

Supplemental Materials and Methods

Strains and culture conditions.

FGSC#4200 was used as the wild-type strain in this study, but in the race tube assays, 87-3 (*ras-1^{bd}*, *a*) was used as the wild-type strain. Calculations of period length were performed as described previously (Liu et al, 1997). Liquid cultures were grown in minimal media (1x Vogel's, 2% glucose). When QA was used to activate the *qa-2* promoter, liquid cultures were grown in 0.01 M QA (pH 5.8), 1x Vogel's, 0.5% glucose, and 0.17% arginine. Race tube media contained 1xVogel's, 0.1% glucose (replaced by 1 mM QA if necessary), 0.17% arginine, 50 ng/ml biotin, and 1.5% agar. For luciferase assays, the AFV (autoclaved FGS-Vogel's) medium was autoclaved with 0.05% fructose, 0.05% glucose, 2% sorbose, 1xVogel's medium, 50 µg/L biotin, and 1.8% agar; firefly luciferin (BioSynt L-8200 D-luciferin firefly [synthetic] potassium salt) was added after autoclaving (final concentration of 50 µM). For rhythmic experiments, the *Neurospora* cultures were transferred from constant light (LL) to DD at time 0 and were harvested after the indicated time in DD.

The *Neurospora* knockout library was obtained from Fungal Genetics Stock Center.

Protein and RNA analyses.

Protein extraction, western blot analyses, RNA extraction, and qRT-PCR were performed as previously described (Cha et al, 2008; Cha et al, 2011). The qRT-PCR results were analyzed by Student's *t*-Test and the significance was indicated with error bars of standard errors. *, P<0.05; **, P<0.01. The data for mRNA levels were normalized with *β-tubulin* and one of the wild-type samples was set as 1. Primer sequences are available upon request. To raise the anti-CATP antibody, the EcoRI fragment of the *catp* ORF was cloned into pGEX-4T-1 and GST-

tagged recombinant was purified from *E. coli*. Antisera were obtained by immunizing rabbits with purified protein (Cocalico Biologicals, Inc.). For quantifications of western results, the bands were scanned and analyzed by ImageJ (<http://rsb.info.nih.gov/ij/>). Representative blots out of more than 2 experiments were shown. Antibodies against histone H3 (ab1791) and the RNA pol II C-terminal domain (phospho S5; ab5131) were purchased from Abcam, and anti-acetyl-histone H3 (Lys14; #06-911) antibody was from Millipore.

Sequences used for the alignment of CATP homologues

Neurospora CATP (GI: 85083472), *Saccharomyces cerevisiae* YTA7 (GI: 1556439), *Mus musculus* ANCCA/ATAD2 (GI: 91199557), *Homo sapiens* homolog (GI: 24497618), *Drosophila melanogaster* TER94-RC (GI: 262272122) and *Arabidopsis thaliana* AT1G05910 (GI: 18390588).

SI Figure legends:

Figure S1. Race tube analysis showing the conidiation rhythms of the *mut10* strain in either wild-type or *ras-1^{bd}* background. Black lines indicate the growth fronts every 24 h.

Figure S2. (A) Densitometric analyses of the western blot analysis in Figure 2B. **(B)** Densitometric analyses of the western blot analysis in Figure 3D.

Figure S3. Light induction of gene expression in the *catp^{KO}* strain is near normal. qRT-PCR results show that light-induced expressions of *frq*, *al-1* and *vvd* in the wild-type and the *catp^{KO}* strains. The wild-type at time 0 (DD24) was set as 1 and error bars indicate SEM (n=3).

**P<0.01 (paired student's *t*-test).

Figure S4. (A) ChIP assays using the antibody against endogenous CATP showed that it binds to the *frq* promoter and ORF regions. The location of each amplicon (distance in bp from AUG) was indicated. **(B)** Histone H3 ChIP showed that the nucleosome occupancy at the *adv-1* and *frh* loci was not affected in the *catp^{KO}* strain. **(C)** qRT-PCR analyses showing that *adv-1* (upper panel) and *frh* (lower panel) mRNA levels were not affected in the *catp^{KO}* strain. Mean with standard errors (n=3). *P<0.05, **P<0.01 (paired student's *t*-test).

Figure S5. Western blot analysis using the CATP antibody showing the expression profile of CATP in the constant darkness at the indicated time points. The asterisk indicates a nonspecific protein band detected by our antibody.

Figure S1

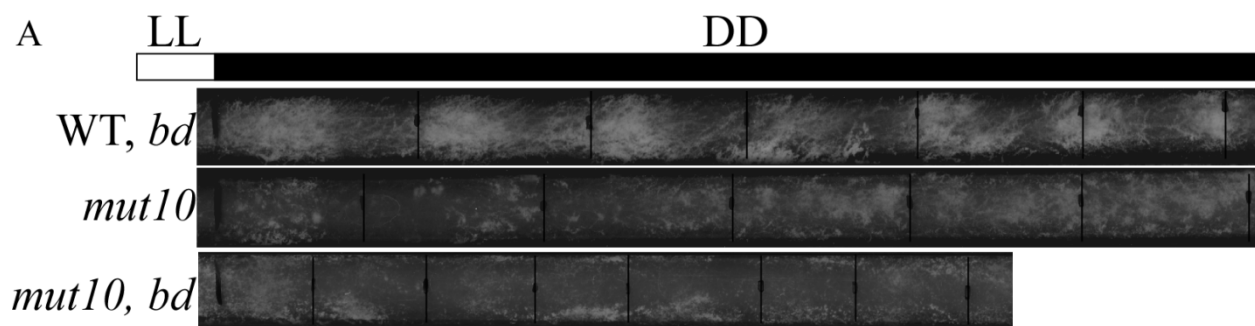
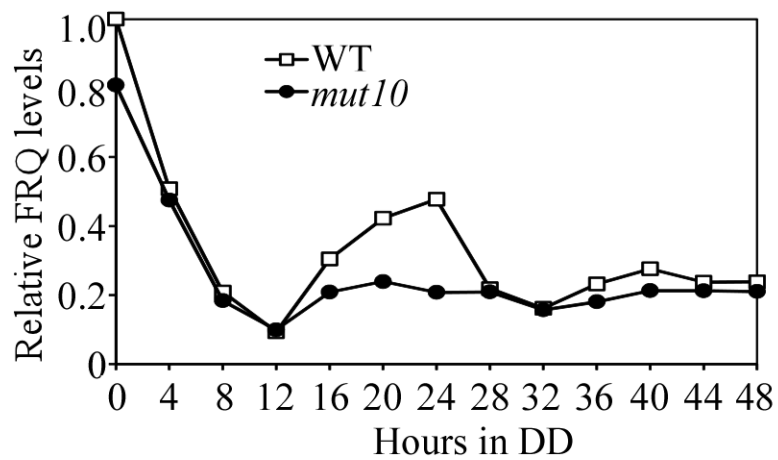


Figure S2

A



B

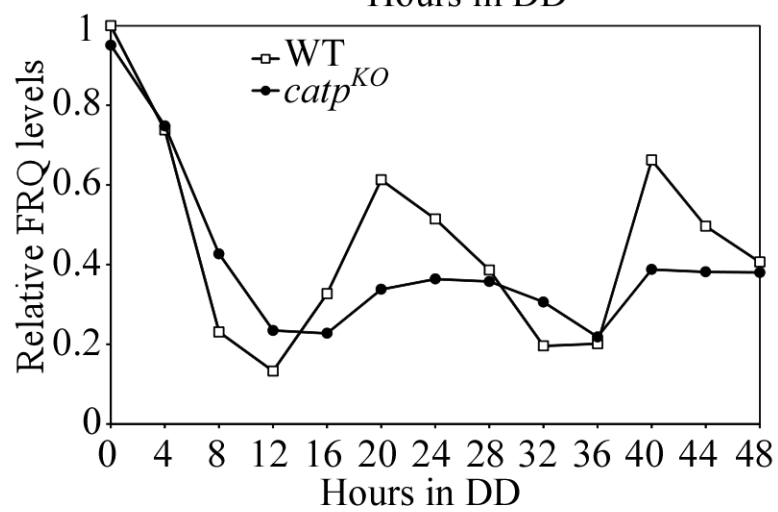


Figure S3

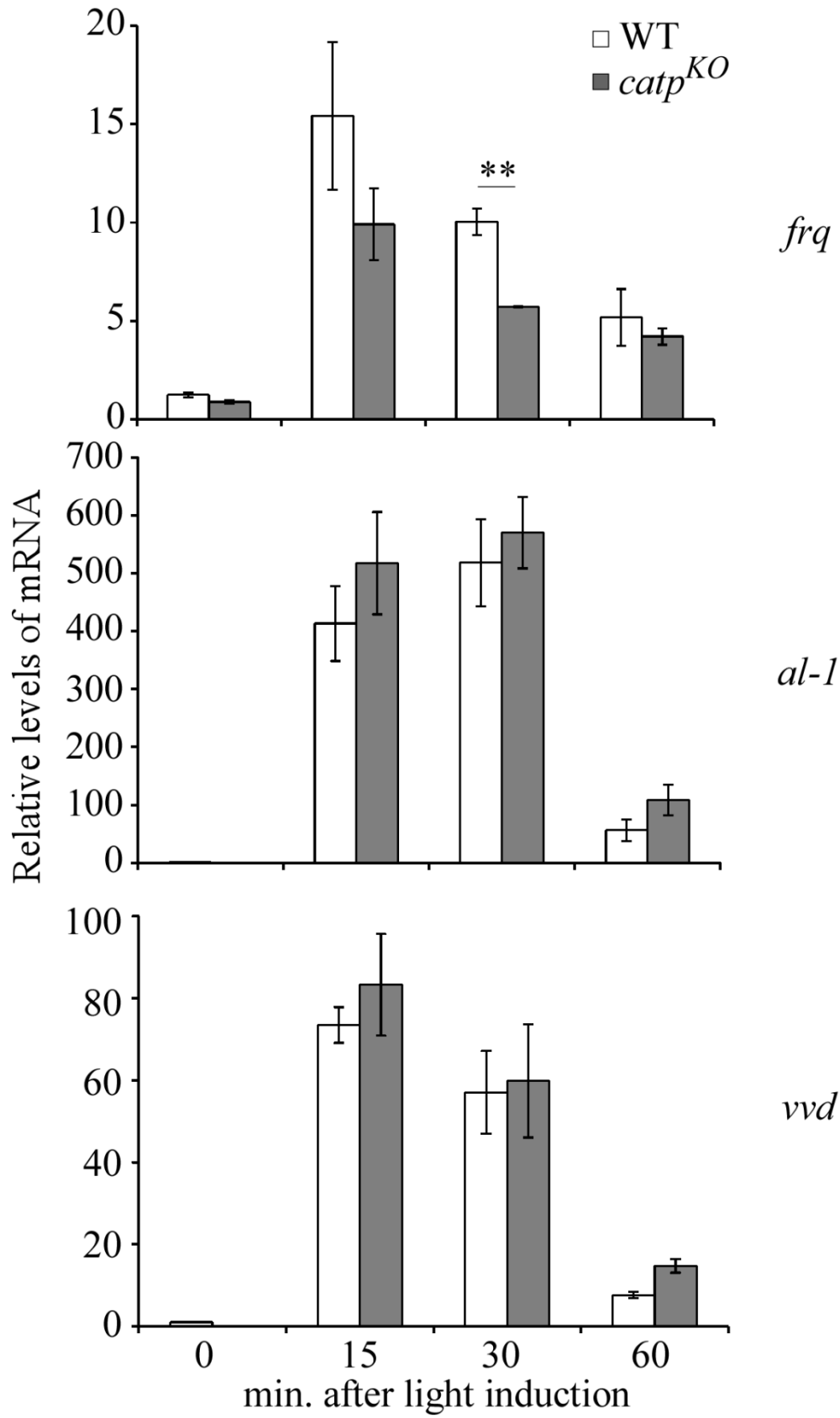
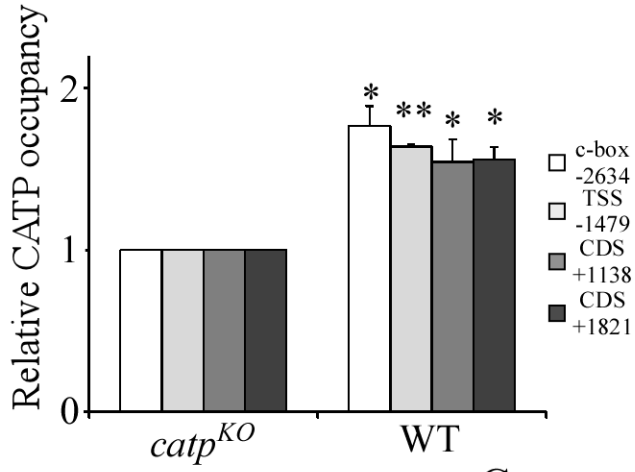
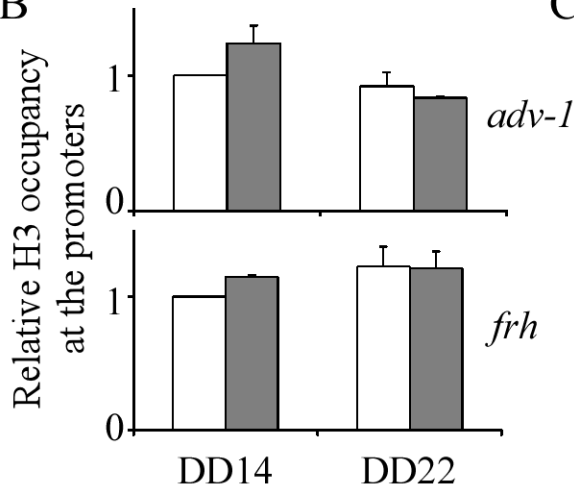


Figure S4

A



B



C

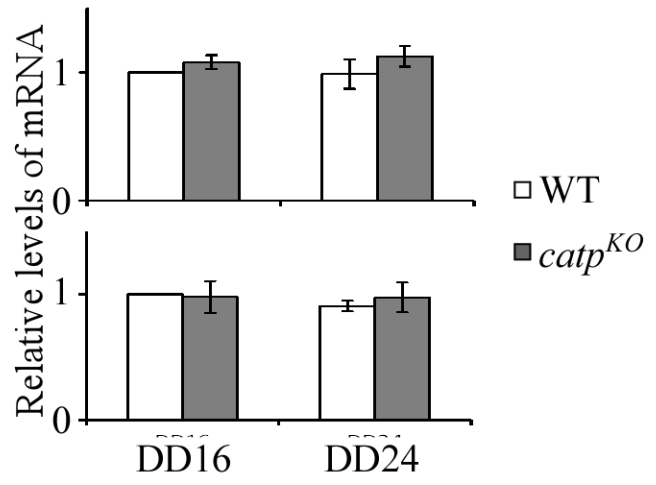


Figure S5

