Supplementary Material for the paper:

Using Strahler's analysis to reduce up to 200-fold the run time of realistic neuron models

by Addolorata Marasco, Alessandro Limongiello and Michele Migliore

- Supplementary Figure 1: The cells used in this paper and the four areas used for synaptic stimulations; projection to the XY plane of 3D reconstructions of Purkinje cells from different animals, as indicated in the table. Colors indicate the different dendritic areas used to stimulate each cell.
- Supplementary Figure 2: I/O properties of a Purkinje cell model; a) representation of the stimulation conditions; colored dendrites represent typical partial and segregated areas; b) (left) average ($n = 5, \pm sd$) APs frequency and ISIs for 1000 synaptic inputs; (right) average firing frequency and ISIs with randomly activated synaptic inputs (Poisson) at 80 Hz; c) (left) raster plots of the spike trains (plots above traces), and somatic potential for cell *e4cb2a2* during simulations with 1000 synapses; grey areas in raster plots indicate regular spiking, with the first ISI of each pattern darker; blank areas indicate pauses; (right) normalized ISIs distribution. Average CV_2 = 0.38 for full and partial, 0.37 for segregated stimulation.
- Supplementary Figure 3: The method is robust for perturbations of peak ionic conductances. (*left*) average trace accuracy for simulations using the original set of values, g_{peak} , for the 12 peak ionic conductances (black) or after 10 random perturbations (red) redrawn from a normal distribution with average g_{peak} and variance 10% g_{peak} , resulting in a maximum perturbation >20% of g_{peak} ; different symbols indicate results obtained for full (circles), partial (triangles), and segregated (squares) stimulations; (*right*) percent of statistically indistinguishable ISIs from simulations using the realistic and the reduced model, compared pairwise (Wilcoxon Signed Rank test); all simulations were carried out using cell *e4cb3a1*.

- Supplementary Figure 4: The reduced model is able to reproduce the differential effect of two synaptic inputs colocalized or at two different locations. (*left*) the full and the reduced morphologies of cell *p19*, and their somatic potential during simulations with two identical synapses targeting the same location (loc.1) or two different branches (loc 1 and 2).
- Supplementary Figure 5: The method is extremely accurate and quite robust for most morphologies and under all conditions of stimulation. Average accuracy and percent of spikes times statistically indistinguishable for each cell as a function of the average synaptic stimulation frequency under different stimulation conditions; average results calculated from 550 (full), 500 (partial), and 600 (segregated) simulations. In all cases the dendrites were clustered using Strahler's order s = 5 (see *Methods*), and statistics was calculated from pairwise comparison of the traces from each simulation.
- Supplementary Figure 6: In several cases, a 3-compartment reduction gave ISIs statistically different from the full model but still high overall trace accuracy. A) Full and reduced model of cells purk2 (*left*) and p20 (*right*); B) (*top*) Somatic membrane potential of cell *purk2* during a simulation with 1000 synchronous synapses activating at a random (Poisson) average frequency of 120 Hz; (*bottom*) Somatic membrane potential of cell *p20* during a simulation with 1000 synchronous synapses activated by a random (Poisson) average frequency of 160 Hz. In both cases, we used a *full* stimulation protocol (see Methods) and the ISIs were statistically different between the full and reduced model (Wilcoxon Rank Sum test, *p*<0.001). The trace accuracy was 0.86 (*purk2*) and 0.89 (*p20*).
- Supplementary Figure 7: The method works also for CA1 pyramidal neurons. a) somatic membrane potential for a full of a hippocampal pyramidal CA1 neuron (*blue* traces, cell *c70863* from Marasco et al., 2012) and the corresponding reduced model (*red* traces) during a simulation activating 140 (*left*) or 400 (*right*) randomly distributed synapses. b) Average number of APs (±sd) elicited in 500 ms long simulations as a function of the number of synaptic inputs activated in the full model (*blue plots*) or reduced (*red plots*) using the

method discussed in this paper (*left*), or a previous method with (middle) or without (*right*) the calculation of the *max Stim* parameter (19); all simulations were carried out as in ref.(19).

Supplementary Table:

	alxP	e1cb4a1	e1cb4a5	e4cb2a2	e4cb3a1	p19	p20	purk1	purk2	purk3
smooth	72	57	56	58	56	82	49	81	56	74
spiny	851	630	633	660	685	1055	650	863	784	760
S1	462	344	345	360	371	568	349	472	420	417
S2	253	193	202	223	221	341	202	266	258	243
S3	136	93	86	77	93	146	99	125	106	100
S4	46	34	30	40	40	53	30	42	30	55
S5	17	14	22	11	14	24	9	26	24	17
S6	9	9	4	7	2	3	8	7	1	1
S7	0	0	0	0	0	1	1	5	1	1
S8	0	0	0	0	0	1	1	1	0	0
R _{in_full}	66	36	74	60	53	21	26	11	13	14
tot. area	9937	24303	10638	11130	9379	39834	24867	55628	39861	46586
3-comp										
R _{in_3comp}	89	37	96	75	73	25	28	17	17	19
soma	553	922	511	990	800	730	750	1521	1521	1521
C 1	850	1787	722	847	1356	2719	1892	5395	5101	4107
C 2	586	1500	723	646	480	2114	1457	3300	2658	3251

Supplementary Table: For each full morphology the following are reported: the number of smooth and spiny dendrites, the number of compartments with different Strahler's number (*S*), the input resistance (R_{in} , in M Ω), and the total membrane area (in μm^2). The bottom part of the Table reports parameters (input resistance, and membrane area) for the 3-compartment reduction for all morphologies; C1 and C2 are the compartments used to implement the smooth and spiny dendrites, respectively, whereas the soma is the same as in the full morphology; highlighted in red are the cells for which a 3-compartment reduction gave the best results.









Full





Partial



avg syn frequency (Hz)





synaptic inputs