

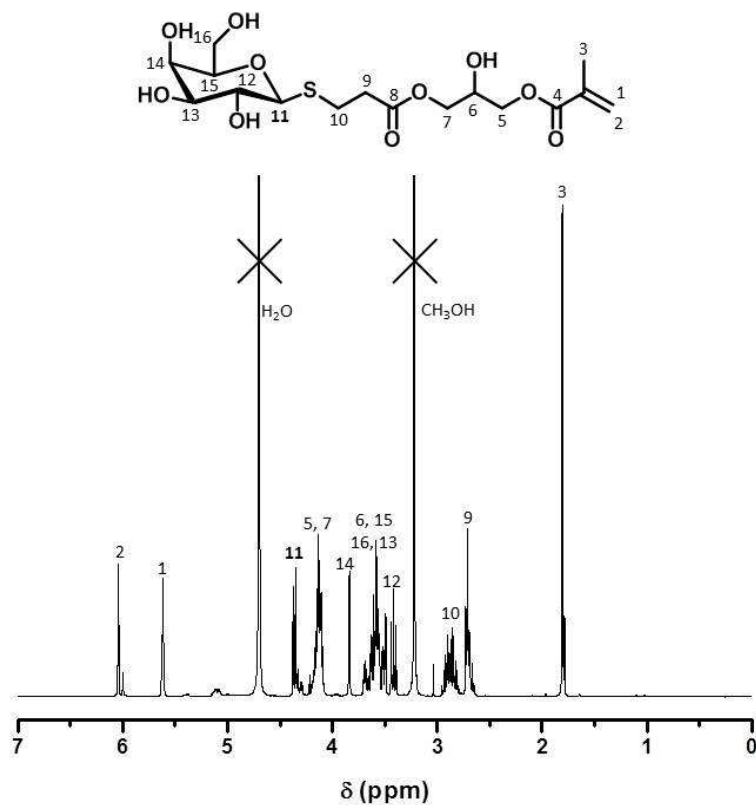
# Polymerization-induced self-assembly of galactose-functionalized biocompatible diblock copolymers for intracellular delivery

Vincent Ladmiral\*, Mona Semsarilar, Irene Canton and Steven P. Armes\*

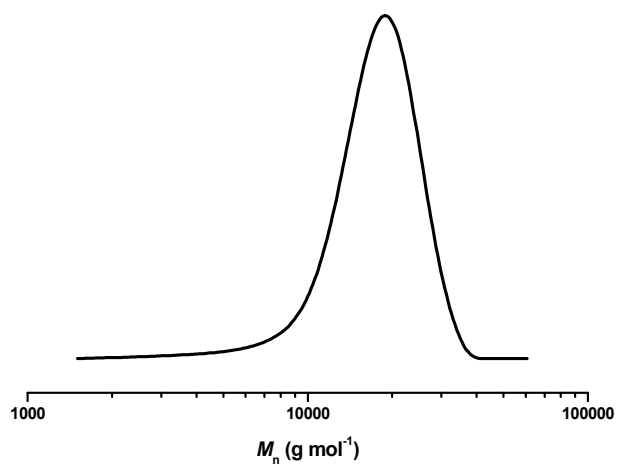
Department of Chemistry, University of Sheffield, Brook Hill, Sheffield, South Yorkshire, S3 7HF, UK.

## Polymerization-induced self-assembly

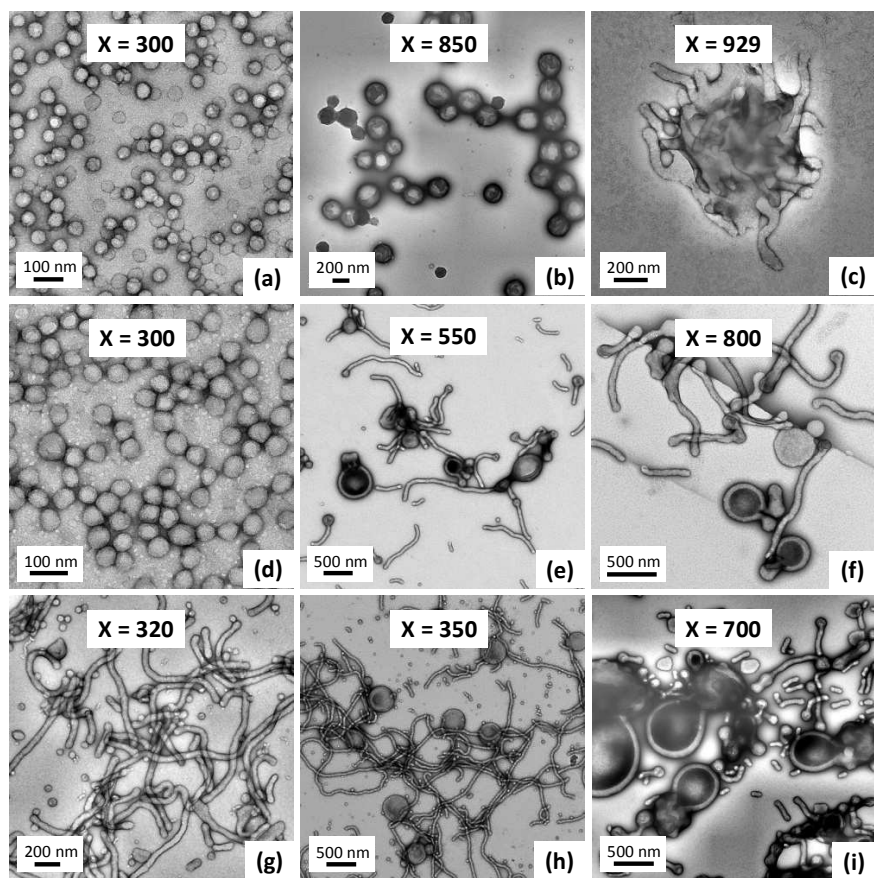
Note: Targeting relatively long PHPMA blocks at 10 % solids produced a mixture of spheres and small vesicles. These spheres are believed to be kinetically-trapped copolymer morphologies. However, no evidence for a sphere-to-vesicle transformation was observed during the long-term storage of these samples at 20 °C for five months.



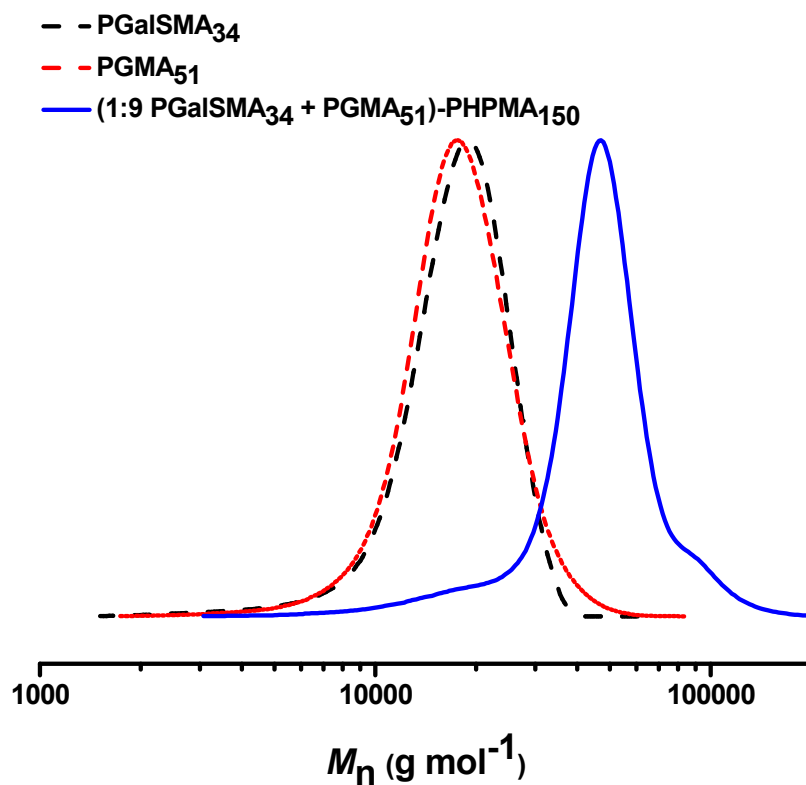
**Figure S1.** Assigned <sup>1</sup>H NMR spectrum recorded for galactose methacrylate monomer dissolved in D<sub>2</sub>O at 20 °C.



**Figure S2.** DMF GPC curve obtained for PGaISMA<sub>34</sub> at 60 °C ( $M_n = 16,300 \text{ g mol}^{-1}$ ,  $M_w = 18,500 \text{ g mol}^{-1}$ ,  $M_w/M_n = 1.13$ ). RAFT synthesis conditions: [GaISMA] : [PETTC] : [ACVA] = 30 : 1 : 0.1; 25 % w/w GaISMA in a 9:1 mixture of PBS buffer (150 mM, pH 7.2) and methanol. Monomer conversion = 97 %.



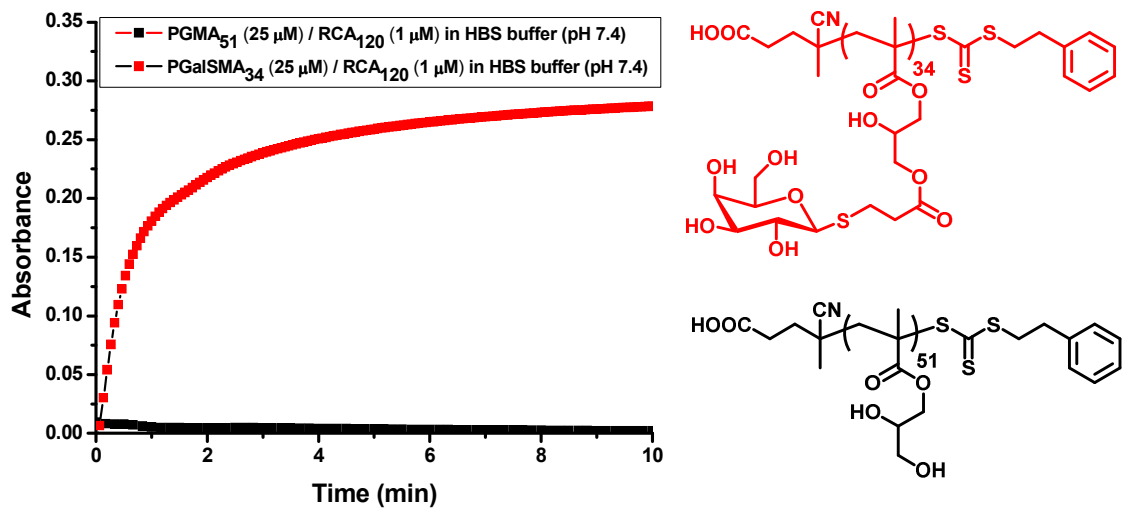
**Figure S3.** Representative TEM images obtained for PGalSMA<sub>34</sub>-PHPMA<sub>x</sub> diblock copolymer nano-objects synthesized by RAFT aqueous dispersion polymerization of HPMA at 70 °C. The target DP (x) for the PHPMA block is indicated on each image. Images (a), (b) and (c) shown the morphologies obtained at 10 % solids; images (d), (e) and (f) refer to morphologies formed at 15 % solids; images (g), (h) and (i) were produced at 20 % solids.



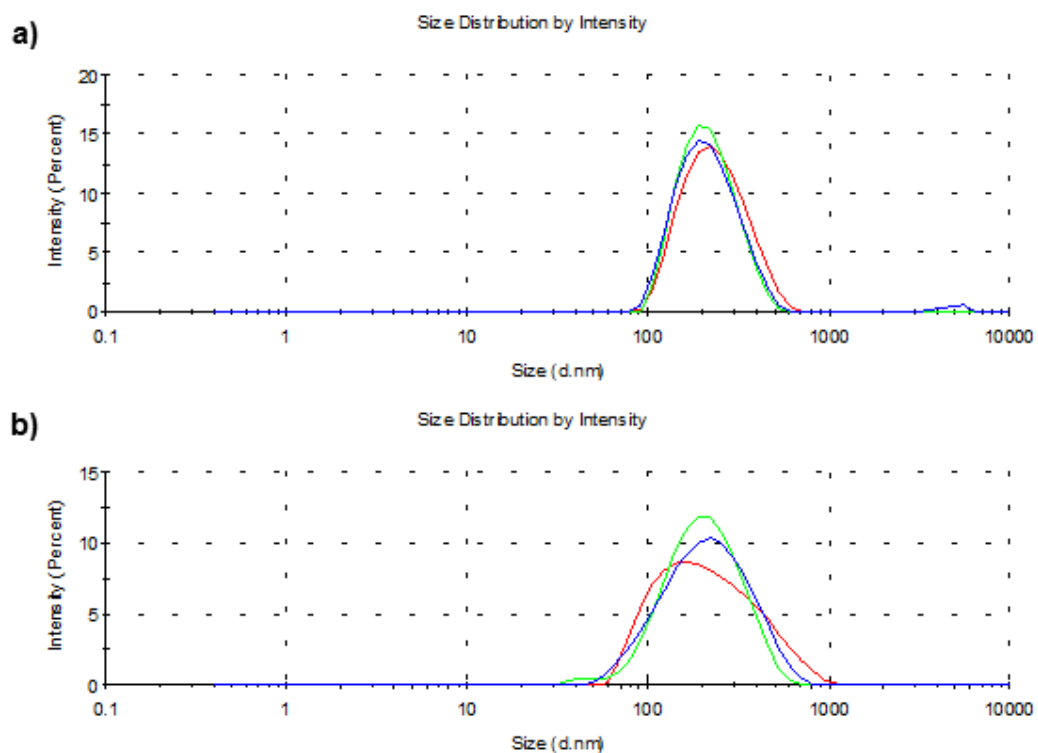
**Figure S4.** Representative DMF GPC traces of the PGalSMA<sub>34</sub> ( $M_n = 16,300 \text{ g mol}^{-1}$ ,  $M_w = 18,500 \text{ g mol}^{-1}$ ,  $M_w/M_n = 1.13$ ) and PGMA<sub>51</sub> ( $M_n = 16,200 \text{ g mol}^{-1}$ ,  $M_w = 18,600 \text{ g mol}^{-1}$ ,  $M_w/M_n = 1.15$ ) macro-CTAs and the corresponding (1:9 PGalSMA<sub>34</sub> + PGMA<sub>51</sub>)-PHPMA<sub>150</sub> ( $M_n = 38,700 \text{ g mol}^{-1}$ ,  $M_w = 47,100 \text{ g mol}^{-1}$ ,  $M_w/M_n = 1.21$ ) diblock copolymer obtained at 20 % solids content.

Solids content (%)	DP <sub>HPMA</sub> Targeted	Conversion (%)	$M_w$ (g mol <sup>-1</sup> )	$M_n$ (g mol <sup>-1</sup> )	$M_w/M_n$
10	90	>99	35,300	30,500	1.15
10	150	>99	48,000	41,200	1.16
10	200	>99	51,100	42,000	1.22
10	270	>99	70,700	56,500	1.25
15	90	>99	35,500	31,000	1.14
15	150	>99	46,100	38,400	1.20
15	200	>99	57,400	48,200	1.19
15	270	>99	72,500	58,500	1.24
20	90	>99	36,400	31,600	1.15
20	150	>99	47,100	38,700	1.21
20	200	>99	57,800	48,000	1.20
20	270	>99	70,300	55,400	1.27

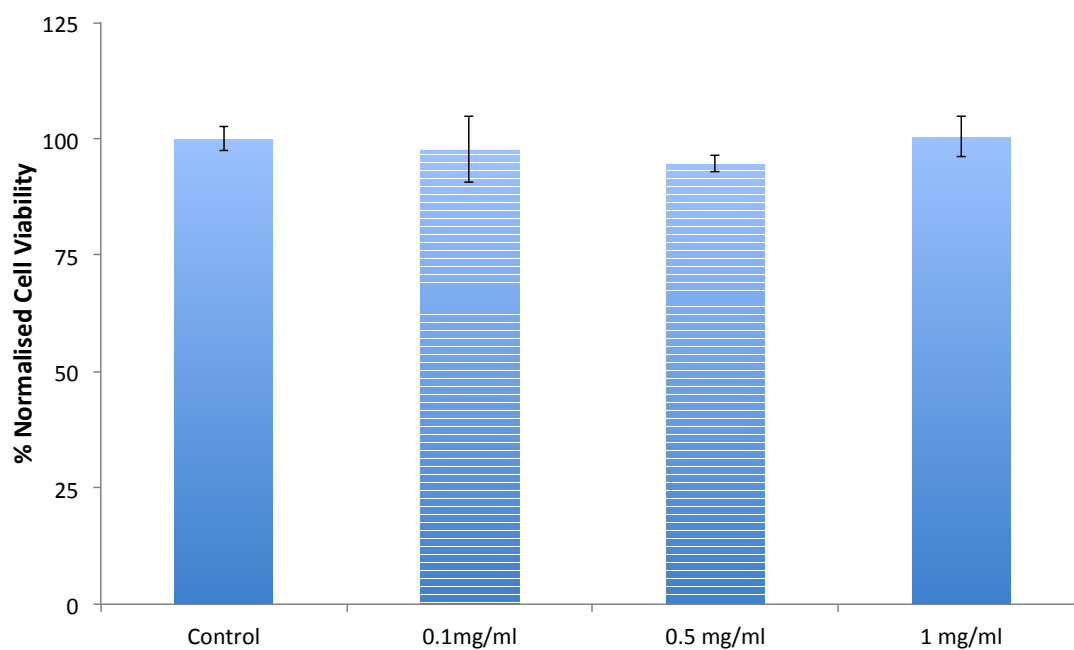
**Table S1.** Molecular weights determined by DMF GPC for (1:9 PGalSMA<sub>34</sub> + PGMA<sub>51</sub>)-PHPMA<sub>x</sub> diblock copolymers synthesized by RAFT aqueous dispersion polymerization.



**Figure S5.** Lectin interaction control experiments. The PGMA<sub>51</sub> macro-CTA (black squares) is not recognized by RCA<sub>120</sub>, hence no binding interaction occurs. In contrast, using PGalSMA<sub>34</sub> (red squares) under the same conditions leads to rapid aggregation in the presence of RCA<sub>120</sub>.



**Figure S6.** Dynamic light scattering particle size distributions for: a) vesicles obtained by PISA (intensity-average diameter,  $d = 231 \pm 13$  nm), b) vesicles obtained by film rehydration ( $d = 236 \pm 15$  nm). These measurements were performed using the same diblock copolymer binary mixtures, i.e. (1:9 PGalSMA<sub>34</sub> + PGMA<sub>51</sub>)-PHPMA<sub>270</sub>. The diameter is calculated from the average of three measurements in each case.



**Figure S7.** Normalized cellular viability of human dermal fibroblasts after incubation with (1:9 PGalSMA<sub>34</sub> + PGMA<sub>51</sub>)-PHPMA<sub>270</sub> vesicles. Cells were incubated in the presence of increasing concentrations of vesicles in cell media over 24 h. Cell viabilities were evaluated using an MTT-ESTA assay and the data were normalized relative to the untreated control (100% viability). N = 3 independent experiments were performed in triplicate wells.