

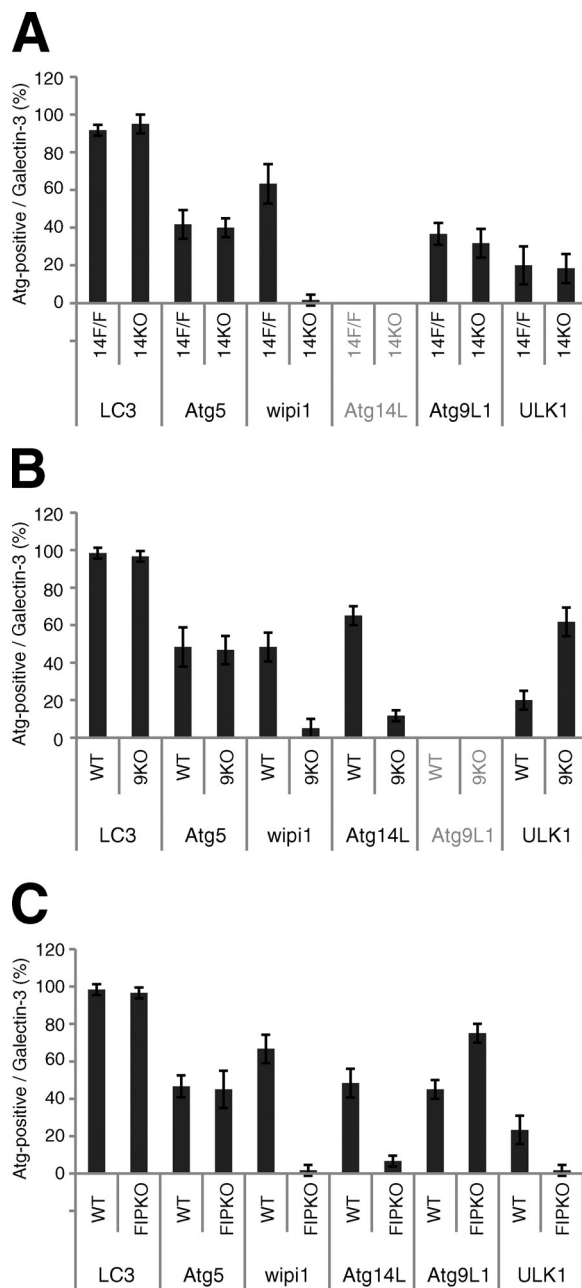
Fujita et al., <http://www.jcb.org/cgi/content/full/jcb.201304188/DC1>

Figure S1. **Hierarchical analysis of Atg proteins in bead-transfected cells.** (A–C) Wild-type and autophagy-deficient cells (Atg14L-KO [A], Atg9L1-KO [B], and FIP200-KO [C]) stably expressing GFP-tagged LC3, Atg5, WIPI-1, Atg14L1, Atg9L1, or ULK1 were transfected with Effectene-coated latex beads for 3 h and subjected to immunocytochemistry for galectin3. The percentages of Atg-positive beads per galectin3-positive beads were enumerated. At least 30 beads were counted ( $n = 3$ ). The values indicate the mean  $\pm$  SD.

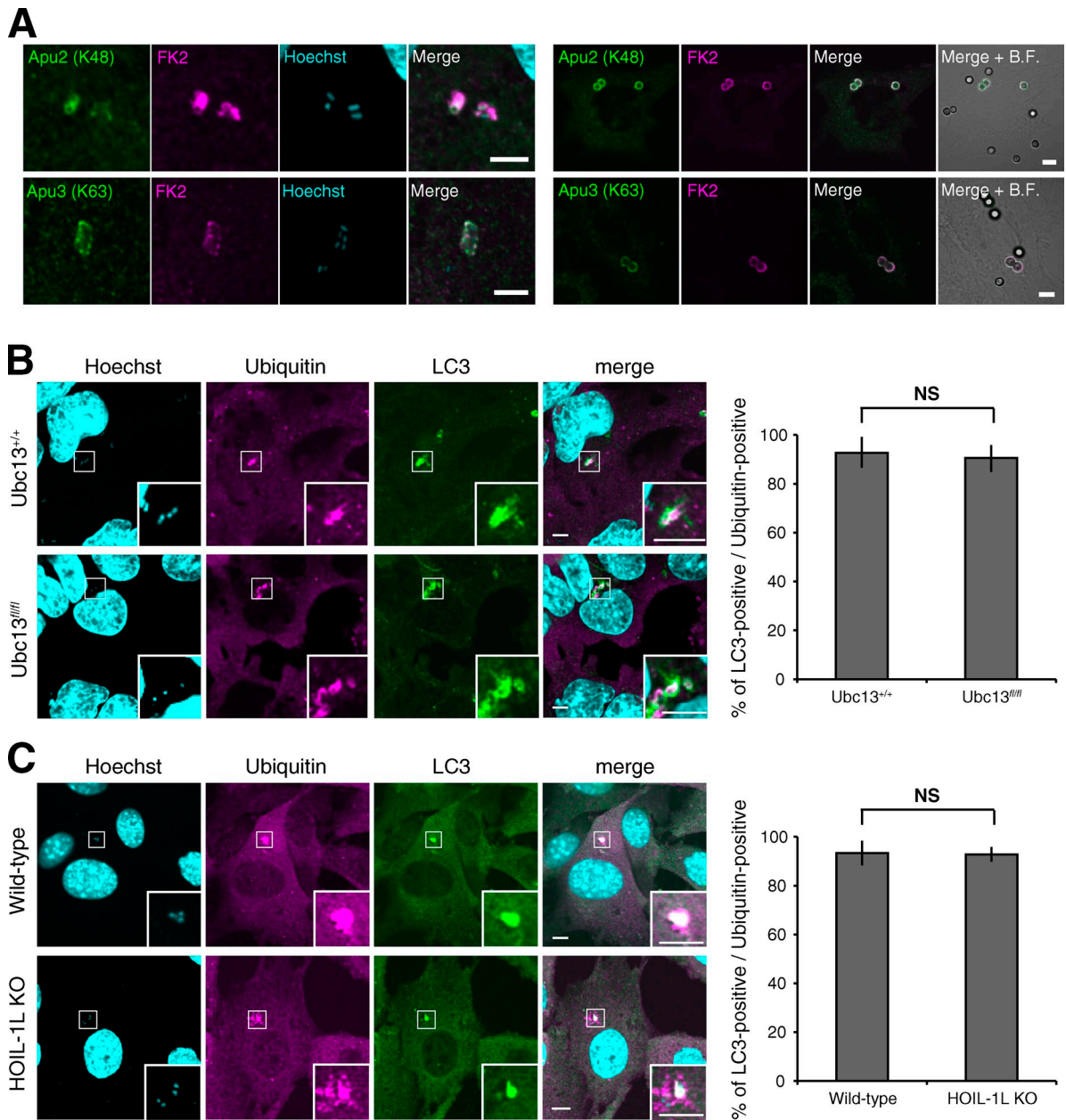


Figure S2. **Ub linkages in selective autophagy against invading *Salmonella* or transfected beads.** (A) HeLa cells were infected with *Salmonella* for 1 h (left) or transfected with Effectene-coated latex beads for 3 h (right), and then subjected to immunocytochemistry for poly-Ub (FK2) and K48-linked Ub (Apu2) or K63-linked Ub (Apu3). Bar, 5  $\mu$ m. (B) Ubc13<sup>+/+</sup> or Ubc13<sup>fl/fl</sup> MEFs were infected with *Salmonella* for 1 h and then subjected to immunocytochemistry for poly-Ub (FK2) and LC3. Bar, 10  $\mu$ m. The percentages of LC3-positive beads per Ub-positive *Salmonella* were enumerated. At least 30 Ub-positive *Salmonella* were counted ( $n = 3$ ). The values are the mean  $\pm$  SD. Statistical analysis was performed using Student's  $t$  test: NS, not significant. (C) Wild-type or HOIL-1L KO MEFs were infected with *Salmonella* for 1 h and then subjected to immunocytochemistry for poly-Ub (FK2) and LC3. Bar, 10  $\mu$ m. The percentages of LC3-positive beads per ubiquitin-positive salmonella were enumerated. At least 50 Ub-positive *Salmonella* were counted ( $n = 3$ ). The values are the mean  $\pm$  SD. Statistical analysis was performed using Student's  $t$  test. NS, not significant.

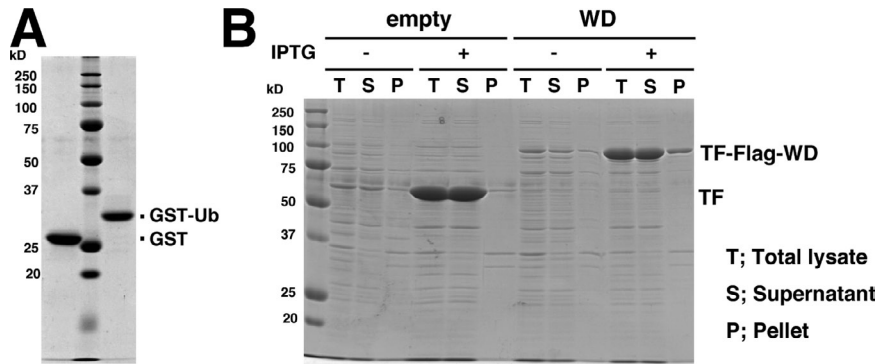


Figure S3. **Bacterially expressed recombinant proteins.** (A) GST or GST-ubiquitin were purified from *E. coli* lysates over glutathione Sepharose 4B and dialyzed with TBS. The recombinant proteins were subjected to SDS-PAGE followed by Coomassie brilliant blue staining. (B) *E. coli* BL21 (DE3) cells were transformed with the pCold-TF control (Empty) or pCold-TF encoding the WD40  $\beta$ -propellers of Atg16L1 (WD). To induce the expression of the recombinant proteins, the bacteria were incubated at 16°C for 12 h in the presence or absence of 100  $\mu$ M IPTG. Total cell lysates (T) were centrifuged at 15,000 rpm for 20 min and then separated into the soluble supernatant (S) and insoluble pellet (P).

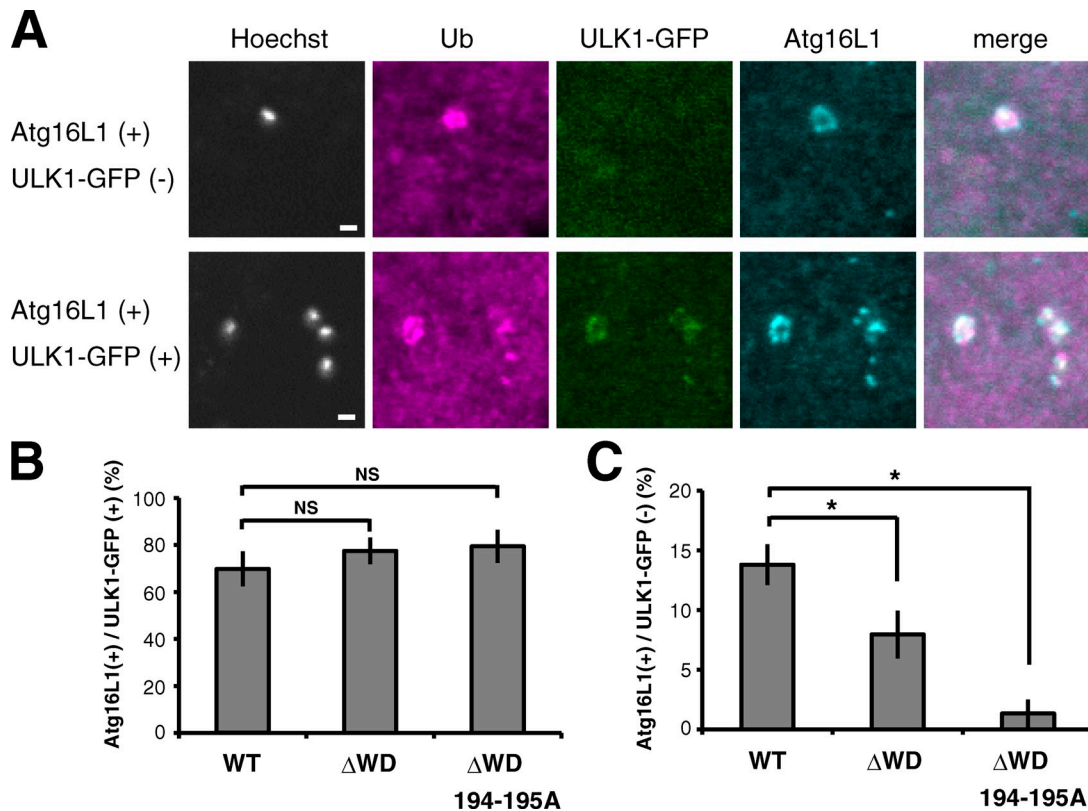


Figure S4. **Effect of deletion of WDR in Atg16L1 on the order of the Atg16L1 and ULK1-FIP200 complex recruitment to Ub-positive *Salmonella*.** Atg16L1- $\Delta/\Delta$  MEFs stably expressing both ULK1-GFP and Atg16L1 full-length,  $\Delta$ WD, or  $\Delta$ WD + 194–195A mutant were infected with *Salmonella* (MOI = 100) for 10 min, chased for another 10 min, and then fixed. The samples were analyzed by immunocytochemistry for Atg16L1 and Ub. Bar: (A) 1  $\mu$ m. The percentage of Atg16L1- and ULK1-GFP-positive bacteria per Ub-positive bacteria was enumerated by fluorescence microscopy (B and C). At least 50 bacteria were counted. The average  $\pm$  SD is shown for three independent experiments. Statistical analysis was performed using Student's *t* test. \*,  $P < 0.05$ ; NS, not significant.

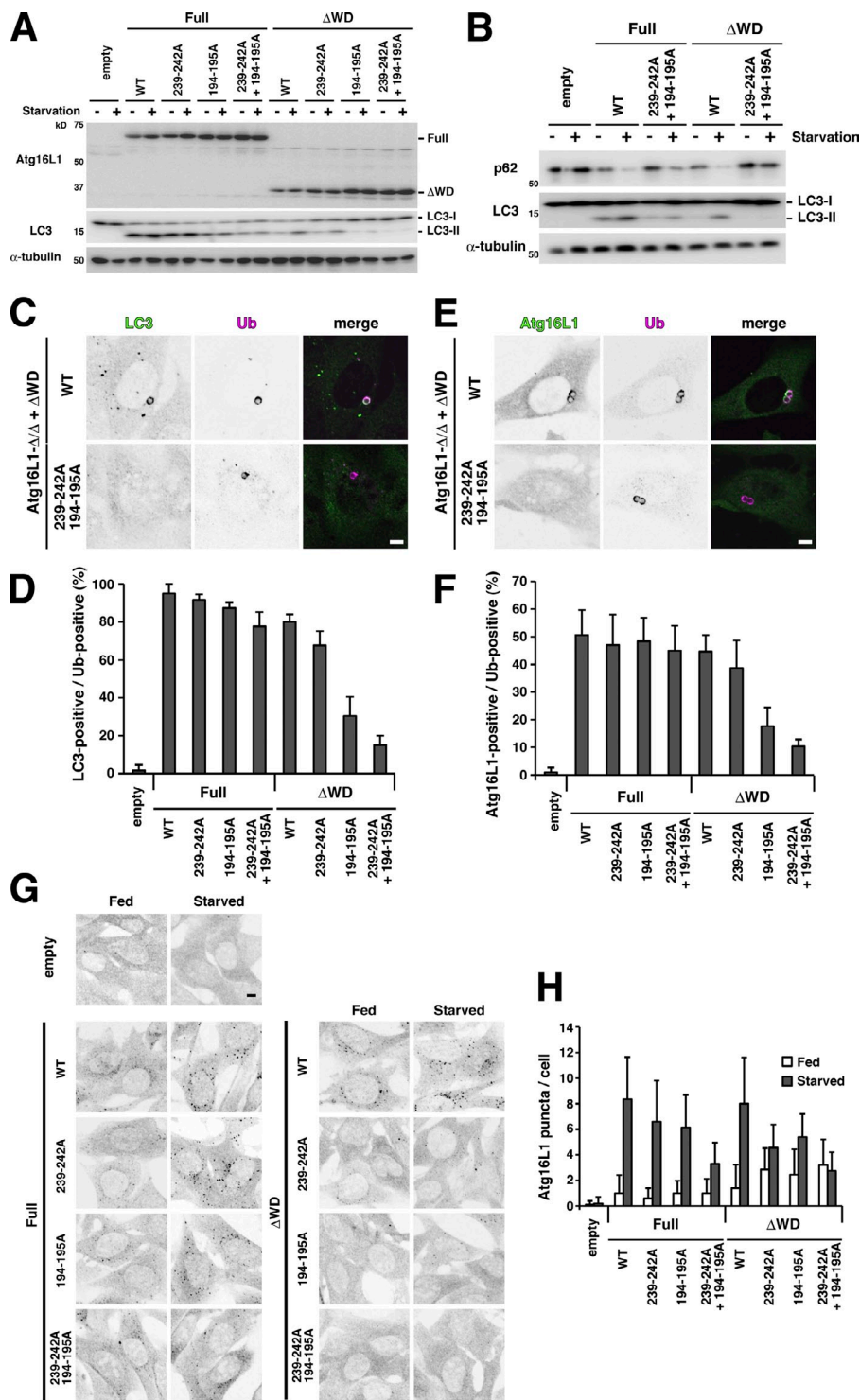


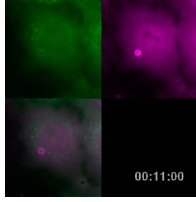
Figure S5. **Effect of Atg16L1 mutations on canonical and selective autophagy against transfected beads.** (A and B) Atg16L1- $\Delta/\Delta$  MEFs stably expressing the indicated constructs were cultured in growth medium (-) or EBSS (+) for 1 h (A) or 2 h (B) and then harvested. Total cell lysates were examined by Western blot analysis using the indicated antibodies. (C-F) Atg16L1- $\Delta/\Delta$  MEFs stably expressing the indicated constructs were transfected with beads for 3 h, and then analyzed by immunocytochemistry for LC3 (C) or Atg16L1 (E). Bar, 5  $\mu$ m. The percentage of LC3- or Atg16L1-positive beads per Ub-positive beads was enumerated by fluorescence microscopy (D and F). At least 50 beads were counted. The average  $\pm$  SD is shown for three independent experiments. (G) Atg16L1- $\Delta/\Delta$  MEFs stably expressing the indicated constructs were cultured in growth medium (Fed) or EBSS (Starved) for 1 h and subjected to immunocytochemistry using an anti-LC3 antibody. Bar, 5  $\mu$ m. (H) Atg16L1- $\Delta/\Delta$  MEFs stably expressing the indicated constructs were cultured in growth medium (Fed) or EBSS (Starved) for 1 h, and subjected to immunocytochemistry using an anti-Atg16L1 antibody. The number of Atg16L1 puncta in each cell was counted for more than 50 cells. The average  $\pm$  SD is shown for three independent experiments.

Table S1. FIP200 peptides detected by affinity purification and mass spectrometry analysis

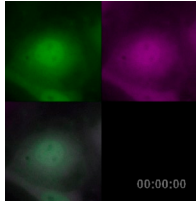
Peptide	Mr(expt)	Mr(calc)	Delta	Miss	Score	Expect
CTAVEIR	847.4356	847.4222	0.0135	0	14	2.6
FKVPLGTK	888.5566	888.5433	0.0134	1	17	1.5
EIVLEDLKK	1085.603	1085.6332	-0.0301	1	22	0.35
ELEDTLQVR	1101.5726	1101.5666	0.0061	0	64	2.10E-05
IPLLECLTR	1113.6266	1113.6216	0.0051	0	47	0.00099
LDSELSALER	1131.598	1131.5771	0.0209	0	64	2.30E-05
ENIINDLSDK	1159.579	1159.5721	0.007	0	47	0.0012
VKHLENQIAK	1178.693	1178.6771	0.0159	1	24	0.22
GELANNLHVR	1192.6408	1192.6312	0.0096	0	77	1.10E-06
EAVICLQNEK	1202.6074	1202.5965	0.011	0	61	3.90E-05
ELAQGFLANQK	1217.641	1217.6404	0.0006	0	50	0.00064
DQCISELISR	1219.5934	1219.5867	0.0068	0	30	0.047
EQCDFNSLTK	1226.5378	1226.5237	0.0141	0	61	3.20E-05
TLQLKEEENK	1230.6574	1230.6455	0.0119	1	36	0.014
KENIINDLSDK	1287.6808	1287.667	0.0138	1	47	0.001
FLEQLEEQEK	1291.6422	1291.6296	0.0127	0	32	0.033
LMSQSMSSVSSR	1298.6078	1298.5959	0.012	0	-74	1.80E-06
SLLEQETENLR	1330.6794	1330.6728	0.0066	0	70	5.30E-06
TTNESLTSFVK	1336.6942	1336.6874	0.0068	0	80	4.00E-07
STELVSPDMR	1343.6868	1343.6755	0.0114	0	52	0.0003
GDSSSLVAELQEK	1361.6722	1361.6674	0.0048	0	96	1.20E-08
AELQSLQSHLK	1381.7353	1381.7201	0.0152	0	14	1.8
TQLALEMYEVAK	1394.7214	1394.7115	0.0099	0	64	1.90E-05
LDSLPEHEDSEK	1397.6436	1397.631	0.0126	0	39	0.0052
ENIINDLSDKLK	1400.7552	1400.7511	0.0042	1	64	1.60E-05
DKDLIESLSEDR	1418.6958	1418.6889	0.007	1	71	3.70E-06
FLEQLEEQEKR	1447.7412	1447.7307	0.0106	1	49	0.00053
KFDCELPDISLK	1463.745	1463.733	0.012	1	17	0.84
ASVSGTSPQSASSPR	1488.7264	1488.7168	0.0096	0	62	3.10E-05
LYALDQMIASCGR	1496.7242	1496.7115	0.0127	0	72	2.70E-06
VTSLHNQAFEIEK	1514.7792	1514.7729	0.0064	0	54	0.00015
GELVCLLEEVQNK	1529.7808	1529.7759	0.0049	0	69	5.50E-06
LKGELVCLLEEVQNK	1770.954	1770.9549	-0.0009	1	46	0.00086
LKGELVCLLEEVQNK	1770.9637	1770.9549	0.0087	1	-33	0.017
IQDNNENYQVGLAELR	1874.92	1874.9122	0.0078	0	93	1.80E-08
TSLIAEQQTNFNTVLR	1935.0058	1935.0061	-0.0003	0	57	6.50E-05

ATG5-ATG12-ATG16L1 protein complexes were affinity purified on Strep-Tactin matrices, and co-purifying proteins were identified by mass spectrometry. In brief, co-purifying proteins were identified after SDS-PAGE separation and band excision. Proteins were digested with trypsin and identified by separating the peptide mixtures by liquid chromatography with online tandem mass spectrometry (LC-MS/MS). Mr(expt), experimental molecular weight (Da); Mr(calc), calculated molecular weight (Da); Delta, mass error (Da); Miss, number of missed cleavages; expect, expectation value.

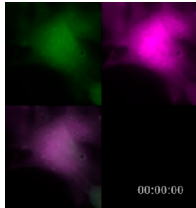




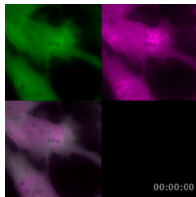
Video 1. **Dynamics of LC3 and galectin3 in autophagy against transfected bead.**



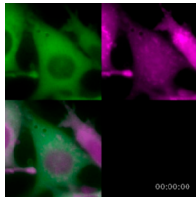
Video 2. **Dynamics of Ub and galectin3 in autophagy against transfected bead.** (S1 and S2) NIH3T3 cells stably expressing mStrawberry (mStr)-Gal3 (magenta) and GFP-LC3 (green, S1) or mStr-Gal3 (magenta) and GFP-Ub (green, S2) were transfected with Effectene-coated latex beads for 30 min and then washed. Live cells were observed at 1-min intervals using a microscope (model IX81; Olympus) equipped with a xenon lamp and cooled charge-coupled device camera (CoolSnap HQ; Roper Scientific) under the control of MetaMorph software. Time is shown in h:min:s.



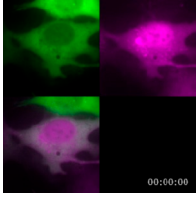
Video 3. **Dynamics of LC3 and Ub in autophagy against transfected bead.**



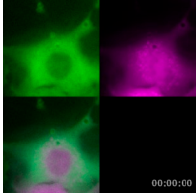
Video 4. **Dynamics of Atg5 and Ub in autophagy against transfected bead.**



Video 5. **Dynamics of WIPI-1 and Ub in autophagy against transfected bead.**



Video 6. **Dynamics of Atg14L and Ub in autophagy against transfected bead.**



Video 7. **Dynamics of ULK1 and Ub in autophagy against transfected bead.** (S3–S7) NIH3T3 cells stably expressing mStr-Ub (magenta) and GFP-tagged (green) LC3 (S3), Atg5 (S4), WIPI-1 (S5), Atg14L1 (S6), or ULK1 (S7) were transfected with Efectene-coated latex beads for 30 min. Live cells were observed at 1-min intervals using a microscope (model IX81; Olympus) equipped with a xenon lamp and cooled charge-coupled device camera (CoolSnap HQ; Roper Scientific) under the control of MetaMorph software. Time is shown in h:min:s. We excluded Atg9L1 in this experiment because Atg9L1 localized to the trans-Golgi network and early endosomes under steady-state conditions, which cannot be distinguished from recruitment to bead-containing endosomes (Young et al., 2006; Kageyama et al., 2011).

## References

- Kageyama, S., H. Omori, T. Saitoh, T. Sone, J.L. Guan, S. Akira, F. Imamoto, T. Noda, and T. Yoshimori. 2011. The LC3 recruitment mechanism is separate from Atg9L1-dependent membrane formation in the autophagic response against *Salmonella*. *Mol. Biol. Cell.* 22:2290–2300. <http://dx.doi.org/10.1091/mbc.E10-11-0893>
- Young, A.R., E.Y. Chan, X.W. Hu, R. Köchl, S.G. Crawshaw, S. High, D.W. Hailey, J. Lippincott-Schwartz, and S.A. Tooze. 2006. Starvation and ULK1-dependent cycling of mammalian Atg9 between the TGN and endosomes. *J. Cell Sci.* 119:3888–3900. <http://dx.doi.org/10.1242/jcs.03172>