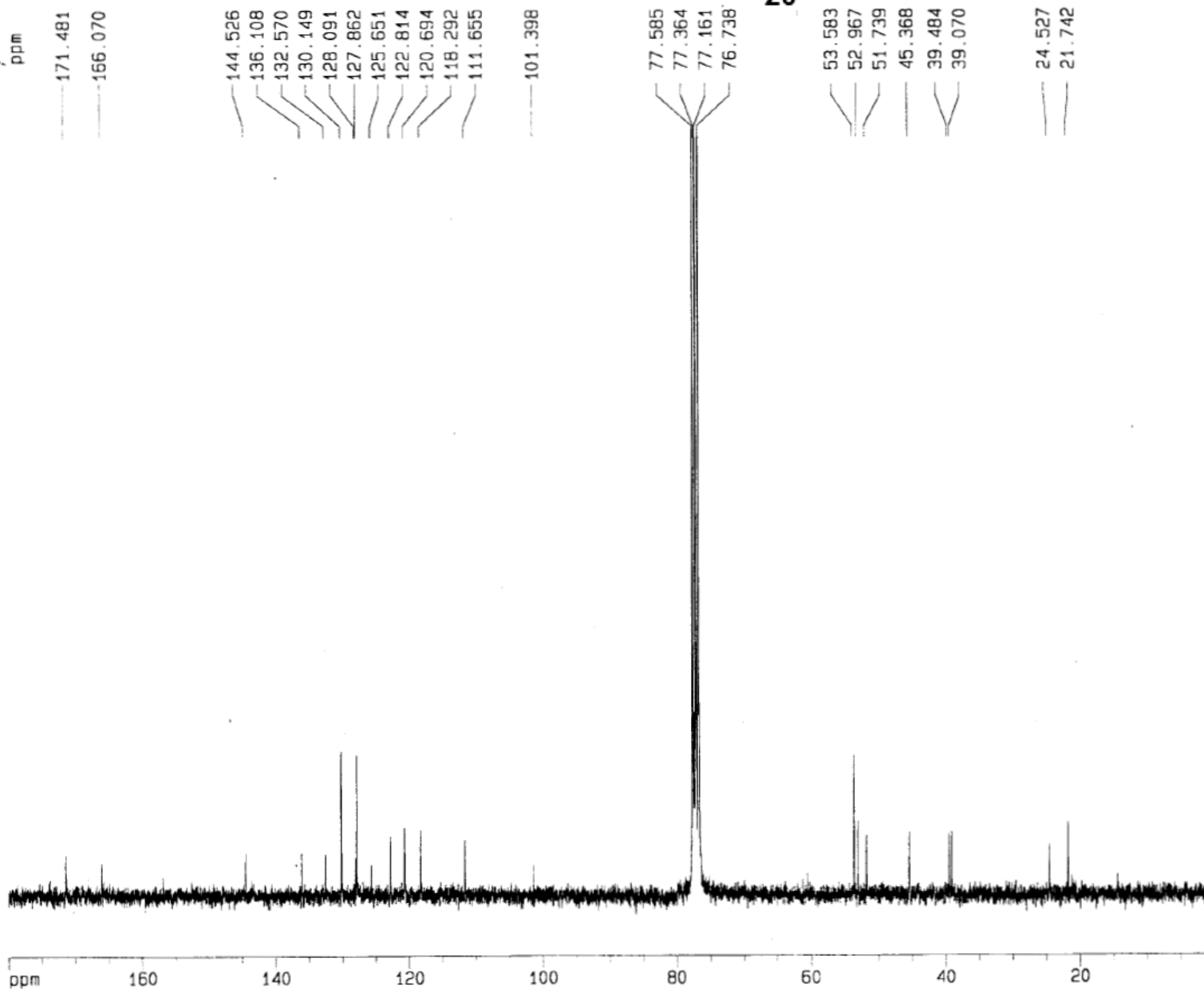
===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



20



Current Data Parameters
 NAME 092312FYGcpd20
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20120924
 Time 9.30
 INSTRUM spect
 PROBHD 5 mm Multinu
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 19456
 DS 4
 SWH 18832.393 Hz
 FIDRES 0.287360 Hz
 AQ 1.7400308 sec
 RG 11585.2
 DW 26.550 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 d12 0.0000200 sec

----- CHANNEL f1 -----
 NUC1 13C
 P1 9.75 usec
 PL1 0.00 dB
 SF01 75.4760200 MHz

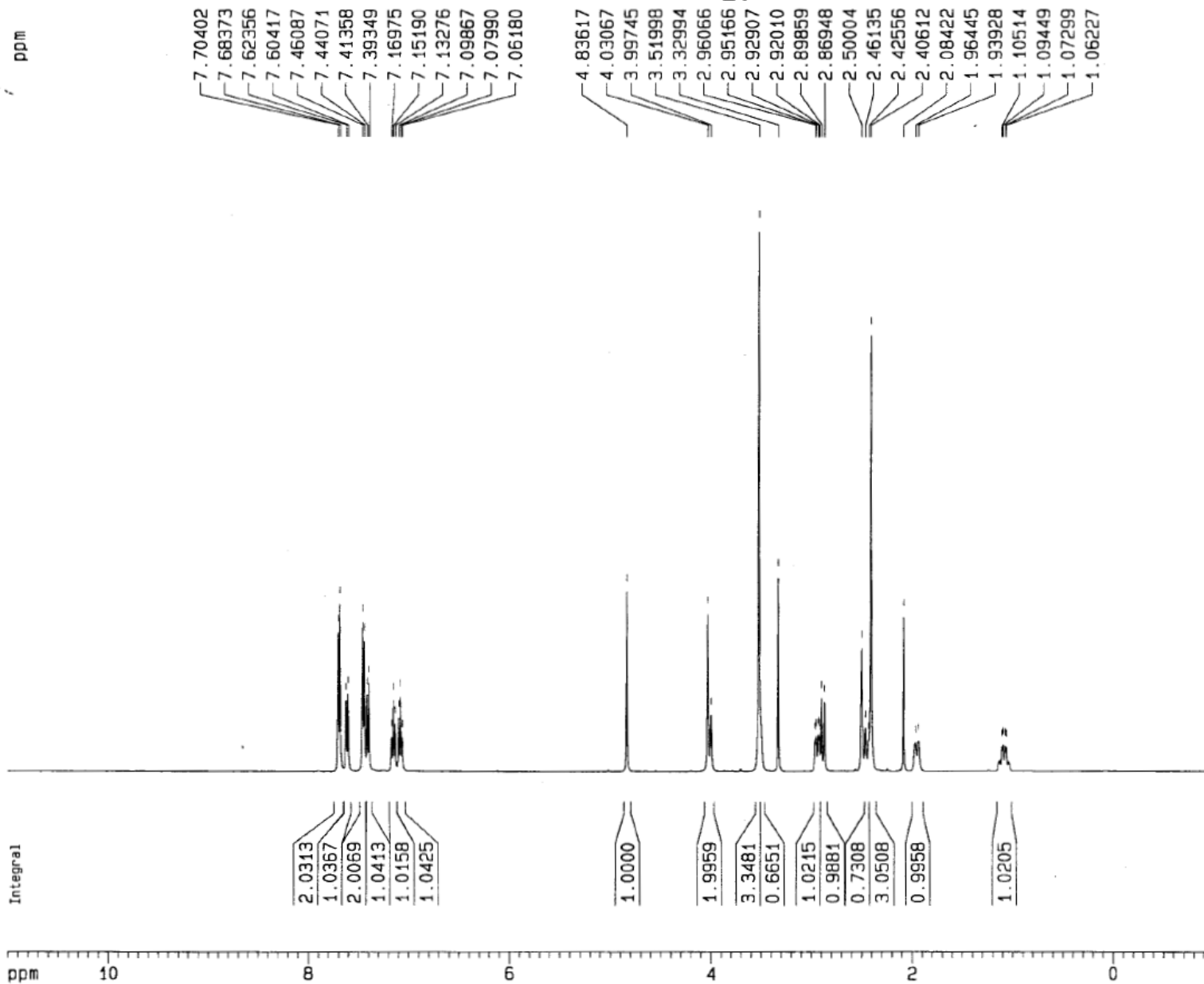
----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 110.00 usec
 PL2 0.00 dB
 PL12 17.50 dB
 PL13 17.50 dB
 SF02 300.1312005 MHz

F2 - Processing parameters

SI 32768
 SF 75.4677382 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters

CX 20.00 cm
 F1P 180.000 ppm
 F1 13584.19 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPNCM 9.00000 ppm/cm
 HZCM 679.20959 Hz/cm



Current Data Parameters

NAME 092612FYQ-21
EXPNO 5
PROCNO 1

F2 - Acquisition Parameters

Date_ 20120926
Time 23.04
INSTRUM spect
PROBHD 5 mm BBI 1H-
PULPROG zg30
TD 65536
SOLVENT DMSO
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 362
DW 60.400 usec
DE 6.00 usec
TE 300.0 K
D1 1.00000000 sec

----- CHANNEL f1 -----

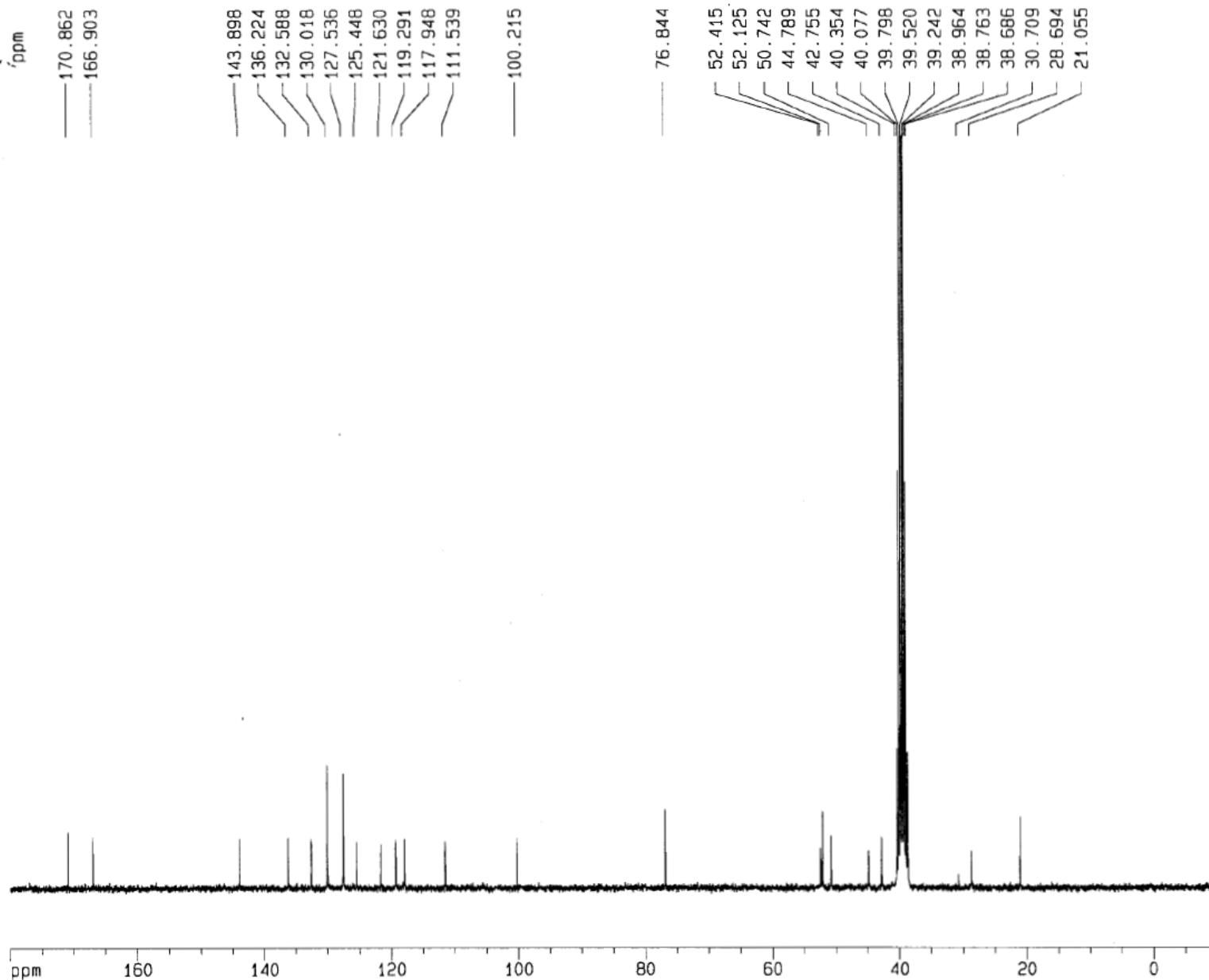
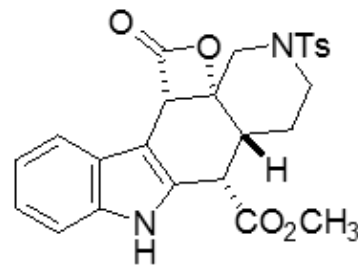
NUC1 1H
P1 6.45 usec
PL1 0.00 dB
SF01 400.1324710 MHz

F2 - Processing parameters

SI 32768
SF 400.1300029 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

1D NMR plot parameters

CX 20.00 cm
F1P 11.000 ppm
F1 4401.43 Hz
F2P -1.000 ppm
F2 -400.13 Hz
PPMCM 0.60000 ppm/cm
HZCM 240.07800 Hz/cm



Current Data Parameters

NAME 092612FYQ-21
EXPNO 2
PROCNO 1

F2 - Acquisition Parameters

Date_ 20120926
Time 23.03
INSTRUM spect
PROBHD 5 mm GNP 1H/1
PULPROG zgpg30
TD 65536
SOLVENT DMSO
NS 1375
DS 4
SWH 18796.992 Hz
FIDRES 0.286819 Hz
AQ 1.7433076 sec
RG 512
DW 26.600 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
D11 0.0300000 sec
D12 0.0000200 sec

===== CHANNEL f1 =====

NUC1 13C
P1 5.25 usec
PL1 -6.00 dB
SFO1 75.4106357 MHz

===== CHANNEL f2 =====

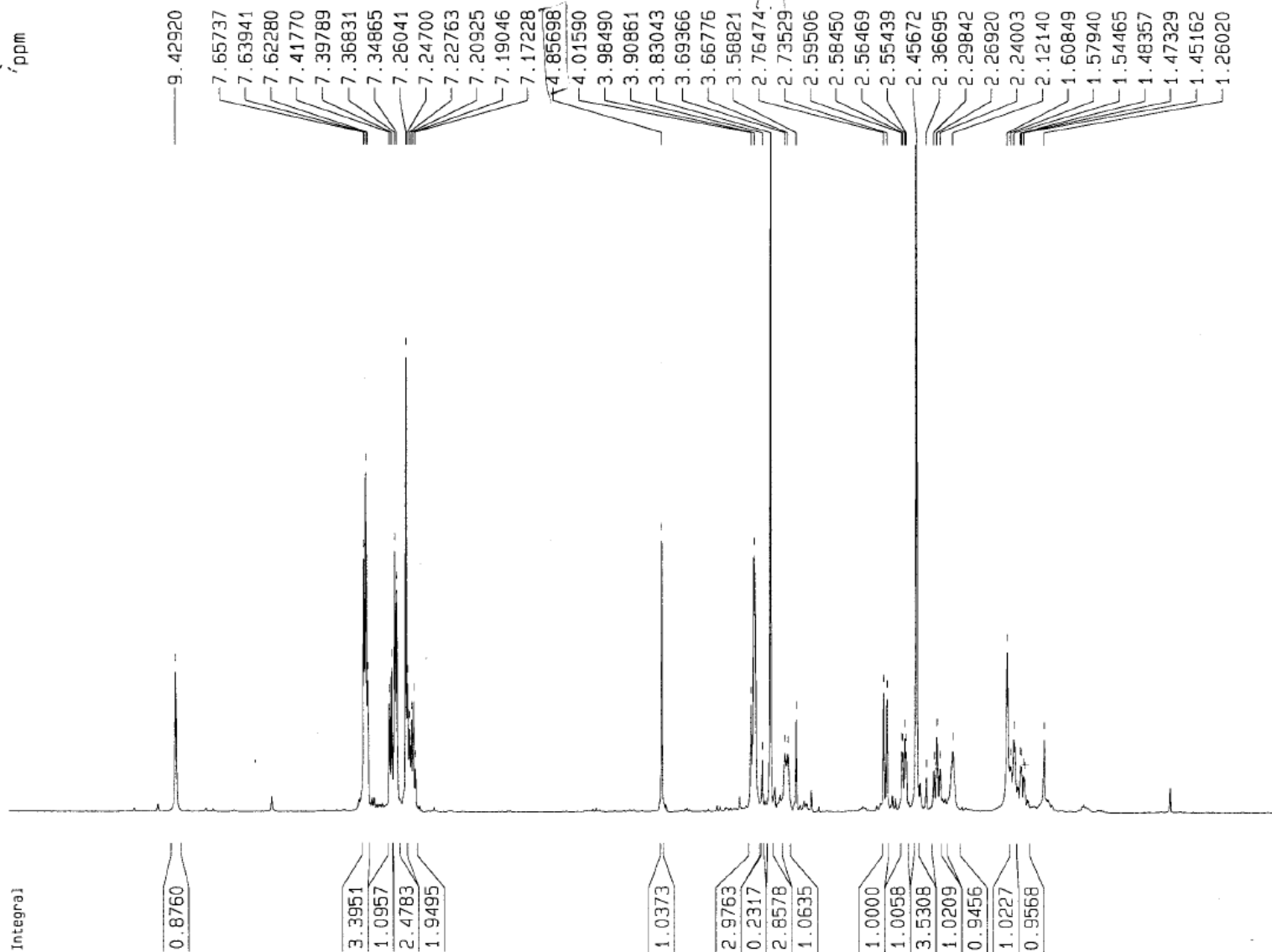
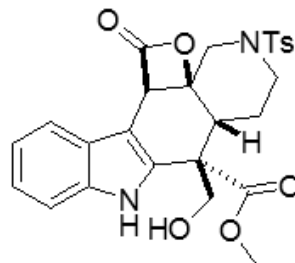
CPDPRG2 waltz16
NUC2 1H
PCPD2 115.00 usec
PL2 0.00 dB
PL12 19.70 dB
PL13 19.70 dB
SFO2 299.8711995 MHz

F2 - Processing parameters

SI 32768
SF 75.4024070 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters

CX 20.00 cm
F1P 180.000 ppm
F1 13572.43 Hz
F2P -10.000 ppm
F2 -754.02 Hz
PPMCM 9.50000 ppm/cm
HZCM 716.32288 Hz/cm



Current Data Parameters
 NAME 080111FJII-052
 EXPNO 2
 PROCNO 1

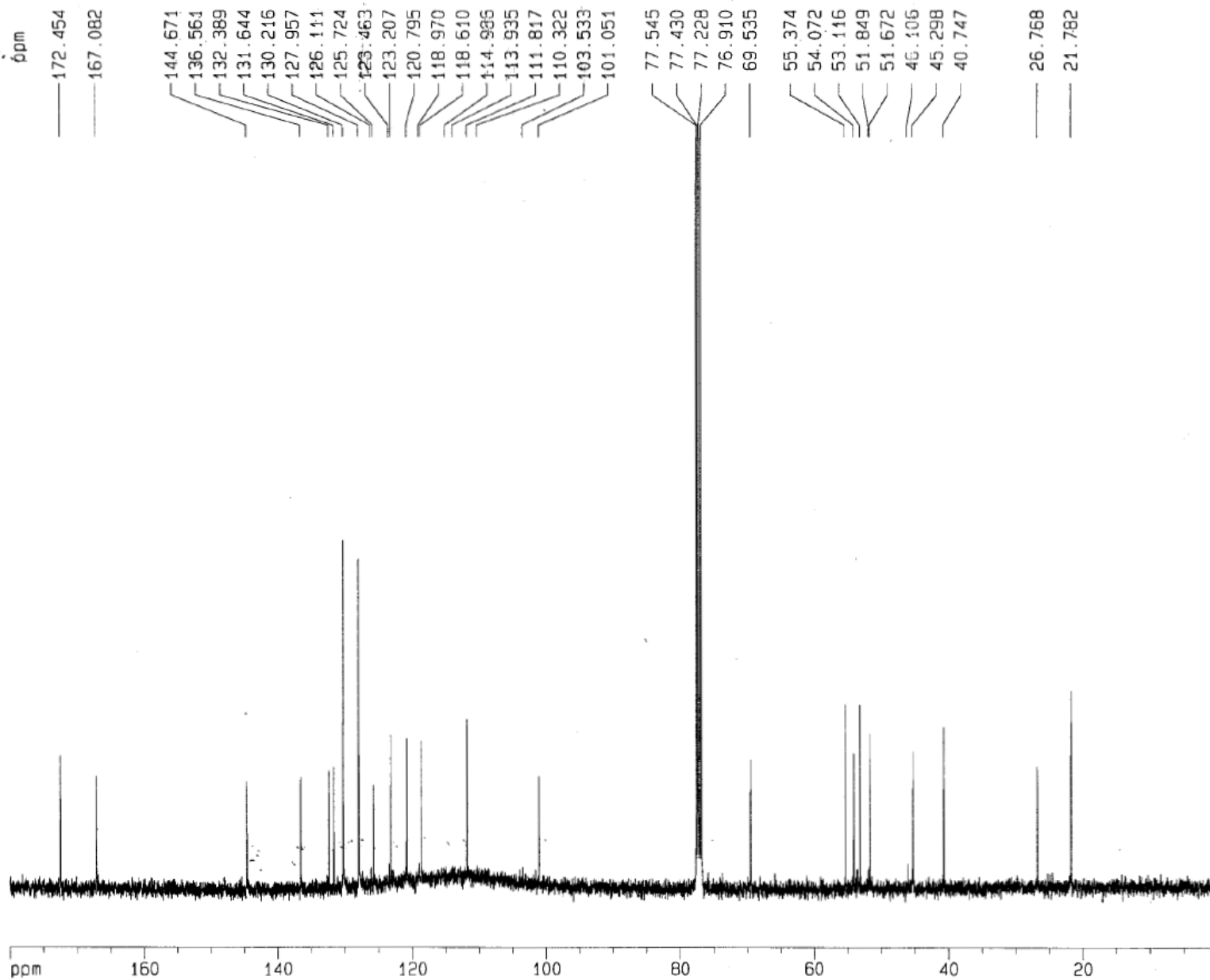
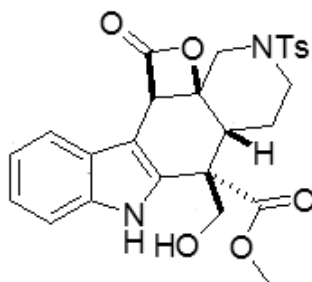
F2 - Acquisition Parameters
 Date_ 20110801
 Time 17.58
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 161.3
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300089 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm

ppm



Current Data Parameters

NAME 080111FJII-052
 EXPNO 7
 PROCNO 1

F2 - Acquisition Parameters

Date_ 20110802
 Time 2.55
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 5120
 DS 4
 SWH 25125.629 Hz
 FIDRES 0.383387 Hz
 AQ 1.3042164 sec
 RG 16384
 DW 19.900 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 d11 0.0300000 sec
 d12 0.00002000 sec

----- CHANNEL f1 -----

NUC1 13C
 P1 16.35 usec
 PL1 -6.00 dB
 SF01 100.6237959 MHz

----- CHANNEL f2 -----

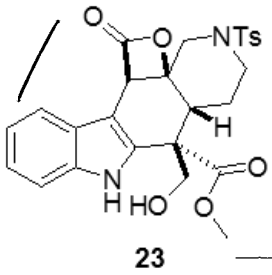
CPDPRG2 waltz16
 NUC2 1H
 PCPD2 114.00 usec
 PL2 0.00 dB
 PL12 24.00 dB
 PL13 24.00 dB
 SF02 400.1316005 MHz

F2 - Processing parameters

SI 32768
 SF 100.5127499 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

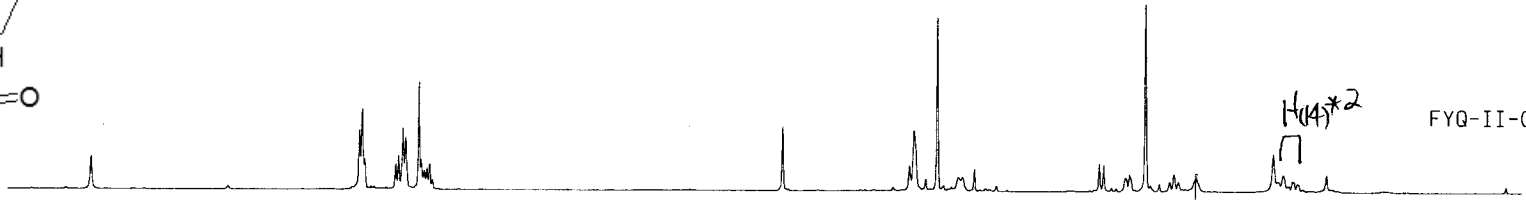
1D NMR plot parameters

CX 20.00 cm
 F1P 180.000 ppm
 F1 18110.29 Hz
 F2P 0.000 ppm
 F2 0.00 Hz
 PPMCM 9.00000 ppm/cm
 HZCM 905.51471 Hz/cm



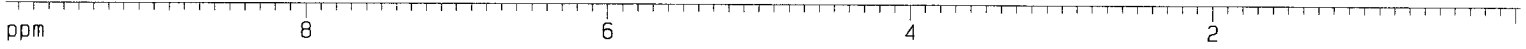
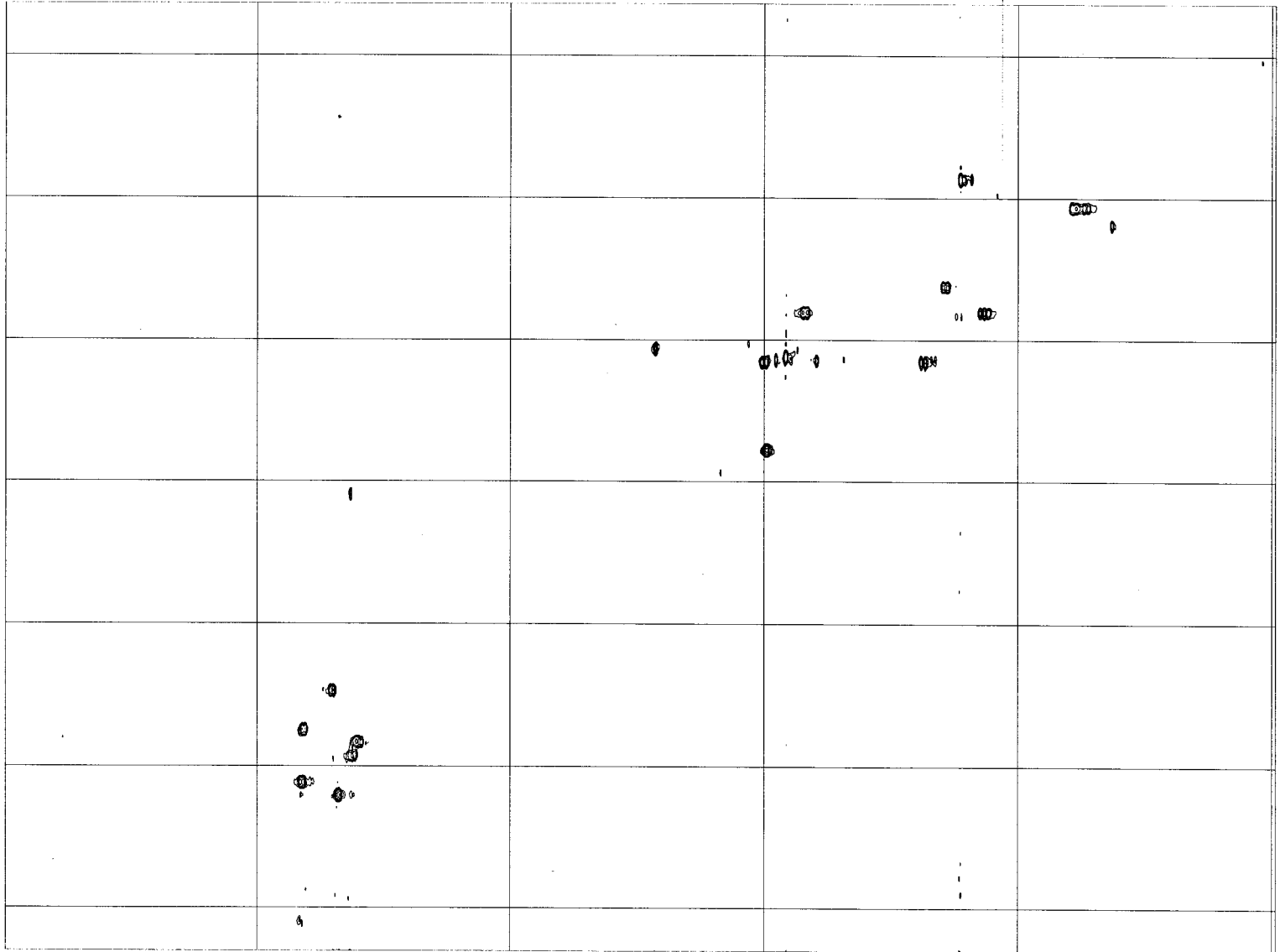
FYQ-II-052053P HMQC

(all)

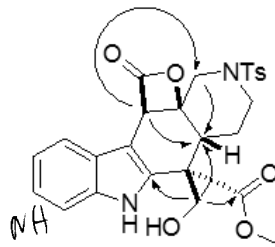


H(4)*2

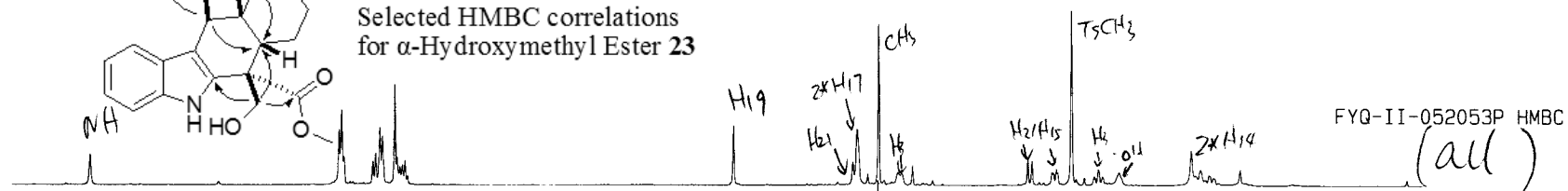
TSC(12) →
 C14 → CH₂
 C15(CH) → (CH)
 C3 → CH₂
 C19 → (CH)
 C20(CH) → (CH)
 C21(CH) → (CH)
 C7 → C16
 CH₂ - C17
 C7 (QC)
 Ar-CH → C9/11, C10/12, C11/13
 C8 (QC) →
 TSC(4) →
 C2 (OC) →
 Ts (OC) →
 C2 →
 TSC(OC) →



0
25
50
75
100
125
150
ppm

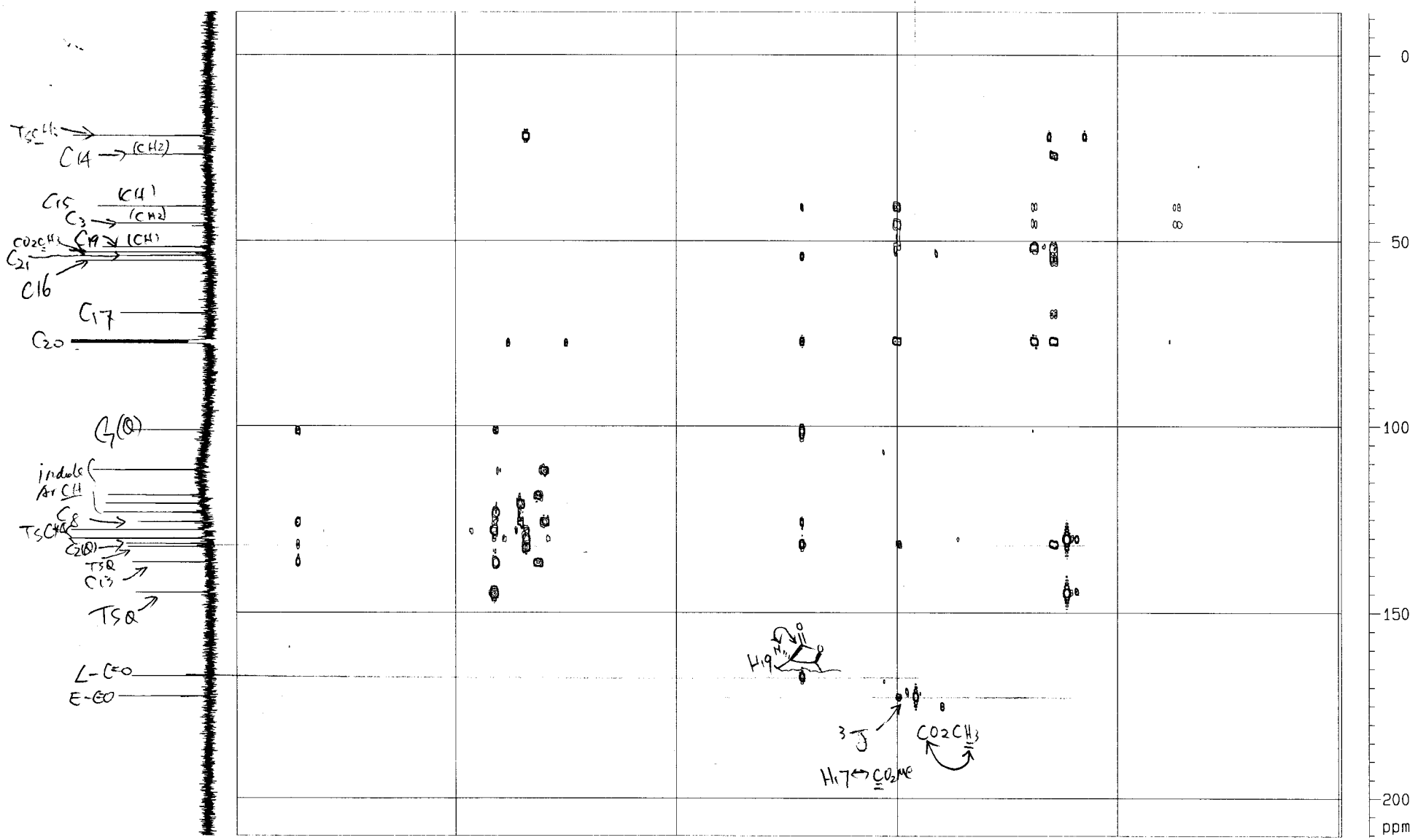


Selected HMBC correlations for α -Hydroxymethyl Ester 23



FYQ-II-052053P HMBC

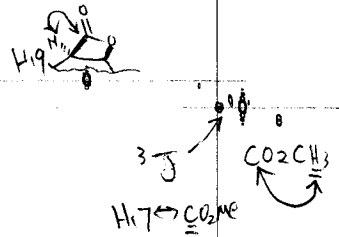
(all)

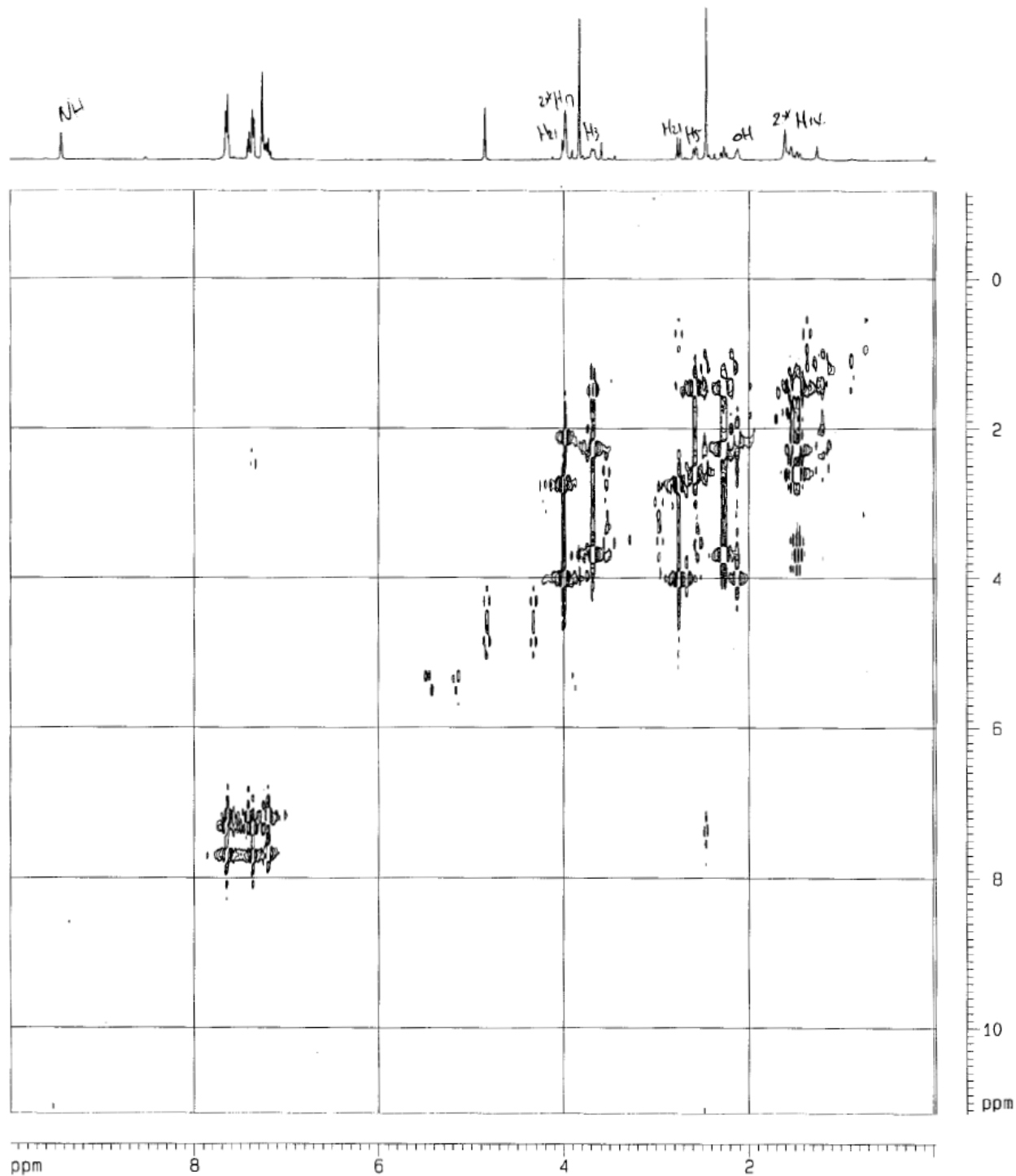


TSC_{CH_3}
 $C_{14} \rightarrow (CH_2)$
 $C_{15} \rightarrow (CH)$
 $C_3 \rightarrow (CH_2)$
 $CO_2CH_3 \rightarrow (CH)$
 $C_{21} \rightarrow (CH)$
 C_{16}
 C_{17}
 C_{20}

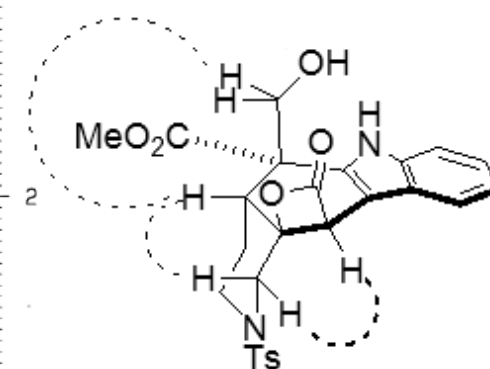
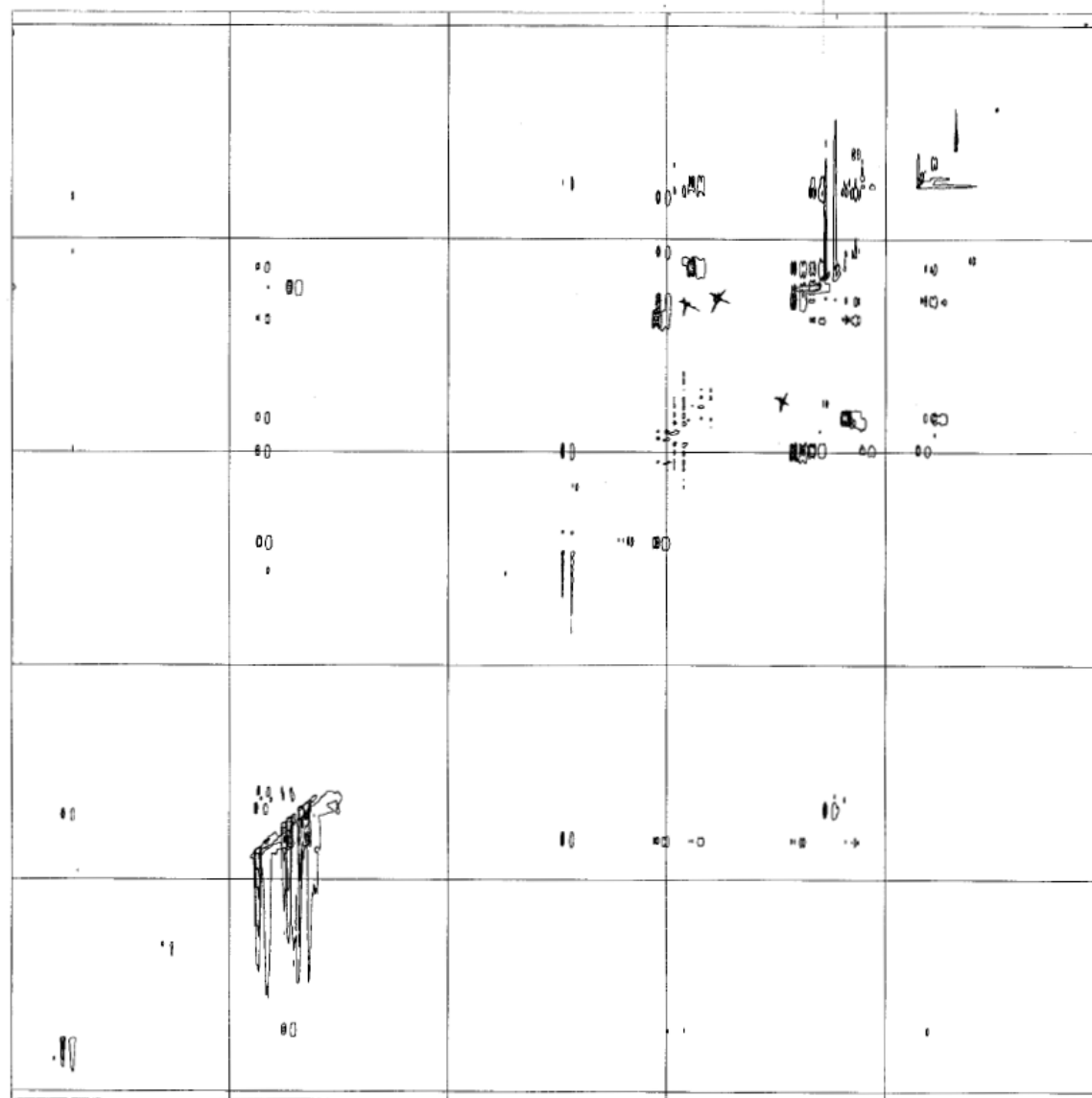
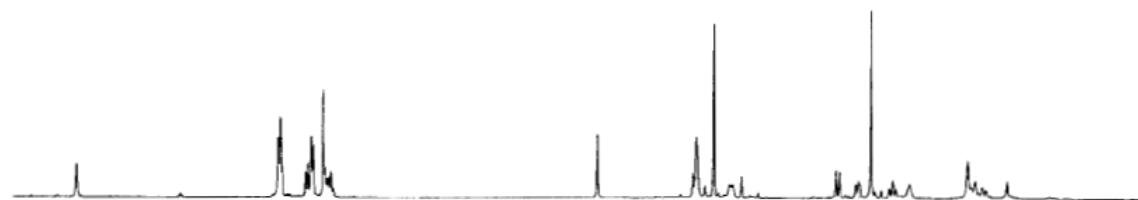
$C_7(O)$
 indols
 Ar-CH
 C_8
 TSC_{CH_2}
 C_{20}
 TSR
 C_{13}
 TSR

$L-C=O$
 $E-C=O$





FYQ-II-052053P NOESY



4 Key nOe correlations
for α -Hydroxymethyl Ester **23**

--- = nOe correlation

6

8

ppm

ppm

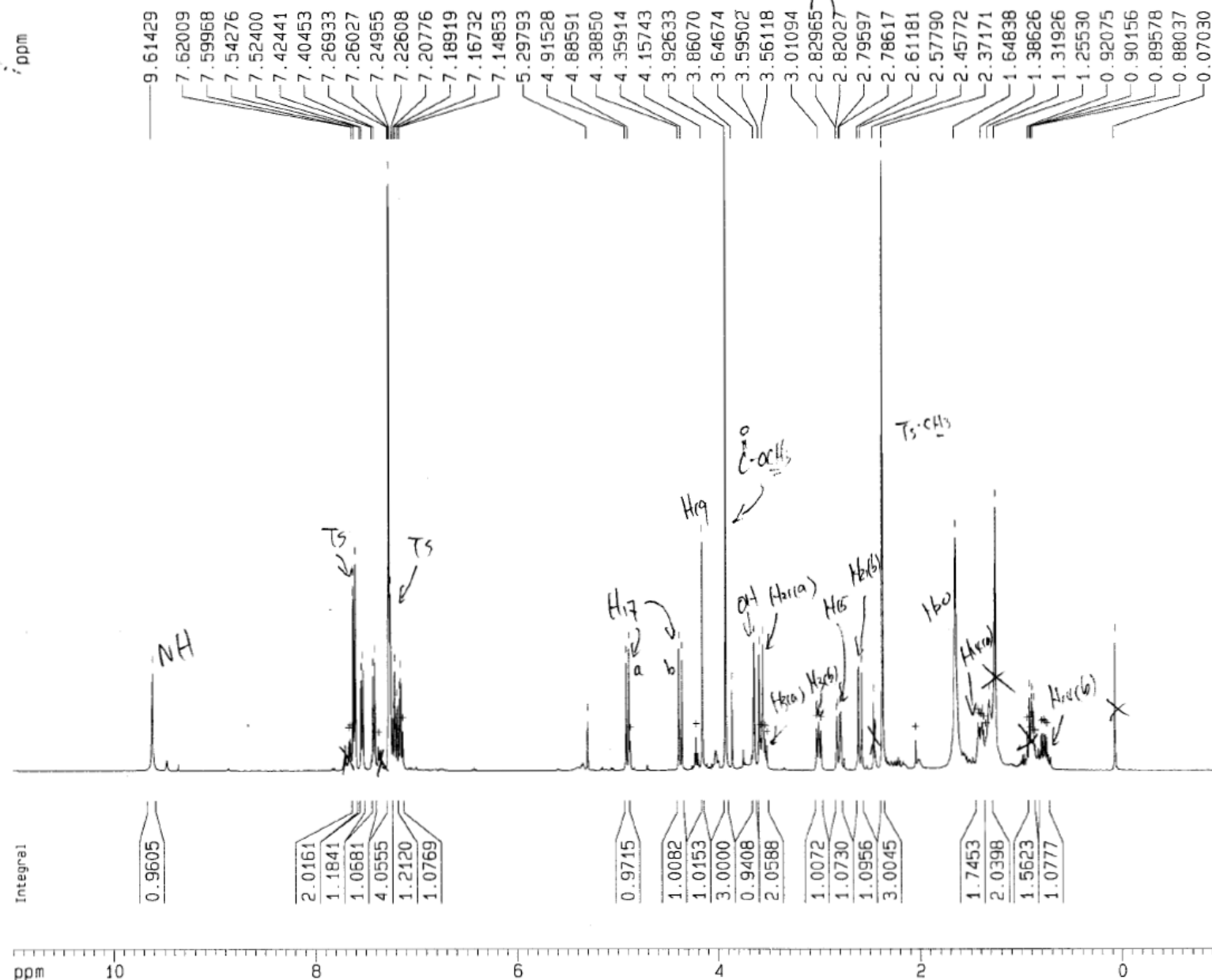
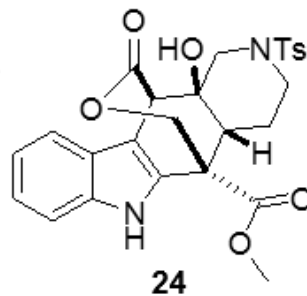
8

6

4

2





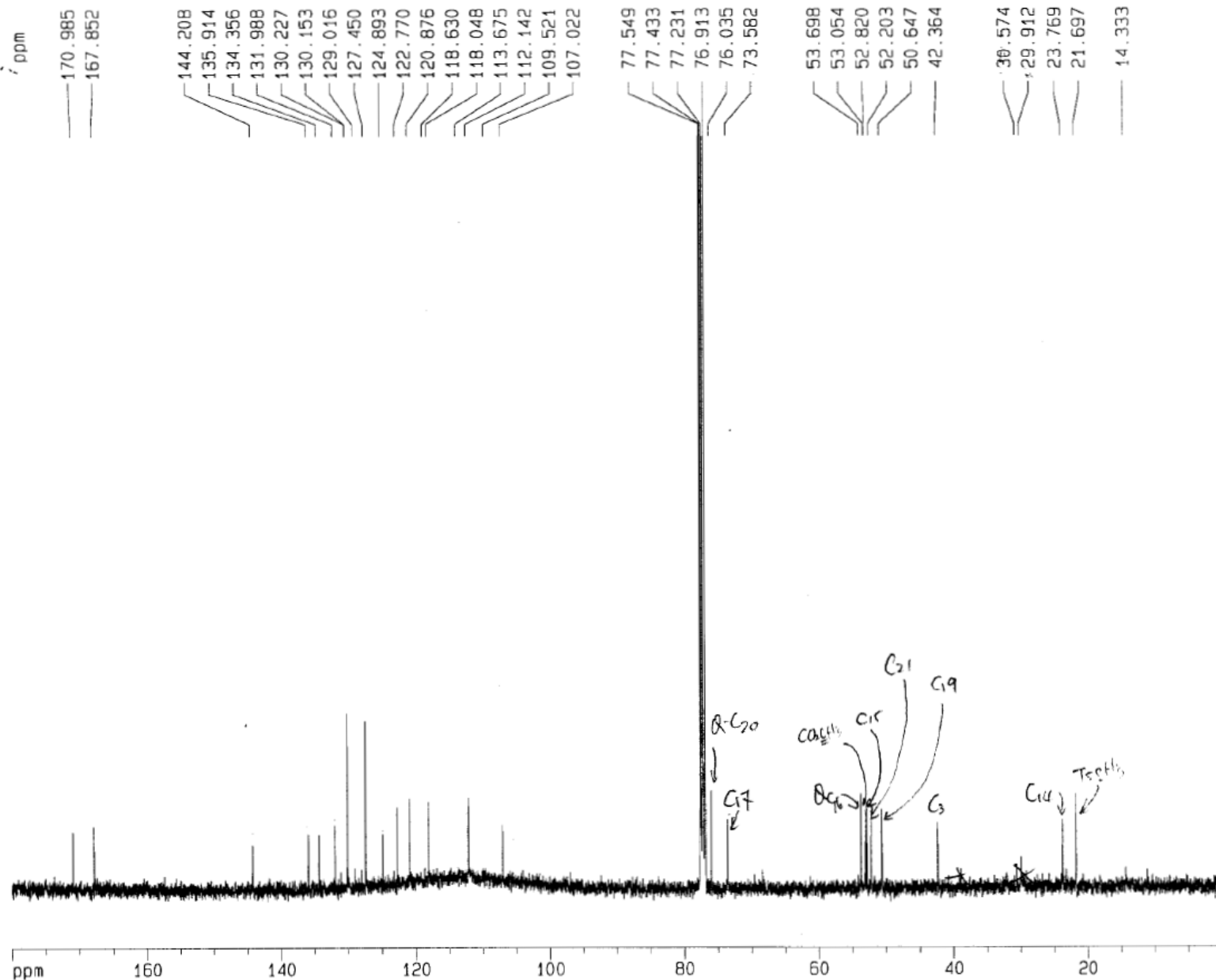
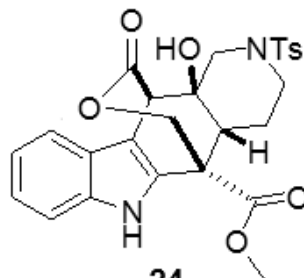
Current Data Parameters
 NAME 080811FJII-064
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20110808
 Time 18.57
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDC13
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 203.2
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300091 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
NAME 080811FJ11-064
EXPNO 7
PROCNO 1

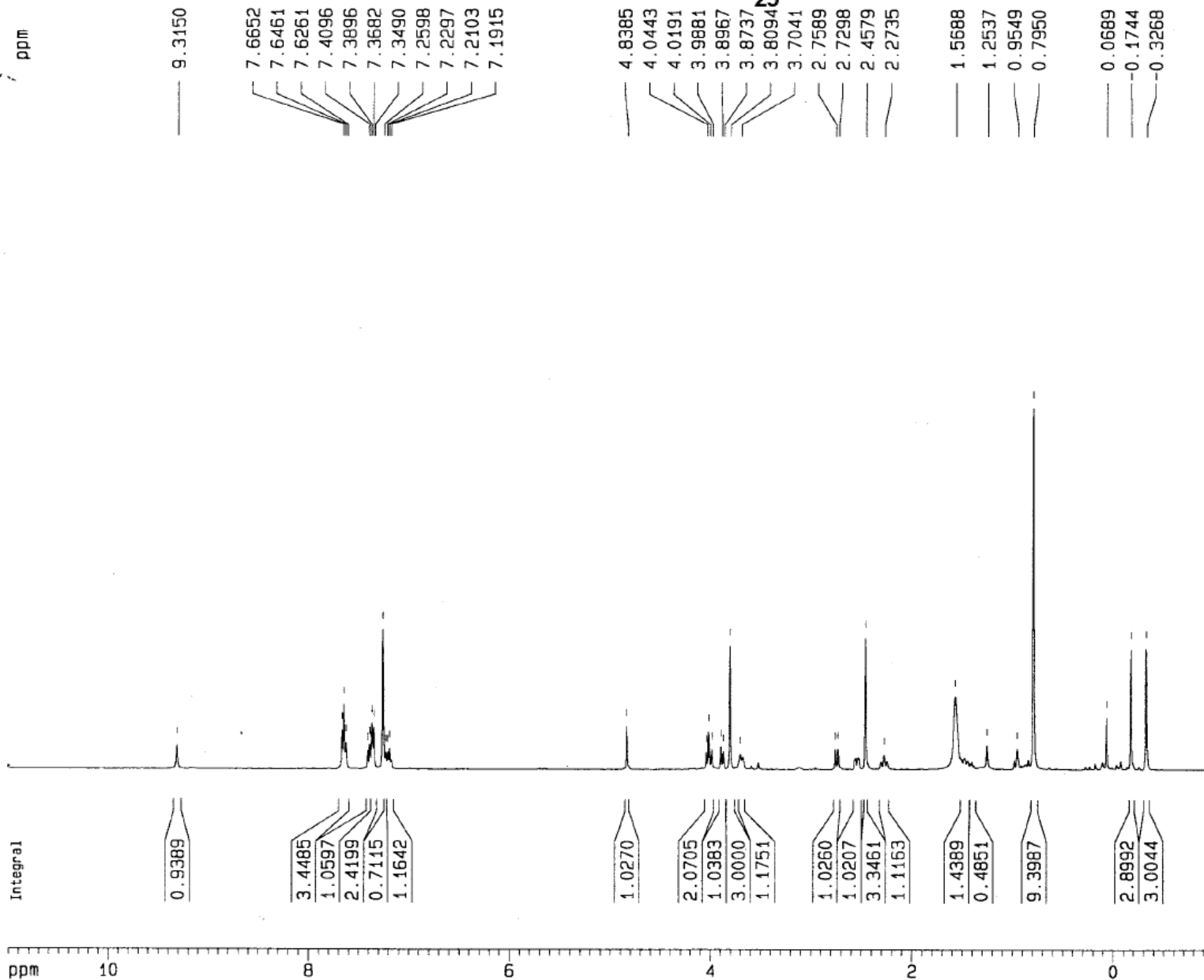
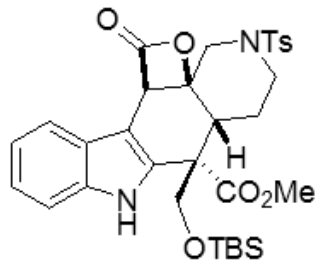
F2 - Acquisition Parameters
Date_ 20110809
Time 4.49
INSTRUM spect
PROBHD 5 mm BBI 1H-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 6144
DS 4
SWH 25125.629 Hz
FIDRES 0.383387 Hz
AQ 1.3042164 sec
RG 16384
DW 19.900 usec
DE 6.00 usec
TE 300.0 K
D1 2.00000000 sec
d11 0.03000000 sec
d12 0.00002000 sec

===== CHANNEL f1 =====
NUC1 13C
P1 16.35 usec
PL1 -6.00 dB
SFO1 100.6237959 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 114.00 usec
PL2 0.00 dB
PL12 24.00 dB
PL13 24.00 dB
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127484 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

1D NMR plot parameters
CX 20.00 cm
F1P 180.000 ppm
F1 18110.29 Hz
F2P 0.000 ppm
F2 0.00 Hz
PRMCM 9.00000 ppm/cm
HZCM 905.51471 Hz/cm



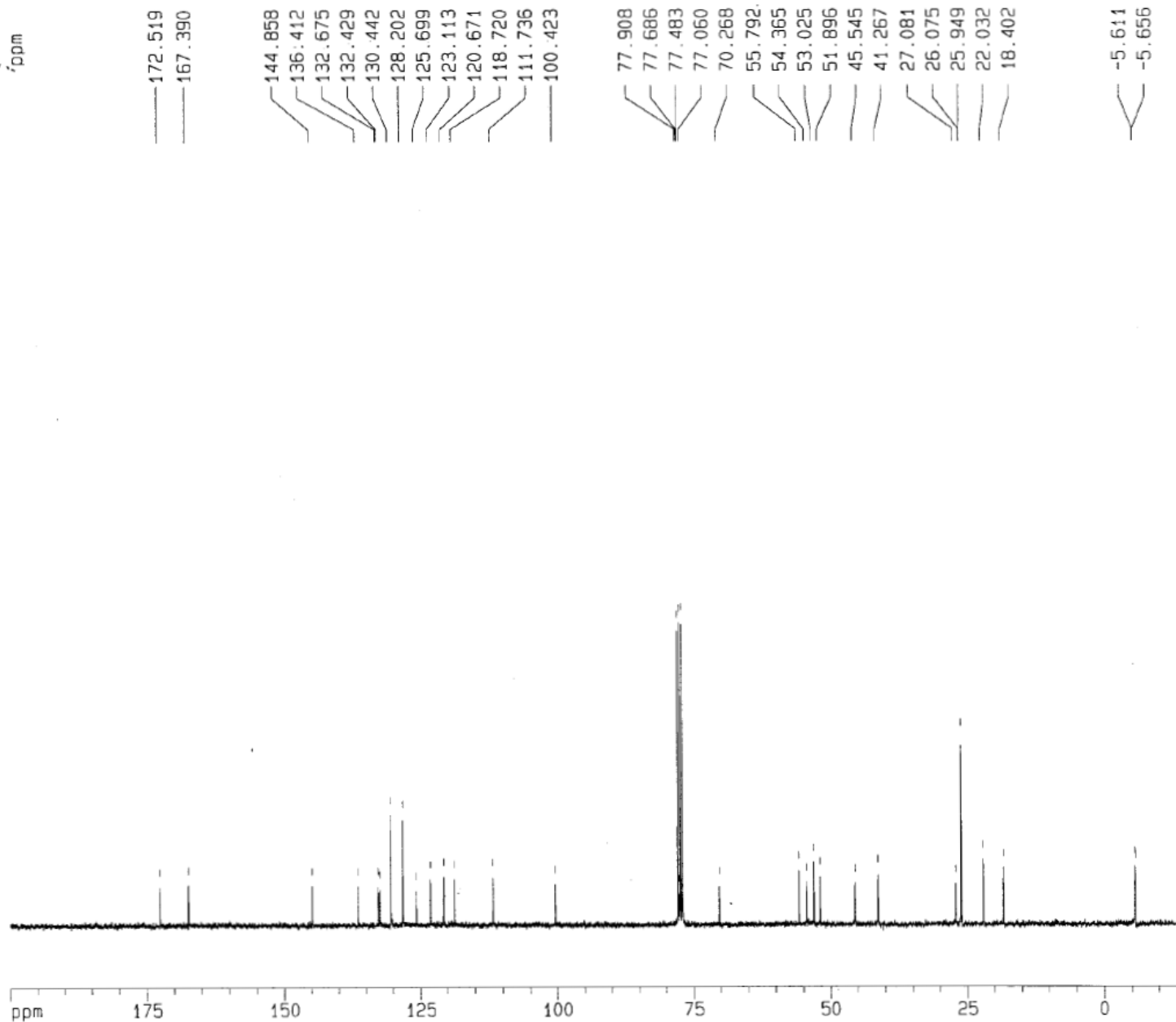
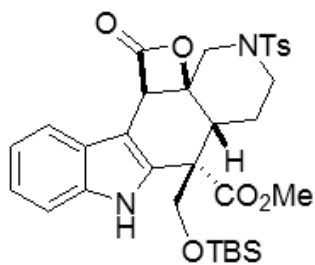
Current Data Parameters
 NAME 090712FJIII291
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120907
 Time 11.52
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 32
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 322.5
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
 NAME 090512FJIII290
 EXPNO 2
 PROCNO 1

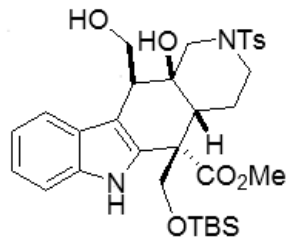
F2 - Acquisition Parameters
 Date_ 20120905
 Time 17.10
 INSTRUM spect
 PROBHD 5 mm GNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 830
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 512
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 D12 0.0000200 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SF01 75.4106357 MHz

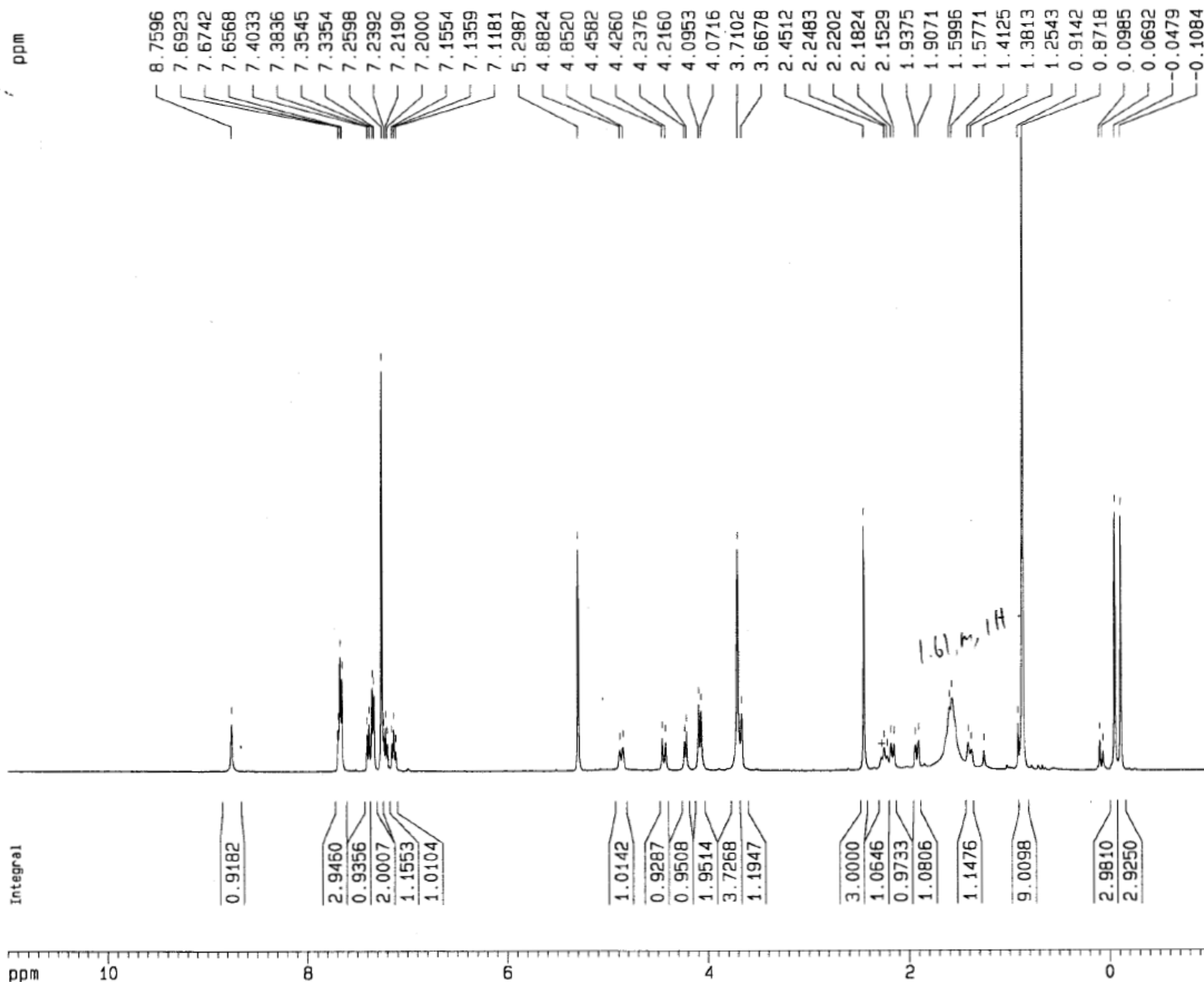
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SF02 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4023410 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 200.000 ppm
 F1 15080.47 Hz
 F2P -30.000 ppm
 F2 -2262.07 Hz
 PPMCM 11.50000 ppm/cm
 HZCM 867.12695 Hz/cm



26



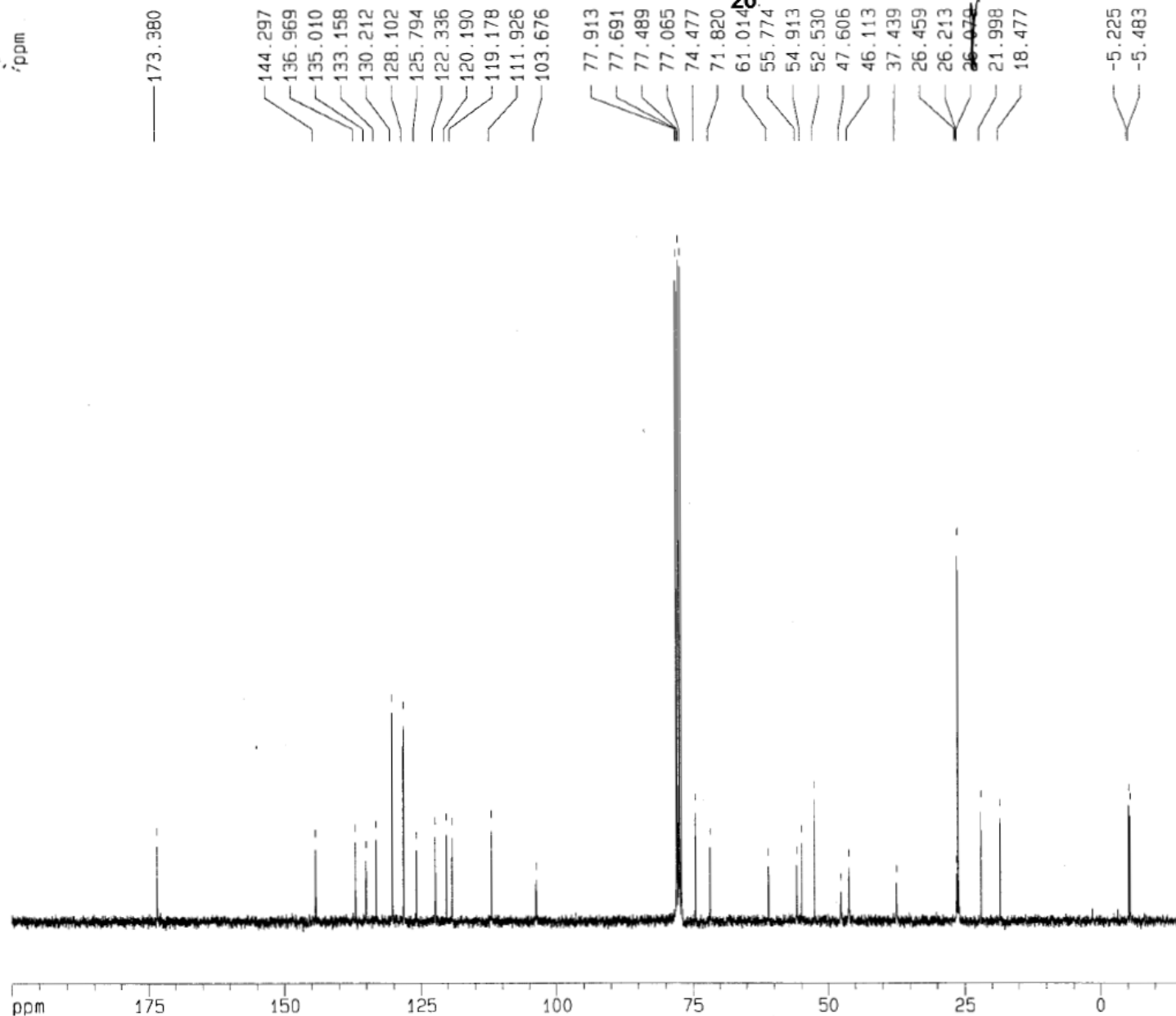
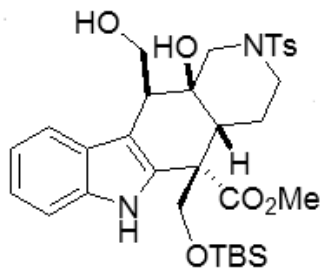
Current Data Parameters
 NAME 090712FJIII292
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120907
 Time 11.45
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 456.1
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SFO1 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300095 MHz
 WDM EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
 NAME 090612FJII292
 EXPNO 2
 PROCNO 1

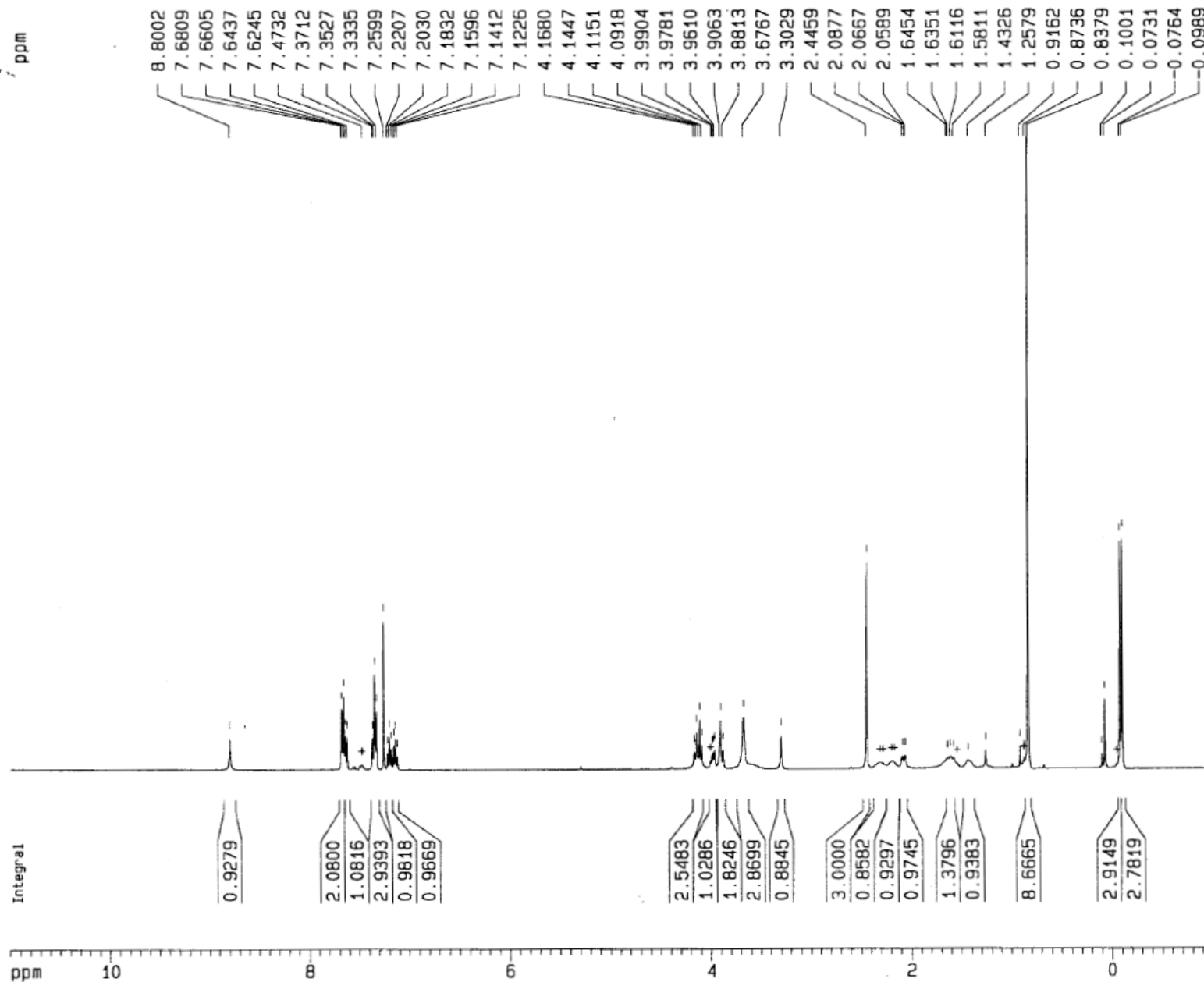
F2 - Acquisition Parameters
 Date_ 20120906
 Time 16.43
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 567
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 1024
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 D12 0.0002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SFO1 75.4106357 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SFO2 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4023410 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 200.000 ppm
 F1 15080.47 Hz
 F2P -30.000 ppm
 F2 -2262.07 Hz
 PPMCM 11.50000 ppm/cm
 HZCM 867.12695 Hz/cm



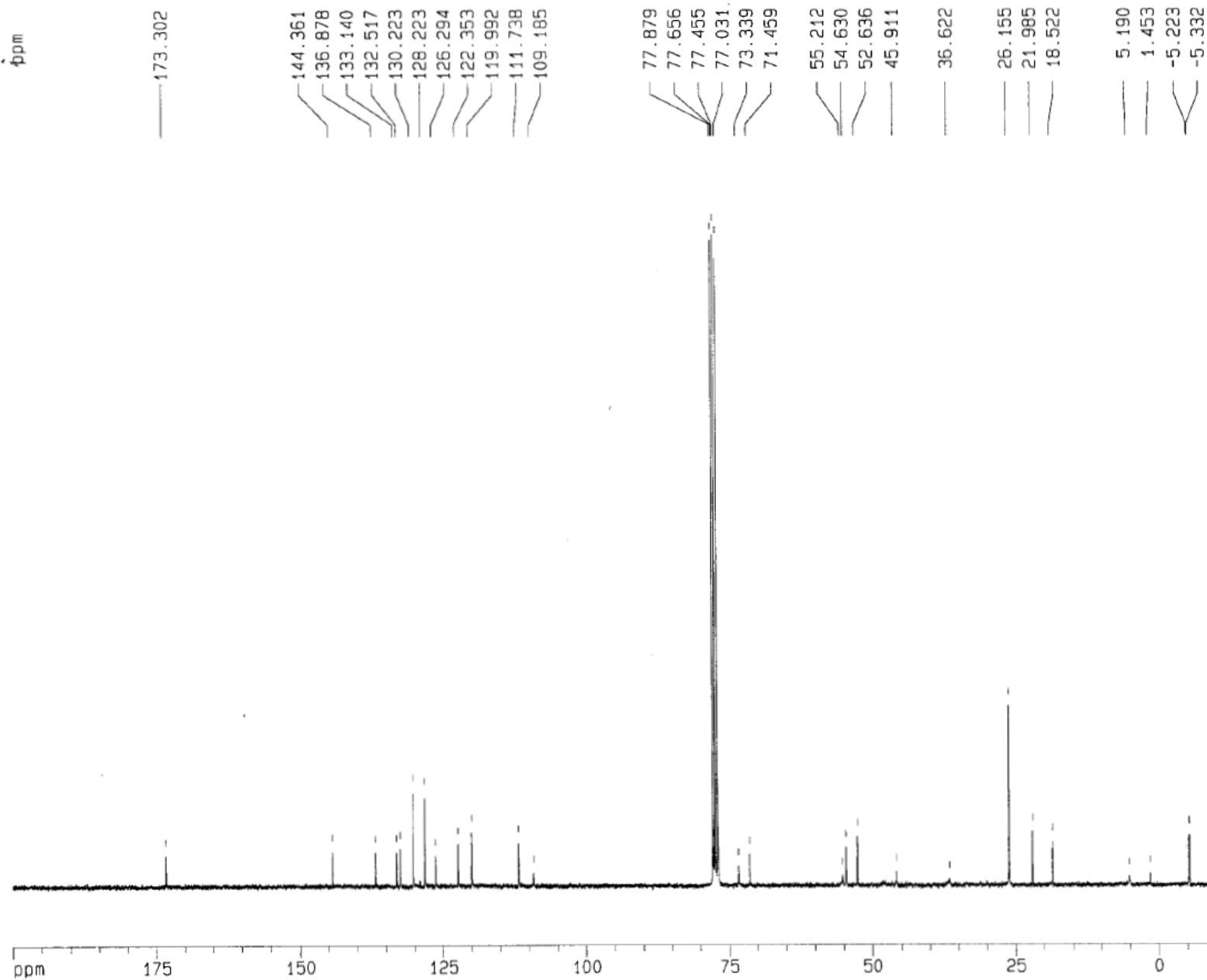
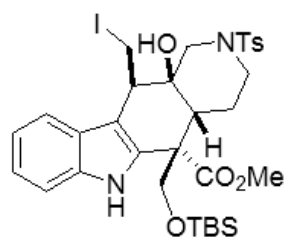
Current Data Parameters
 NAME 090712FJIII294
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120907
 Time 19.09
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 574.7
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

----- CHANNEL f1 -----
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300091 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



Current Data Parameters
 NAME 090812FJII1294
 EXPNO 2
 PROCNO 1

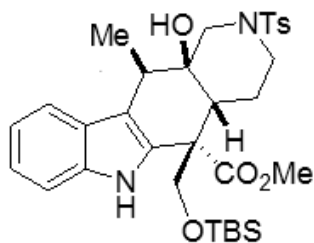
F2 - Acquisition Parameters
 Date_ 20120908
 Time 14.13
 INSTRUM spect
 PROBHD 5 mm GNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 2848
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 1625.5
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 D12 0.0002000 sec

===== CHANNEL f1 =====
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SF01 75.4106357 MHz

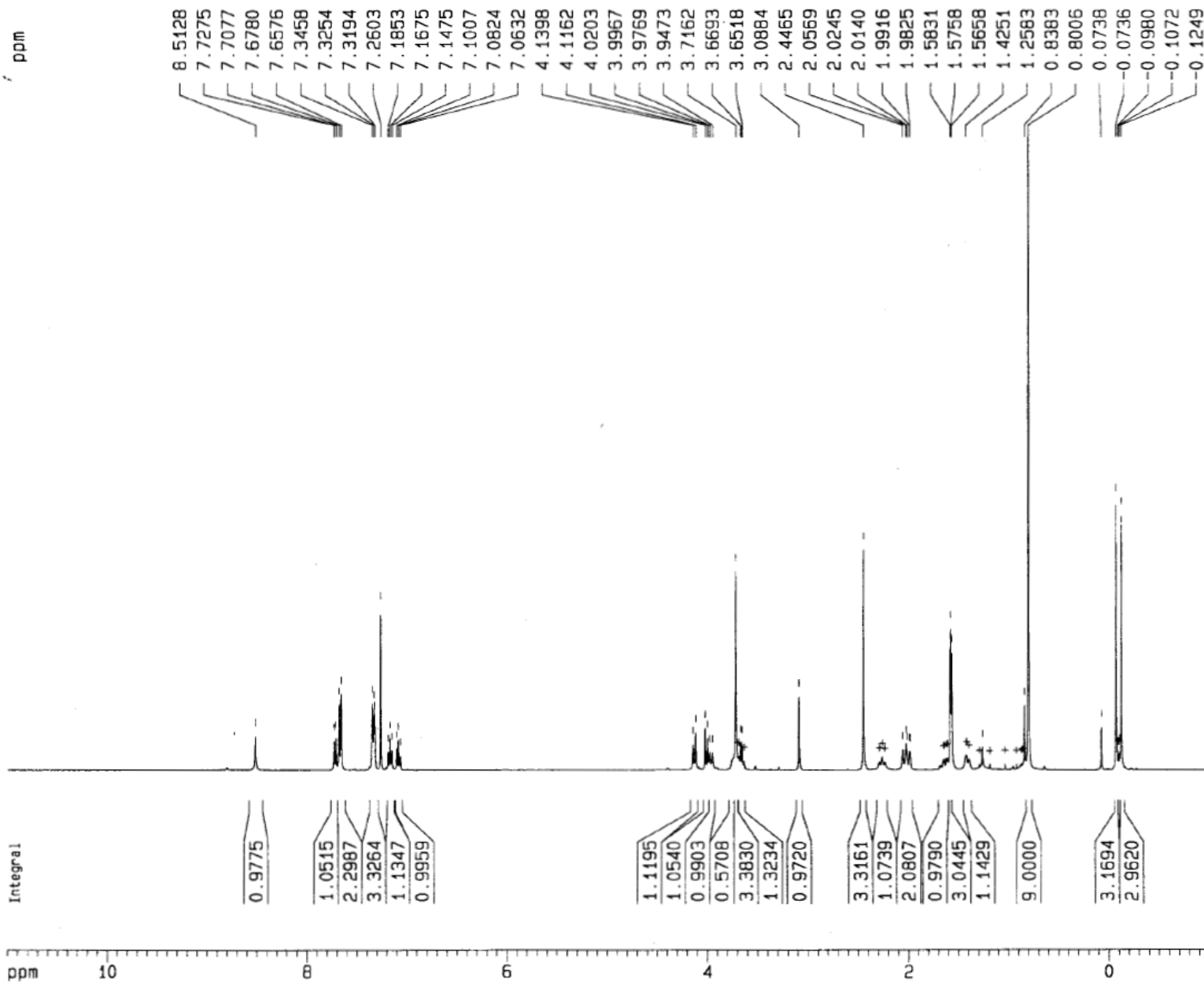
===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SF02 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4023410 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 200.000 ppm
 F1 15080.47 Hz
 F2P -10.000 ppm
 F2 -754.02 Hz
 PPMCM 10.50000 ppm/cm
 HZCM 791.72461 Hz/cm



28



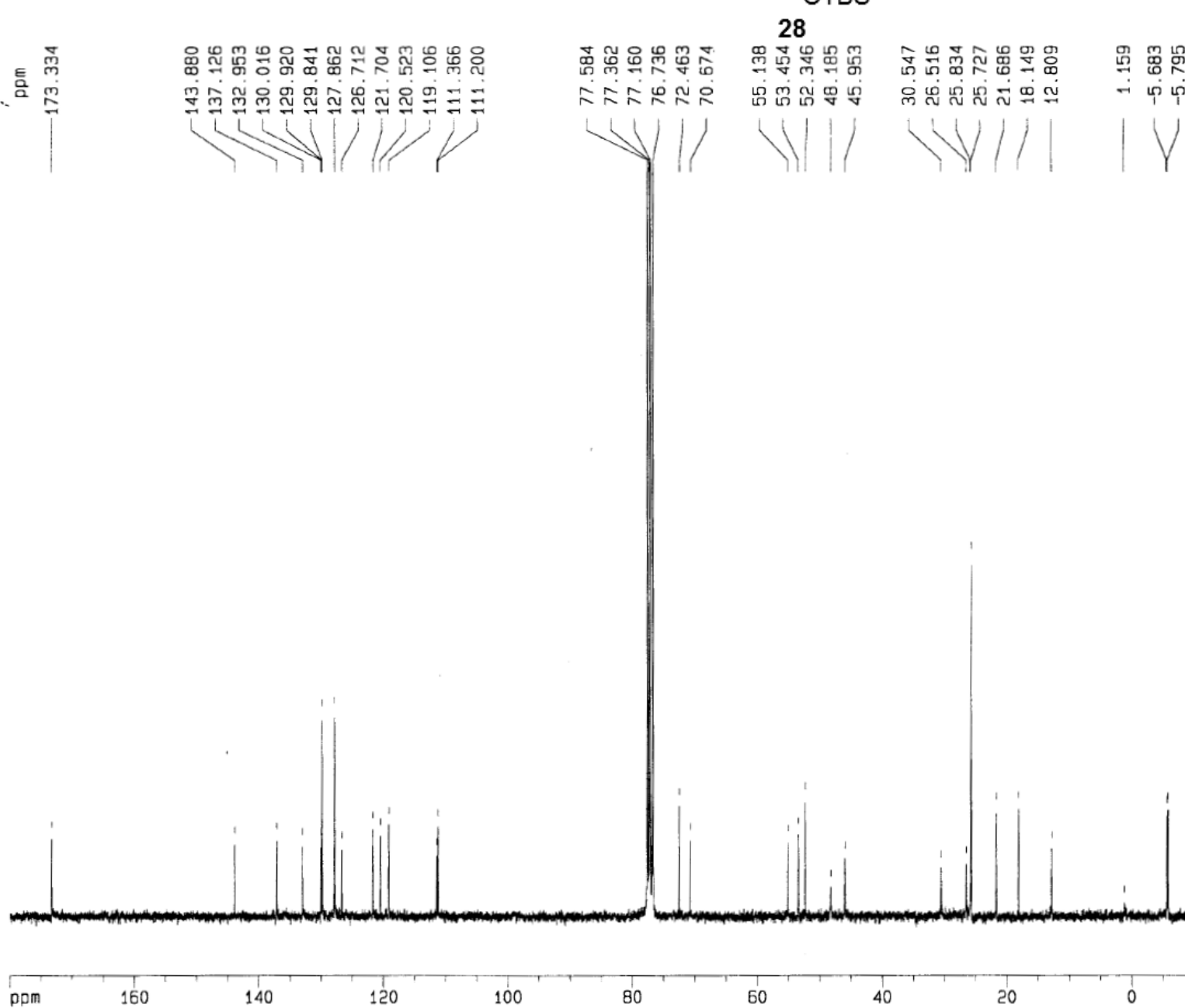
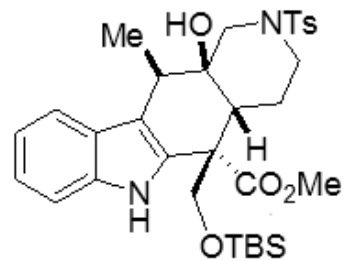
Current Data Parameters
NAME 090912FJIII296
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20120909
Time 22.57
INSTRUM spect
PROBHD 5 mm BBI 1H-
PULPROG zg30
TD 65536
SOLVENT CDC13
NS 16
DS 2
SWH 8278.146 Hz
FIDRES 0.126314 Hz
AQ 3.9584243 sec
RG 724.1
DW 60.400 usec
DE 6.00 usec
TE 300.0 K
D1 1.00000000 sec

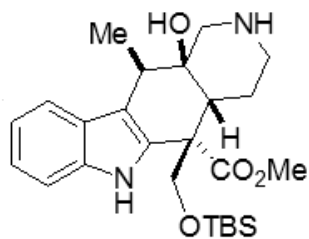
===== CHANNEL f1 =====
NUC1 1H
P1 6.45 usec
PL1 0.00 dB
SF01 400.1324710 MHz

F2 - Processing parameters
SI 32768
SF 400.1300091 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

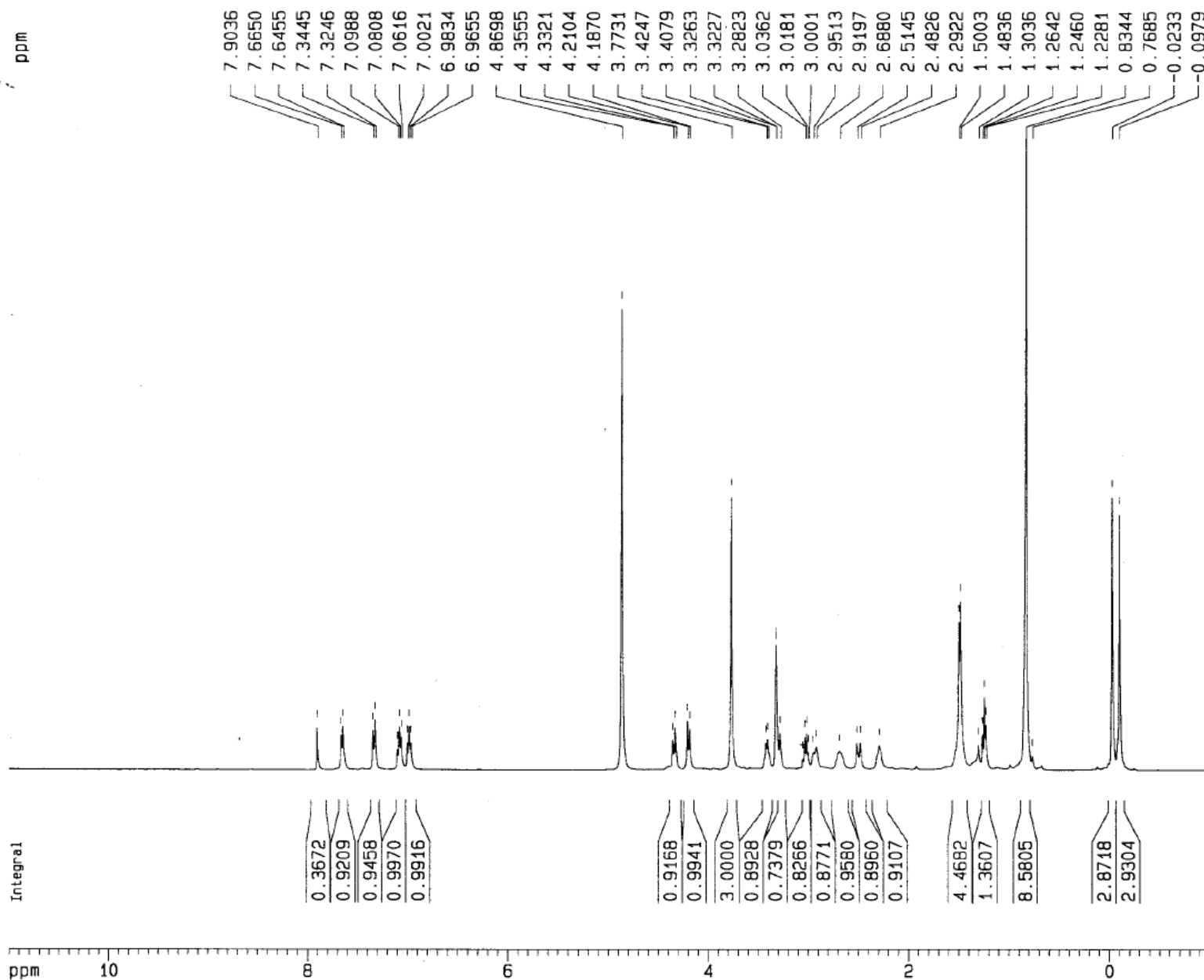
1D NMR plot parameters
CX 20.00 cm
F1P 11.000 ppm
F1 4401.43 Hz
F2P -1.000 ppm
F2 -400.13 Hz
PPMCM 0.60000 ppm/cm
HZCM 240.07800 Hz/cm



Current Data Parameters
NAME 090912FJ111296
EXPNO 1



29



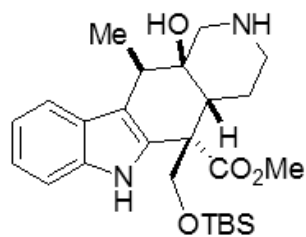
Current Data Parameters
 NAME 091212FJIII299
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120912
 Time 20.11
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT MeOH
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 101.6
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

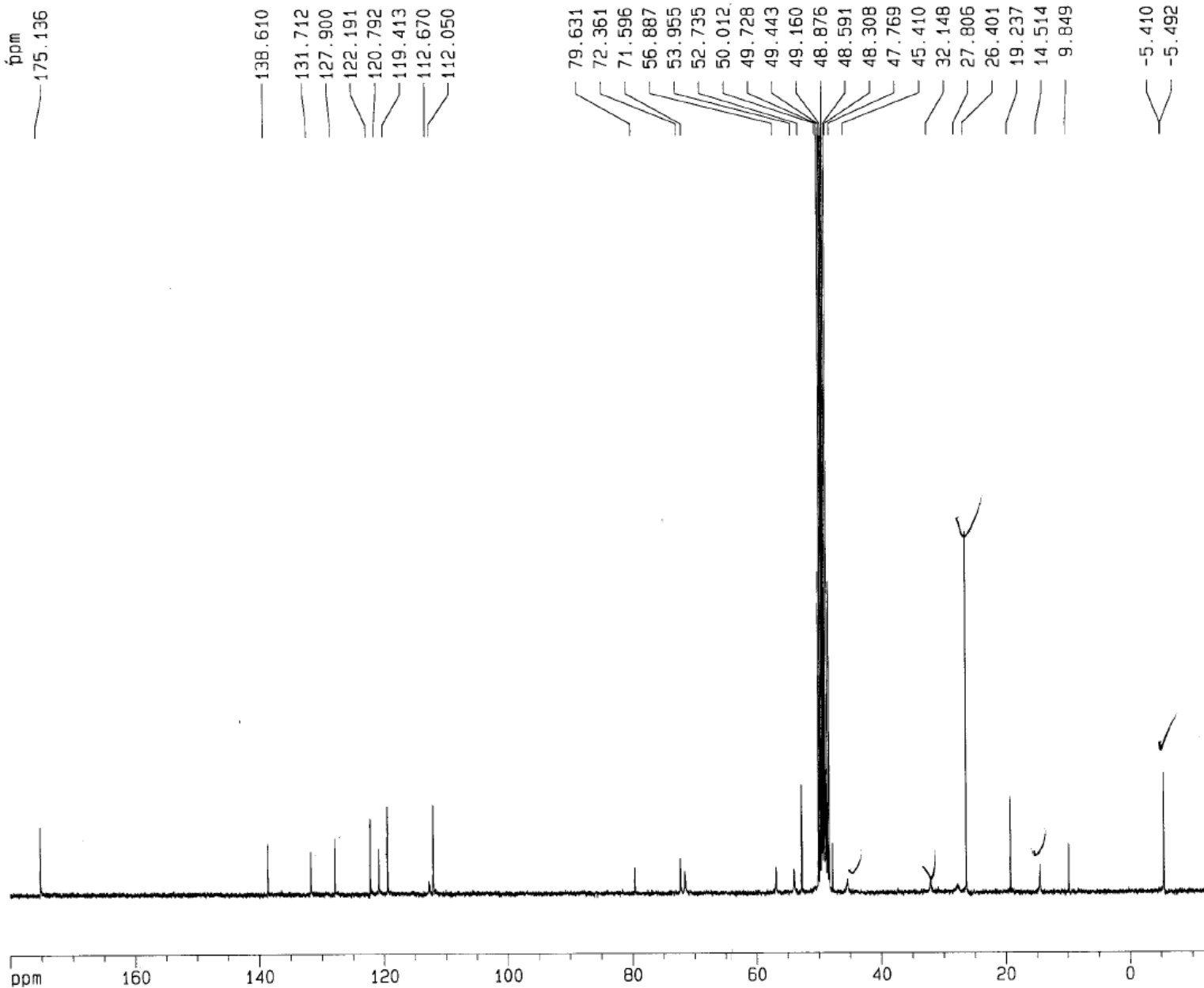
===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300009 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



29



Current Data Parameters
 NAME 091312FJII299
 EXPNO 2
 PROCNO 1

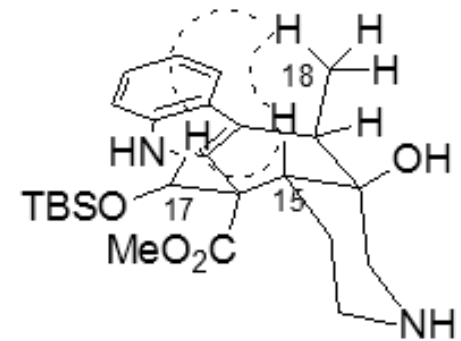
F2 - Acquisition Parameters
 Date_ 20120913
 Time 20.19
 INSTRUM spect
 PROBHD 5 mm GNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOH
 NS 5720
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 512
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec

----- CHANNEL f1 -----
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SFO1 75.4106357 MHz

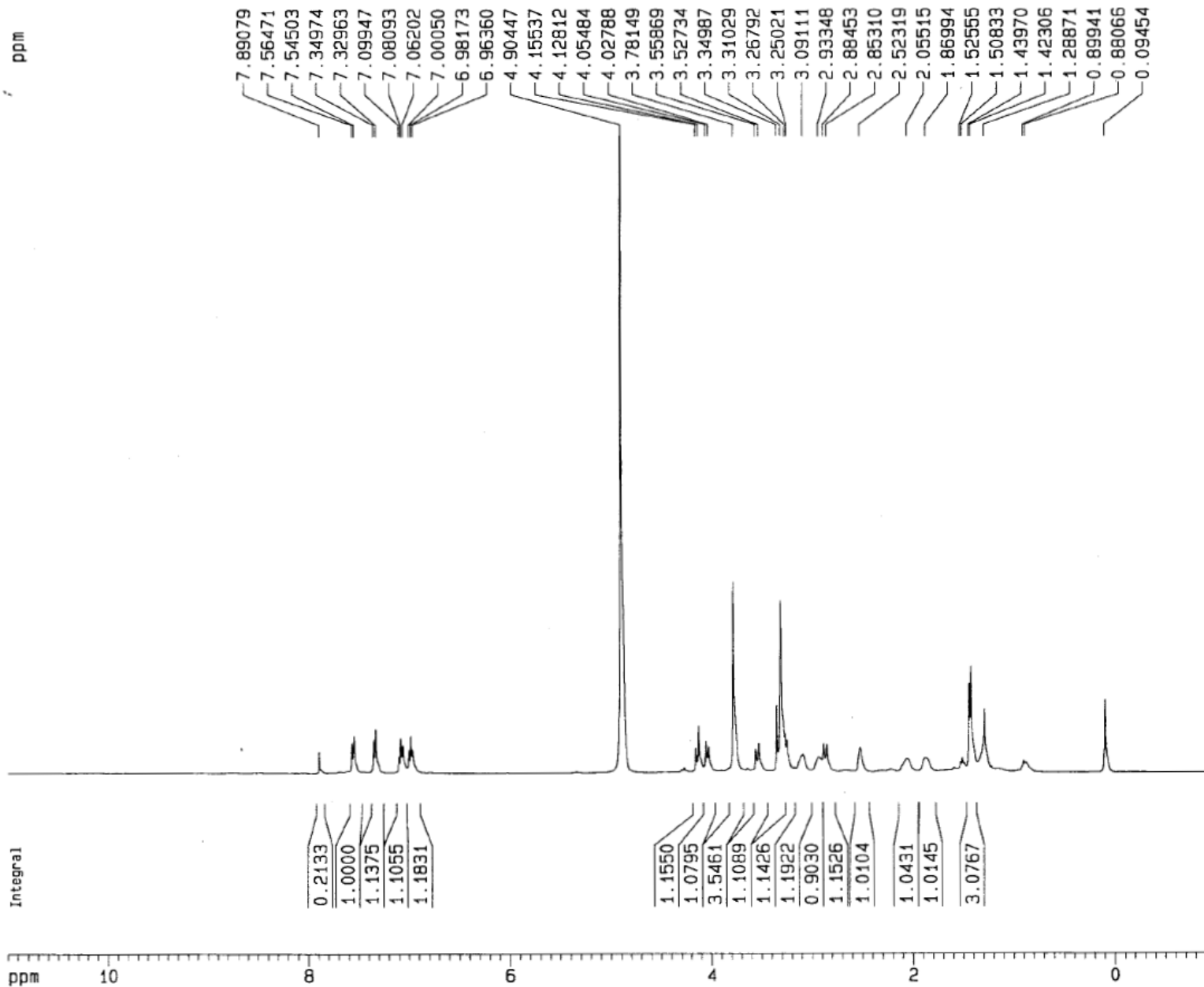
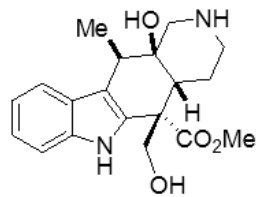
----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SFO2 299.8711995 MHz

F2 - Processing parameters
 SI 32768
 SF 75.4022561 MHz
 HDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 180.000 ppm
 F1 13572.41 Hz
 F2P -15.000 ppm
 F2 -1131.03 Hz
 PPMCM 9.75000 ppm/cm
 HZCM 735.17212 Hz/cm



Key nOe correlations
for piperidine 29



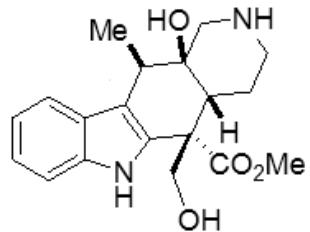
Current Data Parameters
 NAME 092012FJV015
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20120920
 Time 20.21
 INSTRUM spect
 PROBHD 5 mm BBI 1H-
 PULPROG zg30
 TD 65536
 SOLVENT MeOH
 NS 16
 DS 2
 SWH 8278.146 Hz
 FIDRES 0.126314 Hz
 AQ 3.9584243 sec
 RG 143.7
 DW 60.400 usec
 DE 6.00 usec
 TE 300.0 K
 D1 1.00000000 sec

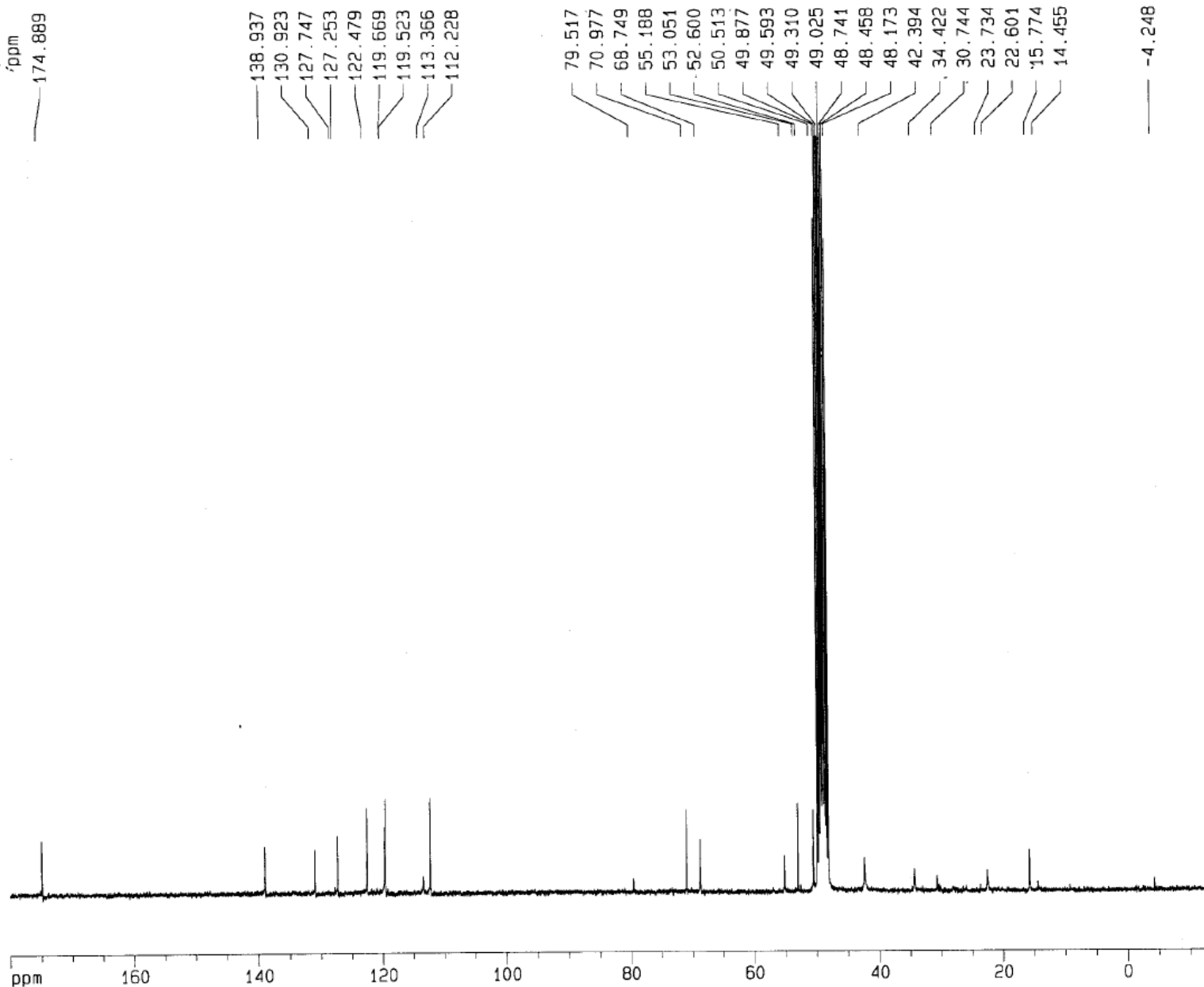
===== CHANNEL f1 =====
 NUC1 1H
 P1 6.45 usec
 PL1 0.00 dB
 SF01 400.1324710 MHz

F2 - Processing parameters
 SI 32768
 SF 400.1300067 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

1D NMR plot parameters
 CX 20.00 cm
 F1P 11.000 ppm
 F1 4401.43 Hz
 F2P -1.000 ppm
 F2 -400.13 Hz
 PPMCM 0.60000 ppm/cm
 HZCM 240.07800 Hz/cm



(+)-3



Current Data Parameters
 NAME 092112FJV015
 EXPNO 2
 PROCNO 1

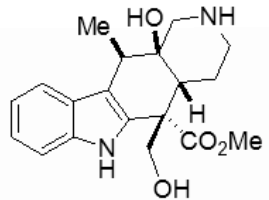
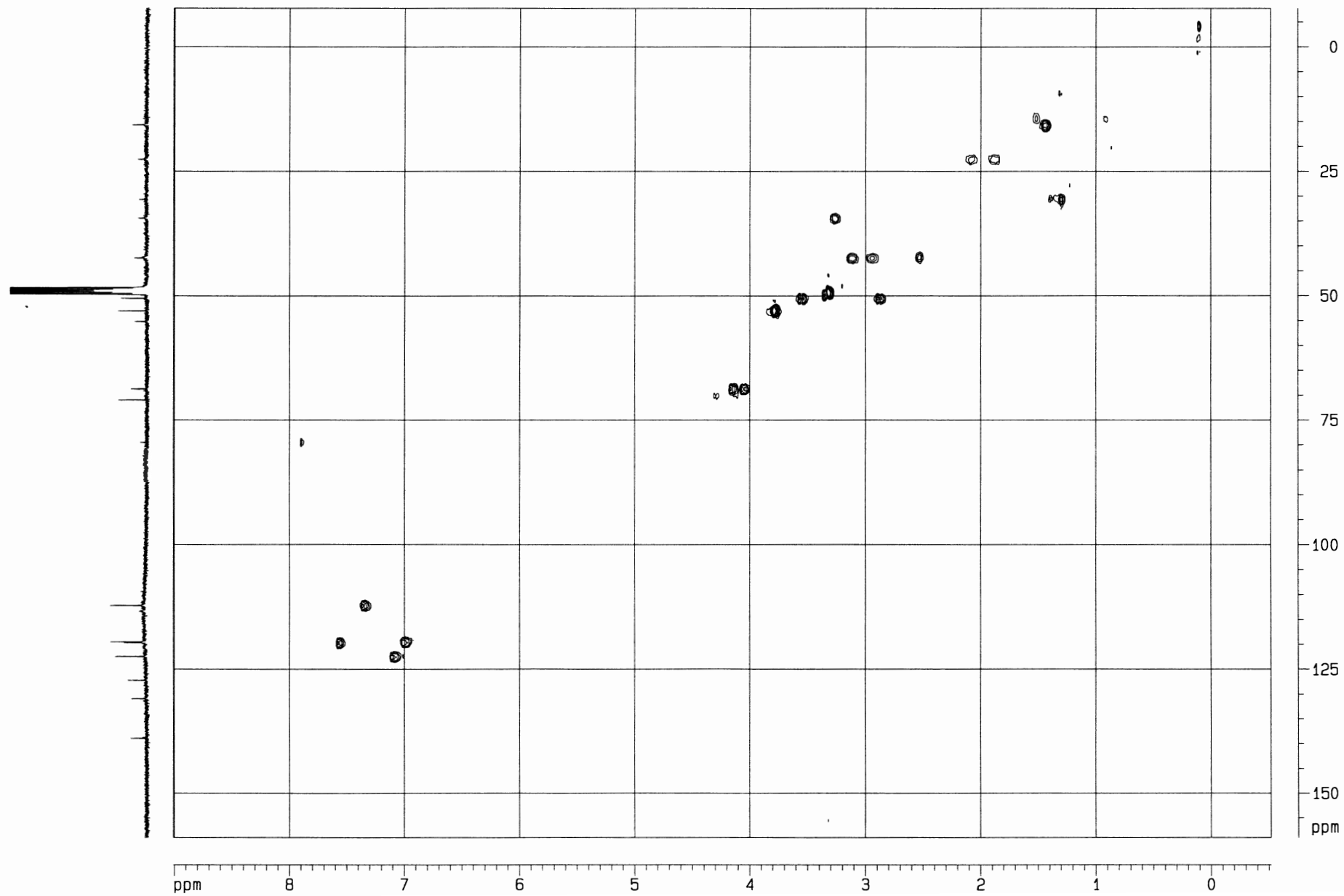
F2 - Acquisition Parameters
 Date_ 20120922
 Time 3.55
 INSTRUM spect
 PROBHD 5 mm QNP 1H/1
 PULPROG zgpg30
 TD 65536
 SOLVENT MeOH
 NS 11264
 DS 4
 SWH 18796.992 Hz
 FIDRES 0.286819 Hz
 AQ 1.7433076 sec
 RG 8192
 DW 26.600 usec
 DE 6.00 usec
 TE 300.0 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 D12 0.00002000 sec

----- CHANNEL f1 -----
 NUC1 13C
 P1 5.25 usec
 PL1 -6.00 dB
 SF01 75.4106357 MHz

----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 115.00 usec
 PL2 0.00 dB
 PL12 19.70 dB
 PL13 19.70 dB
 SF02 299.8711995 MHz

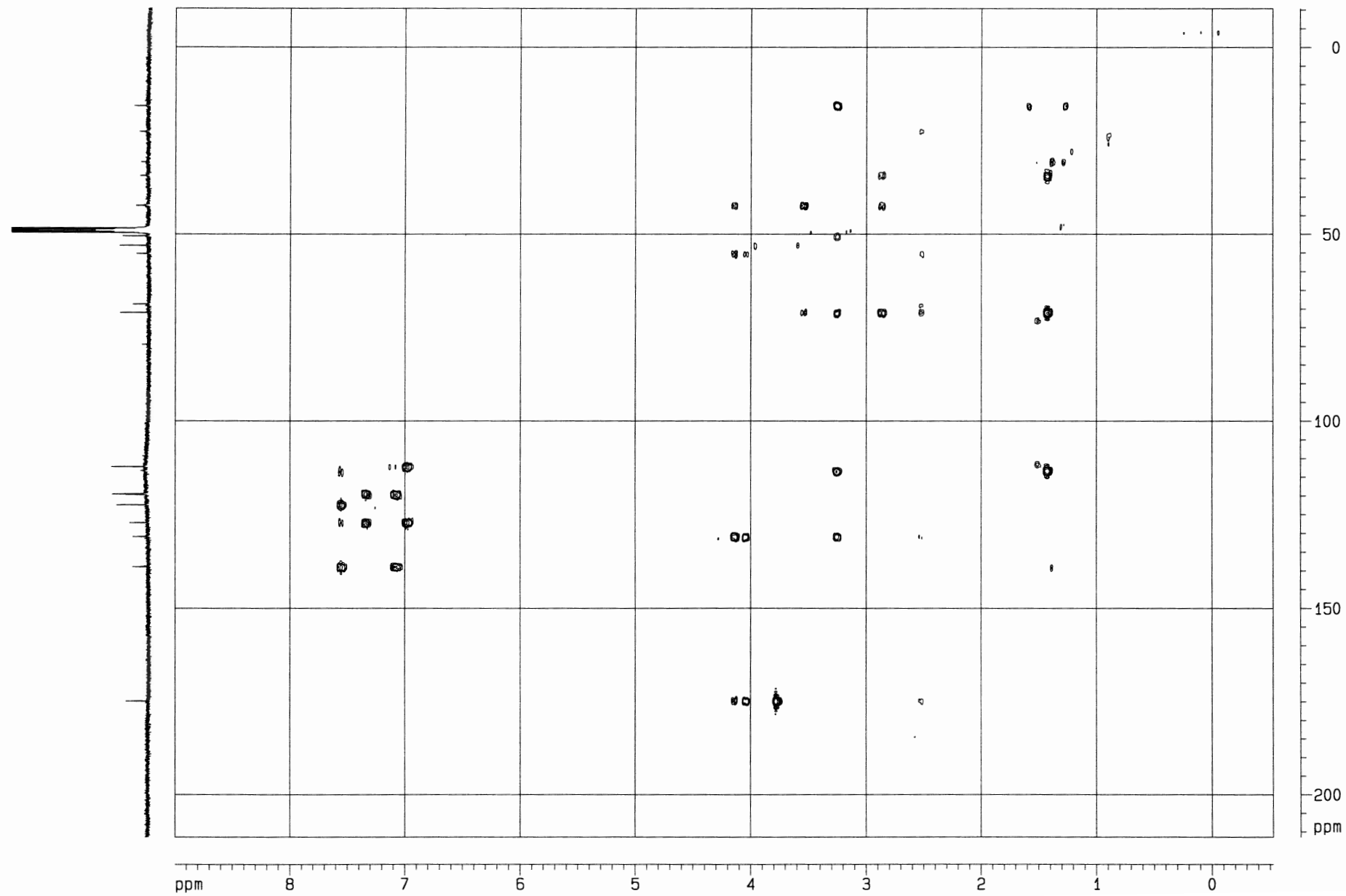
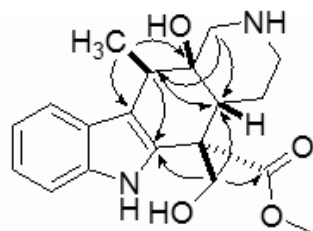
F2 - Processing parameters
 SI 32768
 SF 75.4022676 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

1D NMR plot parameters
 CX 20.00 cm
 F1P 180.000 ppm
 F1 13572.41 Hz
 F2P -15.000 ppm
 F2 -1131.03 Hz
 PPMCM 9.75000 ppm/cm
 HZCM 735.17218 Hz/cm

**(±)-3**

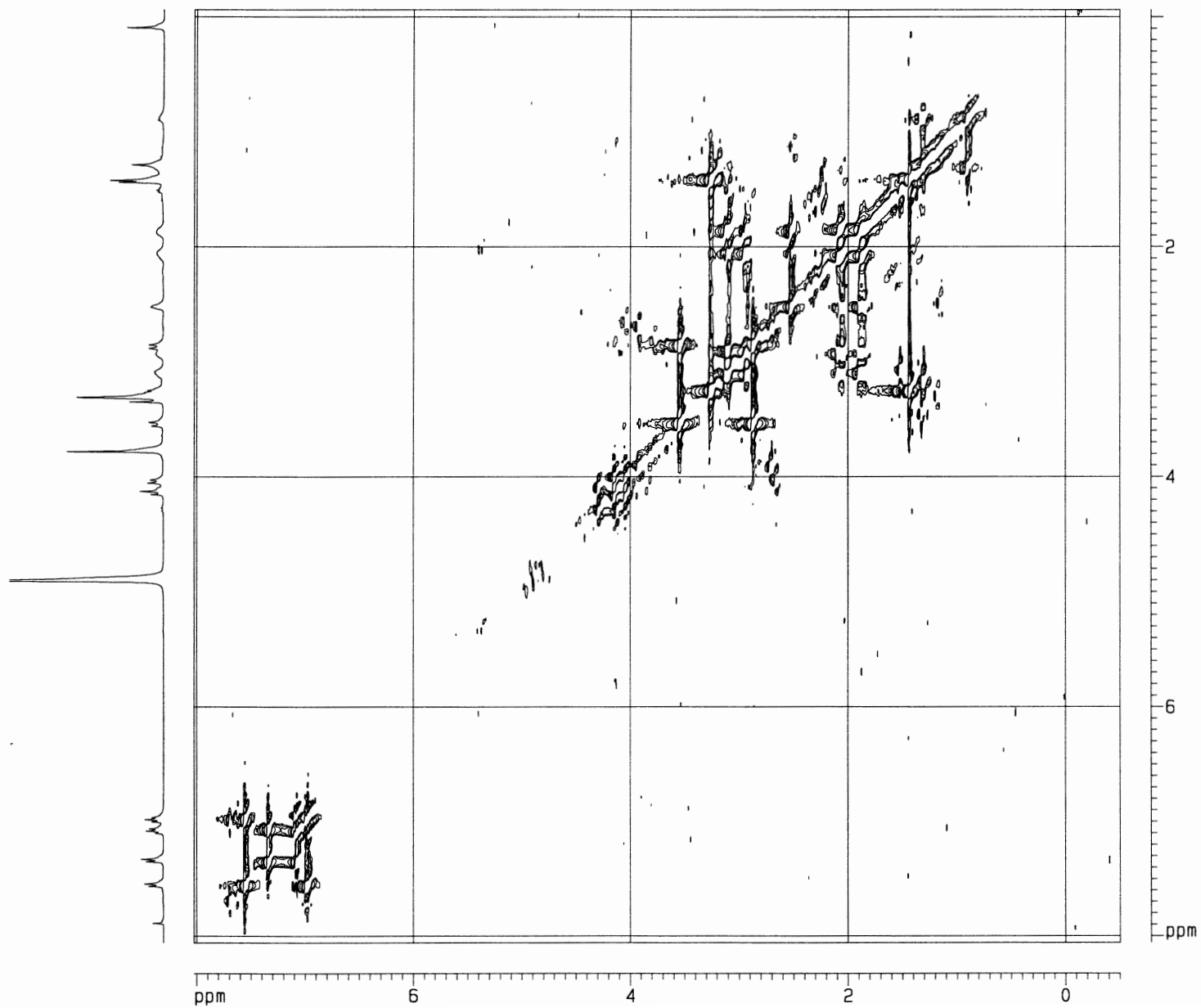
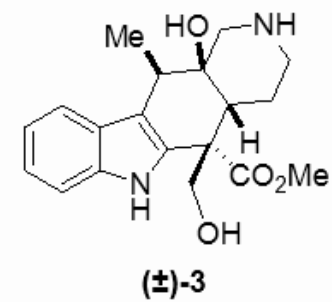
Selected HMBC correlations
for alstilobanine A ((±)-3)

FYQ-V-015 HMBC d4-MeOD

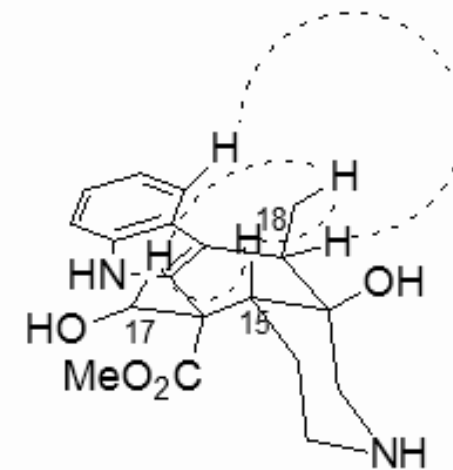
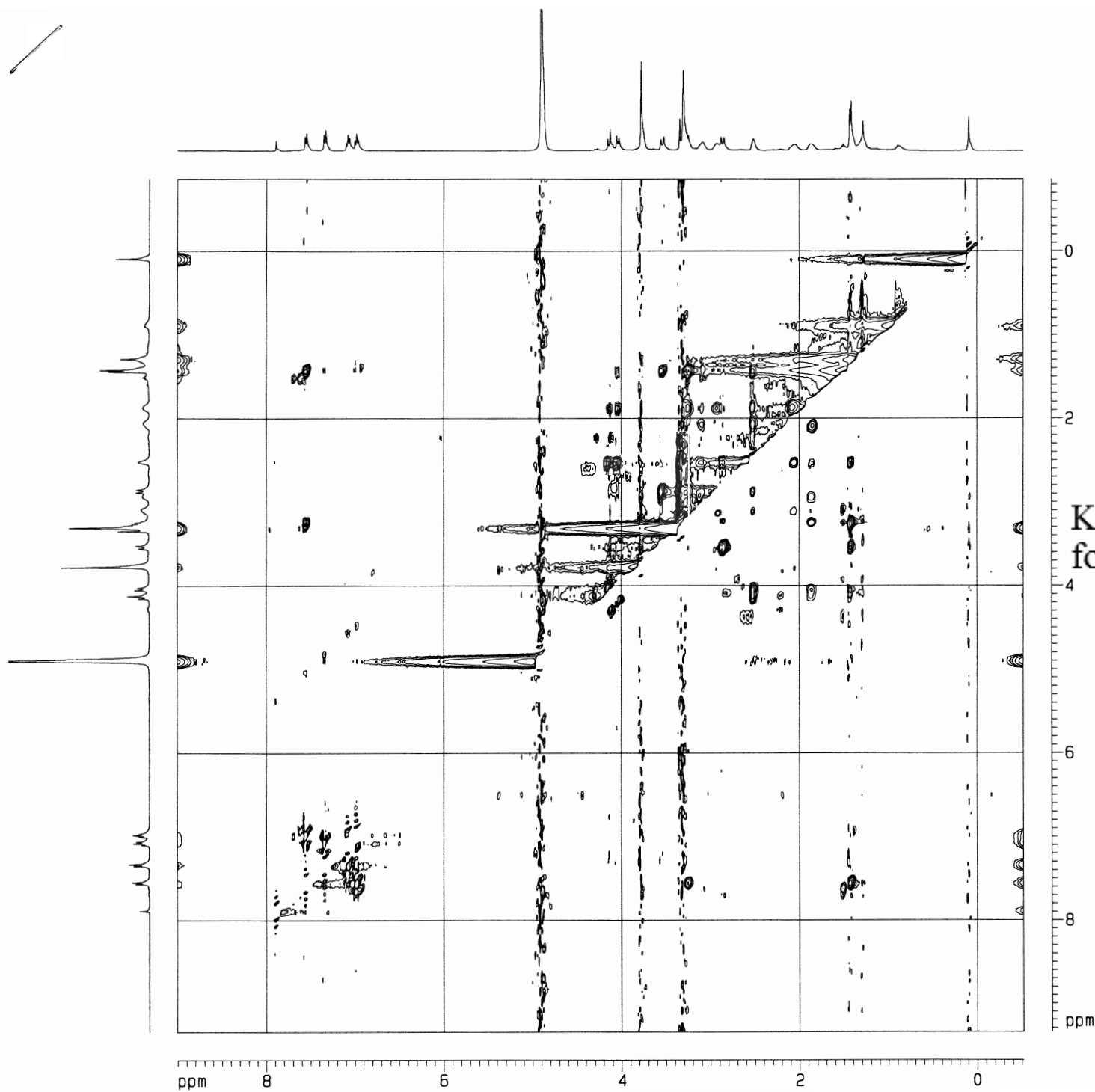


S81

FYQ-V-015 COSY d4-MeOD



FYQ-V-015 NOESY d4-MeOD



Key nOe correlations
for alstilobanine A ((±)-3)