Supporting Information

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Fig. S1. Sequence analyses of carboxyl-terminal peptidase A (AtCtpA) and related protein sequences. (A) Comparison of amino acid sequences of mature proteins: (1) Arabidopsis carboxyl-terminal peptidase (CtpA; At4g17740); (2) Spinacia CtpA; (3) Arabidopsis At3g57680; (4) Arabidopsis At5g46390; and (5) Synechocystis PCC 6803 CtpA. *Conserved residues essential for catalytic activity of CtpA. Identical and similar amino acids are highlighted by black and gray shading, respectively. Alignment of protein sequences was performed with ClustalX program (www.ch.embnet.org/software/ClustalW.html). (B) Phylogeny of mature protein sequences from Arabidopsis CtpA, Spinacia CtpA, Arabidopsis At3g57680, Arabidopsis At5g46390, and Synechocystis PCC 6803 CtpA. The percentage indicates the similarity among different amino acid sequences. DNAMAN program was used in this analysis.

C-terminal sequence in wild type

1451 CAAAGTCGTT TCCGAAAGAT GAAGAGGCGT TCTGTGGATG CCTTAAGGAT 1501 CCTACAGCTG CTTGTTATCT CAATCAAGGC CTACTTTTTT CTAGATGA

- 451 LSDGSGLAVT VARYETPAHT DIDKVGVTPD HPLPKSFPKD EEAFCGCLKD 501 PTAACYLNQG LLFSR
- C-terminal sequence in atctpa-2

1451 CAAAGTCGTT TCCGAAAGAT GAAGAGGCGT TCTGTGGATG C<u>TTAGACAAC</u> 1501 <u>TTAATAACAC ATTGCGGACG TTTTTAA</u>

451 LSDGSGLAVT VARYETPAHT DIDKVGVTPD HPLPKSFPKD EEAFCGC<u>LDN</u> 501 <u>L1THCGRF</u>

Fig. S2. Flanking sequences of transfer DNA (T-DNA) insertion in the *atctpa-2* mutant compared with *AtCtpA* sequence in WT. The T-DNA insertion sequence and corresponding amino acid sequence are underlined. The mutated version of AtCtpA in the *atctpa-2* mutant contains 497 as of AtCtpA plus 11 as from the T-DNA sequence.



Fig. S3. Immunoblot analysis of representative subunits of photosynthetic complexes from wild-type (WT), *atctpa-1*, and *atctpa-2* plants. Chloroplast thylakoid protein samples equivalent to 2 μg chlorophyll were resolved by 10% SDS/PAGE containing 8M urea; blots were probed with the indicated antibodies. The sample preparation and plant growth conditions were the same as that in Fig. 3.



Fig. S4. Nonphotochemical quenching (NPQ) analyses. NPQ kinetics after rapid induction and dark relaxation were measured using intact leaves from 4-wk-old WT and *atctpa-2* plants grown in soil (350 μ E) under long-day condition. Actinic light intensity, 1,800 μ E. Data are presented as means \pm SD (n = 4). WT and *acctpa-2* data are significantly different except for the last three time points (P < 0.05).



Fig. S5. Complementation of *atctpa-2* mutant. (*A*, *Upper*) Immunoblot analysis of AtCtpA in WT, *atctpa-2*, and two independent lines of *atctpa-2* mutant plants complemented with *355::AtCtpA* construct. (*A*, *Lower*) Photographs of corresponding plants of WT, *atctpa-2*, and complemented lines. Plants were grown under 350 μE for 4 wk in soil with long-day cycle. (*B*) D1 status in WT, *atctpa-2*, and *355::AtCtpA/atctpa-2* plants after 2 h high light treatment (1,800 μE). Immunoblot analysis was carried out as that in Fig. 3.

Table S1. Primers used in this study

Primer name	Primer sequence
atctpa-1 left board	GCCTCGAGGAAGAGAAGATTC
atctpa-1 right board	TTCCTTGATGGTGGTTCAGTC
atctpa-2 left board	GTGGAGGTTCTTTCCCAGAAG
atctpa-2 right board	GCTTGCTCTTGGTGATTTTTG
LBb1.3	ATTTTGCCGATTTCGGAAC
P1 F	AGAGTTGGAATTTTCCCGGGGGAA
P1 R	GAAATTTCTGGTTTCGATTAGCTAAAGCTAT
P2 F	ATGGAGGTCCTTGCGAGCTCTTCAT
P2 R	GCATCCACAGAACGCCTCTT
AtCtpA CDS F	ATGGAGGTCCTTGCGAGCTCTTCAT
AtCtpA CDS R	TCATCTAGAAAAAAGTAGGCCTTGATTGAGATAACAAG
Actin2 F	GGAAGGATCTGTACGGTAAC
Actin2 R	TGTGAACGATTCCTGGACCT

DNAS Nd