

Supporting Information

McLaughlin-Drubin et al. 10.1073/pnas.1310432110

SI Materials and Methods

Cells. U2OS-tet on were maintained in DMEM supplemented with 10% (vol/vol) tet system-approved FBS, 50 U/mL penicillin, 50 mg/mL streptomycin, and 250 mg/mL G418. CaSki, SiHa, and HeLa cells (ATCC) were maintained in DMEM with 10% newborn calf serum, 50 U/mL penicillin, and 50 mg/mL streptomycin. Me-180 cells (ATCC) were maintained in McCoy's 5a Medium modified with 10% (vol/vol) FBS, 50 U/mL penicillin, and 50 mg/mL streptomycin. GSK J4 was obtained from Xcess Biosciences.

RNAi. The cells were transfected with the following shRNA constructs for depletion of histone lysine demethylases (KDM)6B-shKDM6B: TRCN0000095268 (Open Biosystems) and MISSION Non-Target shRNA control vector (Sigma). The cells were transfected with the following shRNA constructs for depletion of tumor suppressor p16^{INK4A} – pbabeshp16AB, pbabep16CD, and pbabep16EF or pbabeU6 as a control (a generous gift from James Rocco, Massachusetts General Hospital, Boston, MA).

The cells were transfected with 40 nM of the following ON-TARGET^{plus} SMARTpools: KDM6B-specific ON-TARGET^{plus} SMARTpool (L-023013-01; Thermo Scientific Dharmacon), cyclin-dependent kinases 4 (CDK4)-specific ON-TARGET^{plus}

SMARTpool (L-003238-00; Thermo Scientific Dharmacon), CDK6-specific ON-TARGET^{plus} SMARTpool (L-003240-00; Thermo Scientific Dharmacon), p16^{INK4A}-specific ON-TARGET^{plus} SMARTpool (sequence AACGCACCGAAUAGU-UACGGUUU or AAUAGUUACGGUCGGAGGCCGUU; Thermo Scientific Dharmacon), or ON-TARGET^{plus} Non-Targeting Pool (D-001810-10; Thermo Scientific Dharmacon) using Lipofectamine 2000 (Invitrogen).

Wild-type p16^{INK4A} (missing the first 24 nucleotides; a gift of Robert Weinberg, Whitehead Institute, Massachusetts Institute of Technology, Cambridge MA) (1) cloned into LXS^N (2) was used in rescue experiments with shp16AB. Rescue forms of p16^{INK4A} (resp16CD and resp16EF, resistant to their respective shRNA), were generated by introducing silent mutations in the p16^{INK4A} sequence using the following primers: shp16CD sense 5'-GGGT-CGGGTAGAAAGATGTGCGGGCGCTG-3', shp16CD antisense 5'-CAGCGCCCGCACATCTTCTACCCGACCC-3', shp16EF sense 5'-GGGGCGCTGCCCAAAGCACCGAATAGT-3', and shp16EF antisense 5'-ACTATTCGGTGCATTGGGCAGCGC-CCC-3'. The mutations were constructed by PCR using the QuikChange site-directed mutagenesis kit (Stratagene) according to the manufacturer's instructions.

1. Medema RH, Herrera RE, Lam F, Weinberg RA (1995) Growth suppression by p16^{ink4} requires functional retinoblastoma protein. *Proc Natl Acad Sci USA* 92(14):6289–6293.

2. Piboonniyom SO, Timmermann S, Hinds P, Munger K (2002) Aberrations in the MTS1 tumor suppressor locus in oral squamous cell carcinoma lines preferentially affect the INK4A gene and result in increased cdk6 activity. *Oral Oncol* 38(2):179–186.

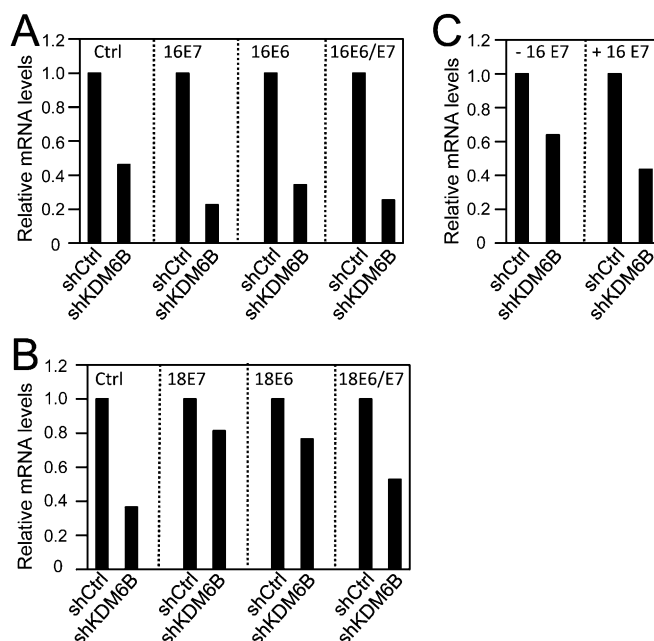


Fig. S1. KDM6B depletion in human foreskin keratinocyte (HFK) populations and U2OS osteosarcoma cells. KDM6B was depleted in: (A) HFKs expressing control vector, human papillomavirus (HPV)16 E7, E6, or E6 and E7; (B) HFKs expressing control vector, HPV18 E7, E6, or E6 and E7; and (C) U2OS-tet on cells with doxycycline-inducible expression of HPV16 E7. mRNA levels were determined by quantitative RT-PCR (qRT-PCR). The graphs depict representative experiments.

